

Total Synthesis of Mycalolides A and B through Olefin Metathesis

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Total Synthesis of Mycalolides A and B via Olefin Metathesis **

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Abstract: *An asymmetric total synthesis of the tris-oxazole marine macrolides, mycalolides A and B is described. This synthesis involves the convergent assembly of highly functionalized C1–C19 tris-oxazole and C20–C35 side-chain segments through the use of olefin metathesis and esterification, as well as Julia–Kocienski olefination and enamide formation as key steps.*

Mycalolides are cytotoxic and antimycotic tris-oxazole macrolides, which was isolated from the marine sponge *Mycale* sp.^[1] They inhibit actomyosin Mg^{2+} –ATPase^[2] and show potent actin-depolymerizing activity by forming a 1:1 complex with monomeric molecule.[3] Mycalolide B (**1**) contains a 2,3-*O*dimethyl-D-glyceryl ester moiety and 13 asymmetric centers as structural features, while a closely related mycalolide A (**2**) contains a ketone functionality at C30. Several tris-oxazole macrolides that are closely related to mycalolides have been isolated, such as ulapualides, $[4]$ halichondramides, $[5]$ jaspisamides,^[6] and kabiramides;^[7] all of these exhibit actindepolymerizing activity and potent cytotoxicity, and some induce apoptosis in tumor cells.^[8] Thus, these agents may be useful for the design and development of novel pharmacological tools for analyzing actin-mediated cell functions, such as muscle contraction, cell motility, and cytokinesis, as well as those of therapeutic agents.[9]

Mycalolides can be divided into two structurally characteristic parts: the C1–C24 tris-oxazole macrolactone and the C25–C35 side-chain functionalized by *N-*methyl enamide moiety. Studies on the structure–activity relationships [10] and photolabeling experiments $[11]$ have established that the sidechain part of mycalolides is important for its ability to bind to and depolymerize actin. In addition, X-ray analyses of the actin– kabiramide $C₁^[12]$ actin–jaspisamide A,^[12] and actin–ulapualide A $R_{\text{complexes}}$ $^{[13]}$ have revealed that their side-chain parts intercalate into the hydrophobic cleft between subdomains 1 and 3 of actin. Meanwhile, we recently synthesized the 19*E*- and 19*Z*-lactone analogs of mycalolides that lack the C25–C35 sidechain; these analogs exhibited moderate cytotoxicity against

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tumor cells (ca. 1/100 of **1**), but did not show actindepolymerizing properties or antimycotic activity against pathogenic fungi.^[14] Thus, both the side-chain and macrolactone moieties were suggested to be essential for the potent biological activities of the parent molecules.

Scheme 1. Strategies for the synthesis of mycalolides A and B.

Due to their extraordinary structures and biological activities, mycalolides and their congeners have received considerable attention in the synthetic community, and several approaches to the construction of conformationally-restricted tris-oxazole macrolactone structures have been described.^[15] To date, total syntheses of mycalolide A (2) ^[16] and ulapualide A ^[17] have been accomplished, in which Yamaguchi lactonization, cyclization of the central oxazole ring, or intramolecular Horner– Wadsworth–Emmons olefination were used to construct macrocycles. However, no total synthesis of mycalolide B has been disclosed to date. We describe here the first total synthesis of (–)-mycalolide B (**1**) and the second synthesis of mycalolide A (**2**) through the use of olefin metathesis as a key step.

Based on the finding that olefin metathesis is a useful method for connecting the C19–C20 double bonds in mycalolide analogs,[10c,18] we designed a plan for the synthesis of **1** (Scheme 1). After disconnection of the C35 *N*-methyl enamide moiety and the C30 ester bond, the macrolactone structure of **1** could be divided into a C1–C19 tris-oxazole segment **3** and a C20–C35 side-chain segment **4**. We expected that the convergent assembly of **3** and **4** via esterification / ring-closing metathesis (RCM) would efficiently afford a key macrolactone.

Instead, cross metathesis of **3** and **4**, in which the carboxyl or hydroxyl groups are protected, and subsequent macrolactonization could also provide the same intermediate. While the side-chain segment 4 was previously synthesized.^[10,18] in this study we planned to modify the synthetic route, which includes the Julia–Kocienski olefination [19] between phenyl tetrazole (PT)-sulfone **5** and aldehyde **6**.

Our synthesis started with the preparation of **5** (Scheme 2). Methylation of the known *syn*-aldol **7** [10b] with methyl trifluoromethanesulfonate (MeOTf) and removal of the chiral auxiliary with LiBH4 yielded primary alcohol **8**. Conversion of **8** into the PT-sulfide with aryl disulfide/ Bu_3P and subsequent oxidation with *m*-CPBA yielded PT-sulfone **5**.

Scheme 2. Synthesis of the C20–C35 segment **4**. Reagents and conditions: a) MeOTf, 2,6-di-tert-butylpyridine, CH₂Cl₂; b) LiBH₄, EtOH, Et₂O-THF, –10 °C; c) 5,5'-dithiobis(1-phenyl-*1H*-tetrazole), tri-*n*-butylphosphine, THF; d) *m*CPBA, NaHCO₃, CH₂Cl₂; e) H₂, Pd(OH)₂/C (20 mol%), NaHCO₃, EtOH; f) 3,4-dimethoxybenzyloxymethyl chloride, *i*-Pr₂NEt, CH₂Cl₂; g) NH₄F, MeOH, 40 °C; h) Dess-Martin periodinane, pyridine, CH₂Cl₂; i) CH₂=CHCH₂MgBr, THF–Et₂O; j) Mel, NaH, THF; k) ⁿBu₄NF, THF, rt to 40 °C.

Next, Julia–Kocienski coupling was examined. Despite the sterically hindered, branched structures of both starting materials, treatment of **5** with LHMDS followed by the addition of aldehyde **6** [20] in THF at –78 °C afforded olefin **9** in 60% yield (condition A, *E*/*Z* = 1.2/1). After several attempts, the yield was improved to 92% (condition B, *E*/*Z* = 1/1.5) with the use of the same base in 1,2-dimethoxyethane (DME) at –55 °C to room temperature. While an excess amount of PT-sulfone **5** (2.5 eq.) was required to complete the reaction, this material was recovered quantitatively and reused.

Catalytic hydrogenation of the C=C double bond and hydrogenolysis of the benzyl group from the *E*/*Z*-mixture of **9** proceeded concurrently with palladium (II) hydroxide on carbon. Subsequent protection of the C30 hydroxy group as a 3,4 dimethoxyphenylmethoxymethyl (DMPMOM) group afforded previously synthesized ether **10**. [10b] Selective deprotection of the TBS group in **10** with NH4F and oxidation of the primary alcohol with Dess–Martin periodinane provided aldehyde **11**. Grignard reaction of **11** with allylmagnesium bromide resulted in a mixture of *S*- and *R*-alcohol **12** (*dr* = 2.7/1), which were separated by column chromatography.^[21] Finally, methylation of the secondary

alcohol in 22*S*-**12** and deprotection of the remaining TBS group with tetra-*n*-butylammonium fluoride (TBAF) gave the C20–C35 segment **4**.

With the side-chain segment **4** in hand, we initially considered the RCM approach to reduce unnecessary protection / deprotection steps (Scheme 3). Condensation of the C1–C19 segment **3** [18] with **4** by the Shiina procedure using 2-methyl-6 nitrobenzoic anhydride (MNBA) $^{[22]}$ afforded the RCM precursor **13**. We previously reported that treatment of **13** with 30 mol% of 2nd-generation Grubbs catalyst (**17a**) [23] in degassed refluxing toluene led to the decomposition of the starting material (entry 1).^[18] However, in refluxing CH₂Cl₂, tris-oxazole lactone **15** was obtained as an *E/Z* mixture (40%, *E*/*Z* = 1.9:1, entry 2), while the reaction did not run to completion.

^a See ref. 18.

s. m. was decomposed and not recovered

Scheme 3. Synthesis of macrolactones **15** and **16** via ring-closing metathesis.

Due to the instability of catalyst **17a** for the slow metathesis reaction of macrocycle precursors, we next examined 2nd-generation Hoveyda–Grubbs (*HG-II*) catalyst (**17b**).[24] We previously reported that treatment of **13** with 30 mol% **17b** in refluxing toluene afforded the RCM product **15** in higher yield but with undesired C19–C20 *Z*-isomer slightly preferred (76%, $E/Z = 1/1.2$, entry 3).^[18] Meanwhile, in the model RCM reactions of C1–C24 macrolactone analogs, the solvent polarity was found to significantly affect the stereoselectivity; the

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Z-isomer was preferred in *n*-hexane and toluene (*E*/*Z* = 1/1.9~2.5), while the *E*-isomer was preferred in CH_2Cl_2 ($E/Z =$ 1.8/1).[14] In fact, for the RCM reaction of **13** with **17b** in refluxing $CH₂Cl₂$, the ratio was improved to 2.0:1, but the reaction did not run to completion, similar to the use of **17a** (entry 4). These results suggested that the C25–C35 segment in **13** minimally affected the stereoselectivity, but decreased the reactivity for RCM reactions, probably due to the steric hindrance in forming the ruthenocyclobutane intermediate. Under refluxing conditions in 1,2-dichloroethane (DCE), the stereoselectivity decreased to 1.0:1 (entry 5). To facilitate the initiation of the catalytic cycle at lower temperature, two highly reactive *HG-II* catalyst derivatives **17c** (Grela catalyst) ^[25] and **17d** (Zhan catalyst 1B) ^[26] were examined, in which nitro or *N*,*N*-dimethylsulfonamide groups are substituted on the 2-isopropoxybenzylidene ligand. Notably, the use of both electron-deficient catalysts similarly increased the yield of **15** to 69–75%, while stereoselectivity was still low (*E*/*Z* = 1.6–1.7/1, entries 6 and 7). We expected that such low stereoselectivity of RCM precursor **13** was due to the presence of structurally hindered C3 TBDPS group. For comparison, a C3 hydroxy analog **14** was prepared from **13** by the treatment with TBAF along with acetic acid (AcOH). With the use of catalyst **17c** in refluxing CH₂Cl₂, the stereoselectivity of C3 hydroxy macrolactone **16** was improved to 2.7:1, but the yield was lower than that of **15** (entry 8).

Scheme 4. Synthesis of **15** via macrolactonization. Reagents and conditions: a) 2,2,2-trichloroethanol, EDCI·HCl, DMAP, CH₂Cl₂; b) TESCl, ImH, DMF, 40 °C; c) **17b** (20 mol%), CH₂Cl₂, reflux; d) AcOH-THF-H₂O; e) Zn, 1 M NH₄OAc aq., THF; f) 2,4,6-trichlorobenzoyl chloride, *i*-Pr₂NEt, benzene, then dropwise addition into DMAP in benzene.

A cross metathesis / macrolactonization approach was examined next (Scheme 4). Condensation of carboxylic acid **3** with 2,2,2-trichloroethanol provided trichloroethyl (TCE) ester **18**. Triethylsilyl (TES) protection of the secondary alcohol in **4** gave silyl ether **19**. In contrast to the RCM reactions, treatment of **18** and **19** (1.2 equiv.) with 20 mol% of *HG-II* catalyst (**17b**) in refluxing CH2Cl2 (13 mM for **18**) preferentially yielded the coupling product **20** in an *E*-selective manner (*E*/*Z* = 5.0:1).[27] After the TES group in (*E*)-**20** was removed under mild acidic conditions, the resultant alcohol was treated with activated zinc in acetate buffer to afford seco acid **21**. Macrolactonization of **21** by the Yamaguchi procedure ^[28] readily proceeded to give the lactone **15**. Due to the higher stereoselectivity, the cross metathesis-macrolactonization approach was preferred to the RCM approach.

The stage was then set for functionalization of the last side-chain part (Scheme 5). Acidic hydrolysis of the C35 methyl acetal in **15** afforded hemiacetal **22**. Selective reductions of the five-membered hemiacetal in **22** using conventional hydride reagents were unsuccessful.^[29] To our delight, however, Luche reduction of **22** at –20 °C exclusively led to 1,2-reduction of the C7 ketone followed by C35 hemiacetal reduction at 0 °C to afford triol **23** quantitatively (*dr* = 10:1 at C7). Next, trityl group protection of the primary alcohol, and chemoselective oxidation of the allylic alcohol with manganese dioxide gave ketone **24**. Subsequent acetylation of the remaining C32 secondary alcohol, removal of the trityl group with formic acid in ether, and oxidation of the primary alcohol with Dess–Martin periodinane gave aldehyde **25**. Dehydrating condensation with *N*methylformamide under acidic conditions,^[30] and deprotection of the DMPMOM group with 2,3-dichloro-5,6-dicyano-*p*benzoquinone (DDQ) afforded secondary alcohol **26**.

Scheme 5. Synthesis of mycalolides A and B. Reagents and conditions: a) 1 M HCl aq., 1,2-dimethoxyethane; b) NaBH₄, CeCl₃·7H₂O, MeOH, -20 to 0 °C; c) TrCl, pyridine; d) MnO₂, CH₂Cl₂; e) Ac₂O, DMAP, pyridine; f) HCOOH, Et₂O; g) Dess-Martin periodinane, pyridine, CH_2Cl_2 ; h) MeNHCHO, PPTS, hydroquinone, MS3A, benzene, reflux; i) DDQ, CH₂Cl₂, ^tBuOH, 1 M phosphate buffer (pH 6.0); j) 2,3-di-*O*-methyl-D-glyceric acid, 2,4,6-trichlorobenzoyl chloride, Et₃N, DMAP, benzene; k) ⁿBu₄NF, AcOH, THF.

Finally, condensation of **26** with 2,3-di-*O*-methyl-D-glyceric acid using the Yamaguchi procedure and removal of the C3 TBDPS group by TBAF along with AcOH furnished mycalolide B (1) in analytically pure form. The ${}^{1}H$ and ${}^{13}C$ NMR spectra of the synthetic mycalolide B are consistent with those of the natural product, along with its specific optical rotation ${[\alpha]_D}^{25}$ –55 (c 0.55, CHCl₃) for synthetic **1**; $[\alpha]_D$ –53 (*c* 1.3, CHCl₃) for natural **1**^[1a]. Synthetic **1** was also identical to an authentic sample on the basis of TLC and HPLC analysis. In addition, oxidation of the secondary alcohol in **26** with Dess–Martin periodinane gave authentic TBDPS-protected mycalolide A,^[16] and removal of the TBDPS group afforded mycalolide A (2), whose ¹H NMR data coincided with the reported one.^[1a,16]

In summary, we have developed a convergent approach for the synthesis of the tris-oxazole marine macrolides, and completed total synthesis of mycalolides A and B. The key elements in this synthesis include the use of RCM / cross metathesis and esterification as fragment coupling technology for complex building blocks that possess a variety of functional groups. Further studies on the synthesis and structure-activity relationships of mycalolides and related actin-targeting natural products, as well as on their mechanisms of action, are currently underway.

Keywords: marine natural product • total synthesis • olefin metathesis • macrolactonization

- [1] a) N. Fusetani, K. Yasumuro, S. Matsunaga, K. Hashimoto, *Tetrahedron Lett.* **1989**, *30*, 2809–2812; b) S. Matsunaga, P. Liu, C. A. Celatka, J. S. Panek, N. Fusetani, *J. Am. Chem. Soc.* **1999**, *121*, 5605– 5606; c) S. Wada, S. Matsunaga, S. Saito, N. Fusetani, S. Watabe, *J. Biochem.* **1998**, *123*, 946–952; d) P. Phuwapraisirisan, S. Matsunaga, R. W. M. van Soest, N. Fusetani, *J. Nat. Prod.* **2002**, *65*, 942–943; e) S. Tsukamoto, K. Koimaru, T. Ohta, *Mar. Drugs* **2005**, *3*, 29–35.
- [2] a) S. Matsunaga, T. Sugawara, N. Fusetani, *J. Nat. Prod.* **1998**, *61*, 1164–1167; b) M. A. Rashid, K. R. Gustafson, J. H. Cardeilina II, M. R. Boyd, *J. Nat. Prod.* **1995**, *58*, 1120–1125.
- [3] a) M. Hori, S. Saito, Y. Shin, H. Ozaki, N. Fusetani, H. Karaki, *FEBS Lett.* **1993**, *322*, 151–154; b) S. Saito, S. Watabe, H. Ozaki, N. Fusetani, H. Karaki, *J. Biol. Chem.* **1994**, *269*, 29710–29714.
- [4] J. A. Roesener, P. J. Scheuer, *J. Am. Chem. Soc.* **1986**, *108*, 846–847.
- [5] a) S. Matsunaga, N. Fusetani, K. Hashimoto, K. Koseki, M. Noma, H. Noguchi, U. Sankawa, *J. Org. Chem.* **1989**, *54*, 1360–1363; b) M. R. Kernan, T. F. Molinski, D. J. Faulkner, *J. Org. Chem.* **1988**, *53*, 5014– 5020; c) S. –C. Chung, S. –H. Lee, K. H. Jang, W. Park, J. –E. Jeon, H. Oh, J. Shin, K. –B. Oh, *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3198–3201.
- [6] J. Kobayashi, O. Murata, H. Shigemori, *J. Nat. Prod.* **1993**, *56*, 787– 791.
- [7] a) S. Matsunaga, N. Fusetani, J. Hashimoto, *J. Am. Chem. Soc.* **1986**, *108*, 847–849; b) J. Tanaka, Y. Yan, J. Choi, J. Bai, V. A. Klenchin, I. Rayment, G. Marriott, *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 13851– 13856.
- [8] a) M. Kita, K. Yoneda, Y. Hirayama, K. Yamagishi, Y. Saito, Y. Sugiyama, Y. Miwa, O. Ohno, M. Morita, K. Suenaga, H. Kigoshi, *ChemBioChem* **2012**, *13*, 1754–1758; b) O. Ohno, M. Morita, K. Kitamura, T. Teruya, K. Yoneda, M. Kita, H. Kigoshi, K. Suenaga, *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1467–1471; c) S. Y. Bae, G. D. Kim, J. Jeon, J. Shin, S. K. Lee, *Toxicology in Vitro* **2013**, *27*, 694–699.
- [9] Reviews: a) K. Yamada, M. Ojika, H. Kigoshi, K. Suenaga, *Nat. Prod. Rep.* **2009**, *26*, 27–43; b) K. Yamada, M. Ojika, H. Kigoshi, K. Suenaga, *Proc. Jpn. Acad. Ser. B* **2010**, *86*, 176–189; c) M. Kita, H. Kigoshi, *Nat. Prod. Rep.* **2015**, *32*, 534–542.
- [10] a) K. Suenaga, S. Miya, T. Kuroda, T. Handa, K. Kanematsu, A. Sakakura, H. Kigoshi, *Tetrahedron Lett.* **2004**, *45*, 5383–5386; b) K. Suenaga, T. Kimura, T. Kuroda, K. Matsui, S. Miya, S. Kuribayashi, A. Sakakura, H. Kigoshi, *Tetrahedron* **2006**, *62*, 8278–8290; c) T. Kimura, S. Kuribayashi, T. Sengoku, K. Matsui, S. Ueda, I. Hayakawa, K. Suenaga, H. Kigoshi, *Chem. Lett.* **2007**, *36*, 1490–1491; d) K. Kitamura, T. Teruya, T. Kuroda, H. Kigoshi, K. Suenaga, *Bioorg. Med. Chem. Lett.* **2009**, *19*, 1896–1898; e) K. Kobayashi, Y. Fujii, Y. Hirayama, S.

Kobayashi, I. Hayakawa, H. Kigoshi, *Org. Lett.* **2012**, *14*, 1290–1293; f) I. Hayakawa, H. Kigoshi, *Heterocycles* **2015**, *91*, 1137–1155.

- [11] a) T. Kuroda, K. Suenaga, A. Sakakura, T. Handa, K. Okamoto, H. Kigoshi, *Bioconjugate Chem.* **2006**, *17*, 524–529; b) M. Kita, Y. Hirayama, K. Yamagishi, K. Yoneda, R. Fujisawa, H. Kigoshi, *J. Am. Chem. Soc.* **2012**, *134*, 20314–20317.
- [12] V. A. Klenchin, J. S. Allingham, R. King, J. Tanaka, G. Marriott, I. Rayment, I. *Nat. Struct. Biol.* **2003**, *10*, 1058–1063.
- [13] J. S. Allingham, J. Tanaka, G. Marriott, I. Rayment, *Org. Lett.* **2004**, *6*, 597–599.
- [14] M. Kita, H. Oka, A. Usui, T. Ishitsuka, Y. Mogi, H. Watanabe, H. Kigoshi, *Tetrahedron* **2012**, *68*, 8753–8760.
- [15] Review: K. S. Yeung, I. Paterson, *Angew. Chem.* **2002**, *114*, 4826– 4847; *Angew. Chem. Int. Ed.* **2002**, *41*, 4632–4653.
- [16] a) P. Liu, J. S. Panek, *J. Am. Chem. Soc.* **2000**, *122*, 1235–1236; b) J. S. Panek, P. Liu, *J. Am. Chem. Soc.* **2000**, *122*, 11090–11097.
- [17] a) S. K. Chattopadhyay, G. Pattenden, *Tetrahedron Lett.* **1998**, *39*, 6095–6098; b) S. K. Chattopadhyay, J. Kempson, A. McNeil, G. Pattenden, M. Reader, D. E. Rippon, D. Waite, *J. Chem. Soc. Perkin Trans. 1* **2000**, 2415–2428; c) S. K. Chattopadhyay, G. Pattenden, *J. Chem. Soc. Perkin Trans. 1* **2000**, 2429–2454; d) G. Pattenden, N. J. Ashweek, C. A. G. Baker–Glenn, G. M. Walker, J. G. K. Yee, *Angew. Chem.* **2007**, *119*, 4437–4441; *Angew. Chem. Int. Ed.* **2007**, *46*, 4359– 4363; e) G. Pattenden, N. J. Ashweek, C. A. G. Baker–Glenn, J. Kempson, G, M. Walker, J. G. K. Yee, *Org. Biomol. Chem.* **2008**, *6*, 1478–1497.
- [18] M. Kita, H. Watanabe, T. Ishitsuka, Y. Mogi, H. Kigoshi, *Tetrahedron Lett.* **2010**, *51*, 4882–4885.
- [19] a) P. R. Blakemore, W. J. Cole, P. J. Kocienski, A. Morley, *Synlett*, **1998**, 26–28; b) P. R. Blakemore, *J. Chem. Soc. Perkin Trans. 1* **2002**, 2563–2585.
- [20] H. Kigoshi, K. Suenaga, T. Mutou, T. Ishigaki, T. Atsumi, H. Ishiwata, A. Sakakura, T. Ogawa, M. Ojika, K. Yamada, *J. Org. Chem.* **1996**, *61*, 5326–5351.
- [21] 22*R*-**12** was converted to the 22*S* isomer in 64% yield (2 steps). See the Supporting Information for details.
- [22] a) I. Shiina, M. Kubota, R. Ibuka, *Tetrahedron Lett.* **2002**, *43*, 7535– 7539; b) I. Shiina, M. Kubota, H. Oshiumi, M. Hashizume, *J. Org. Chem.* **2004**, *69*, 1822–1825.
- [23] a) M. Scholl, S. Ding, C. W. Lee, R. H. Grubbs, *Org. Lett.* **1999**, *1*, 953– 956; b) A. K. Chatterjee, T. –L. Choi, D. P. Sanders, R. H. Grubbs, *J. Am. Chem. Soc.* **2003**, *125*, 11360–11370.
- [24] a) J. S. Kingsbury, J. P. A. Harrity, P. J. Bonitatebus, A. H. Hoveyda, *J. Am. Chem. Soc.* **1999**, *121*, 791–799; b) S. B. Garber, J. S. Kingsbury, B. L. Gray, A. H. Hoveyda, *J. Am. Chem. Soc.* **2000**, *122*, 8168–8179.
- [25] a) K. Grela, S. Harutyunyan, A. Michrowska, *Angew. Chem.* **2002**, *114*, 4210–4212; *Angew. Chem. Int. Ed.* **2002**, *41*, 4038–4040; b) A. Michrowska, R. Bujok, S. Harutyunyan, V. Sashuk, G. Dolgonos, K. Grela. *J. Am. Chem. Soc.* **2004**, *126*, 9318–9325.
- [26] Z. –Y. Zhan, WO Patent 2007003135, 2007.
- [27] A homodimer of **19** was also formed (*E*/*Z* ~4/1, 8%), which could also be subjected to cross metathesis with **18**, or converted to **20** by ethenolysis. See the Supporting Information for details.
- [28] J. Inanaga, K. Hirata, H. Saeki, T. Kabuki, M. Yamaguchi, *Bull. Chem. Soc. Jpn.* **1979**, *52*, 1989–1993.
- [29] For example, sodium borohydride reduction of **22** at 0 °C preferred the 1,4-reduction of conjugated ketone (quant.), and sodium trimethoxyborohydride reduction at room temperature resulted in a mixture of unreacted conjugated ketone (45%) and α , β -saturated C7 alcohol (36%), while the hemiacetal was completely converted to 1,4 diol in both cases.
- [30] To avoid the formation of several β-eliminated products including the demethoxy, deacetoxy, and de-DMPMOM ether groups, the reaction was stopped before the completion, and unreacted aldehyde **25** was recovered (33%).

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An asymmetric total synthes is of the tris -oxazole macrolides , mycalolides A and B, is described. This synthesis involves the convergent assembly of C1 –C19 tris -oxazole and C20 –C35 side -chain segments through the use of olefin metathesis and esterification as key steps .

Mycalolide B (1)

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