

1 Review

2 Mesoporous Silica Nanoparticles for the Treatment of 3 Complex Bone Diseases: Bone Cancer, Bone Infection 4 and Osteoporosis

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13 Received: date; Accepted: date; Published: date

14 **Abstract:** Bone diseases, such as bone cancer, bone infection and osteoporosis, constitute a major
15 issue for modern societies as a consequence of their progressive ageing. Even though these
16 pathologies can be currently treated in the clinic, some of those treatments present drawbacks that
17 may lead to severe complications. For instance, chemotherapy lacks of great tumor tissue selectivity,
18 affecting healthy and diseased tissues. In addition, the inappropriate use of antimicrobials is leading
19 to the appearance of drug-resistance bacteria and persistent biofilms, rendering current antibiotics
20 useless. Furthermore, current antiosteoporotic treatments present many side effects as a
21 consequence of their poor bioavailability and the need to use higher doses. In view of the exposed
22 evidences, the encapsulation and selective delivery to the diseased tissues of the different
23 therapeutic compounds seem highly convenient. In this sense, silica-based mesoporous
24 nanoparticles offer great loading capacity within their pores, the possibility of modifying the surface
25 to target the particles to the malignant areas and great biocompatibility. This manuscript is intended
26 to be a comprehensive review of the available literature on complex bone diseases treated with
27 silica-based mesoporous nanoparticles, whose further development and eventual translation into
28 the clinic could bring significant benefits for our future society.

29 **Keywords:** Mesoporous Silica Nanoparticles; Mesoporous Bioactive Glasses; Bone cancer; Bone
30 infection; Bone Regeneration; Osteoporosis; Stimuli-Responsive Drug Delivery; Targeted Drug
31 Delivery.

33 1. Introduction

34 In the last decades, nanotechnology has been applied to a variety of fields, ranging from novel
35 electronic devices to the study of biological processes [1–4]. In particular, the application of
36 nanotechnology to medicine, the so-called nanomedicine, has attracted the attention of many
37 researchers, and it is expected to revolutionize the pharmaceutical and biotechnological fields in the
38 near future [5–7].

39 The first developments in the field of nanomedicine were reported in the early 60's when
40 liposomes were first proposed as carriers [8,9]. Since then, scientists have engineered many different
41 nanocarriers to address effective delivery of therapeutics. Those nanoparticles can be classified as
42 either organic or inorganic. Examples of organic nanocarriers include liposomes, which are
43 amphiphilic lipids that rearrange in water to yield vesicles with an inner aqueous compartment
44 surrounded by lipid bilayers [10]; polymeric nanoparticles produced from polymer chains showing

45 different functionalities [11] or polymeric micelles composed by amphiphilic block copolymers able
46 to rearrange in aqueous media [12]. Examples of inorganic nanocarriers include metal nanoparticles
47 synthesized from noble metals, such as gold or silver [13]; carbon nanoparticles such as carbon
48 nanotubes, fullerenes or mesoporous carbon nanoparticles [14] or silica-based mesoporous
49 nanoparticles, which have been extensively studied owing to their capacity to load large amounts of
50 therapeutic molecules [15]. The main advantages of silica-based mesoporous nanoparticles over other
51 types of particles include the robustness of the silica framework, that allows the use of harsh reaction
52 conditions for their modification, and their excellent textural properties. In fact, conventional
53 polymeric nanoparticles usually present low drug capacity, usually less than 5% of total weight,
54 whereas these silica-based mesoporous nanoparticles offer greater values [16,17]. The main
55 disadvantage over other formulations would be the fact that the translation of these type of particles
56 remains challenging. However, it should be mentioned that silica is “generally recognized as safe”
57 by the US FDA, and it is often used as excipient in drug formulations and as dietary supplement
58 [18,19]. In this sense, the administration of fenofibrate-loaded ordered mesoporous silica materials in
59 men was found to be safe, and the doses were well tolerated by the patients [20]. In addition, small
60 silica nanoparticles (c-dots, 7 nm) for imaging purposes were approved by FDA for a human clinical
61 trial, demonstrating that they were well tolerated by the patients and accumulated in the tumor site
62 [21]. In consequence, silica-based nanoparticles constitute a powerful and promising tool that might
63 be promptly translated into the clinic.

64 This review will cover the application of silica-based mesoporous nanoparticles for the treatment
65 of complex bone diseases, such as bone cancer, bone infection and osteoporosis. These pathologies
66 are predominantly found in elderly people, who will constitute a quarter of the European population
67 by 2020 [22]. Then, bone diseases will definitely entail a significant impact on the health care systems
68 and, consequently, bone-targeted nanomedicines, *i.e.*, nanomedicines able to specifically reach bone
69 diseases, could bring significant benefits for our future society.

70 2. Mesoporous Silica Materials

71 2.1. The Beginning of a New Era: Ordered Mesoporous Silica Materials

72 Ordered mesoporous silica materials were first reported in the early 90's by Mobil Oil
73 Corporation researchers [23] and scientists from Waseda university [24]. These bulk mesoporous
74 materials have attracted great attention because they present (1) tunable and narrow pore size
75 distributions (2-30 nm); (2) adjustable porous structures; (3) high specific surface areas (up to 1500
76 cm²/g); (4) high pore volumes (*ca.* 1 cm³/g) and (5) high silanol density on the surface that allows
77 further modifications [25,26]. Owing to their exquisite physico-chemical properties, mesoporous
78 silica materials have been broadly applied in a number fields, including heavy metal adsorption
79 [27,28], catalysis [29,30] or energy storage [31,32], among others.

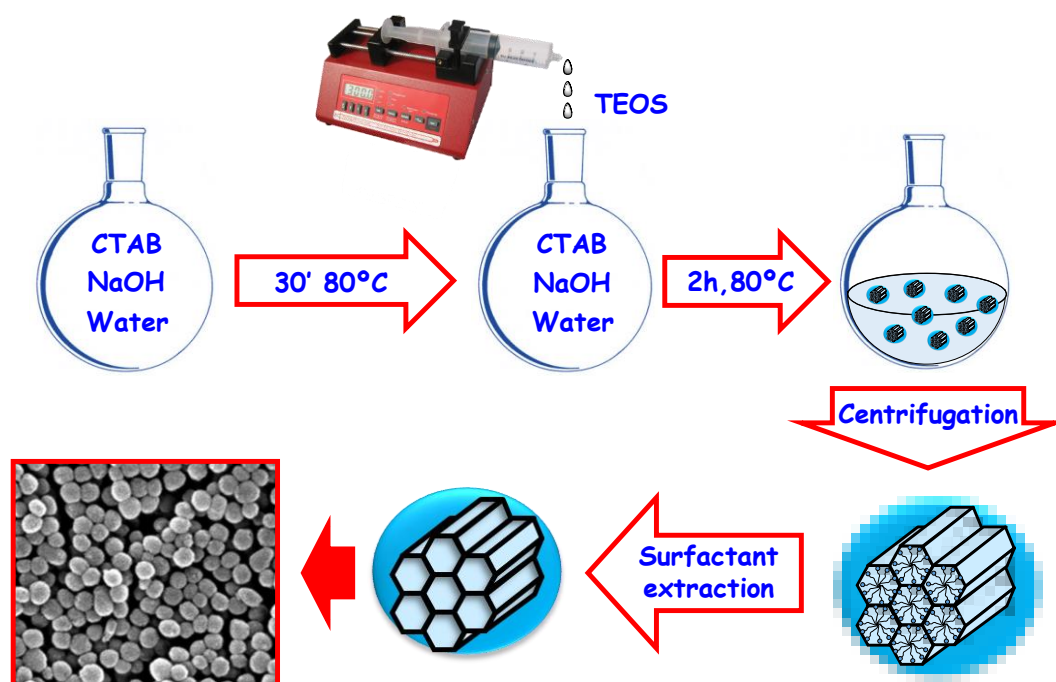
80 In addition, these materials find broad application within the field of biomaterials, owing to their
81 ability to adsorb molecules within their pores and release them in a sustained fashion. In fact, these
82 materials have been widely studied since Prof. Vallet-Regí and coworkers first reported their
83 suitability as drug delivery systems back in 2001 [33].

84 In light of their great properties and their potential biomedical application, researchers focused
85 their efforts on translating those excellent features of bulk materials to the nanoscale dimension. As
86 a result, mesoporous silica nanoparticles (MSNs) were developed soon after, opening the gates to
87 multiple biomedical applications, such as controlled drug delivery [34,35], efficient gene transfection
88 [36–38], antibacterial treatment [39,40] or bone tissue regeneration [41,42], among others.

89 2.2. Synthesis and Functionalization of Mesoporous Silica Nanoparticles

90 The synthesis of MSNs is based on a modification of the Stöber method, which initially yielded
91 micron-sized monodispersed and non-porous silica spheres [43]. In this sense, the addition of
92 surfactants as structure-directing agents results in silica nanoparticles with excellent physico-
93 chemical properties and showing porosity. This methodology allows to obtain homogenous

94 nanoparticles within the range 50-300 nm [25]. The morphology and dimensions of these surfactant-
 95 templated mesoporous silicas can be tailored by controlling the reaction conditions (e.g., pH,
 96 temperature, surfactant concentration or silica precursor) [44]. As an example, a synthetic protocol
 97 for the synthesis of MCM-41 (*Mobil Composition of Matter*) MSNs is depicted in Figure 1.



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Figure 1. Synthesis of MCM-41 MSNs using a modification of the Stöber method. The surfactant molecules self-assemble forming rod-like micelles around which the silica precursors polymerize, leading to the formation of a silica backbone with hexagonally ordered mesopores. TEOS: Tetraethyl ortosilicate; CTAB: Cetyltrimethylammonium bromide.

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The positively charged polar heads of the surfactant molecules interact with the negatively charged silica precursors, leading to the formation of the silica framework by means of the hydrolysis and condensation of the silica precursor onto the self-assembled rod-like surfactant micelles. Then, the organic template is removed using a solvent extraction method, yielding MSNs with empty pores ready to be filled with therapeutic molecules. This method is usually preferred over calcination, since the latter may cause irreversible aggregation of the particles and cytotoxic byproducts, limiting their potential application [45,46].

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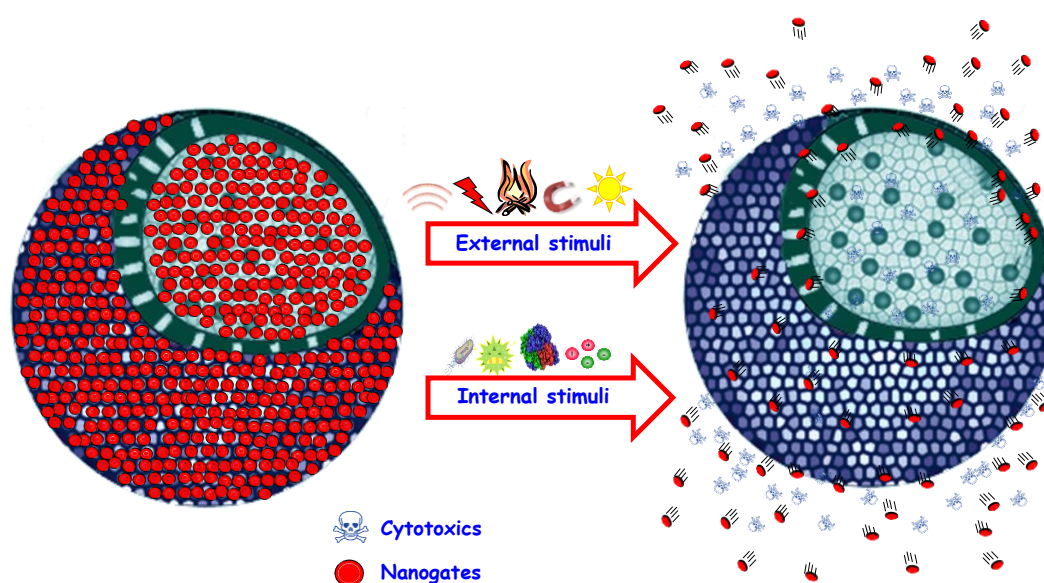
One of the most remarkable features of MSNs is their high density of silanol groups on the surface. These chemical groups allow the easy functionalization of the nanoparticles surface, usually using organosilanes bearing different functionalities (amine, carboxylic acid, thiol...), to increase the versatility of the produced nanocarriers. The particular organosilane employed allows to tune the interactions between the payload and the silica matrix, which might be beneficial for particular diseases [47,48]. The functionalization can be accomplished through two different approximations: post-synthesis or co-condensation. The post-synthesis method involves the modification of the surface after the synthesis. This approximation can lead to different groups inside and outside the pores, depending on whether the process is performed before or after removing the template. The co-condensation approach consists in the simultaneous addition of the silica precursor and the functional organosilane during the formation of the particles. This approximation can yield nanoparticles bearing various functional groups homogenously distributed throughout the silica backbone or biodegradable periodic nanoparticles with labile bonds within the silica framework [25].

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2.3. Mesoporous Silica Nanoparticles as Smart Drug Delivery Systems

124 Aside from being biocompatible, any nanoparticle intended to be employed as drug delivery
 125 system should fulfill some basic requirements, such as maximizing the amount of therapeutics
 126 loaded, minimizing premature release, reaching the target area and releasing the cargo on-demand
 127 only where needed. In this sense, the extraordinary textural properties of MSNs endow them with
 128 great loading capacities, being able to load huge amounts of therapeutic molecules within their pores,
 129 as demonstrated by Scanning Transmission Electron Microscopy [49]. In addition to serving as drug
 130 reservoir, the silica matrix provides a protective shell for the molecules against potential pH- or
 131 enzymatic-mediated drug degradation in the organism.

132 The loading of therapeutic molecules within MSNs can be easily accomplished as consequence
 133 of their open porous structure. However, this also means that the cargo molecules might easily diffuse
 134 out of the pores before reaching the target area. This premature release can be minimized using the
 135 so-called stimuli-responsive gatekeepers, which are structures able to open and close the pore
 136 entrances on-demand in response to certain stimuli [50–53]. In this manner, premature and
 137 nonspecific drug release would be minimized and the release would only take place upon application
 138 of a convenient stimulus at the diseased area (Figure 2).
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141 **Figure 2.** Schematic representation of stimuli-responsive MSNs. In response to the stimulus, the
 142 gatekeeper opens the pore entrances, triggering the drug release. The origin of the stimulus can be
 143 internal (pH, enzymes, redox species, etc) or external (magnetic fields, light, ultrasounds, etc).

144 Stimuli can be applied from inside or outside the organism. The use of internal stimuli is
 145 interesting because of the significant variations of various relevant biomarkers that can be found in
 146 some diseases. For instance, the pH of some subcellular organelles and that of the tumoral matrix are
 147 more acidic compared to the physiological value [54], and analogous behavior is observed in bacterial
 148 infections [55]. These pH variations have been employed to trigger the release from pH-responsive
 149 smart MSNs [56–59]. In addition, some enzymes, which have been observed to be overexpressed in
 150 osteoporotic [60] or tumoral scenarios [61,62], have the ability to cleave very specific peptidic
 151 sequences. In this sense, it is possible to use those peptides to close the pore entrances of MSNs and
 152 trigger the drug release only in those situations, where the enzymes are overexpressed [63–65].
 153 Another relevant example of internal stimulus is the overexpression of redox species in the cytoplasm
 154 of tumoral cells compared to the extracellular fluids [66,67], which has been employed to initiate the
 155 drug release from different redox-responsive MSNs [68–70].

156 External stimuli, which should be innocuous to the organism, have also attracted great attention.
 157 Their main advantage is that they would allow the application of the stimulus directly by the
 158 clinician, thereby providing a much higher control of the release kinetics. For instance, the generation

159 of heat through the application of alternating magnetic fields has been employed trigger the drug
160 release from MSNs and generate hyperthermia-mediated cell death [71–73]. The use of light
161 (ultraviolet, visible, near-infrared) has also attracted the attention of many researchers, and
162 constitutes a non-invasive method to trigger the release from MSNs [74–76]. Another relevant
163 example of non-invasive and innocuous stimulus are ultrasounds, which have been successfully
164 employed to externally trigger the payload release from MSNs [77–79].

165 2.4. Biodistribution and Biodegradation of Mesoporous Silica Nanoparticles

166 The most common routes of administration of the above-mentioned smart nanoparticles are
167 intravenous, subcutaneous or localized injections in the target area. In particular, the intravenous
168 administration leads to the rapid delivery and distribution of the particles throughout the organism,
169 albeit it entails challenging issues. For instance, the particles administration leads to the formation of
170 a protein corona around them that defines their biological entity. This protein coating might limit the
171 functionality of the nanoparticles and enables their recognition by the organism, triggering their
172 removal by the mononuclear phagocyte system and decreasing the efficiency of the treatments [80].
173 An effective approximation to overcome that issue would be the modification of the nanoparticles
174 with hydrophilic polymers, such as poly(ethylene glycol), which might help reduce the amount of
175 proteins adsorbed onto the nanoparticles by creating a hydrophilic layer, enhancing their colloidal
176 stability and increasing their circulating half-life [81–83]. In this sense, it has been shown in murine
177 models that non-PEGylated MSNs rapidly accumulate in the lung, liver and spleen while their
178 PEGylated counterparts show increased circulating half-life [84,85].

179 Besides the effect of PEGylation on the particles biodistribution, there are other relevant
180 parameters that influence the final fate of MSNs. For instance, it has been shown *in vivo* that the larger
181 the nanoparticles the faster their excretion [85]. In addition, it has been observed that, unlike spherical
182 particles, those presenting elongated or cylindrical shapes undergo faster clearance from the
183 bloodstream [86]. Finally, the surface charge is a key parameter since it determines the interaction of
184 the particles with the surrounding media. In this sense, it has been shown that positively charged
185 nanoparticles are more prone to undergo opsonization and subsequent clearance than their slightly
186 negative or neutral counterparts [87,88].

187 Aside from achieving effective accumulation at the diseased area, it would be desirable that the
188 MSNs degrade somehow to facilitate their excretion after exerting their therapeutic activity. In this
189 sense, the dissolution rate of the silica backbone is a key factor for their elimination. Silica-based
190 mesoporous nanoparticles are composed of polycondensed silica tetrahedrons (SiO₄) interconnected
191 by siloxane bonds (-Si-O-Si-) and presenting silanol groups (-Si-OH) on the surface. The silica
192 dissolution is consequence of the nucleophilic attack of water to the siloxane and silanol groups,
193 generating biocompatible silicic acid as by-product that can be excreted through the urine [89]. The
194 dissolution rate depends on the particular characteristics of the particles and can be tuned through
195 the introduction of organic modifications on the surface. Those modifications have been shown not
196 to affect the biodistribution and biocompatibility of the MSNs [86].

197 3. Mesoporous Silica Nanoparticles for the Treatment of Bone Cancer

198 3.1. General Concepts on Bone Cancer and Bone Metastasis

199 Cancer is the term given to a group of diseases sharing an unstoppable cell division and with
200 potential to spread in other organs and tissues. It is a leading cause of mortality worldwide and its
201 prevalence is progressively increasing, with 1.7 million of estimated new cases and 600,000 cases of
202 estimated deaths only in the United States in 2019 [90].

203 Bone-related tumors fall into primary bone tumors and metastatic bone tumors. They are
204 considered to be highly deadly even though chemotherapy has improved the patient survival for
205 sarcomas [91]. The most common malignant primary bone tumors are osteosarcoma,
206 chondrosarcoma and Ewing sarcoma, which account for 70% of such malignancies. They originate in
207 the bone, where mesenchymal stem cells behave both as ontogenic progenitor tumor cells and

208 stromal cells that contribute to tumor development. The stroma of these tumors comprises
209 osteoblasts, osteoclasts, endothelial and immune cells and mesenchymal stem cells. In particular,
210 osteoclast have grabbed great attention because their activity (bone destruction) can be metabolically
211 enhanced directly by tumor cells and, reversibly, the presence of osteoclasts boosts the aggressiveness
212 of cancer cells [92].

213 Metastasis is the spread of cancer cells from a primary tumor to distant sites to create secondary
214 tumors. It is a stage of the disease usually considered to be incurable and whose treatments are mainly
215 palliative [93]. Its origin is the pre-metastatic niche, which is an environment in a secondary organ
216 induced by the primary cancer cells that provides favorable conditions for the growth of tumoral cells
217 [94]. The exact mechanism of that metastatic organotropism remains unclear but it is thought to be
218 related with tumor-derived exosomes. Exosomes are nanometric membrane-bound vesicles secreted
219 by tumors cells that contain functional biomolecules, such as proteins, RNA, DNA and lipids [95]. In
220 this sense, it has been reported that tumor exosome integrins can determine organotropic metastasis
221 by fusing with organ-specific resident cells to establish the pre-metastatic niche. Once uptaken, they
222 induce cellular changes in the target organ (through the activation of Src phosphorylation and pro-
223 inflammatory S100), thus promoting cancer cell colonization and organ-specific metastasis [96].

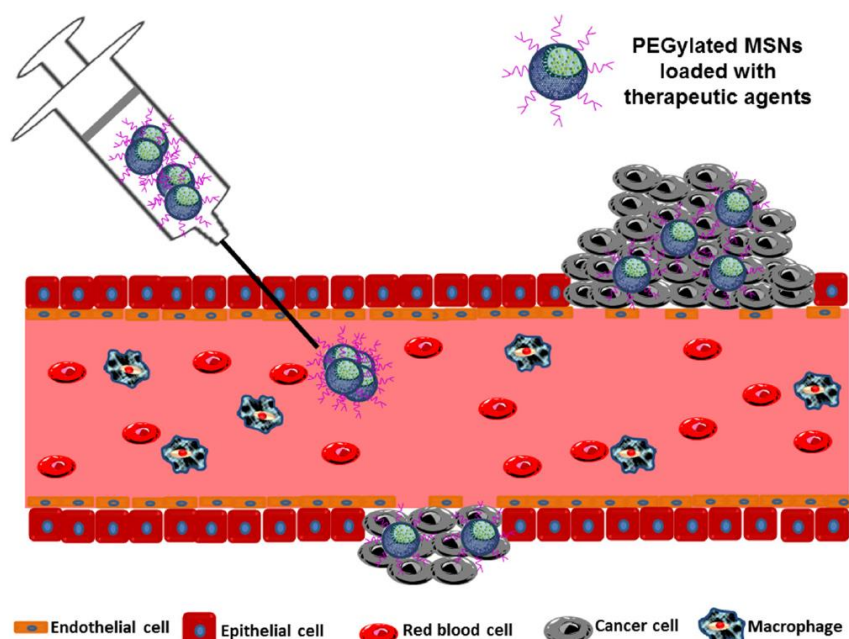
224 A characteristic feature of this disease is that some types of cancer cells preferentially migrate
225 and induce metastasis to specific organs [95]. In this sense, breast and prostate tumors normally lead
226 to bone metastases, which are secondary tumors formed when primary tumor cells home to the
227 skeleton [97,98]. Cancer cells can leave the primary tumor site owing to the poor adhesion among
228 each other in the tumoral matrix [99]. Once colonized the bone, tumor cells secrete proteins that
229 interact with resident cells in the bone marrow to induce the differentiation, recruitment and
230 activation of osteoblasts and osteoclasts. Then, during the bone resorption the calcium ions and the
231 growth factors secreted from the mineralized bone matrix promote tumor cell growth, leading to
232 vicious cycle that supports tumor growth in bone and subsequent fatal outcome [94].

233 It is believed that, when primary tumor cells migrate, the interaction of these disseminated cells
234 with the new microenvironment determines whether they will proliferate to form a secondary tumor
235 or undergo growth arrest and subsequent dormancy. Dormant cells are cells that stop dividing but
236 still survive in a quiescent state, waiting for the appropriate environmental conditions to re-enter the
237 cell cycle again [100]. These cells are clinically undetectable and, consequently, constitute a major
238 issue for future tumor recurrence and metastases [101]. Current pharmacological approximations are
239 aimed at maintaining cancer cells in the dormant state; reactivating dormant cells to increase their
240 susceptibility to drugs; and eliminating cancer cells. Those strategies rely on the modulation of certain
241 factors present on or secreted by the dormant cells in such a way that their overexpression of
242 inhibition affects the fate of those dormant cells [102]. In this sense, the use of mesoporous silica
243 nanoparticles might be interesting to enhance those treatments, as they could be employed to load
244 therapeutic agents able to modulate the expression of those factors. In addition, they could be
245 employed to co-load those agents with antitumoral drugs, consequently enhancing the efficacy of the
246 treatments and minimizing tumor recurrence.

247 3.2. Nanotechnology for Cancer Treatment

248 Current anticancer treatments mainly rely on chemotherapy, radiotherapy and/or surgery [103–
249 105]. Those treatments, yet effective in many cases, present several drawbacks. In particular,
250 chemotherapy lacks of a great tumor tissue selectivity, leading to nonspecific drug distribution and
251 side effects. In this sense, nanoparticles have emerged as a powerful tool to encapsulate drugs and
252 reduce side effects [106–108].

253 The rationale behind the use of nanoparticles in cancer treatment relies on the Enhanced
254 Permeability and Retention effect (EPR effect), which is the basis of some commercialized
255 nanomedicines [109]. The EPR effect, first reported by Maeda and coworkers [110], promotes the
256 passive accumulation of nanoparticles in solid tumors as a result of the hypervascularity, the
257 enhanced permeability and the poor lymphatic drainage found in many tumors (Figure 3).



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Figure 3. EPR effect. Nanoparticles passively accumulate in the tumor owing to the presence of fenestration in the tumor blood vessels. Once there, the particles remain in the tissue for long periods of time as a consequence of the poor lymphatic drainage. Reproduced from Ref [111] with permission of MDPI.

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Owing to the uncontrolled angiogenesis, the newly formed vessels present an abnormal architecture, including wide fenestrations (200-2000 nm endothelial cell-cell gaps), irregular vascular alignment or lack of smooth muscle layer, among others. As a result, molecules larger than 40 kDa leak out from them and accumulate in the extravascular tumoral tissues. On the contrary, healthy tissues do not show this abnormal development and no accumulation is observed, thus creating a differential selectivity for cancer tissues [112]. In addition, unlike normal tissues where the extracellular fluid is constantly removed, tumors present defective lymphatic drainage and the accumulated macromolecules tend to remain in the tumoral mass for longer periods of time [113].

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The magnitude of the EPR effect in humans highly depends on the particularities of the patient and the tumor [114] although some alternative strategies, such as tumor-homing peptides or some types of cells, are currently being explored to overcome the lack of EPR effect.

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These alternative approximations have successfully been evaluated using *in vivo* tumor models, demonstrating the suitability of using MSNs for tumor drug delivery. In this sense, tumor-homing peptides (e.g., iRGD, iNGR) not only induce spontaneous accumulation of nanoparticles in the tumor tissues, but also enhance their diffusion into the tumoral mass [115,116]. In addition, there are certain types of cells with migratory properties that can transport nanoparticles directly to tumors tissues. For instance, nanoparticles can be attached to hypoxic bacteria that migrate to the hypoxic areas of tumors [117,118]. In addition, mesenchymal stem cells have been shown to migrate to tumors in response to the secretion of various signaling molecules. Then, a smart strategy is to induce the internalization of drug-loaded nanoparticles within these cells to then delivering them specifically to tumor tissues [119–122].

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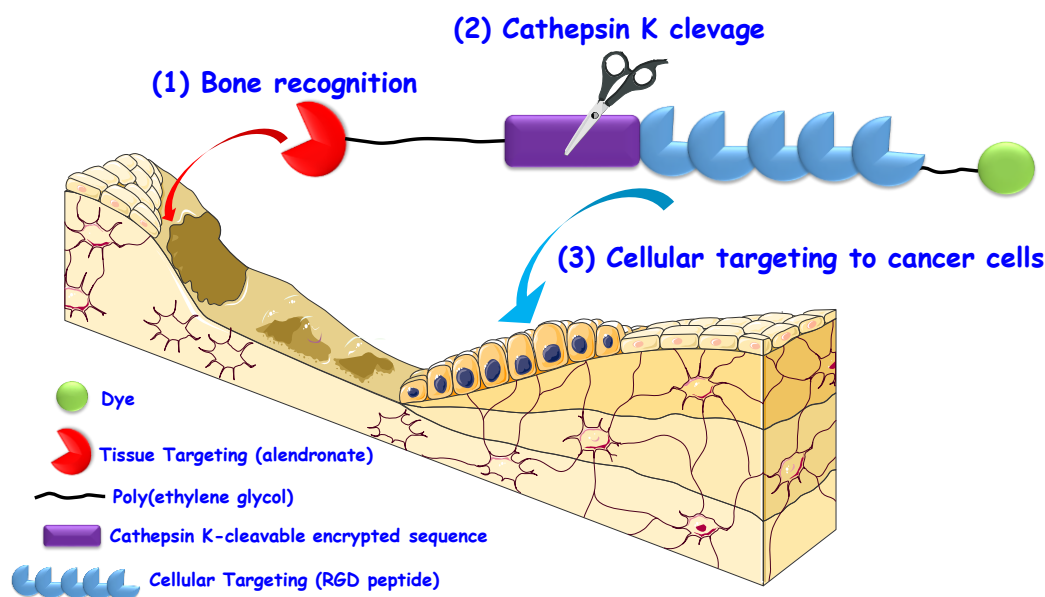
Besides delivering the nanoparticles to malignant tissues, the carriers can be engineered so that they preferentially recognize cancer cells over healthy cells. This targeting strategy relies on the overexpression of some receptors only on the membrane of tumoral cells. Examples of this approach include the functionalization of the particles with antibodies [123,124], proteins [70,125], small molecules [126–130] or peptides [131–133], among others.

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3.3. Targeting Bone-Localized Tumors with Mesoporous Silica Nanoparticles

290 Addressing nanoparticles to bone metastases is challenging, as small metastases are poorly
 291 vascularized and, consequently, the magnitude of the EPR effect is low compared to big solid tumors
 292 [134]. A smart approximation would be the modification of the particles with targeting molecules
 293 with high affinity towards calcium phosphate surfaces (bone tissue), such as bisphosphonates [135],
 294 to complement the EPR effect. In this sense, the surface modification with the bisphosphonate
 295 zoledronate has been proved to be effective in delivering MSNs to bone metastases originated from
 296 lung [136] and breast cancer [137].

297 Besides targeting the particles to bone tissue, it would be desirable for the nanomedicines to be
 298 subsequently internalized only by the tumoral cells. In this sense, our group recently reported a smart
 299 approximation for the sequential targeting of bone tumors or bone metastases that could be easily
 300 implemented into any nanomedicine (Figure 4) [138].



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302 **Figure 4.** Encrypted approach for the sequential targeting of bone cancer tissue and cancer cells. (1)
 303 The presence of a bone targeting agent (alendronate) would help accumulate the nanomedicines in
 304 the bone tumor tissue; (2) Once there, the overexpressed cathepsin K would cleave a specific peptidic
 305 sequence, (3) exposing the RGD (arginine-glycine-aspartic) motif, which is able to promote the
 306 selective uptake of nanomedicine by sarcoma tumoral cells.

307 As observed in Figure 4, the system is composed of two targeting agents and employs PEG
 308 chains to mimic a nanocarrier. The first one is the bisphosphonate alendronate, which can bind bone
 309 tissue. Then, there is a peptidic fragment containing a cathepsin K-cleavable sequence followed by
 310 the RGD motif, which is able to promote the selective internalization in osteosarcoma cells thanks to
 311 the overexpression of $\alpha\beta$ integrins. In this manner, the alendronate molecule would help the EPR
 312 effect to accumulate the nanomedicines in the bone tumor tissue. Once there, cathepsin K, which is
 313 overexpressed in bone tumors and bone metastases, would cleave the encrypting sequence, thereby
 314 exposing the RGD motif and triggering the preferential uptake of the nanomedicines.

315 As it happens with many other cancer cells, bone tumoral cells overexpress specific receptors
 316 that can be targeted using conveniently engineered MSNs. Aside from targeting MSNs to
 317 osteosarcoma [139], the RGD motif can also be employed to recognize endothelial cells, which can
 318 help MSNs target the tumor endothelium of fibrosarcoma to then eliminate the cancerous cells using
 319 multimodal therapy [140]. In this sense, folic acid can be employed to target overexpressed folate
 320 receptors in fibrosarcoma [141] and osteosarcoma cells [142]. In addition, the modification of MSNs
 321 with a glucose analog enhances their accumulation in bone tumor cells, as a consequence of their
 322 great glucose consumption due to the high metabolic demand of tumors [143]. Some surface
 323 receptors, such as the CD11c, can also be targeted using specific antibodies, which are able to trigger
 324 the selective internalization of MSNs in osteosarcoma [144].

325 The decoration of MSNs with proteins can also increase their cellular uptake. For instance, the
326 lectin concanavalin A binds overexpressed sialic acid residues to promote the cellular uptake of pH-
327 responsive MSNs in osteosarcoma cells [145]. Transferrin receptors are overexpressed in
328 fibrosarcoma cells and, consequently, the protein transferrin can be employed to enhance the uptake
329 of MSNs in those bone tumoral cells [146].

330 Besides employing active targeting moieties, MSNs can be internalized *via* electrostatic
331 interactions with the negatively charged cell membrane. The positively charged surface can be
332 shielded using PEG, which can be detached using a cleavable bond. The charge is exposed again
333 upon application of ultrasounds, which triggers the nanoparticles uptake after the accumulation in
334 the solid bone tumor *via* EPR effect [147].

335 3.4. Controlled Release of Therapeutics in Bone Tumors with Mesoporous Silica Nanoparticles

336 There are various examples of the suitability of using silica-based mesoporous nanomatrices for
337 the delivery of antitumoral [148–152] or imaging agents [139,153,154] to bone cancer cells. Moreover,
338 researchers have taken advantage of the features of the bone tumoral environment to design stimuli-
339 responsive MSNs for the treatment of sarcomas. Among the internal stimuli, the acidic environment
340 of the lysosomes can be employed to trigger the drug release from pH-responsive polymer-coated
341 MSNs [145] or pulsatile on-off MSNs whose pore entrances are sealed with pH-responsive
342 nanovalves [155]. In addition, it is possible to load immunotherapy agents within the pores of pH-
343 responsive lipid-coated MSNs for synergistic chemo-immunotherapy [156]. In addition to pH
344 variations, the enzyme alkaline phosphatase, which is characteristic bone-related tumors, can be
345 employed to degrade the gatekeepers of silica-based mesoporous glasses [157]. Moreover, the
346 esterase enzymes can also be employed to cleave the nanocaps of MSNs [142].

347 There are some examples of the use of light to trigger the drug release from MSNs in bone tumor
348 scenarios. For instance, ultraviolet light can be employed to cleave light-responsive bonds connected
349 to transferrin, which acts as both gatekeeper and targeting agent, triggering the drug release [146]. In
350 addition, porphyrins can be engineered as gatekeepers using a linker cleavable in the presence of
351 singlet oxygen, which are self-produced by the porphyrin caps upon application of visible light [158].

352 Aside from delivering small therapeutic molecules, MSNs allow the effective delivery of
353 proteins [159] or DNA strands [160] into bone cancer cells. There is a type of nucleic acids, small
354 interfering RNA (siRNA), that triggers the knockdown of specific and relevant proteins, which makes
355 them useful for the treatment of various diseases [161]. Unfortunately, siRNAs have short half-life,
356 poor penetration through cell membranes and easily degrade upon RNase action in the organism
357 [162]. For that reason, the use of MSNs as protective shell for these nucleic acids have been widely
358 explored. In this sense, the polo-like kinase 1, which is an essential gene for the correct execution of
359 cell division [163], is overexpressed in bone tumors and has been targeted with great efficacy using
360 siRNA-loaded MSNs [164–167].

361 A summary of all the nanocarriers here described for bone tumors is summarized in Table 1.

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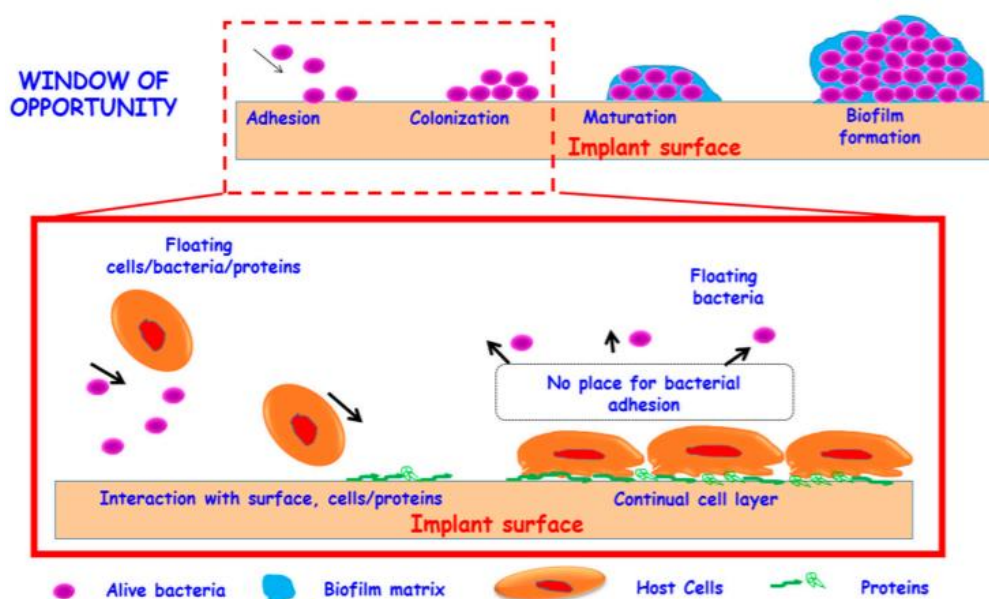
Table 1. Summary of the different silica-based nanocarriers applied for the treatment of bone tumors

Cell line	Description	Reference
Osteosarcoma		
MG-63	MSNs loaded with ammonia borate as negative computed tomography contrast agents for the diagnosis of osteosarcoma	[154]
	Silica-based mesoporous glass nanospheres for the delivery of alendronate against osteosarcoma cells and osteoclasts	[150]
	Silica-based mesoporous glasses with osteogenic properties for the release of alendronate against osteosarcoma cells	[149]
	Eu-doped silica-based mesoporous glass nanospheres with osteogenic properties for the release of doxorubicin	[148]
	Influence of the different functionalizations of MSNs on their uptake by osteosarcoma cells	[144]
KHOS	Poly-L-lysine-coated MSNs for the delivery of siRNA to knockdown polo-like kinase 1	[165]
	MSNs with large mesopores for the delivery of siRNA to knockdown polo-like kinase 1	[164]
	Co-loading of topotecan and siRNA to knockdown polo-like kinase 1 in dendrimer-like MSNs	[166]
	PEI-coated MSNs for the delivery of siRNA to knockdown polo-like kinase 1	[167]
HOS	Stimuli-responsive silica-based mesoporous glasses responsive to alkaline phosphatase overexpressed in bone tumors	[157]
	Dendrimer-coated MSNs for the delivery of non-viral oligonucleotides	[160]
	MSNs functionalized with singlet oxygen-sensitive porphyrin caps for release of topotecan	[168]
	MSNs engineered for ultrasound-induced cellular uptake through the detachment of a shielding PEG layer	[147]
	Concanavalin A-targeted and pH-responsive MSNs for the delivery of doxorubicin	[169]
HTB-85	Silica-based mesoporous glass nanospheres with osteogenic properties for the release of doxorubicin	[151]
U2Os	Folic acid-targeted MSNs for enzyme-responsive release of camptothecin	[142]
UMR-106	RGD-targeted and Bi-doped MSNs for chemo-photothermal therapy and imaging	[139]
Fibrosarcoma		
L-929	Ultrasound, pH and magnetically-responsive on-off gated MSNs for the delivery of doxorubicin	[155]
	Gd-doped MSNs for magnetic resonance imaging of fibrosarcoma	[153]
	pH-responsive MSNs for the intracellular delivery of proteins	[159]
	pH-responsive MSNs for combined chemo-immunotherapy	[156]
HT-1080	Influence of MSNs size on the doxorubicin release and the uptake of the particles by fibrosarcoma cells	[152]
	MSNs decorated through an ultraviolet light-responsive linker with transferrin acting as gatekeeper and targeting agent	[125]
	RGD-targeted MSNs for multimodal treatment of fibrosarcoma in a chicken embryo model	[140]

365 **4. Mesoporous Silica Nanoparticles for the Treatment of Bone Infection**

366 *4.1. General Concepts on Bacterial Bone Infections*

367 Bone infection is a major issue for health care systems and entails important socioeconomic
 368 implications [170]. The appearance of bone infections is directly related with the progressive ageing
 369 of current society and, consequently, the increased use of implantable medical devices and their
 370 potential bacterial contamination. These infections are mainly caused by *Staphylococcus epidermis*,
 371 *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa* [40]. Regular bacteria can be
 372 relatively easy eliminated using antibiotics. However, the inappropriate use of those antimicrobials
 373 is progressively leading to more cases of drug-resistant bacteria, which are expected to cause more
 374 than 10 million deaths by 2050 [171]. This antimicrobial resistance induces uncontrolled bacterial
 375 growth and formation of persistent biofilms. Biofilms are communities of microorganisms embedded
 376 in a self-produced polysaccharide matrix [172]. This protective matrix endows them with resistance
 377 to antibiotics and host immune systems that, otherwise, would eliminate bacteria in their planktonic
 378 state (free-floating bacteria) [173]. The biofilm—related antimicrobial resistance relies, not only on
 379 the physical hindrance of the matrix, but also on (1) the presence of bacterial and host DNA and
 380 proteins that may increase the shielding capacity of the matrix [174]; (2) the presence of bacteria with
 381 different acquired resistances and antibiotic sensitivities [175]; (3) the development of efflux pumps
 382 [176]; (4) the presence of enzymes able to degrade antimicrobials [177] and (5) the establishment of
 383 quorum sensing (bacteria-bacteria communication) [178]. The process of biofilm formation is
 384 depicted in Figure 3.



385

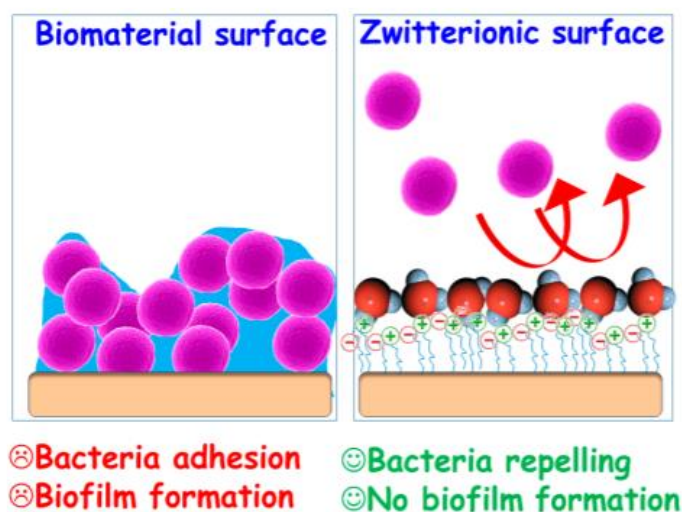
386 **Figure 3.** Schematic representation of biofilm formation on an implant surface. The process involves
 387 4 steps: (1) bacterial adhesion, (2) bacterial growth, (3) maturation and (4) biofilm formation. In
 388 addition, bacteria may leak out from the matrix and lead to bacterial dispersion. The first stages
 389 constitute a *window of opportunity*, in which it is still possible to prevent biofilm formation.
 390 Reproduced from Ref [40] with permission of MDPI.

391 The formation of the biofilm comprises 4 steps: (1) adhesion of bacteria to the implant surface;
 392 (2) bacterial growth in multiple bacterial layers; (3) maturation and (4) final biofilm formation. In
 393 addition, bacteria detach from the biofilm to then colonize other areas and induce further infections
 394 [179]. As observed in Figure 3, during the first phases of biofilm formation the individual
 395 microorganisms are floating on the implant, reversibly interacting with the surface. In consequence,

396 these stages constitute a *window of opportunity* that clinicians should take advantage of to prevent
397 irreversible biofilm formation and subsequent resistance [40].

398 4.2. Preventing Protein and Bacterial Adhesion and Biofilm Formation: Zwitterionic Mesoporous Silica 399 Nanoparticles.

400 In view of the evidences exposed in the previous subsection, avoiding bacterial contamination
401 of implants constitutes a major concern. In this sense, the development of the so-called *zwitterionic*
402 materials has fueled the design of antifouling nanostructured materials able to prevent protein
403 adsorption, bacterial adhesion and biofilm formation (Figure 4).
404



405

406 Figure 4. Schematic representation of bacterial colonization in standard surfaces *vs.* *zwitterionic*
407 surfaces. Unlike in unmodified surfaces, *zwitterionic* materials create a hydration layer that prevents
408 bacterial adhesion and biofilm formation. Reproduced from Ref [40] with permission of MDPI.

409 *Zwitterionic* surfaces are characterized by an equal number of negative and positive charges, so
410 the net charge is expected to be neutral. This neutrality leads to the formation of a hydration layer
411 onto the surface that physically hampers adhesion and biofilm formation [180]. In fact, owing to the
412 reduced protein adsorption, *zwitterionic* functionalizations have also been postulated as substitutes
413 for PEGylation [181], which might be beneficial to overcome the growing appearance of anti-PEG
414 antibodies [182].

415 The first example of mesoporous silica materials with *zwitterionic* behavior was reported by our
416 group back in 2010, using SBA-15 mesoporous materials modified with randomly distributed amino
417 and carboxylic acid short chains on the surface that resulted in significantly lower protein adhesion
418 [183]. A similar approach using amino and phosphonate groups was recently reported, yielding
419 MSNs with extremely low protein adsorption and excellent antibacterial properties. In addition, the
420 nanoparticles showed great biocompatibility with preosteoblasts, assuring their biocompatibility for
421 the treatment of bone infection [184]. Interestingly, this *zwitterionic* approach using two small
422 molecules can be employed to design pH-responsive gatekeepers by taking advantage of the
423 interaction between both short chains, which interact at physiological pH and experience repulsion
424 forces at acid pH [185].

425 Aside from merging molecules with opposite charges, there are molecules that are *zwitterionic*
426 in nature. In this sense, the modification of MSNs with phosphorylcholine groups yields
427 nanoparticles showing reduced protein adsorption and able to provide sustained drug release in
428 response to changes in pH [186]. An analogous approximation is the modification of MSNs with
429 sulfobetaine groups to prevent protein adhesion [187]. Moreover, it is possible to polymerize this
430 kind of *zwitterionic* molecules to yield polymer-coated nanoparticles with low protein binding affinity
431 [188]. In addition, there are some amino acids that are useful for the design of this kind of surfaces.
432 For instance, the amino acid lysine presents this behavior owing to the $-NH_3^+/COO^-$ pairs and has

433 been grafted to MSNs [189] and silica-based mesoporous bioactive glasses [190], leading to reduced
434 bacterial adhesion and biofilm formation. A similar approach consists in using the amino acid
435 cysteine to obtain neutral surfaces, yielding MSNs with high stability in human serum [191].

436 4.3. Addressing Bone Infections with Mesoporous Silica Nanoparticles

437 Besides preventing biofilm formation, it is still necessary the elimination of the infection. In this
438 sense, it is possible to engineer multifunctional mesoporous silica nanomatrices able to prevent
439 bacterial adhesion and biofilm formation and to release antimicrobials in a controlled manner only
440 in infected bone tissues [192,193]. In addition, there are examples of stimuli-responsive mesoporous
441 bioactive silica-based nanomatrices able to trigger the release only in the presence of proteolytic
442 enzymes characteristics of infected bone tissue scenarios [157,194].

443 In an effort to increase the efficiency of the delivery and, consequently, a reduction of the dose,
444 the research efforts have been headed towards the development of bacteria-targeted MSNs. In this
445 sense, the presence of positive charges on the surface of the particles increases their affinity to the
446 negatively charged biofilm and bacteria wall. In this manner, it is easier for the particles to diffuse
447 into the biofilm to then interact with bacteria and exert their therapeutic effect. Examples of this
448 approach include the use of short positively charged alkoxy silanes [195] or third-generation
449 dendrimers, whose great number of positive charges is able to permeate the bacteria wall and induce
450 the MSNs internalization [196]. Besides using positively charged MSNs, lectins have been shown to
451 be effective in targeting and promoting internalization of MSNs into the biofilm, as a consequence of
452 the presence of glycan-type polysaccharides in this protective matrix. In fact, the lectin concanavalin
453 A is able to trigger this internalization and exert antibacterial effect by itself, which is even more
454 emphasized when loading an antibiotic in the mesopores [197].

455 A smart approximation to enhance the possibilities that mesoporous silica materials may offer
456 against bone infection is the incorporation of the particles within scaffolds. In the context of bone
457 diseases, scaffolds are materials that are intended to mimic bone tissue and contribute to its
458 regeneration. The advantages over using bare scaffolds are increased antibiotic loading capacity or
459 controlled drug release, among others [198]. Examples of this approximation are the incorporation of
460 silica-based mesoporous glasses in PLGA (poly-(L-lactic-co-glycolic acid)) [199] or MSNs in porous
461 collagen gelatin [200] for the controlled release of vancomycin against bone infection. In addition,
462 MSNs-loaded scaffolds allow the co-delivery of therapeutic compounds. In this sense, it is possible
463 to load cephalexin within the mesopores and vascular endothelial growth factors in the scaffold
464 structure to achieve bacteria elimination and bone reconstruction [201].

465 A summary of different materials for the treatment of bone infection can be found in Table 2.

466
467



Table 2. Summary of mesoporous silica-based materials against bone infection

Bacteria	Description	Reference
	Pronase-responsive gatekeepers for levofloxacin-loaded silica-based mesoporous glasses	[202]
	Levofloxacin-loaded <i>Zwitterionic</i> MSNs with reduced protein adhesion	[184]
	Lysine-coated MSNs to inhibit <i>e.coli</i> adhesion	[189]
<i>E.coli</i>	Acid phosphatase-responsive gatekeepers for levofloxacin-loaded silica-based mesoporous glasses	[203]
	Positively charge MSNs target the bacteria wall of <i>e.coli</i>	[195]
	Levofloxacin-loaded MSNs coated with polycationic dendrimers destroys biofilm and internalize in bacteria	[204]
	Levofloxacin-loaded MSNs decorated with concanavalin A targets and internalize the biofilm	[197]
	Levofloxacin-loaded <i>Zwitterionic</i> MSNs with reduced protein adhesion	[184]
	Lysine-coated <i>zwitterionic</i> MSNs to inhibit <i>s.aureus</i> adhesion and <i>s.aureus</i> biofilm formation	[189]
	Lysine-coated <i>zwitterionic</i> silica-based mesoporous glasses to prevent <i>s.aureus</i> adhesion	[190]
<i>S.aureus</i>	Levofloxacin-loaded and positively charged MSNs targets and destroy <i>s.aureus</i> biofilm and bacteria	[195]
	MSNs-loaded scaffolds for the co-delivery of cephalexin and vascular endothelial growth factor	[201]
	Vancomycin-loaded silica-based mesoporous glasses contained in PLGA scaffolds	[199]
	Vancomycin-loaded MSNs contained in collagen gelatin scaffolds	[200]

470 5. Mesoporous Silica Nanoparticles for the Treatment of Osteoporosis

471 5.1. General Concepts on Osteoporosis

472 Osteoporosis is the most frequent metabolic disease affecting bone tissue. It is characterized by
473 reduced bone mass and microarchitectural deterioration and results in more than 9 million fractures
474 annually worldwide (one osteoporotic fracture every 3 seconds) [205], with special incidence in aged
475 women [206]. Its origin relies on the alteration of the bone remodeling process, which consists in the
476 removal of old bone (osteoclast) to then create new one (osteoblasts). The imbalance of this process
477 leads to reduced bone mass and, consequently, osteoporosis.

478 Current osteoporosis treatments, which are not fully satisfactory, are limited to anti-resorptive
479 drugs and anabolic agents [207,208]. Anti-resorptive drugs decrease the excess of bone resorption by
480 targeting osteoclast activity. Examples of these compounds include bisphosphonates [209], raloxifene
481 [210] or denosumab [211]. The excess of bone resorption can be counteracted using anabolic agents,
482 which are compounds able to stimulate bone formation. Examples of these drugs are human
483 parathyroid hormone [212], growth factors or siRNA [213].

484 Unfortunately, current treatments present some drawbacks. For instance, bisphosphonates are
485 known to induce gastric side effects or fractures after long use. Raloxifene may cause venous
486 thromboembolism. Moreover, cases of hypocalcemia, anaphylaxis or atrial fibrillation have been
487 associated to denosumab. In addition, anabolic agents, such as siRNA, might be easily degraded by
488 the harsh environment present in the organism [212]. These issues might be addressed by delivering
489 the antiosteoporotic agents specifically to the diseased bone tissues and, consequently, the use of
490 nanoparticles seems highly appealing.

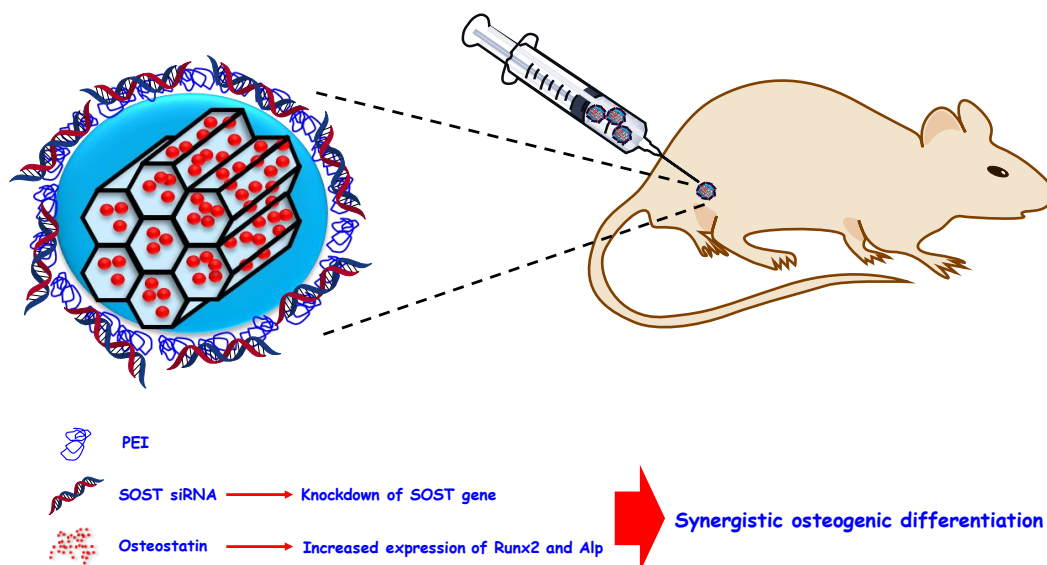
491 5.2. Addressing Osteoporosis with Mesoporous Silica Nanoparticles

492 The first example of mesoporous silica materials applied for the controlled release of anti-
493 resorptive molecules was reported by our group back in 2006, when MCM-41 and SBA-15 materials
494 were employed for the loading and controlled release of alendronate [214]. In this sense, the
495 introduction of phosphorous groups in SBA-15 mesoporous silica nanomatrices enhanced the
496 loading of alendronate and induced the formation of apatite, a component of bone, making these
497 materials promising candidates for the treatment of osteoporosis [215]. Additional examples of anti-
498 resorptive molecules loaded in mesoporous silica-based nanoparticles are ipriflavone [216], salmon
499 calcitonin [217] or zoledronic acid [218], showing all of them promising results in terms of anti-
500 osteoclast activity and osteogenesis.

501 A great feature of MSNs is that they allow the loading of hydrophobic compounds, consequently
502 enhancing their bioavailability. In this sense, they allow the incorporation within their mesopores of
503 sparingly soluble anabolic agents able to induce bone formation. Examples are the loading of
504 dexamethasone, which induces bone regeneration through the stimulation of bone mesenchymal
505 stem cells [219], or estradiol, which enhances the biological functions of osteoblast and inhibits the
506 proliferation of osteoclasts [220].

507 Osteostatin, a C-terminal peptide from a parathyroid hormone-related protein, induces strong
508 bone anabolism through a great stimulation of osteoblastogenesis [221]. It has been shown that
509 osteostatin-loaded SBA-15 greatly stimulate osteoblastic growth *in vitro* [222]. Furthermore, these
510 osteostatin-loaded mesoporous materials have been proved to be effective in regenerating bone
511 defects *in vivo* [212,223]. In addition to osteostatin, the bone morphogenic protein-2 (BMP-2) is
512 considered to be one of the most effective growth factors to induce osteoblast differentiation and
513 boost bone regeneration. In this sense, MSNs are useful for the co-delivery of dexamethasone and
514 BMP-2 to achieve great bone regeneration *in vivo* [224]. Moreover, the residues 73-92 of BMP-2 not
515 only promote osteogenesis and bone regeneration but also increase the internalization of bone
516 mesenchymal stem cells of MSNs decorated with this peptidic fragment. [225].

517 Aside from being useful for bone cancer treatment, siRNA molecules also find application in the
 518 treatment of osteoporosis. In this sense, the localized release of siRNA able to knockdown RANK
 519 from silica-based mesoporous bioactive glasses has been shown to be highly effective in
 520 suppressing osteoclastogenesis and, consequently, osteoporosis [226]. A similar approach against
 521 osteoporosis was recently reported by our group using an *in vivo* model of ovariectomized mice
 522 (Figure 5).



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Figure 5. PEI-coated MSNs as anti-osteoporotic nanocarrier. Osteostatin was loaded in the mesopores and a siRNA able to knockdown SOST gene was introduced within the polymeric mesh. The co-delivery of both therapeutic agents resulted in synergistic osteogenesis in ovariectomized mice. PEI: Polyethyleneimine.

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MSNs can load therapeutic compounds not only in their mesopores but also within polymeric coatings through electrostatic interaction. In this sense, Figure 5 shows MSNs carrying the anabolic agent osteostatin in the pores and a specific siRNA able to knockdown SOST gene interacting with a PEI coating. This gene encodes the protein sclerostin, which can inhibit the Wnt/ β -catenin pathway, a major signaling carrier that regulates bone development and remodeling. Based on this, the siRNA and osteostatin-loaded nanoparticles were administered to osteoporotic ovariectomized mice, showing synergistic effects on all the bone regeneration biomarkers studied [227].

There are some metal ion species known to induce osteogenesis. For instance, copper ions enhance bone density by inhibiting bone resorption, and their incorporation in mesoporous silica nanospheres has been proved to be effective in stimulating the differentiation of bone mesenchymal stem cells into the osteogenic lineage [228]. Moreover, impregnating silica-based mesoporous bioactive glasses with Ga (III) leads to the formation of apatite together with the disruption of osteoclastogenesis and early differentiation of pre-osteoblast towards osteoblastic phenotype [229]. In addition, the osteogenic ability of Zn^{2+} ions is enhanced when the ions are co-delivered with osteostatin from silica-based mesoporous bioactive glasses [230]. Furthermore, there are nanoparticles able to stimulate bone regeneration *per se*. Examples of these kind of behavior are Au nanoparticles supported on MSNs that increase the osteogenic capability of preosteoblastic cells [231] or silica-based mesoporous bioactive glasses that are capable of reducing the bone-resorbing capability of osteoclasts [232].

A summary of all the above-described materials for the treatment of osteoporosis can be found in Table 3.

Table 3. Summary of silica-based mesoporous materials for the treatment of osteoporosis

Therapeutic agent	Description	Reference
Anti-resorptive treatment		
Alendronate	First example of controlled release of bisphosphonates from mesoporous silica materials (MCM-41 and SBA-15)	[214]
	Phosphorus-containing SBA-15 mesoporous silica materials for bone regeneration and release of alendronate	[215]
Ipriflavone	Silica-based mesoporous nanospheres for the release of ipriflavone without affecting osteoblast viability	[216]
Zoledronic acid	Zoledronic acid-loaded MSNs/hydroxyapatite coatings on implants with enhanced inhibition of osteoclasts activity	[218]
Salmon calcitonin	MSNs for the release of salmon calcitonin with significant therapeutic effects <i>in vivo</i>	[217]
siRNA (RANK)	Silica-based mesoporous glass nanospheres to deliver of siRNA to knockdown RANK and inhibit osteoclastogenesis	[226]
Ions	Mesoporous silica-based nanospheres for the delivery of Cu ions able to inhibit osteoclastogenesis	[228]
	Silica-based mesoporous glasses for the release of Ga ions able to disturb osteoclastogenesis	[229]
Particle	Silica-based mesoporous glasses reduce the bone-resorbing capability of osteoclasts <i>per se</i>	[232]
	Au nanoparticles supported on MSNs increases the osteogenic capability of pre-osteoblastic cells	[231]
Anabolic treatment		
Dexamethasone	Alendronate-targeted MSNs for the delivery of dexamethasone to bone tissue	[219]
Estradiol	Multilayered-coated MSNs for the delivery of estradiol from titanium substrates	[220]
Osteostatin	Osteostatin-loaded SBA-15 mesoporous silica materials stimulates the growth and differentiations of osteoblasts	[222]
	Osteostatin-loaded SBA-15 mesoporous materials regenerates bone in a rabbit femur cavity defect	[212]
	Osteostatin-loaded SBA-15 mesoporous silica materials increase the early repair response in bone after local injury	[233]
BMP-2 and dexamethasone	pH-responsive co-delivery of dexamethasone and BMP-2 protein for synergistic osteogenic effect	[224]
	BMP-2 derived peptide-decorated MSNs for enhanced uptake in bone mesenchymal stem cells and synergistic effect of the peptidic fragment and dexamethasone	[225]
Osteostatin and siRNA (SOST)	Enhanced osteogenic expression through MSNs co-delivering osteostatin and siRNA able to knockdown SOST gene	[227]
Zn ions and osteostatin	Co-delivery of osteogenic Zn ions and osteostatin from mesoporous silica-based glasses induces high osteogenic response	[230]

552 **6. Conclusions**

553 Complex bone diseases, such as bone cancer, bone infection and osteoporosis, constitute a major
554 concern for our progressively aged modern societies. Most of the current treatments present several
555 drawbacks, leading to the deterioration of the patient health and the subsequent socioeconomic
556 impact. In this sense, the use of nanoparticles, in particular mesoporous silica-based nanoparticles,
557 has emerged as a powerful approximation to reduce the different side effects. This type of
558 nanoparticles present high loading capacities, biocompatibility and can be engineered to prevent
559 premature drug release and address the particles to the affected tissues. The different nanosystems
560 here presented constitute reliable approximations for the treatment of bone diseases and,
561 consequently, current research should be headed towards the effective translation of these
562 nanomaterials into the clinic.

563 **Author Contributions:** All authors have contributed equally.

564 **Funding:** The research was funded by the European Research Council through ERC-2015-AdG-694160 (VERDI)
565 grant.

566 **Acknowledgments:** The authors acknowledge financial support from European Research Council through ERC-
567 2015-AdG-694160 (VERDI) project.

568 **Conflicts of Interest:** The authors declare no conflict of interest.

569 **References**

- 570 1. Appell, D. Wired for success. *Nature* **2002**, *419*, 553–555.
- 571 2. Serrano, E.; Rus, G.; García-Martínez, J. Nanotechnology for sustainable energy. *Renew. Sustain. Energy*
572 *Rev.* **2009**, *13*, 2373–2384.
- 573 3. Tong, H.; Ouyang, S.; Bi, Y.; Umezawa, N.; Oshikiri, M.; Ye, J. Nano-photocatalytic Materials:
574 Possibilities and Challenges. *Adv. Mater.* **2012**, *24*, 229–251.
- 575 4. Nie, S.; Xing, Y.; Kim, G.J.; Simons, J.W. Nanotechnology Applications in Cancer. *Annu. Rev. Biomed.*
576 *Eng.* **2007**, *9*, 257–288.
- 577 5. Wicki, A.; Witzigmann, D.; Balasubramanian, V.; Huwyler, J. Nanomedicine in cancer therapy:
578 Challenges, opportunities, and clinical applications. *J. Control. Release* **2015**, *200*, 138–157.
- 579 6. Zhu, X.; Radovic-Moreno, A.F.; Wu, J.; Langer, R.; Shi, J. Nanomedicine in the management of microbial
580 infection – Overview and perspectives. *Nano Today* **2014**, *9*, 478–498.
- 581 7. Doane, T.L.; Burda, C. The unique role of nanoparticles in nanomedicine: imaging, drug delivery and
582 therapy. *Chem. Soc. Rev.* **2012**, *41*, 2885.
- 583 8. Bangham, A.D.; Standish, M.M.; Weissmann, G. The Action of Steroids and Streptolysin S on the
584 Permeability of Phospholipid Structures to Cations. *J. Mol. Biol.* **1965**, *13*, 253–259.
- 585 9. Bangham, A.D.; Horne, R.W. Negative Staining of Phospholipids and their Structural Modification by
586 Surface-active Agents as observed in the Electron Microscope. *J. Mol. Biol.* **1964**, *8*, 660–668.
- 587 10. Bozzuto, G.; Molinari, A. Liposomes as nanomedical devices. *Int. J. Nanomedicine* **2015**, *10*, 975–999.
- 588 11. Kumari, A.; Yadav, S.K.; Yadav, S.C. Biodegradable polymeric nanoparticles based drug delivery
589 systems. *Colloids Surfaces B Biointerfaces* **2010**, *75*, 1–18.
- 590 12. Oerlemans, C.; Bult, W.; Bos, M.; Storm, G.; Nijsen, J.F.W.; Hennink, W.E. Polymeric Micelles in
591 Anticancer Therapy: Targeting, Imaging and Triggered Release. *Pharm. Res.* **2010**, *27*, 2569–2589.

- 592 13. Azharuddin, M.; Zhu, G.H.; Das, D.; Ozgur, E.; Uzun, L.; Turner, A.P.F.; Patra, H.K. A repertoire of
593 biomedical applications of noble metal nanoparticles. *Chem. Commun.* **2019**, *55*, 6964–6996.
- 594 14. Maiti, D.; Tong, X.; Mou, X.; Yang, K. Carbon-Based Nanomaterials for Biomedical Applications: A
595 Recent Study. *Front. Pharmacol.* **2019**, *9*, 1401.
- 596 15. Manzano, M.; Vallet-Regí, M. New developments in ordered mesoporous materials for drug delivery. *J.*
597 *Mater. Chem.* **2010**, *20*, 5593–5604.
- 598 16. Argyo, C.; Weiss, V.; Bräuchle, C.; Bein, T. Multifunctional Mesoporous Silica Nanoparticles as a
599 Universal Platform for Drug Delivery Multifunctional Mesoporous Silica Nanoparticles as a Universal
600 Platform for Drug Delivery. *Chem. Mater.* **2014**, *26*, 435–451.
- 601 17. Zhang, Y.; Zhi, Z.; Jiang, T.; Zhang, J.; Wang, Z.; Wang, S. Spherical mesoporous silica nanoparticles for
602 loading and release of the poorly water-soluble drug telmisartan. *J. Control. Release* **2010**, *145*, 257–263.
- 603 18. Narayan, R.; Nayak, U.Y.; Raichur, A.M.; Garg, S. Mesoporous silica nanoparticles: A comprehensive
604 review on synthesis and recent advances. *Pharmaceutics* **2018**, *10*.
- 605 19. Rosenholm, J.M.; Mamaeva, V.; Sahlgren, C.; Lindén, M. Nanoparticles in targeted cancer therapy:
606 Mesoporous silica nanoparticles entering preclinical development stage. *Nanomedicine* **2012**, *7*, 111–120.
- 607 20. Bukara, K.; Schueller, L.; Rosier, J.; Martens, M.A.; Daems, T.; Verheyden, L.; Eelen, S.; Van Speybroeck,
608 M.; Libanati, C.; Martens, J.A.; et al. Ordered mesoporous silica to enhance the bioavailability of poorly
609 water-soluble drugs: Proof of concept in man. *Eur. J. Pharm. Biopharm.* **2016**, *108*, 220–225.
- 610 21. Phillips, E.; Penate-Medina, O.; Zanzonico, P.B.; Carvajal, R.D.; Mohan, P.; Ye, Y.; Humm, J.; Gönen, M.;
611 Kalaigian, H.; Schöder, H.; et al. Clinical translation of an ultrasmall inorganic optical-PET imaging
612 nanoparticle probe. *Sci. Transl. Med.* **2014**, *6*, 1–10.
- 613 22. Sobczak, D. *Population ageing in Europe: facts, implications and policies*; Publications Office of the European
614 Union, 2014;
- 615 23. Kresge, C.T.; Leonowicz, M.E.; Roth, W.J.; Vartuli, J.C.; Beck, J.S. Ordered mesoporous molecular sieves
616 synthesized by a liquid-crystal template mechanism. *Nature* **1992**, *359*, 710–712.
- 617 24. Yanagisawa, T.; Shimizu, T.; Kuroda, K.; Kato, C. The preparation of alkyltrimethylammonium-
618 kanemite complexes and their conversion to microporous materials. *Bull. Chem. Soc. Jpn.* **1990**, *63*, 988–
619 992.
- 620 25. Hoffmann, F.; Cornelius, M.; Morell, J.; Fröba, M. Silica-based mesoporous organic-inorganic hybrid
621 materials. *Angew. Chemie - Int. Ed.* **2006**, *45*, 3216–3251.
- 622 26. Vallet-Regí, M.; Balas, F.; Arcos, D. Mesoporous materials for drug delivery. *Angew. Chemie - Int. Ed.*
623 **2007**, *46*, 7548–7558.
- 624 27. Walcarius, A.; Mercier, L. Mesoporous organosilica adsorbents : nanoengineered materials for removal
625 of organic and inorganic pollutants. *J. Mater. Chem.* **2010**, *20*, 4478–4511.
- 626 28. Sangvanich, T.; Morry, J.; Fox, C.; Ngamcherdtrakul, W.; Goodyear, S.; Castro, D.; Fryxell, G.E.;
627 Addleman, R.S.; Summers, A.O.; Yantasee, W. Novel Oral Detoxification of Mercury, Cadmium, And
628 Lead with Thiol-Modified Nanoporous Silica. *ACS Appl. Mater. Interfaces* **2014**, *6*, 5483–5493.
- 629 29. Yan, Z.; Meng, H.; Shi, L.; Li, Z.; Kang, P. Synthesis of mesoporous hollow carbon hemispheres as highly
630 efficient Pd electrocatalyst support for ethanol oxidation. *Electrochem. commun.* **2010**, *12*, 689–692.
- 631 30. Serrano, E.; Linares, N.; García-Martínez, J.; Berenguer, J.R. Sol–Gel Coordination Chemistry: Building
632 Catalysts from the Bottom-Up. *ChemCatChem* **2013**, *5*, 844–860.

- 633 31. Zhang, Y.; Zheng, S.; Zhu, S.; Ma, J.; Sun, Z.; Farid, M. Evaluation of paraffin infiltrated in various porous
634 silica matrices as shape-stabilized phase change materials for thermal energy storage. *Energy Convers.*
635 *Manag.* **2018**, *171*, 361–370.
- 636 32. Mitran, R.A.; Berger, D.; Munteanu, C.; Matei, C. Evaluation of Different Mesoporous Silica Supports for
637 Energy Storage in Shape-Stabilized Phase Change Materials with Dual Thermal Responses. *J. Org. Chem.*
638 *C* **2015**, *119*, 15177–15184.
- 639 33. Vallet-Regí, M.; Rámila, A.; Del Real, R.P.; Pérez-Pariente, J. A new property of MCM-41: Drug delivery
640 system. *Chem. Mater.* **2001**, *13*, 308–311.
- 641 34. Wang, Y.; Zhao, Q.; Han, N.; Bai, L.; Li, J.; Liu, J.; Che, E.; Hu, L.; Zhang, Q.; Jiang, T.; et al. Mesoporous
642 silica nanoparticles in drug delivery and biomedical applications. *Nanomedicine Nanotechnology, Biol.*
643 *Med.* **2015**, *11*, 313–327.
- 644 35. Giret, S.; Wong Chi Man, M.; Carcel, C. Mesoporous-Silica-Functionalized Nanoparticles for Drug
645 Delivery. *Chem. – A Eur. J.* **2015**, *21*, 13850–13865.
- 646 36. Trewyn, B.G.; Nieweg, J.A.; Zhao, Y.; Lin, V.S. Biocompatible mesoporous silica nanoparticles with
647 different morphologies for animal cell membrane penetration. *Chem. Eng. J.* **2008**, *137*, 23–29.
- 648 37. Slowing, I.I.; Vivero-Escoto, J.L.; Wu, C.-W.; Lin, V.S. Mesoporous silica nanoparticles as controlled
649 release drug delivery and gene transfection carriers. *Adv. Drug Deliv. Rev.* **2008**, *60*, 1278–1288.
- 650 38. Paris, J.L.; De la Torre, P.; Cabañas, M.V.; Manzano, M.; Flores, A.I.; Vallet-Regí, M. Suicide-gene
651 transfection of tumor-tropic placental stem cells employing ultrasound-responsive nanoparticles. *Acta*
652 *Biomater.* **2019**, *83*, 372–378.
- 653 39. Vallet-Regí, M.; Colilla, M.; Izquierdo-Barba, I. Bioactive Mesoporous Silicas as Controlled Delivery
654 Systems: Application in Bone Tissue Regeneration. *J. Biomed. Nanotechnol.* **2008**, *4*, 1–13.
- 655 40. Colilla, M.; Izquierdo-Barba, I.; Vallet-Regí, M. The Role of Zwitterionic Materials in the Fight against
656 Proteins and Bacteria. *Medicines* **2018**, *5*, 125.
- 657 41. Jia, Y.; Zhang, P.; Sun, Y.; Kang, Q.; Xu, J.; Zhang, C.; Chai, Y. Regeneration of large bone defects using
658 mesoporous silica coated magnetic nanoparticles during distraction osteogenesis. *Nanomedicine*
659 *Nanotechnology, Biol. Med.* **2019**, *21*, 102040.
- 660 42. Arcos, D.; Vallet-Regí, M. Sol–gel silica-based biomaterials and bone tissue regeneration. *Acta Biomater.*
661 **2010**, *6*, 2874–2888.
- 662 43. Stöber, W.; Fink, A.; Bohn, E. Controlled Growth of Monodisperse Silica Spheres in the Micron Size
663 Range. *J. Colloid Interface Sci.* **1968**, *26*, 62–69.
- 664 44. Wu, S.H.; Lin, H.P. Synthesis of mesoporous silica nanoparticles. *Chem. Soc. Rev.* **2013**, *42*, 3862–3875.
- 665 45. Möller, K.; Bein, T. Talented Mesoporous Silica Nanoparticles. *Chem. Mater.* **2017**, *29*, 371–388.
- 666 46. Kecht, J.; Bein, T. Oxidative removal of template molecules and organic functionalities in mesoporous
667 silica nanoparticles by H₂O₂ treatment. *Microporous Mesoporous Mater.* **2008**, *116*, 123–130.
- 668 47. Wang, G.; Otuonye, A.N.; Blair, E.A.; Denton, K.; Tao, Z.; Asefa, T. Functionalized mesoporous materials
669 for adsorption and release of different drug molecules: A comparative study. *J. Solid State Chem.* **2009**,
670 *182*, 1649–1660.
- 671 48. Nieto, A.; Balas, F.; Manzano, M.; Vallet-Regí, M. Functionalization degree of SBA-15 as key factor to
672 modulate sodium alendronate dosage. *Microporous Mesoporous Mater.* **2008**, *116*, 4–13.
- 673 49. Vallet-Regí, M.; Manzano, M.; González-Calbet, J.M.; Okunishi, E. Evidence of drug confinement into

- 674 silica mesoporous matrices by STEM spherical aberration corrected microscopy. *Chem. Commun.* **2010**,
675 46, 2956–8.
- 676 50. Moreira, A.F.; Dias, D.R.; Correia, I.J. Stimuli-responsive mesoporous silica nanoparticles for cancer
677 therapy: A review. *Microporous Mesoporous Mater.* **2016**, 236, 141–157.
- 678 51. Gisbert-Garzarán, M.; Manzano, M.; Vallet-Regí, M. pH-Responsive Mesoporous Silica and Carbon
679 Nanoparticles for Drug Delivery. *Bioengineering* **2017**, 4.
- 680 52. Argyo, C.; Weiss, V.; Bräuchle, C.; Bein, T. Multifunctional mesoporous silica nanoparticles as a
681 universal platform for drug delivery. *Chem. Mater.* **2014**, 26, 435–451.
- 682 53. Colilla, M.; González, B.; Vallet-Regí, M. Mesoporous silica nanoparticles for the design of smart delivery
683 nanodevices. *Biomater. Sci.* **2013**, 1, 114–134.
- 684 54. Kato, Y.; Ozawa, S.; Miyamoto, C.; Maehata, Y.; Suzuki, A.; Maeda, T.; Baba, Y. Acidic extracellular
685 microenvironment and cancer. *Cancer Cell Int.* **2013**, 13, 89.
- 686 55. Hu, D.; Li, H.; Wang, B.; Ye, Z.; Lei, W.; Jia, F.; Jin, Q.; Ren, K.-F.; Ji, J. Surface-Adaptive Gold
687 Nanoparticles with Effective Adherence and Enhanced Photothermal Ablation of Methicillin-Resistant
688 *Staphylococcus aureus* Biofilm. *ACS Nano* **2017**, 11, 9330–9339.
- 689 56. Gisbert-Garzarán, M.; Lozano, D.; Vallet-Regí, M.; Manzano, M. Self-Immolative Polymers as novel pH-
690 responsive gate keepers for drug delivery. *RSC Adv.* **2017**, 7, 132–136.
- 691 57. Juárez, L.A.; Añón, E.; Giménez, C.; Sancenón, F.; Martínez-Mañez, R.; Costero, A.M.; Gaviña, P.; Parra,
692 M.; Bernardos, A. Self-Immolative Linkers as Caps for the Design of Gated Silica Mesoporous Supports.
693 *Chem. - A Eur. J.* **2016**, 22, 14126–14130.
- 694 58. Muhammad, F.; Guo, M.; Qi, W.; Sun, F.; Wang, A.; Guo, Y.; Zhu, G. pH-Triggered Controlled Drug
695 Release from Mesoporous Silica Nanoparticles via Intracellular Dissolution of ZnO Nanolids. *J. Am.*
696 *Chem. Soc.* **2011**, 133, 8778–8781.
- 697 59. Chen, X.; Liu, Y.; Lin, A.; Huang, N.; Long, L.; Gang, Y.; Liu, J. Folic acid-modified mesoporous silica
698 nanoparticles with pH-responsiveness loaded with Amp for an enhanced effect against anti-drug-
699 resistant bacteria by overcoming efflux pump systems. *Biomater. Sci.* **2018**, 6, 1923–1935.
- 700 60. Costa, A.G.; Cusano, N.E.; Silva, B.C.; Cremers, S.; Bilezikian, J.P. Cathepsin K: its skeletal actions and
701 role as a therapeutic target in osteoporosis. *Nat. Rev. Rheumatol.* **2011**, 7, 447–456.
- 702 61. Husmann, K.; Muff, R.; Bolander, M.E.; Sarkar, G.; Born, W.; Fuchs, B. Cathepsins and osteosarcoma:
703 Expression analysis identifies cathepsin K as an indicator of metastasis. *Mol. Carcinog.* **2008**, 47, 66–73.
- 704 62. Gondi, C.S.; Rao, J.S. Cathepsin B as a cancer target. *Expert Opin. Ther. Targets* **2013**, 17, 281–91.
- 705 63. Liu, Y.; Ding, X.; Li, J.; Luo, Z.; Hu, Y.; Liu, J.; Dai, L.; Zhou, J.; Hou, C.; Cai, K. Enzyme responsive drug
706 delivery system based on mesoporous silica nanoparticles for tumor therapy in vivo. *Nanotechnology*
707 **2015**, 26, 145102–145116.
- 708 64. Cheng, Y.J.; Luo, G.F.; Zhu, J.Y.; Xu, X.D.; Zeng, X.; Cheng, D.B.; Li, Y.M.; Wu, Y.; Zhang, X.Z.; Zhuo,
709 R.X.; et al. Enzyme-induced and tumor-targeted drug delivery system based on multifunctional
710 mesoporous silica nanoparticles. *ACS Appl. Mater. Interfaces* **2015**, 7, 9078–9087.
- 711 65. Zhao, Q.; Liu, J.; Zhu, W.; Sun, C.; Di, D.; Zhang, Y.; Wang, P.; Wang, Z.; Wang, S. Dual-stimuli
712 responsive hyaluronic acid-conjugated mesoporous silica for targeted delivery to CD44-overexpressing
713 cancer cells. *Acta Biomater.* **2015**, 23, 147–156.
- 714 66. Traverso, N.; Ricciarelli, R.; Nitti, M.; Marengo, B.; Furfaro, A.L.; Pronzato, M.A.; Marinari, U.M.;
715 Domenicotti, C. Role of Glutathione in Cancer Progression and Chemoresistance. *Oxid. Med. Cell. Longev.*

- 716 2013, 972913.
- 717 67. Guo, X.; Cheng, Y.; Zhao, X.; Luo, Y.; Chen, J.; Yuan, W.E. Advances in redox-responsive drug delivery
718 systems of tumor microenvironment. *J. Nanobiotechnology* **2018**, *16*, 74–83.
- 719 68. Li, Z.-Y.; Hu, J.-J.; Xu, Q.; Chen, S.; Jia, H.-Z.; Sun, Y.-X.; Zhuo, R.-X.; Zhang, X.-Z. A redox-responsive
720 drug delivery system based on RGD containing peptide-capped mesoporous silica nanoparticles. *J.*
721 *Mater. Chem. B* **2015**, *3*, 39–44.
- 722 69. Zhang, B.; Luo, Z.; Liu, J.; Ding, X.; Li, J.; Cai, K. Cytochrome c end-capped mesoporous silica
723 nanoparticles as redox-responsive drug delivery vehicles for liver tumor-targeted triplex therapy in vitro
724 and in vivo. *J. Control. Release* **2014**, *192*, 192–201.
- 725 70. Chen, X.; Sun, H.; Hu, J.; Han, X.; Liu, H.; Hu, Y. Transferrin gated mesoporous silica nanoparticles for
726 redox-responsive and targeted drug delivery. *Colloids Surfaces B Biointerfaces* **2017**, *152*, 77–84.
- 727 71. Guisasola, E.; Baeza, A.; Talelli, M.; Arcos, D.; Moros, M.; De La Fuente, J.M.; Vallet-Regí, M. Magnetic-
728 Responsive Release Controlled by Hot Spot Effect. *Langmuir* **2015**, *31*, 12777–12782.
- 729 72. Guisasola, E.; Asín, L.; Beola, L.; De La Fuente, J.M.; Baeza, A.; Vallet-Regí, M. Beyond Traditional
730 Hyperthermia: In Vivo Cancer Treatment with Magnetic-Responsive Mesoporous Silica Nanocarriers.
731 *ACS Appl. Mater. Interfaces* **2018**, *10*, 12518–12525.
- 732 73. Xing, R.; Lin, H.; Jiang, P.; Qu, F. Biofunctional mesoporous silica nanoparticles for magnetically oriented
733 target and pH-responsive controlled release of ibuprofen. *Colloids Surfaces A Physicochem. Eng. Asp.* **2012**,
734 403, 7–14.
- 735 74. Martínez-Carmona, M.; Lozano, D.; Baeza, A.; Colilla, M.; Vallet-Regí, M. A novel visible light
736 responsive nanosystem for cancer treatment. *Nanoscale* **2017**, *9*, 15967–15973.
- 737 75. Villaverde, G.; Gómez-Graña, S.; Guisasola, E.; García, I.; Hanske, C.; Liz-Marzán, L.M.; Baeza, A.; Vallet-
738 Regí, M. Targeted Chemo-Photothermal Therapy: A Nanomedicine Approximation to Selective
739 Melanoma Treatment. *Part. Part. Syst. Charact.* **2018**, *35*, 1800148–1800158.
- 740 76. Wang, D.; Wu, S. Red-Light-Responsive Supramolecular Valves for Photocontrolled Drug Release from
741 Mesoporous Nanoparticles. *Langmuir* **2016**, *32*, 632–636.
- 742 77. Paris, J.L.; Cabanas, M.V.; Manzano, M.; Vallet-Regí, M. Polymer-Grafted Mesoporous Silica
743 Nanoparticles as Ultrasound-Responsive Drug Carriers. *ACS Nano* **2015**, *9*, 11023–11033.
- 744 78. Paris, J.L.; Villaverde, G.; Cabañas, M.V.; Manzano, M.; Vallet-Regí, M. From proof-of-concept material
745 to PEGylated and modularly targeted ultrasound-responsive mesoporous silica nanoparticles. *J. Mater.*
746 *Chem. B* **2018**, *6*, 2785–2794.
- 747 79. Lee, S.F.; Zhu, X.M.; Wang, Y.X.J.; Xuan, S.H.; You, Q.; Chan, W.H.; Wong, C.H.; Wang, F.; Yu, J.C.;
748 Cheng, C.H.K.; et al. Ultrasound, pH and magnetically responsive crown-ether-coated core/shell
749 nanoparticles as drug encapsulation and release systems. *ACS Appl. Mater. Interfaces* **2013**, *5*, 1566–1574.
- 750 80. Croissant, J.G.; Fatieiev, Y.; Almalik, A.; Khashab, N.M. Mesoporous Silica and Organosilica
751 Nanoparticles: Physical Chemistry, Biosafety, Delivery Strategies, and Biomedical Applications. *Adv.*
752 *Healthc. Mater.* **2018**, *7*, 1–75.
- 753 81. Jokerst, J. V.; Lobovkina, T.; Zare, R.N.; Gambhir, S.S. Nanoparticle PEGylation for imaging and therapy.
754 *Nanomedicine* **2011**, *6*, 715–728.
- 755 82. Amoozgar, Z.; Yeo, Y. Recent advances in stealth coating of nanoparticle drug delivery systems. *Wiley*
756 *Interdiscip. Rev. Nanomedicine Nanobiotechnology* **2012**, *4*, 219–233.
- 757 83. Clemments, A.M.; Muniesa, C.; Landry, C.C.; Botella, P. Effect of surface properties in protein corona

- 758 development on mesoporous silica nanoparticles. *RSC Adv.* **2014**, *4*, 29134–29138.
- 759 84. Kramer, L.; Winter, G.; Baur, B.; Kuntz, A.J.; Kull, T.; Solbach, C.; Beer, A.J.; Lindén, M. Quantitative and
760 correlative biodistribution analysis of ⁸⁹Zr-labeled mesoporous silica nanoparticles intravenously
761 injected into tumor-bearing mice. *Nanoscale* **2017**, *9*, 9743–9753.
- 762 85. He, Q.; Zhang, Z.; Gao, F.; Li, Y.; Shi, J. In vivo Biodistribution and Urinary Excretion of Mesoporous
763 Silica Nanoparticles: Effects of Particle Size and PEGylation. *Small* **2011**, *7*, 271–280.
- 764 86. Manzano, M.; Vallet-Regí, M. Mesoporous silica nanoparticles for drug delivery. *Adv. Funct. Mater.* **2019**,
765 1902634.
- 766 87. Gustafson, H.H.; Holt-Casper, D.; Grainger, D.W.; Ghandehari, H. Nanoparticle Uptake: The Phagocyte
767 Problem. *Nano Today* **2015**, *10*, 487–510.
- 768 88. Arvizo, R.R.; Miranda, O.R.; Moyano, D.F.; Walden, C.A.; Giri, K.; Robertson, J.D.; Rotello, V.M.; Reid,
769 J.M.; Mukherjee, P. Modulating Pharmacokinetics, Tumor Uptake and Biodistribution by Engineered
770 Nanoparticles. *PLoS One* **2011**, *6*, 24374.
- 771 89. Paris, J.L.; Colilla, M.; Izquierdo-barba, I.; Manzano, M.; Vallet-Regí, M. Tuning mesoporous silica
772 dissolution in physiological environments: a review. *J. Mater. Sci.* **2017**, *52*, 8761–8771.
- 773 90. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2019. *CA. Cancer J. Clin.* **2019**, *69*, 7–34.
- 774 91. Lewis, V.O. What's new in musculoskeletal oncology. *J. Bone Jt. Surg. - Ser. A* **2009**, *91*, 1546–1556.
- 775 92. Cortini, M.; Baldini, N.; Avnet, S. New advances in the study of bone tumors: A lesson from the 3D
776 environment. *Front. Physiol.* **2019**, *10*.
- 777 93. Fornetti, J.; Welm, A.L.; Stewart, S.A. Understanding the Bone in Cancer Metastasis. *J. Bone Miner. Res.*
778 **2018**, *33*, 2099–2113.
- 779 94. Weilbaecher, K.N.; Guise, T.A.; Mccauley, L.K. Cancer to bone : a fatal attraction. *Nat. Publ. Gr.* **2011**, *11*,
780 411–425.
- 781 95. Liu, Y.; Cao, X. Organotropic metastasis: Role of tumor exosomes. *Cell Res.* **2016**, *26*, 149–150.
- 782 96. Hoshino, A.; Costa-Silva, B.; Shen, T.L.; Rodrigues, G.; Hashimoto, A.; Tesic Mark, M.; Molina, H.;
783 Kohsaka, S.; Di Giannatale, A.; Ceder, S.; et al. Tumour exosome integrins determine organotropic
784 metastasis. *Nature* **2015**, *527*, 329–335.
- 785 97. Coleman, R.E. Metastatic bone disease: clinical features, pathophysiology and treatment strategies.
786 *Cancer Treat. Rev.* **2001**, *27*, 165–176.
- 787 98. Mundy, G.R. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat. Rev. Cancer*
788 **2002**, *2*, 584–593.
- 789 99. Oppenheimer, S.B. Cellular basis of cancer metastasis: A review of fundamentals and new advances.
790 *Acta Histochem.* **2006**, *108*, 327–334.
- 791 100. Aguirre-Ghiso, J.A. Models, mechanisms and clinical evidence for cancer dormancy. *Nat. Rev. Cancer*
792 **2007**, *7*, 834–846.
- 793 101. Jahanban-Esfahlan, R.; Seidi, K.; Manjili, M.H.; Jahanban-Esfahlan, A.; Javaheri, T.; Zare, P. Tumor Cell
794 Dormancy: Threat or Opportunity in the Fight against Cancer. *Cancers (Basel)*. **2019**, *11*.
- 795 102. Recasens, A.; Munoz, L. Targeting Cancer Cell Dormancy. *Trends Pharmacol. Sci.* **2019**, *40*, 128–141.
- 796 103. Wyld, L.; Audisio, R.A.; Poston, G.J. The evolution of cancer surgery and future perspectives. *Nat. Rev.*

- 797 *Clin. Oncol.* **2015**, *12*, 115–124.
- 798 104. Baskar, R.; Lee, K.A.; Yeo, R.; Yeoh, K.-W. Cancer and Radiation Therapy: Current Advances and Future
799 Directions. *Int. J. Med. Sci.* **2012**, *9*, 193–199.
- 800 105. Chabner, B.A.; Roberts, T.G. Chemotherapy and the war on cancer. *Nat. Rev. Cancer* **2005**, *5*, 65–72.
- 801 106. Cho, K.; Wang, X.; Nie, S.; Chen, Z.; Shin, D.M. Therapeutic nanoparticles for drug delivery in cancer.
802 *Clin. Cancer Res.* **2008**, *14*, 1310–1316.
- 803 107. Baeza, A.; Manzano, M.; Colilla, M.; Vallet-Regí, M. Recent advances in mesoporous silica nanoparticles
804 for antitumor therapy: Our contribution. *Biomater. Sci.* **2016**, *4*, 803–813.
- 805 108. Mura, S.; Nicolas, J.; Couvreur, P. Stimuli-responsive nanocarriers for drug delivery. *Nat. Mater.* **2013**,
806 *12*, 991–1003.
- 807 109. Grodzinski, P.; Kircher, M.; Goldberg, M.; Gabizon, A. Integrating Nanotechnology into Cancer Care.
808 *ACS Nano* **2019**, *13*, 7370–7376.
- 809 110. Matsumura, Y.; Maeda, H. A New Concept for Macromolecular Therapeutics in Cancer Chemotherapy:
810 Mechanism of Tumorotropic Accumulation of Proteins and the Antitumor Agent Smancs. *Cancer Res.*
811 **1986**, *46*, 6387–6392.
- 812 111. Martínez-Carmona, M.; Colilla, M.; Vallet-Regí, M. Smart Mesoporous Nanomaterials for Antitumor
813 Therapy. *Nanomaterials* **2015**, *5*, 1906–1937.
- 814 112. Fang, J.; Nakamura, H.; Maeda, H. The EPR effect: Unique features of tumor blood vessels for drug
815 delivery, factors involved, and limitations and augmentation of the effect. *Adv. Drug Deliv. Rev.* **2011**, *63*,
816 136–151.
- 817 113. Maeda, H.; Nakamura, H.; Fang, J. The EPR effect for macromolecular drug delivery to solid tumors:
818 Improvement of tumor uptake, lowering of systemic toxicity, and distinct tumor imaging in vivo. *Adv.*
819 *Drug Deliv. Rev.* **2013**, *65*, 71–79.
- 820 114. Natfji, A.A.; Ravishankar, D.; Osborn, H.M.I.; Greco, F. Parameters affecting the Enhanced Permeability
821 and Retention Effect: The need for patient selection. *J. Pharm. Sci.* **2017**, *106*, 3179–3187.
- 822 115. Liu, X.; Jiang, J.; Ji, Y.; Lu, J.; Chan, R.; Meng, H. Targeted drug delivery using iRGD peptide for solid
823 cancer treatment. *Mol. Syst. Des. Eng.* **2017**, *2*, 370–379.
- 824 116. Ruoslahti, E. Tumor penetrating peptides for improved drug delivery. *Adv. Drug Deliv. Rev.* **2017**, *110*–
825 *111*, 3–12.
- 826 117. Das, S.; Raj, R. Prospects of Bacteriotherapy with Nanotechnology in Nanoparticledrug Conjugation
827 Approach for Cancer Therapy. *Curr. Med. Chem.* **2016**, *23*, 1477–1494.
- 828 118. Suh, S.B.; Jo, A.; Traore, M.A.; Zhan, Y.; Coutermarsh-Ott, S.L.; Ringel-Scaia, V.M.; Allen, I.C.; Davis,
829 R.M.; Behkam, B. Nanoscale Bacteria-Enabled Autonomous Drug Delivery System (NanoBEADS)
830 Enhances Intratumoral Transport of Nanomedicine. *Adv. Sci.* **2019**, *6*, 1801309.
- 831 119. Paris, J.L.; de la Torre, P.; Cabañas, M.V.; Manzano, M.; Grau, M.; Flores, A.I.; Vallet-Regí, M.
832 Vectorization of ultrasound-responsive nanoparticles in placental mesenchymal stem cells for cancer
833 therapy. *Nanoscale* **2017**, *9*, 5528–5537.
- 834 120. Paris, J.L.; Torre, P. de la; Manzano, M.; Cabañas, M.V.; Flores, A.I.; Vallet-Regí, M. Decidua-derived
835 mesenchymal stem cells as carriers of mesoporous silica nanoparticles. In vitro and in vivo evaluation
836 on mammary tumors. *Acta Biomater.* **2016**, *33*, 275–282.
- 837 121. Wang, X.; Gao, J.; Ouyang, X.; Wang, J.; Sun, X.; Lv, Y. Mesenchymal stem cells loaded with paclitaxel–

- 838 poly(Lactic-co-glycolic acid) nanoparticles for glioma-targeting therapy. *Int. J. Nanomedicine* **2018**, *13*,
839 5231–5248.
- 840 122. Layek, B.; Sadhukha, T.; Panyam, J.; Prabha, S. Nano-Engineered Mesenchymal Stem Cells Increase
841 Therapeutic Efficacy of Anticancer Drug Through True Active Tumor Targeting. *Mol. Cancer Ther.* **2018**,
842 *17*, 1196–1206.
- 843 123. Lurje, G.; Lenz, H.-J. EGFR Signaling and Drug Discovery. *Oncology* **2009**, *77*, 400–410.
- 844 124. Villegas, M.R.; Baeza, A.; Noureddine, A.; Durfee, P.N.; Butler, K.S.; Agola, J.O.; Brinker, C.J.; Vallet-
845 Regí, M. Multifunctional Protocells for Enhanced Penetration in 3D Extracellular Tumoral Matrices.
846 *Chem. Mater.* **2018**, *30*, 112–120.
- 847 125. Martínez-Carmona, M.; Baeza, A.; Rodríguez-Milla, M.A.; García-Castro, J.; Vallet-Regí, M. Mesoporous
848 silica nanoparticles grafted with a light-responsive protein shell for highly cytotoxic antitumoral
849 therapy. *J. Mater. Chem. B* **2015**, *3*, 5746–5752.
- 850 126. Liong, M.; Zink, J.I.; Lu, J.; Tamanoi, F.; Kovochich, M.; Xia, T.; Nel, A.E.; Ruehm, S.G. Multifunctional
851 inorganic nanoparticles for imaging, targeting, and drug delivery. *ACS Nano* **2008**, *2*, 889–896.
- 852 127. Porta, F.; Lamers, G.E.M.; Morrhayim, J.; Chatzopoulou, A.; Schaaf, M.; den Dulk, H.; Backendorf, C.;
853 Zink, J.I.; Kros, A. Folic Acid-Modified Mesoporous Silica Nanoparticles for Cellular and Nuclear
854 Targeted Drug Delivery. *Adv. Healthc. Mater.* **2013**, *2*, 281–286.
- 855 128. Lv, G.; Qiu, L.; Liu, G.; Wang, W.; Li, K.; Zhao, X.; Lin, J. pH sensitive chitosan-mesoporous silica
856 nanoparticles for targeted delivery of a ruthenium complex with enhanced anticancer effects. *Dalt. Trans.*
857 **2016**, *45*, 18147–18155.
- 858 129. Ren, W.X.; Han, J.; Uhm, S.; Jang, Y.J.; Kang, C.; Kim, J.H.; Kim, J.S. Recent development of biotin
859 conjugation in biological imaging, sensing, and target delivery. *Chem. Commun.* **2015**, *51*, 10403–10418.
- 860 130. Russell-Jones, G.; Mctavish, K.; Mcewan, J.; Rice, J.; Nowotnik, D. Vitamin-mediated targeting as a
861 potential mechanism to increase drug uptake by tumours. *J. Inorg. Biochem.* **2004**, *98*, 1625–1633.
- 862 131. Wu, X.; Han, Z.; Schur, R.M.; Lu, Z. Targeted Mesoporous Silica Nanoparticles Delivering Arsenic
863 Trioxide with Environment Sensitive Drug Release for E ffective Treatment of Triple Negative Breast
864 Cancer. *ACS Biomater. Sci.* **2016**, *2*, 501–507.
- 865 132. Xu, H.; Wang, Z.; Li, Y.; Guo, Y.; Zhou, H.; Li, Y.; Wu, F.; Zhang, L.; Yang, X.; Lu, B.; et al. Preparation
866 and characterization of a dual-receptor mesoporous silica nanoparticle–hyaluronic acid–RGD peptide
867 targeting drug delivery system. *RSC Adv.* **2016**, *6*, 40427–40435.
- 868 133. Paris, J.L.; Villaverde, G.; Cabañas, M.V.; Manzano, M.; Vallet-Regí, M. From proof-of-concept material
869 to PEGylated and modularly targeted ultrasound-responsive mesoporous silica nanoparticles. *J. Mater.*
870 *Chem. B* **2018**, *6*, 2785–2794.
- 871 134. Schroeder, A.; Heller, D.A.; Winslow, M.M.; Dahlman, J.E.; Pratt, G.W.; Langer, R.; Jacks, T.; Anderson,
872 D.G. Treating metastatic cancer with nanotechnology. *Nat. Rev. Cancer* **2012**, *12*, 39–50.
- 873 135. Henneman, Z.J.; Nancollas, G.H.; Ebetino, F.H.; Russell, R.G.G.; Phipps, R.J. Bisphosphonate binding
874 affinity as assessed by inhibition of carbonated apatite dissolution in vitro. *J. Biomed. Mater. Res. A* **2008**,
875 *85*, 993–1000.
- 876 136. Sun, W.; Han, Y.; Li, Z.; Ge, K.; Zhang, J. Bone-Targeted Mesoporous Silica Nanocarrier Anchored by
877 Zoledronate for Cancer Bone Metastasis. *Langmuir* **2016**, *32*, 9237–9244.
- 878 137. Sun, W.; Ge, K.; Jin, Y.; Han, Y.; Zhang, H.; Zhou, G.; Yang, X.; Liu, D.; Liu, H.; Liang, X.J.; et al. Bone-
879 Targeted Nanoplatform Combining Zoledronate and Photothermal Therapy to Treat Breast Cancer Bone
880 Metastasis. *ACS Nano* **2019**, *13*, 7556–7567.

- 881 138. Villaverde, G.; Nairi, V.; Baeza, A.; Vallet-Regí, M. Double Sequential Encrypted Targeting Sequence: A
882 New Concept for Bone Cancer Treatment. *Chem. - A Eur. J.* **2017**, *23*, 7174–7179.
- 883 139. Lu, Y.; Li, L.; Lin, Z.; Li, M.; Hu, X.; Zhang, Y.; Peng, M. Enhancing Osteosarcoma Killing and CT Imaging
884 Using Ultrahigh Drug Loading and NIR-Responsive Bismuth Sulfide@Mesoporous Silica Nanoparticles.
885 *Adv. Healthc. Mater.* **2018**, *7*, 1800602.
- 886 140. Paris, J.L.; Villaverde, G.; Gómez-Graña, S.; Vallet-Regí, M. Nanoparticles for Multimodal Antivascular
887 Therapeutics: Dual Drug Release, Photothermal and Photodynamic Therapy. *Acta Biomater.* **2020**, *101*,
888 459–468.
- 889 141. Lin, H.M.; Lin, H.Y.; Chan, M.H. Preparation, characterization, and in vitro evaluation of folate-modified
890 mesoporous bioactive glass for targeted anticancer drug carriers. *J. Mater. Chem. B* **2013**, *1*, 6147–6156.
- 891 142. Porta, F.; Lamers, G.E.M.; Morrhayim, J.; Chatzopoulou, A.; Schaaf, M.; den Dulk, H.; Backendorf, C.;
892 Zink, J.I.; Kros, A. Folic Acid-Modified Mesoporous Silica Nanoparticles for Cellular and Nuclear
893 Targeted Drug Delivery. *Adv. Healthc. Mater.* **2013**, *2*, 281–286.
- 894 143. Mehravi, B.; Ahmadi, M.; Amanlou, M.; Mostaar, A.; Ardestani, M.S.; Ghalandarlaki, N. Conjugation of
895 glucosamine with Gd 3+ -based nanoporous silica using a heterobifunctional ANB-NOS crosslinker for
896 imaging of cancer cells. *Int. J. Nanomedicine* **2013**, *8*, 3383–3394.
- 897 144. Shahabi, S.; Döscher, S.; Bollhorst, T.; Treccani, L.; Maas, M.; Dringen, R.; Rezwani, K. Enhancing Cellular
898 Uptake and Doxorubicin Delivery of Mesoporous Silica Nanoparticles via Surface Functionalization:
899 Effects of Serum. *ACS Appl. Mater. Interfaces* **2015**, *7*, 26880–26891.
- 900 145. Martínez-Carmona, M.; Lozano, D.; Colilla, M.; Vallet-Regí, M. Lectin-conjugated pH-responsive
901 mesoporous silica nanoparticles for targeted bone cancer treatment. *Acta Biomater.* **2018**, *65*, 393–404.
- 902 146. Martínez-Carmona, M.; Baeza, A.; Rodríguez-Milla, M.A.; García-Castro, J.; Vallet-Regí, M. Mesoporous
903 Silica Nanoparticles Grafted with Light-Responsive Protein Shell for Highly Cytotoxic Antitumoral
904 Therapy. *RSC Adv.* **2015**, *3*, 5746–5752.
- 905 147. Paris, J.L.; Manzano, M.; Cabañas, M.V.; Vallet-Regí, M. Mesoporous silica nanoparticles engineered for
906 ultrasound-induced uptake by cancer cells. *Nanoscale* **2018**, *10*, 6402–6408.
- 907 148. Zhang, Y.; Hu, M.; Wang, X.; Zhou, Z.; Liu, Y. Design and evaluation of europium containing
908 mesoporous bioactive glass nanospheres: Doxorubicin release kinetics and inhibitory effect on
909 osteosarcoma MG 63 cells. *Nanomaterials* **2018**, *8*.
- 910 149. Ravanbakhsh, M.; Labbaf, S.; Karimzadeh, F.; Pinna, A.; Houreh, A.B.; Nasr-Esfahani, M.H. Mesoporous
911 bioactive glasses for the combined application of osteosarcoma treatment and bone regeneration. *Mater.*
912 *Sci. Eng. C* **2019**, *104*, 109994.
- 913 150. Boanini, E.; Panseri, S.; Arroyo, F.; Montesi, M.; Rubini, K.; Tampieri, A.; Covarrubias, C.; Bigi, A.
914 Alendronate functionalized mesoporous bioactive glass nanospheres. *Materials (Basel)*. **2016**, *9*.
- 915 151. Wu, C.; Fan, W.; Chang, J. Functional mesoporous bioactive glass nanospheres: Synthesis, high loading
916 efficiency, controllable delivery of doxorubicin and inhibitory effect on bone cancer cells. *J. Mater. Chem.*
917 *B* **2013**, *1*, 2710–2718.
- 918 152. Bouchoucha, M.; Côté, M.F.; C-Gaudreault, R.; Fortin, M.A.; Kleitz, F. Size-Controlled Functionalized
919 Mesoporous Silica Nanoparticles for Tunable Drug Release and Enhanced Anti-Tumoral Activity. *Chem.*
920 *Mater.* **2016**, *28*, 4243–4258.
- 921 153. Shao, Y.; Tian, X.; Hu, W.; Zhang, Y.; Liu, H.; He, H. Biomaterials The properties of Gd₂O₃-assembled
922 silica nanocomposite targeted nanoprobe and their application in MRI. *Biomaterials* **2012**, *33*, 6438–6446.
- 923 154. Meng, X.; Zhang, H.; Zhang, M.; Wang, B.; Liu, Y.; Wang, Y. Negative CT Contrast Agents for the

- 924 Diagnosis of Malignant Osteosarcoma. *Adv. Sci.* **2019**, *6*, 1901214.
- 925 155. Lee, S.F.; Zhu, X.M.; Wang, Y.X.J.; Xuan, S.H.; You, Q.; Chan, W.H.; Wong, C.H.; Wang, F.; Yu, J.C.;
926 Cheng, C.H.K.; et al. Ultrasound, pH, and magnetically responsive crown-ether-coated core/shell
927 nanoparticles as drug encapsulation and release systems. *ACS Appl. Mater. Interfaces* **2013**, *5*, 1566–1574.
- 928 156. Kong, M.; Tang, J.; Qiao, Q.; Wu, T.; Qi, Y.; Tan, S.; Gao, X.; Zhang, Z. Biodegradable hollow mesoporous
929 silica nanoparticles for regulating tumor microenvironment and enhancing antitumor efficiency.
930 *Theranostics* **2017**, *7*, 3276–3292.
- 931 157. Polo, L.; Gómez-Cerezo, N.; Aznar, E.; Vivancos, J.L.; Sancenón, F.; Arcos, D.; Vallet-Regí, M.; Martínez-
932 Máñez, R. Molecular gates in mesoporous bioactive glasses for the treatment of bone tumors and
933 infection. *Acta Biomater.* **2017**, *50*, 114–126.
- 934 158. Martínez-Carmona, M.; Lozano, D.; Baeza, A.; Colilla, M.; Vallet-Regí, M. A novel visible light
935 responsive nanosystem for cancer treatment. *Nanoscale* **2017**, *9*, 15967–15973.
- 936 159. Wu, X.; Wu, S.; Yang, L.; Han, J.; Han, S. Cytosolic delivery of proteins mediated by aldehyde-displaying
937 silica nanoparticles with pH-responsive characteristics. *J. Mater. Chem.* **2012**, *22*, 17121–17127.
- 938 160. Martínez, Á.; Fuentes-Paniagua, E.; Baeza, A.; Sánchez-Nieves, J.; Cicuéndez, M.; Gómez, R.; De La Mata,
939 F.J.; González, B.; Vallet-Regí, M. Mesoporous Silica Nanoparticles Decorated with Carboxilane
940 Dendrons as New Non-viral Oligonucleotide Delivery Carriers. *Chem. - A Eur. J.* **2015**, *21*, 15651–15666.
- 941 161. Vaishnav, A.K.; Gollob, J.; Gamba-Vitalo, C.; Hutabarat, R.; Sah, D.; Meyers, R.; de Fougerolles, T.;
942 Maraganore, J. A status report on RNAi therapeutics. *Silence* **2010**, *1*, 14.
- 943 162. Lee, S.H.; Kang, Y.Y.; Jang, H.-E.; Mok, H. Current preclinical small interfering RNA (siRNA)-based
944 conjugate systems for RNA therapeutics. *Adv. Drug Deliv. Rev.* **2016**, *104*, 78–92.
- 945 163. de Cárcer, G. The Mitotic Cancer Target Polo-Like Kinase 1: Oncogene or Tumor Suppressor? *Genes*
946 *(Basel)*. **2019**, *10*, 208.
- 947 164. Xiong, L.; Bi, J.; Tang, Y.; Qiao, S.Z. Magnetic Core-Shell Silica Nanoparticles with Large Radial
948 Mesopores for siRNA Delivery. *Small* **2016**, *12*, 4735–4742.
- 949 165. Hartono, S.B.; Gu, W.; Kleitz, F.; Liu, J.; He, L.; Middelberg, A.P.J.; Yu, C.; Lu, G.Q.; Qiao, S.Z. Poly-L-
950 lysine functionalized large pore cubic mesostructured silica nanoparticles as biocompatible carriers for
951 gene delivery. *ACS Nano* **2012**, *6*, 2104–2117.
- 952 166. Du, X.; Shi, B.; Liang, J.; Bi, J.; Dai, S.; Qiao, S.Z. Developing functionalized dendrimer-like silica
953 nanoparticles with hierarchical pores as advanced delivery nanocarriers. *Adv. Mater.* **2013**, *25*, 5981–5985.
- 954 167. Hartono, S.B.; Yu, M.; Gu, W.; Yang, J.; Strounina, E.; Wang, X.; Qiao, S.; Yu, C. Synthesis of multi-
955 functional large pore mesoporous silica nanoparticles as gene carriers. *Nanotechnology* **2014**, *25*.
- 956 168. Martínez-Carmona, M.; Lozano, D.; Baeza, A.; Colilla, M.; Vallet-Regí, M. A novel visible light
957 responsive nanosystem for cancer treatment. *Nanoscale* **2017**, *9*, 15967–15973.
- 958 169. Martínez-Carmona, M.; Lozano, D.; Colilla, M.; Vallet-Regí, M. Lectin-Conjugated pH-Responsive
959 Mesoporous Silica Nanoparticles for Targeted Bone Cancer Treatment. *Acta Biomater.* **2018**, *65*, 393–404.
- 960 170. Taubes, G. The Bacteria Fight Back. *Science (80-.)*. **2008**, *321*, 356 LP – 361.
- 961 171. de Kraker, M.E.A.; Stewardson, A.J.; Harbarth, S. Will 10 Million People Die a Year due to Antimicrobial
962 Resistance by 2050? *PLOS Med.* **2016**, *13*, e1002184.
- 963 172. Davies, D. Understanding biofilm resistance to antibacterial agents. *Nat. Rev. Drug Discov.* **2003**, *2*, 114–
964 122.

- 965 173. Vallet-Regí, M.; González, B.; Izquierdo-Barba, I. Nanomaterials as promising alternative in the infection
966 treatment. *Int. J. Mol. Sci.* **2019**, *20*.
- 967 174. Spear, M. The Biofilm Challenge: Breaking Down the Walls. *Plast. Surg. Nurs.* **2011**, *31*, 117–120.
- 968 175. Webb, R.; Cutting, K. Biofilm the challenge. *J. Wound Care* **2014**, *23*, 519.
- 969 176. Alibert, S.; N'gompaza Diarra, J.; Hernandez, J.; Stutzmann, A.; Fouad, M.; Boyer, G.; Pagès, J.-M.
970 Multidrug efflux pumps and their role in antibiotic and antiseptic resistance: a pharmacodynamic
971 perspective. *Expert Opin. Drug Metab. Toxicol.* **2017**, *13*, 301–309.
- 972 177. Nicoloff, H.; Andersson, D.I. Indirect resistance to several classes of antibiotics in cocultures with
973 resistant bacteria expressing antibiotic-modifying or -degrading enzymes. *J. Antimicrob. Chemother.* **2015**,
974 *71*, 100–110.
- 975 178. Mooney, J.A.; Pridgen, E.M.; Manasherob, R.; Suh, G.; Blackwell, H.E.; Barron, A.E.; Bollyky, P.L.;
976 Goodman, S.B.; Amanatullah, D.F. Periprosthetic bacterial biofilm and quorum sensing. *J. Orthop. Res.*
977 **2018**, *36*, 2331–2339.
- 978 179. Stewart, P.S.; William Costerton, J. Antibiotic resistance of bacteria in biofilms. *Lancet* **2001**, *358*, 135–138.
- 979 180. Chen, S.; Li, L.; Zhao, C.; Zheng, J. Surface hydration : Principles and applications toward low-fouling /
980 nonfouling biomaterials. *Polymer (Guildf)*. **2010**, *51*, 5283–5293.
- 981 181. García, K.P.; Zarschler, K.; Barbaro, L.; Barreto, J.A.; O'Malley, W.; Spiccia, L.; Stephan, H.; Graham, B.
982 Zwitterionic-coated “stealth” nanoparticles for biomedical applications: Recent advances in countering
983 biomolecular corona formation and uptake by the mononuclear phagocyte system. *Small* **2014**, *10*, 2516–
984 2529.
- 985 182. Zhang, P.; Sun, F.; Liu, S.; Jiang, S. Anti-PEG antibodies in the clinic: Current issues and beyond
986 PEGylation. *J. Control. Release* **2016**, *244*, 184–193.
- 987 183. Colilla, M.; Izquierdo-Barba, I.; Sánchez-Salcedo, S.; Fierro, J.L.G.; Hueso, J.L.; Vallet-Regí, M. Synthesis
988 and characterization of zwitterionic SBA-15 nanostructured materials. *Chem. Mater.* **2010**, *22*, 6459–6466.
- 989 184. Encinas, N.; Angulo, M.; Astorga, C.; Colilla, M.; Izquierdo-Barba, I.; Vallet-Regí, M. Mixed-charge
990 pseudo-zwitterionic mesoporous silica nanoparticles with low-fouling and reduced cell uptake
991 properties. *Acta Biomater.* **2019**, *84*, 317–327.
- 992 185. Khatoon, S.; Han, H.S.; Lee, M.; Lee, H.; Jung, D.W.; Thambi, T.; Ikram, M.; Kang, Y.M.; Yi, G.R.; Park,
993 J.H. Zwitterionic mesoporous nanoparticles with a bioresponsive gatekeeper for cancer therapy. *Acta*
994 *Biomater.* **2016**, *40*, 282–292.
- 995 186. Nechikkattu, R.; Park, S.S.; Ha, C.S. Zwitterionic functionalised mesoporous silica nanoparticles for
996 alendronate release. *Microporous Mesoporous Mater.* **2019**, *279*, 117–127.
- 997 187. Estephan, Z.G.; Jaber, J.A.; Schlenoff, J.B. Zwitterion-stabilized silica nanoparticles: Toward nonstick
998 nano. *Langmuir* **2010**, *26*, 16884–16889.
- 999 188. Zhu, Y.; Sundaram, H.S.; Liu, S.; Zhang, L.; Xu, X.; Yu, Q.; Xu, J.; Jiang, S. A robust graft-to strategy to
1000 form multifunctional and stealth zwitterionic polymer-coated mesoporous silica nanoparticles.
1001 *Biomacromolecules* **2014**, *15*, 1845–1851.
- 1002 189. Villegas, M.F.; Garcia-Uriostegui, L.; Rodríguez, O.; Izquierdo-Barba, I.; Salinas, A.J.; Toriz, G.; Vallet-
1003 Regí, M.; Delgado, E. Lysine-grafted MCM-41 silica as an antibacterial biomaterial. *Bioengineering* **2017**,
1004 *4*.
- 1005 190. Sánchez-Salcedo, S.; García, A.; Vallet-Regí, M. Prevention of bacterial adhesion to zwitterionic
1006 biocompatible mesoporous glasses. *Acta Biomater.* **2017**, *57*, 472–486.

- 1007 191. Rosen, J.E.; Gu, F.X. Surface functionalization of silica nanoparticles with cysteine: A low-fouling
1008 zwitterionic surface. *Langmuir* **2011**, *27*, 10507–10513.
- 1009 192. Colilla, M.; Martínez-Carmona, M.; Sánchez-Salcedo, S.; Ruiz-González, M.L.; González-Calbet, J.M.;
1010 Vallet-Regí, M. A novel zwitterionic bioceramic with dual antibacterial capability. *J. Mater. Chem. B* **2014**,
1011 *2*, 5639–5651.
- 1012 193. Izquierdo-Barba, I.; Sánchez-Salcedo, S.; Colilla, M.; Feito, M.J.; Ramírez-Santillán, C.; Portolés, M.T.;
1013 Vallet-Regí, M. Inhibition of bacterial adhesion on biocompatible zwitterionic SBA-15 mesoporous
1014 materials. *Acta Biomater.* **2011**, *7*, 2977–2985.
- 1015 194. Polo, L.; Gómez-Cerezo, N.; García-Fernández, A.; Aznar, E.; Vivancos, J.L.; Arcos, D.; Vallet-Regí, M.;
1016 Martínez-Mañez, R. Mesoporous Bioactive Glasses Equipped with Stimuli-Responsive Molecular Gates
1017 for Controlled Delivery of Levofloxacin against Bacteria. *Chem. - A Eur. J.* **2018**, *24*, 18944–18951.
- 1018 195. Pedraza, D.; Díez, J.; Barba, I.I.; Colilla, M.; Vallet-Regí, M. Amine-functionalized mesoporous silica
1019 nanoparticles: A new nanoantibiotic for bone infection treatment. *Biomed. Glas.* **2018**, *4*.
- 1020 196. González, B.; Colilla, M.; Díez, J.; Pedraza, D.; Guembe, M.; Izquierdo-Barba, I.; Vallet-Regí, M.
1021 Mesoporous silica nanoparticles decorated with polycationic dendrimers for infection treatment. *Acta*
1022 *Biomater.* **2018**, *68*, 261–271.
- 1023 197. Martínez-Carmona, M.; Izquierdo-Barba, I.; Colilla, M.; Vallet-Regí, M. Concanavalin A-targeted
1024 mesoporous silica nanoparticles for infection treatment. *Acta Biomater.* **2019**, *96*, 547–556.
- 1025 198. Vallet-Regí, M.; Ruiz-Hernández, E. Bioceramics: From bone regeneration to cancer nanomedicine. *Adv.*
1026 *Mater.* **2011**, *23*, 5177–5218.
- 1027 199. Cheng, T.; Qu, H.; Zhang, G.; Zhang, X. Osteogenic and antibacterial properties of vancomycin-laden
1028 mesoporous bioglass/PLGA composite scaffolds for bone regeneration in infected bone defects. *Artif.*
1029 *Cells, Nanomedicine Biotechnol.* **2018**, *46*, 1935–1947.
- 1030 200. Zhou, X.; Weng, W.; Chen, B.; Feng, W.; Wang, W.; Nie, W.; Chen, L.; Mo, X.; Su, J.; He, C. Mesoporous
1031 silica nanoparticles/gelatin porous composite scaffolds with localized and sustained release of
1032 vancomycin for treatment of infected bone defects. *J. Mater. Chem. B* **2018**, *6*, 740–752.
- 1033 201. Paris, J.L.; Lafuente-Gómez, N.; Cabañas, M.V.; Román, J.; Peña, J.; Vallet-Regí, M. Fabrication of a
1034 nanoparticle-containing 3D porous bone scaffold with proangiogenic and antibacterial properties. *Acta*
1035 *Biomater.* **2019**, *86*, 441–449.
- 1036 202. Polo, L.; Gómez-Cerezo, N.; Aznar, E.; Vivancos, J.-L.; Sancenón, F.; Arcos, D.; Vallet-Regí, M.; Martínez-
1037 mañez, R. Molecular gates in mesoporous bioactive glasses for the treatment of bone tumors and
1038 infection. *Acta Biomater.* **2017**, *50*, 114–126.
- 1039 203. Polo, L.; Gómez-Cerezo, N.; García-Fernández, A.; Aznar, E.; Vivancos, J.L.; Arcos, D.; Vallet-Regí, M.;
1040 Martínez-Mañez, R. Mesoporous bioactive glasses equipped with stimuli-responsive molecular gates for
1041 the controlled delivery of levofloxacin against bacteria. *Chem. - A Eur. J.* **2018**, *24*, 18944–18951.
- 1042 204. González, B.; Díez, J.; Pedraza, D.; Guembe, M.; Izquierdo-Barba, I.; Vallet-Regí, M. Mesoporous silica
1043 nanoparticles decorated with polycationic dendrimers for infection treatment. *Acta Biomater.* **2018**, *68*,
1044 261–271.
- 1045 205. Johnell, O.; Kanis, J.A. An estimate of the worldwide prevalence and disability associated with
1046 osteoporotic fractures. *Osteoporos. Int.* **2006**, *17*, 1726–1733.
- 1047 206. Vondracek, S.F.; Linnebur, S.A. Diagnosis and management of osteoporosis in the older senior. *Clin.*
1048 *Interv. Aging* **2009**, *4*, 121–136.
- 1049 207. Mora-Raimundo, P.; Manzano, M.; Vallet-Regí, M. Nanoparticles for the treatment of osteoporosis.

- 1050 *AIMS Bioeng.* **2017**, *4*, 259–274.
- 1051 208. Arcos, D.; Boccaccini, A.R.; Bohner, M.; Díez-Pérez, A.; Epple, M.; Gómez-Barrena, E.; Herrera, A.;
1052 Planell, J.A.; Rodríguez-Mañas, L.; Vallet-Regí, M. The relevance of biomaterials to the prevention and
1053 treatment of osteoporosis. *Acta Biomater.* **2014**, *10*, 1793–1805.
- 1054 209. Brown, J.P.; Morin, S.; Leslie, W.; Papaioannou, A.; Cheung, A.M.; Davison, K.S.; Goltzman, D.; Hanley,
1055 D.A.; Hodsman, A.; Josse, R.; et al. Bisphosphonates for treatment of osteoporosis: expected benefits,
1056 potential harms, and drug holidays. *Can. Fam. Physician* **2014**, *60*, 324–333.
- 1057 210. D’Amelio, P.; Isaia, G.C. The use of raloxifene in osteoporosis treatment. *Expert Opin. Pharmacother.* **2013**,
1058 *14*, 949–956.
- 1059 211. Zaheer, S.; LeBoff, M.; Lewiecki, E.M. Denosumab for the treatment of osteoporosis. *Expert Opin. Drug*
1060 *Metab. Toxicol.* **2015**, *11*, 461–470.
- 1061 212. Trejo, C.G.; Lozano, D.; Manzano, M.; Doadrio, J.C.; Salinas, A.J.; Dapía, S.; Gómez-Barrena, E.; Vallet-
1062 Regí, M.; García-Honduvilla, N.; Buján, J.; et al. The osteoinductive properties of mesoporous silicate
1063 coated with osteostatin in a rabbit femur cavity defect model. *Biomaterials* **2010**, *31*, 8564–8573.
- 1064 213. Tokatlian, T.; Segura, T. siRNA applications in nanomedicine. *WIREs Nanomedicine and*
1065 *Nanobiotechnology* **2010**, *2*, 305–315.
- 1066 214. Balas, F.; Manzano, M.; Horcajada, P.; Vallet-Regí, M. Confinement and controlled release of
1067 bisphosphonates on ordered mesoporous silica-based materials. *J. Am. Chem. Soc.* **2006**, *128*, 8116–8117.
- 1068 215. Colilla, M.; Izquierdo-Barba, I.; Vallet-Regí, M. Phosphorus-containing SBA-15 materials as
1069 bisphosphonate carriers for osteoporosis treatment. *Microporous Mesoporous Mater.* **2010**, *135*, 51–59.
- 1070 216. Casarrubios, L.; Gómez-Cerezo, N.; Feito, M.J.; Vallet-Regí, M.; Arcos, D.; Portolés, M.T. Incorporation
1071 and effects of mesoporous SiO₂-CaO nanospheres loaded with ipriflavone on osteoblast/osteoclast
1072 cocultures. *Eur. J. Pharm. Biopharm.* **2018**, *133*, 258–268.
- 1073 217. Yu, P.; Chen, Y.; Wang, Y.; Liu, Y.; Zhang, P.; Guo, Q.; Li, S.; Xiao, H.; Xie, J.; Tan, H.; et al. Pentapeptide-
1074 decorated silica nanoparticles loading salmon calcitonin for in vivo osteoporosis treatment with
1075 sustained hypocalcemic effect. *Mater. Today Chem.* **2019**, *14*, 100189.
- 1076 218. Zhu, M.; Zhu, Y.; Ni, B.; Xie, N.; Lu, X.; Shi, J.; Zeng, Y.; Guo, X. Mesoporous silica
1077 nanoparticles/hydroxyapatite composite coated implants to locally inhibit osteoclastic activity. *ACS*
1078 *Appl. Mater. Interfaces* **2014**, *6*, 5456–5466.
- 1079 219. Ren, H.; Chen, S.; Jin, Y.; Zhang, C.; Yang, X.; Ge, K.; Liang, X.J.; Li, Z.; Zhang, J. A traceable and bone-
1080 targeted nanoassembly based on defect-related luminescent mesoporous silica for enhanced osteogenic
1081 differentiation. *J. Mater. Chem. B* **2017**, *5*, 1585–1593.
- 1082 220. Hu, Y.; Cai, K.; Luo, Z.; Jandt, K.D. Layer-by-layer assembly of β -estradiol loaded mesoporous silica
1083 nanoparticles on titanium substrates and its implication for bone homeostasis. *Adv. Mater.* **2010**, *22*,
1084 4146–4150.
- 1085 221. Esbrit, P.; Alcaraz, M.J. Current perspectives on parathyroid hormone (PTH) and PTH-related protein
1086 (PTHrP) as bone anabolic therapies. *Biochem. Pharmacol.* **2013**, *85*, 1417–1423.
- 1087 222. Lozano, D.; Manzano, M.; Carlos, J.; Salinas, A.J.; Vallet-Regí, M.; Gómez-barrena, E.; Esbrit, P.
1088 Osteostatin-loaded bioceramics stimulate osteoblastic growth and differentiation. *Acta Biomater.* **2010**, *6*,
1089 797–803.
- 1090 223. Lozano, D.; Trejo, C.G.; Gómez-Barrena, E.; Manzano, M.; Doadrio, J.C.; Salinas, A.J.; Vallet-Regí, M.;
1091 García-Honduvilla, N.; Esbrit, P.; Buján, J. Osteostatin-loaded onto mesoporous ceramics improves the
1092 early phase of bone regeneration in a rabbit osteopenia model. *Acta Biomater.* **2012**, *8*, 2317–2323.

- 1093 224. Gan, Q.; Zhu, J.; Yuan, Y.; Liu, H.; Qian, J.; Li, Y.; Liu, C. A dual-delivery system of pH-responsive
1094 chitosan-functionalized mesoporous silica nanoparticles bearing BMP-2 and dexamethasone for
1095 enhanced bone regeneration. *J. Mater. Chem. B* **2015**, *3*, 2056–2066.
- 1096 225. Zhou, X.; Feng, W.; Qiu, K.; Chen, L.; Wang, W.; Nie, W.; Mo, X.; He, C. BMP-2 Derived Peptide and
1097 Dexamethasone Incorporated Mesoporous Silica Nanoparticles for Enhanced Osteogenic Differentiation
1098 of Bone Mesenchymal Stem Cells. *ACS Appl. Mater. Interfaces* **2015**, *7*, 15777–15789.
- 1099 226. Kim, T.; Singh, R.K.; Kang, M.S.; Kim, J.; Kim, H.-W. Inhibition of osteoclastogenesis through siRNA
1100 delivery with tunable mesoporous bioactive nanocarriers. *Acta Biomater.* **2016**, *29*, 352–364.
- 1101 227. Mora-Raimundo, P.; Lozano, D.; Manzano, M.; Vallet-Regí, M. Nanoparticles to Knockdown
1102 Osteoporosis-Related Gene and Promote Osteogenic Marker Expression for Osteoporosis Treatment.
1103 *ACS Nano* **2019**, *13*, 5451–5464.
- 1104 228. Shi, M.; Chen, Z.; Farnaghi, S.; Friis, T.; Mao, X.; Xiao, Y.; Wu, C. Copper-doped mesoporous silica
1105 nanospheres, a promising immunomodulatory agent for inducing osteogenesis. *Acta Biomater.* **2016**, *30*,
1106 334–344.
- 1107 229. Gómez-cerezo, N.; Verron, E.; Montouillout, V.; Fayon, F.; Lagadec, P.; Bouler, J.M.; Bujoli, B. The
1108 response of pre-osteoblasts and osteoclasts to gallium containing mesoporous bioactive glasses. *Acta*
1109 *Biomater.* **2018**, *76*, 333–343.
- 1110 230. Rebeca, P.; Sanchez-salcedo, S.; Lozano, D.; Heras, C.; Esbrit, P.; Vallet-Regí, M.; Salinas, A.J. Osteogenic
1111 Effect of ZnO-Mesoporous Glasses Loaded with Osteostatin. *Nanomaterials* **2018**, *8*.
- 1112 231. Liang, H.; Jin, C.; Ma, L.; Feng, X.; Deng, X.; Wu, S.; Liu, X.; Yang, C. Accelerated Bone Regeneration by
1113 Gold-Nanoparticle-Loaded Mesoporous Silica through Stimulating Immunomodulation. *ACS Appl.*
1114 *Mater. Interfaces* **2019**, *11*, 41758–41769.
- 1115 232. Gómez-cerezo, N.; Casarrubios, L.; Morales, I.; Feito, M.J.; Vallet-Regí, M.; Arcos, D.; Portolés, M.T.
1116 Effects of a mesoporous bioactive glass on osteoblasts, osteoclasts and macrophages. *J. Colloid Interface*
1117 *Sci.* **2018**, *528*, 309–320.
- 1118 233. Lozano, D.; Trejo, C.G.; Gómez-barrena, E.; Manzano, M.; Doadrio, J.C.; Salinas, A.J.; Vallet-Regí, M.;
1119 García-Honduvilla, N.; Esbrit, P.; Buján, J. Osteostatin-loaded onto mesoporous ceramics improves the
1120 early phase of bone regeneration in a rabbit osteopenia model. *Acta Biomater.* **2012**, *8*, 2317–2323.

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