

REVIEW

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Advances in prevention and therapy of neonatal dairy calf diarrhoea: a systematical review with emphasis on colostrum management and fluid therapy

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Abstract

Neonatal calf diarrhoea remains the most common cause of morbidity and mortality in preweaned dairy calves worldwide. This complex disease can be triggered by both infectious and non-infectious causes. The four most important enteropathogens leading to neonatal dairy calf diarrhoea are *Escherichia coli*, rota- and coronavirus, and *Cryptosporidium parvum*. Besides treating diarrhoeic neonatal dairy calves, the veterinarian is the most obvious person to advise the dairy farmer on prevention and treatment of this disease. This review deals with prevention and treatment of neonatal dairy calf diarrhoea focusing on the importance of a good colostrum management and a correct fluid therapy.

Keywords: Neonatal calf diarrhoea, Fluid therapy, Colostrum management

Introduction

Neonatal calf diarrhoea (NCD), defined in this paper as diarrhoea in calves aged 1-month-old or younger is a complex disease that can be triggered by both infectious and non-infectious causes. The prevalence and incidence risk for NCD in dairy cattle herds has recently been reported to be 19.1 and 21.2%, respectively [1,2]. In calves aged <31 days, enteritis is the most common cause of death with a case fatality risk for NCD of 4.9% and a peak probability of dying due to enteritis during the second week of life [1,3]. Enterotoxigenic *Escherichia coli* K99/F5, rota- and coronavirus, and *Cryptosporidium* spp. (≥85% *C. parvum*) are the 4 most important enteropathogens causing NCD worldwide with rotavirus and *C. parvum* most frequently identified in faecal samples from young calves [2,4-7]. *Escherichia coli* K99/F5 typically causes diarrhoea in calves 1-4 days old and the other three cause diarrhoea most often in 1 to 3-week-old calves. Neonatal calf diarrhoea prevalence for *E. coli*, rota- and

coronavirus, and *C. parvum* ranges from 2.6-45.1%, 17.7-79.9%, 3.1-21.6% and 27.8-63.0%, respectively [2,5,7-12]. However, the enteropathogens most commonly implicated in NCD outbreaks can also be found in faecal samples from healthy calves, meaning that the presence of a pathogen is not always causative [2,7]. Less common enteropathogens causing NCD are *Salmonella* spp., attaching and effacing *E. coli* among which enteropathogenic *E. coli*, enterohemorrhagic *E. coli* and Shiga toxin producing *E. coli*, *Clostridium difficile*, *Clostridium perfringens*, and torovirus [2,13-16]. The veterinarian has an important advisory role in the daily prevention and treatment challenge in practice. The aim of this review is therefore to provide a critical analysis of recent literature on prevention and treatment of NCD with emphasis on colostrum management and fluid therapy.

Search strategy and inclusion and exclusion criteria for references

Web of Science (<https://webofknowledge.com/>) was searched in September 2014 with 'calf' AND 'neonatal' AND 'diarrhoea' AND 'fluid therapy' (46 references) followed by 'calf' AND 'neonatal' AND 'diarrhoea' AND 'colostrum' (136 references) as subject-specific terms.

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Articles published in peer reviewed journals of the past decade (2004-present) were considered to retain reliable and recent advances on prevention and treatment of NCD (93 references). Literature reviews and case reports were not included but were scanned for missed references (190 references). Relevance screening was conducted by the first author withholding only English papers dealing with colostrum management or fluid therapy in conjunction with neonatal diarrhoea in dairy calves (105 references). There were no publication status restrictions.

Review

Prevention

Dairy farms with a confirmed NCD diagnosis should consult a veterinarian. Together they should go through the herd anamnesis addressing the young stock management creating a list with possible critical control points. Key questions in this anamnesis are: colostrum management, housing and hygiene, feeding of the calves, possible periods of stress, drugs used, and prevention of immunomodulating infectious diseases (e.g. bovine viral diarrhoea virus or infectious bovine rhinotracheitis virus).

Colostrum management is the most important preventive measure that should be addressed and this will be discussed in further detail later in this review.

In view of the increasing pressure in the veterinary field to lower the use of antimicrobials (AB), using AB as a prophylactic or metaphylactic measure is debatable. Metaphylactic use of AB can only be recommended for a short period on herds actively struggling with *E. coli* diarrhoea problems. Furthermore, calves receiving prophylactic AB in their milk for the first 2 weeks of life have a 28% greater risk for diarrhoea compared with calves receiving no prophylactic AB in their milk [17].

In a Swedish study in 2010 disinfection of single pens between calves was more common in herds suffering from NCD when compared to herds without problems [7]. This can be explained by herds starting with disinfecting procedures if suffering from NCD. Single pens should be disinfected before moving any new calf and not only when an outbreak of NCD occurs.

The control of *C. parvum* can only be achieved by combining a good hygiene management and effective preventive drugs [18-20]. Newborn calves should not be mixed with older calves since the age of the calf is an important risk factor for shedding of *C. parvum* oocysts [18,21-23]. In Europe, halofuginone lactate is the only registered product for prevention and treatment of *C. parvum*. Different studies find a delayed and lower oocyst output peak the first 2 weeks after birth in halofuginone treated groups as compared to placebo groups [21,24-27].

Calves born to dams vaccinated against *E. coli*, rotavirus and coronavirus also shed less *C. parvum* oocysts [6]. This

lower shedding of *C. parvum* oocyst is most likely a reflection of a generally higher standard of herd management in these herds, rather than a direct protective effect. A vaccine against *C. parvum* has not been developed yet.

The literature is contradictory or inconclusive concerning ancillary preventive products such as phytopharmaceuticals (e.g. clinoptilitezeolite) and probiotics (e.g. *E. coli* Nissle 1917) [28-30]. The herd veterinarian is advised to consult results of peer-reviewed controlled trials for separate products.

Colostrum management

Although the importance of a good colostrum management leading to an adequate passive transfer is undebatable in the prevention of NCD [17,31-33], there are studies showing no significant effect of colostrum feeding routines on the risk of diarrhoea or on the risk of shedding *C. parvum* [4,6]. This lack of a significant effect can be explained by a high number of diarrhoea cases caused by *C. parvum*, for which colostrum IgG is less protective [6]. An FPT prevalence <10% is considered as a rational and achievable goal when using a cut-off value of 10 g IgG/l [34]. However, several studies proposed serum IgG concentrations up to 15 g/l as cut-off points for defining failure of passive transfer (FPT) [1,35,36]. Percentages of FPT range from 8.4 to >90.5% depending on the cut-off value, the population studied, and/or the method used to estimate calf serum IgG levels [1,12,17,37-39]. The odds of FPT are higher when there is no on-farm routine screening [38]; therefore, IgG concentration of 48-hours-old calves should be estimated regularly to test the compliance of the colostrum management and is most practical in the field by determining serum total protein using a refractometer.

Colostrum also contains other beneficial constituents in higher concentrations than normal milk: immunologically active leukocytes, fat, protein, fat-soluble vitamins (e.g. retinol, tocopherol, β -carotene), water-soluble vitamins (e.g. niacin, thiamine, riboflavin, vitamin B12, pyridoxal, pyridoxamine, pyridoxine), minerals (e.g. Ca, P, Mg, Na, K, Zn, Fe, Cu, S, Mn) and, non-specific antimicrobial factors (e.g. lactoferrin) [40].

The functional importance of colostrum leukocytes is not yet fully understood. Some studies show that they enhance lymphocyte responses to nonspecific mitogens and specific antigens and increase antigen-presenting capacity [41-44], while others suggest that the role of fresh colostrum leukocytes may not be as important as once thought [45]. As protection against *C. parvum* is mainly cell-mediated, further research into the importance of colostrum leukocytes is warranted.

Colostrum quality

IgG concentration Colostrum IgG concentration is an important factor that affects whether calves receive

sufficient passive immunity from colostrum [46]. Unfortunately, the amount of IgG in maternal colostrum varies dramatically among cows (<1-235 g/l) with 29.4-57.8% of samples that do not reach the desired amount of 50 g IgG/l [34,47-49]. Colostral quality is difficult to estimate by the farmer based on produced volume or appearance of the colostrum. Use of weight at first milking as a screening test to identify bovine colostrum with inadequate IgG concentration is not justified because of the low sensitivity [50]. Somatic cell count, measured after calving, was significantly higher in cows producing colostrum of inferior quality compared with those producing high-quality colostrum [47]. For all these reasons, it is better to measure colostral IgG content indirectly by using a colostrometer (50 g IgG/l = density of 1045) or a brix refractometer (50 g IgG/l = 21-22 Brix) or directly by a cow-side immunoassay kit (single line = IgG concentration >50 g IgG/l [50-53]). To measure colostrum quality it should be avoided to use forestripping samples for testing purposes, as these samples may overestimate the IgG concentration [54]. Colostrum collected more than 2 hours after calving significantly lowers the colostral IgG concentration [55,56], probably because of dilutional effects and because colostral Ig diffuse passively into the cow's systemic circulation if colostrum is not milked out as soon as possible after calving. Colostrum from cows without a dry period has a lower IgG concentration compared with colostrum from cows having a dry period of 28 or 56 days [57]. Pooling of colostrum is not advised since this is a risk factor for FPT [38]. Cows in their 3rd or 4th parity or older usually have significantly higher levels of IgG per liter colostrum than heifers or 2nd parity cows [34,47,48] which is reflected in calves born to heifers having a greater risk for FPT than calves born to multiparous cows [32]. However, this correlation between parity and colostrum IgG concentration is not always evident [36]. Discarding colostrum from heifers is thus not advisable.

Colostrum replacement products have a highly variable performance [49,58-65], and the herd veterinarian is advised to consult results of peer-reviewed controlled trials for separate products. Colostrum supplemented in milk replacer during the first 2 weeks of life (10 g IgG, bid) reduces NCD rates because of the intestinal activity of colostral antibodies and epithelial growth promoting substances [66].

Bacteriological quality Critical control points for bacterial contamination of colostrum are the harvest and storage process. Storing colostrum at ambient temperatures results in a significant increase of bacteria [67]. Fresh colostrum can be stored up to 96 hours at 4°C when potassium sorbate is added as a preservative [67]. Industry recommendations for bacterial load are limited to 100,000 colony forming units (cfu)/ml. Depending on the time of

sampling or used storage method, 0-43% of colostrum samples exceed the maximum advised bacterial count of 100,000 cfu/ml [48,67].

Pasteurizing colostrum (60°C, 60 min) can reduce bacterial load while the viscosity and the colostral IgG concentration remain within acceptable limits for feeding [68-72]. Higher-quality batches of colostrum suffer a significantly greater magnitude of loss of IgG as compared with lower- or intermediate- quality batches of colostrum [68,71]. Calves fed pasteurized colostrum had significantly higher serum IgG concentrations, serum protein values, and/or a greater apparent efficiency of absorption, compared with calves fed unpasteurized colostrum [46,72-75], probably explained by bacteria that bind free IgG in the gut lumen and block uptake and transport of IgG molecules across intestinal epithelial cells. Moreover, calves fed pasteurized colostrum had a significantly decreased risk for treatment for diarrhoea [46].

Colostrum feeding practices

Calves allowed to nurse their dam have higher odds of FPT than calves separated from their dam within 3 hours of birth [37,38]. When allowed to nurse, calves drink too late and too little colostrum. Feeding calves as much colostrum as they want by nipple bottle within 1-4 hours after birth and at 12 hours of age substantially reduces the probability of FPT [34,37,76]. Bottle fed calves that do not ingest colostrum voluntarily, should be tube fed [34,76]. Feeding at least 150 to 200 g of colostral IgG is required for adequate passive transfer of colostral IgG when colostrum is administered once by oesophageal intubation <2 hours after birth [36]. To estimate the exact amount of IgG given to a calf, colostrum quality should be measured as explained higher: if colostrum contains 50 g IgG/L, feeding 4 L of colostrum suffices to provide 200 g IgG to the calf. There is no added benefit in feeding 4 L of colostrum compared to 3 L when colostrum of comparable quality is fed once using an oesophageal tube [77]. Larger IgG intakes are required by calves being fed >2 hours after birth [36]. Hand feeding colostrum >4 hours after birth is a risk factor for FPT [38]. A slight delay in the increase of serum Ig concentration was obvious in calves receiving 4 L of colostrum by oesophageal intubation compared with bottle-fed calves receiving 2 L. However, calves receiving colostrum by oesophageal intubation reached significantly higher Ig concentrations compared with bottle-fed calves [78]. Because of the difference in administered volume (4 L versus 2 L) between the two methods (oesophageal intubation versus bottle-fed), it can only be concluded from this study that an appropriate use of an oesophageal tube to feed 4 L colostrum is a safe and reliable method for an adequate passive immune transfer in healthy newborn calves. This study does not prove colostrum fed by

oesophageal intubation to be better than bottle-feeding colostrum. Further research is needed to reveal if the rapid passage of Ig from the reticulorumen to the abomasum also occurs in weak born calves. Moreover, tube feeding can cause moderate depression or a more difficult adaptation to feeding with a nipple bucket [78].

Calves cared for by female workers are less likely to develop FPT [37], probably because women are more patient with newborn calves that refuse to suckle. The odds of FPT are higher for calves experiencing dystocia [32] which can be explained by drinking too little colostrum and/or a lower apparent efficiency of absorption.

Treatment

Fluid therapy

Diarrhoea leads to dehydration, acidosis, electrolyte imbalance, and hypoglycemia, all of which should be addressed by a well-executed fluid therapy management [31,79,80].

Metabolic acidosis in diarrhoeic calves arises not only from fecal bicarbonate loss and hyper-L-lactatemia, but mainly from a hyper-D-lactatemia. Hyper-L-lactatemia is a result of dehydration and decreased tissue perfusion. Hyper-D-lactatemia is caused by absorption of D-lactic acid produced in the gastro-intestinal tract by fermentation of malabsorbed carbohydrates [81,82]. Variations in behaviour, posture and palpebral reflex are more closely correlated with elevations of serum D-lactate concentrations than with decreases in base excess (BE) [79,83,84]. However, BE values can be estimated based on posture, behaviour, and palpebral reflex (Table 1) with calves aged ≥ 7 days having lower BE values than younger calves [31,84,85]. Impairment of a good sucking reflex is more related to the degree of dehydration than it reflects acidosis [79,83,86]. There is even no obvious correlation between the serum levels of D-lactate and dehydration [82]. Remarkably, ability to stand was maintained in quite some calves despite BE values below -20 mmol/l, demonstrating the risk of undercorrection when based on this ability [85].

High potassium levels are more closely correlated to dehydration than to parameters indicative of metabolic acidosis [87,88]. Therefore, K^+ levels should be decreased to normal by correction of the acidosis and hypoglycaemia but more importantly by improving tissue perfusion.

Oral rehydration therapy Administration of an oral rehydration solution (ORS) should be commenced as soon as diarrhoea starts and should be continued as long as the calf has diarrhoea.

The osmolality of an ORS is determined primarily by the concentrations of sodium and glucose. Calves fed hyperosmotic ORS solutions (600–717 mOsm/l) (HORS) have a slower abomasal emptying rate compared with

Table 1 Guide to assessing the degree of acidosis based on posture, behaviour and palpebral reflex of the calf (adapted from [85])

	Category	BE (mmol/l)
Posture	Calf standing up by itself	-2.2
	Calf standing up after encouragement	0.0
	Standing steadily after lifting	2.8
	Standing unsteadily, able to correct position if forced	-11.7
	Standing unsteadily, unable to correct position if forced	-20.6
	Unable to stand, sternal recumbency	-20.9
Behaviour	Unable to stand, lateral recumbency	-25.4
	Adequate reaction to acoustic and optical stimuli, very bright and alert	2.5
	Adequate reaction to acoustic and optical stimuli	1.8
	Delayed reaction to acoustic and optical stimuli	-14.6
	Calf reacts only to painful stimuli	-21.0
	No reaction to painful stimuli	-25.4
Palpebral reflex	Eyelids are closed immediately and fully	0.7
	Eyelids are closed immediately but not fully	-7.6
	Eyelids are closed with delay and not fully	-19.9
	Eyelids are not closed at all	-24.3

calves fed iso-osmotic ORS solutions (300–360 mOsm/l) (IORS). This slower emptying rate increases the risk for bloat or abomasitis and produces a slower rate of plasma volume expansion [80,89,90]. Unless previously assumed, the volume present in the reticulorumen of healthy calves after intubation appears to be predominantly due to reflux from the abomasum, rather than spillage from incomplete closure of the oesophageal groove. The calculated change in abomasal volume of intubated calves approximated that one of calves that suckled their ORS [80]. In contrast, the delivery of HORS to the small intestine is slower after intubation indicating a different effect of intubation on coordinating motility between the reticulorumen, abomasum and duodenum than does suckling [80]. These effects taken together, suckling an IORS provides the fastest rate of solution delivery to the small intestine and a slightly faster rate of plasma volume expansion than does suckling or oesophageal intubation of a HORS. However, suckling or oesophageal intubation of a HORS provides the most appropriate oral solution for treating hypoglycaemic calves, because the HORS produces a larger and more sustained increase in plasma glucose concentration [80].

Alkalinizing agents commonly used in commercial ORS are bicarbonate and bicarbonate precursors, mainly

acetate, propionate, citrate and phosphate. The higher assumed alkalinisation of ORS products containing bicarbonate (BORS) or citrate (CORS) when compared to solutions containing other bicarbonate precursors is confirmed by most recent studies in absolute numbers, but never significantly different [90-93]. High SID-fluids (≥ 79 mmol/l) are recommended to treat acidosis because the SID value of an ORS determines the degree of abomasal and serum alkalinisation [91,94]. Prolongation of the *in vivo* clotting time is more likely to occur when larger volumes of an ORS in cow's milk are fed, because luminal pH will initially be greater and remain greater for a longer period of time [90,95]. From these studies it can be carefully concluded that feeding a BORS or CORS does not affect milk clotting *in vivo* and disagrees with general recommendations to not feed BORS or CORS concurrently with or short before/after cow's milk to calves. The SID value and fed amount of an ORS seem to play a more important role, both in correcting dehydration and metabolic acidosis [91,94].

It should be noted that most of the research discussed above was executed in a small number of healthy, non-diarrhoeic calves. Milk clotting, abomasal luminal pH and abomasal emptying rates could differ between diarrhoeic and non-diarrhoeic calves.

Intravenous fluid therapy Intravenous (IV) fluid therapy should be implemented if the calf is severely dehydrated (>8%), depressed, has a weakened/absent suckle reflex or suffers from a dilated abomasum and/or intestinal hypomotility. The fluids should always be warmed e.g. by adding a coil to the IV fluid line and placing this in a bucket of hot water so that the calf does not need to use extra energy to bring the given fluid to body temperature. To sustain improved clinical status, an IV fluid therapy needs to be followed by continued ORS therapy [31,96]. Both the jugular as the auricular veins can be catheterized for IV rehydration, but by using the ear vein, larger quantities of fluid can be given during a longer period of time in an on farm setting.

Bicarbonate requirements for IV correction of acidosis are still being calculated as follows: bicarbonate (g) = body weight (kg) \times BE (mmol/l) \times 0.6 (l/kg) \times 0.084 (g/mmol) [86,96]. When using this formula, metabolic acidosis was not corrected in more than half of the calves and the risk of failure to correct acidosis increased with D-lactate concentrations [86]. Calves with distinct changes in posture and demeanour thus seem to need higher doses of bicarbonate than calculated with the factor of 0.6 in the formula mentioned. However, in this study no follow-up therapy with ORS was offered, which could explain calves becoming acidotic again after 24 h. Other studies reporting treatment failures had a more severe metabolic acidosis before treatment compared with

successfully treated calves [31,96]. The formula mentioned above should only be used for estimation of buffer required for correction of incurred losses and overdosing seems more desirable than underdosing [86]. Bicarbonate can be given IV as an isotonic (IBS) or hypertonic solution (HBS). In the field it is more practical to use a HBS compared with an IBS. Rapid IV administration of an 8.4% bicarbonate formulation at 5–10 ml/kg provided an effective and safe method to improve acid–base abnormalities [31,96-98]. There were no direct indications in these studies of potential adverse effects related to electrolyte concentrations, oxygen-haemoglobin dissociation curve, hypercapnia or paradoxical intracellular acidosis, as previously assumed. It can be concluded from these studies that HBS can be safely used if the speed of an IV administration does not exceed 1.25 ml/kg/min in calves not suffering from respiratory problems.

Hypoglycaemia is best addressed by giving 150 mg glucose/kg. Larger quantities of glucose should be avoided because of the risk of glycosuria. Calves that are cachectic and reluctant to drink can also be given a constant glucose infusion using the ear vein approach as discussed earlier.

Other important treatment measures

Routine use of AB in diarrhoeic calves cannot be recommended due to increased levels of antibiotic resistance. However, systemically ill calves (depression, anorexia, fever) often suffer from *E. coli* septicaemia and thus parenteral Gram-negative-spectrum AB are advised to treat these calves. Antimicrobial susceptibility testing methods should be performed on the herd level at a regular basis [99,100]. As mentioned, halofuginone lactate is the only registered product for prevention and treatment of *C. parvum* in Europe. In 2009 a meta-analysis for the effects of therapeutic halofuginone treatment was conducted [21]. Their results were considered as uninterpretable because of a lack of sufficient data. Azithromycin (1,500 mg/d, 7 days, per oral (PO) and lasalocid (8 mg/kg, sid, 3 days, PO) reduce oocyst shedding [101,102]. Therapeutic use of nitazoxanide (15 mg/kg, bid, PO, 10 days) did not improve the clinical appearance, nor the intensity of oocyst excretion [103]. In contrast, calves of the therapeutic group showed a longer diarrhoeic episode compared to the untreated control group. Antiviral drugs to treat rota- and coronavirus are not commercially available. Non-steroidal anti-inflammatory drugs (NSAID) like meloxicam (0.5 mg/kg, one-shot, SC) improve appetite and growth rate, probably because NCD is accompanied by malaise and gastrointestinal discomfort [39]. Despite their widespread use, there is no valid recent research available to recommend products reducing intestinal motility (e.g. hyoscine N-butylbromide).

The literature is contradictory or inconclusive concerning other ancillary treatments such as phytopharmaceuticals

and probiotics [104,105]. For example, the number of days having diarrhoea, severity of diarrhoea and mortality rates were similar between an oral treatment with neomycin or dried oregano leaves [105]. However, a negative control group receiving no treatment was not included and they also defined calves older than 4 days with diarrhoea to suffer from colibacillosis. *Lactobacillus rhamnosus* GG did not affect the outcome of therapy in diarrhoeic calves [104]. The herd veterinarian is advised to consult results of peer-reviewed controlled trials for separate products.

Overall, the herd veterinarian should focus on the prevention of NCD.

Conclusions

Neonatal calf diarrhoea is a complex disease with a high morbidity and mortality in dairy cattle herds. Colostrum management is the most important preventive measure but is often neglected. Also, in the therapy management a good fluid therapy is indispensable. However, mistakes are often made. The herd veterinarian should be up-to-date when it comes to prevention and therapy of NCD. He or she is the most obvious person to communicate these items to the dairy farmer.

Abbreviations

BORS: Bicarbonate containing oral rehydration solution; cfu: Colony forming units; *C. parvum*: *Cryptosporidium parvum*; CORS: Citrate containing oral rehydration solution; *E. coli*: *Escherichia coli*; FPT: Failure of passive transfer; HBS: Hypertonic bicarbonate solution for intravenous use; IBS: Isotonic bicarbonate solution for intravenous use; IgG: Immunoglobulin G; IV: Intravenous; NCD: Neonatal calf diarrhoea; ORS: Oral rehydration solution; PO: Per oral; SID: Strong ion difference.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

VM performed the literature review and drafted the manuscript. GH and GO contributed to the final manuscript. All authors have read and approved the final version of the manuscript.

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References

1. Windeyer MC, Leslie KE, Godden SM, Hodgins DC, Lissemore KD, LeBlanc SJ: Factors associated with morbidity, mortality, and growth of dairy heifer calves up to 3 months of age. *Prev Vet Med* 2014, **113**:231–240.
2. Bartels CJM, Holzhauer M, Jorritsma R, Swart WAJM, Lam TJGM: Prevalence, prediction and risk factors of enteropathogens in normal and non-normal faeces of young Dutch dairy calves. *Prev Vet Med* 2010, **93**:162–169.
3. Svensson C, Linder A, Olsson SO: Mortality in Swedish dairy calves and replacement heifers. *J Dairy Sci* 2006, **89**:4769–4777.
4. Gulliksen SM, Jor E, Lie KI, Hamnes IS, Løken T, Åkerstedt J, Østerås O: Enteropathogens and risk factors for diarrhea in Norwegian dairy calves. *J Dairy Sci* 2009, **92**:5057–5066.
5. Santin M, Trout JM, Xiao L, Zhou L, Greiner E, Fayer R: Prevalence and age-related variation of *Cryptosporidium* species and genotypes in dairy calves. *Vet Parasitol* 2004, **122**:103–117.
6. Trotz-Williams LA, Martin SW, Leslie KE, Duffield T, Nydam DV, Peregrine AS: Calf-level risk factors for neonatal diarrhea and shedding of *Cryptosporidium parvum* in Ontario dairy calves. *Prev Vet Med* 2007, **82**:12–28.
7. Silverlås C, de Verdier K, Emanuelson U, Mattsson JG, Björkman C: *Cryptosporidium* infection in herds with and without calf diarrhoeal problems. *Parasitol Res* 2010, **107**:1435–1444.
8. Geurden T, Claerebout E, Verducruysse J, Berkvens D: A Bayesian evaluation of four immunological assays for the diagnosis of clinical cryptosporidiosis in calves. *Vet J* 2008, **176**:400–402.
9. Izzo MM, Kirkland PD, Mohler VL, Perkins NR, Gunn AA, House JK: Prevalence of major enteric pathogens in Australian dairy calves with diarrhoea. *Austral Vet J* 2011, **89**:167–173.
10. Ok M, Güler L, Turgut K, Ok Ü, Sen I, Gündüz IK, Birdane MF, Güzelbektes H: The studies on the aetiology of diarrhoea in neonatal calves and determination of virulence gene markers of *Escherichia coli* strains by multiplex PCR. *Zoonoses Public Health* 2009, **56**:94–101.
11. Trotz-Williams LA, Jarvie BD, Martin SW, Leslie KE, Peregrine AS: Prevalence of *Cryptosporidium parvum* infection in south-western Ontario and its association with diarrhea in neonatal dairy calves. *Can Vet J* 2005, **46**:349–351.
12. Uhde FL, Kaufmann T, Sager H, Albini S, Zanonni R, Schelling E, Meylan M: Prevalence of four enteropathogens in the faeces of young diarrhoeic dairy calves in Switzerland. *Vet Rec* 2008, **163**:362–366.
13. Wieler LH, Sobjinski G, Schlapp T, Failing K, Weiss R, Menge C, Baljer G: Longitudinal prevalence study of diarrheagenic *Escherichia coli* in dairy calves. *Berl Munch Tierarztl Wochenschr* 2007, **120**:296–306.
14. Rodriguez-Palacios A, Stämpfli HR, Stalker M, Duffield T, Weese JS: Natural and experimental infection of neonatal calves with *Clostridium difficile*. *Vet Microbiol* 2007, **124**:166–172.
15. Berge ACB, Moore DA, Sischo WM: Prevalence and antimicrobial resistance patterns of *Salmonella enterica* in preweaned calves from dairies and calf ranches. *Am J Vet Res* 2006, **67**:1580–1588.
16. Kirisawa R, Takeyama A, Koiwa M, Iwai H: Detection of bovine torovirus in fecal specimens of calves with diarrhea in Japan. *J Vet Med Sci* 2007, **69**:471–476.
17. Berge ACB, Moore DA, Besser TE, Sischo WM: Targeting therapy to minimize antimicrobial use in preweaned calves: Effects on health, growth, and treatment costs. *J Dairy Sci* 2009, **92**:4707–4714.
18. De Waele V, Speybroeck N, Berkvens D, Mulcahy G, Murphy TM: Control of cryptosporidiosis in neonatal calves: Use of halofuginone lactate in two different calf rearing systems. *Prev Vet Med* 2010, **96**:143–151.
19. Trotz-Williams LA, Martin SW, Leslie KE, Duffield T, Nydam DV, Peregrine AS: Association between management practices and within-herd prevalence of *Cryptosporidium parvum* shedding on dairy farms in southern Ontario. *Prev Vet Med* 2008, **83**:11–23.
20. Maddox-Hyttel C, Langkjaer RB, Enemark HL, Vigre H: *Cryptosporidium* and *Giardia* in different age groups of Danish cattle and pigs-occurrence and management associated risk factors. *Vet Parasitol* 2006, **141**:48–59.
21. Silverlås C, Björkman C, Egenvall A: Systematic review and meta-analyses of the effects of halofuginone against calf cryptosporidiosis. *Prev Vet Med* 2009, **91**:73–84.
22. Santin M, Trout JM, Fayer R: A longitudinal study of cryptosporidiosis in dairy cattle from birth to 2 years of age. *Vet Parasitol* 2008, **155**:15–23.
23. Kváč M, Kouba M, Vítovec J: Age-related and housing-dependence of *Cryptosporidium* infection of calves from dairy and beef herds in South Bohemia, Czech Republic. *Vet Parasitol* 2006, **137**:202–209.
24. Jarvie BD, Trotz-Williams LA, McKnight DR, Leslie KE, Wallace MM, Todd CG, Sharpe PH, Peregrine AS: Effect of halofuginone lactate on the occurrence of *Cryptosporidium parvum* and growth of neonatal dairy calves. *J Dairy Sci* 2005, **88**:1801–1806.
25. Trotz-Williams LA, Jarvie BD, Peregrine AS, Duffield TF, Leslie KE: Efficacy of halofuginone lactate in the prevention of cryptosporidiosis in dairy calves. *Vet Rec* 2011, **168**:509.
26. Klein P: Preventive and therapeutic efficacy of halofuginone-lactate against *Cryptosporidium parvum* in spontaneously infected calves: A centralised, randomised, double-blind, placebo-controlled study. *Vet J* 2008, **177**:429–431.
27. Lallemond M, Villeneuve A, Belda J, Dubreuil P: Field study of the efficacy of halofuginone and decoquinat in the treatment of cryptosporidiosis in veal calves. *Vet Rec* 2006, **159**:672–677.
28. Galvão KN, Santos JEP, Coscioni A, Villaseñor M, Sischo WM, Berge ACB: Effect of feeding live yeast products to calves with failure of passive transfer on performance and patterns of antibiotic resistance in fecal *Escherichia coli*. *Reprod Nutr Dev* 2005, **45**:427–440.

29. Sadeghi AA, Shawrang P: **Effects of natural zeolite clinoptilolite on passive immunity and diarrhea in newborn Holstein calves.** *Livest Sci* 2008, **113**:307–310.
30. Von Buenau R, Jaekel L, Schubotz E, Schwarz S, Stroff T, Krueger M: ***Escherichia coli* strain Nissle 1917: significant reduction of neonatal calf diarrhea.** *J Dairy Sci* 2005, **88**:317–323.
31. Koch A, Kaske M: **Clinical efficacy of intravenous hypertonic saline solution or hypertonic bicarbonate solution in the treatment of inappetent calves with neonatal diarrhea.** *J Vet Intern Med* 2008, **22**:202–211.
32. Furman-Fratczak K, Rzasa A, Stefaniak T: **The influence of colostral immunoglobulin concentration in heifer calves' serum on their health and growth.** *J Dairy Sci* 2011, **94**:5536–5543.
33. Chand N, Pandey NN, Mondal DB: **Effect of timing and frequency of colostrum feeding on immunoglobulin G status and susceptibility to colibacillic diarrhoea in neonatal calves.** *Indian J of Anim Sci* 2009, **79**:653–657.
34. Chigerwe M, Tyler JW, Summers MK, Middleton JR, Schultz LG, Nagy DW: **Evaluation of factors affecting serum IgG concentrations in bottle-fed calves.** *J Am Vet Med Assoc* 2009, **234**:785–789.
35. Güngör Ö, Bastan A, Erbil MK: **The usefulness of the γ -glutamyltransferase activity and total proteinemia in serum for detection of the failure of immune passive transfer in neonatal calves.** *Rev Med Vet* 2004, **155**:27–30.
36. Chigerwe M, Tyler JW, Schultz L, Middleton JR, Stevens BJ, Spain JN: **Effect of colostrum administration by use of oesophageal intubation on serum IgG concentrations in Holstein bull calves.** *Am J Vet Res* 2008, **69**:1158–1163.
37. Trotz-Williams LA, Leslie KE, Peregrine AS: **Passive immunity in ontario dairy calves and investigation of its association with calf management practices.** *J Dairy Sci* 2008, **91**:3840–3849.
38. Beam AL, Lombard JE, Kopral CA, Garber LP, Winter AL, Hicks JA, Schlater JL: **Prevalence of failure of passive transfer of immunity in newborn heifer calves and associated management practices on US dairy operations.** *J Dairy Sci* 2009, **92**:3973–3980.
39. Todd CG, Millman ST, McKnight DR, Duffield TF, Leslie KE: **Nonsteroidal anti-inflammatory drug therapy for neonatal calf diarrhea complex: Effects on calf performance.** *J Anim Sci* 2010, **88**:2019–2028.
40. Kehoe SI, Jayarao BM, Heinrichs AJ: **A survey of bovine colostrum composition and colostrum management practices on pennsylvania dairy farms.** *J Dairy Sci* 2007, **90**:4108–4116.
41. Donovan DC, Reber AJ, Gabbard JD, Aceves-Avila M, Galland KL, Holbert KA, Ely LO, Hurley DJ: **Effect of maternal cells transferred with colostrum on cellular responses to pathogen antigens in neonatal calves.** *Am J Vet Res* 2007, **68**:778–782.
42. Reber AJ, Hippen AR, Hurley DJ: **Effects of the ingestion of whole colostrum or cell-free colostrum on the capacity of leukocytes in newborn calves to stimulate or respond in one-way mixed leukocyte cultures.** *Am J Vet Res* 2005, **66**:1854–1860.
43. Reber AJ, Donovan DC, Gabbard J, Galland K, Aceves-Avila M, Holbert KA, Marshall L, Hurley DJ: **Transfer of maternal colostral leukocytes promotes development of the neonatal immune system I. Effects on monocyte lineage cells.** *Vet Imm Immunopathol* 2008, **123**:186–196.
44. Reber AJ, Donovan DC, Gabbard J, Galland K, Aceves-Avila M, Holbert KA, Marshall L, Hurley DJ: **Transfer of maternal colostral leukocytes promotes development of the neonatal immune system Part II. Effects on neonatal lymphocytes.** *Vet Imm Immunopathol* 2008, **123**:305–313.
45. Stielor A, Bernardo BS, Donovan GA: **Neutrophil and monocyte function in neonatal dairy calves fed fresh or frozen colostrum.** *J Appl Res Vet Med* 2012, **10**:328–334.
46. Godden SM, Smolenski DJ, Donahue M, Oakes JM, Bey R, Wells S, Sreevatsan S, Stabel J, Fetrow J: **Heat-treated colostrum and reduced morbidity in preweaned dairy calves: Results of a randomized trial and examination of mechanisms of effectiveness.** *J Dairy Sci* 2012, **95**:4029–4040.
47. Gulliksen SM, Lie KI, Sølverød L, Østerås O: **Risk factors associated with colostrum quality in Norwegian dairy cows.** *J Dairy Sci* 2008, **91**:704–712.
48. Morrill KM, Conrad E, Lago A, Campbell J, Quigley JD, Tyler H: **Nationwide evaluation of quality and composition of colostrum on dairy farms in the United States.** *J Dairy Sci* 2012, **95**:3997–4005.
49. Swan H, Godden S, Bey R, Wells S, Fetrow J, Chester-Jones H: **Passive transfer of immunoglobulin G and preweaning health in Holstein calves fed a commercial colostrum replacer.** *J Dairy Sci* 2007, **90**:3857–3866.
50. Chigerwe M, Tyler JW, Middleton JR, Spain JN, Dill JS, Stevens BJ: **Comparison of four methods to assess colostral IgG concentration in dairy cows.** *J Am Vet Med Assoc* 2008, **233**:761–766.
51. Biemann V, Gillan J, Perkins NR, Skidmore AL, Godden S, Leslie KE: **An evaluation of Brix refractometry instruments for measurement of colostrum quality in dairy cattle.** *J Dairy Sci* 2010, **93**:3713–3721.
52. Quigley JD, Lago A, Chapman C, Erickson P, Polo J: **Evaluation of the Brix refractometer to estimate immunoglobulin G concentration in bovine colostrum.** *J Dairy Sci* 2013, **96**:1148–1155.
53. Chigerwe M, Dawes ME, Tyler JW, Middleton JR, Moore MP, Nagy DM: **Evaluation of a cow-side immunoassay kit for assessing IgG concentration in colostrum.** *J Am Vet Med Assoc* 2005, **227**:129–131.
54. Godden SM, Hazel A: **Relationship between milking fraction and immunoglobulin G concentration in first milking colostrum from holstein cows.** *Bovine Pr* 2011, **45**:64–69.
55. Morin DE, Nelson SV, Reid ED, Nagy DW, Dahl GE, Constable PD: **Effect of colostrum volume, interval between calving and first milking, and photoperiod on colostral IgG concentrations in dairy cows.** *J Am Vet Med Assoc* 2010, **237**:420.
56. Moore M, Tyler JW, Chigerwe M, Dawes ME, Middleton JR: **Effect of delayed colostrum collection on colostral IgG concentration in dairy cows.** *J Am Vet Med Assoc* 2005, **226**:1375–1377.
57. Rastani RR, Grummer RR, Bertics SJ, Gümen A, Wiltbank MC, Mashek DG, Schwab MC: **Reducing dry period length to simplify feeding transition cows: milk production, energy balance, and metabolic profiles.** *J Dairy Sci* 2005, **88**:1004–1014.
58. Fidler AP, Fidler ML, Alley GW, Smith: **Short communication: Serum immunoglobulin G and total protein concentrations in dairy calves fed a colostrum-replacement product.** *J Dairy Sci* 2011, **94**:3609–3612.
59. Godden SM, Haines DM, Hagman D: **Improving passive transfer of immunoglobulins in calves. I: Dose effect of feeding a commercial colostrum replacer.** *J Dairy Sci* 2009, **92**:1750–1757.
60. Poulsen K, Foley AJ, Collins MT, McGuirk SM: **Comparison of passive transfer of immunity in neonatal dairy calves fed colostrum or bovine serum-based colostrum replacement and colostrum supplement products.** *J Am Vet Med Assoc* 2010, **237**:949–954.
61. Priestley D, Bittar JH, Ibarbia L, Risco CA, Galvão KN: **Effect of feeding maternal colostrum or plasma-derived or colostrum-derived colostrum replacer on passive transfer of immunity, health, and performance of preweaning heifer calves.** *J Dairy Sci* 2013, **96**:3247–3256.
62. Foster DM, Smith GW, Sanner TR, Busso GV: **Serum IgG and total protein concentrations in dairy calves fed two colostrum replacement products.** *J Am Vet Med Assoc* 2006, **229**:1282–1285.
63. Jones CM, James RE, Quigley JD, McGilliard ML: **Influence of pooled colostrum or colostrum replacement on IgG and evaluation of animal plasma in milk replacer.** *J Dairy Sci* 2004, **87**:1806–1814.
64. Smith GW, Foster DM: **Short Communication: Absorption of protein and immunoglobulin G in calves fed a colostrum replacer.** *J Dairy Sci* 2007, **90**:2905–2908.
65. Aly SS, Pithua P, Champagne JD, Haines DM: **A randomized controlled trial on preweaning morbidity, growth and mortality in Holstein heifers fed a lacteal-derived colostrum replacer or pooled maternal colostrum.** *BMC Vet Res* 2013, **9**:168.
66. Berge ACB, Besser TE, Moore DA, Sischo WM: **Evaluation of the effects of oral colostrum supplementation during the first fourteen days on the health and performance of preweaned calves.** *J Dairy Sci* 2009, **92**:286–295.
67. Stewart S, Godden S, Bey R, Rapnicki P, Fetrow J, Farnsworth R, Scanlon M, Arnold Y, Clow L, Mueller K, Ferrouillet C: **Preventing bacterial contamination and proliferation during the harvest, storage, and feeding of fresh bovine colostrum.** *J Dairy Sci* 2005, **88**:2571.
68. Donahue M, Godden SM, Bey R, Wells S, Oakes JM, Sreevatsan S, Stabel J, Fetrow J: **Heat treatment of colostrum on commercial dairy farms decreases colostrum microbial counts while maintaining colostrum immunoglobulin G concentrations.** *J Dairy Sci* 2012, **95**:2697–2702.
69. Elizondo-Salazar JA, Jayarao BM, Heinrichs AJ: **Effect of heat treatment of bovine colostrum on bacterial counts, viscosity, and immunoglobulin G concentration.** *J Dairy Sci* 2010, **93**:961–967.
70. Godden S, McMartin S, Feirtag J, Stabel J, Bey R, Goyal S, Metzger L, Fetrow J, Wells S, Chester-Jones H: **Heat treatment of bovine colostrum. II: Effects of heating duration on pathogen viability and immunoglobulin G.** *J Dairy Sci* 2006, **89**:3476–3483.
71. McMartin S, Godden S, Metzger L, Feirtag J, Bey R, Stabel J, Goyal S, Fetrow J, Wells S, Chester-Jones H: **Heat treatment of bovine colostrum. I: Effects of**

- temperature on viscosity and immunoglobulin G level. *J Dairy Sci* 2006, **89**:2110–2118.
72. Johnson JL, Godden SM, Molitor T, Ames T, Hagman D: Effects of feeding heat-treated colostrum on passive transfer of immune and nutritional parameters in neonatal dairy calves. *J Dairy Sci* 2007, **90**:5189–5198.
73. Elizondo-Salazar JA, Heinrichs AJ: Feeding heat-treated colostrum to neonatal dairy heifers: Effects on growth characteristics and blood parameters. *J Dairy Sci* 2009, **92**:3265–3273.
74. Gelsinger SL, Gray SM, Jones CM, Heinrichs AJ: Heat treatment of colostrum increases immunoglobulin G absorption efficiency in high-, medium-, and low-quality colostrum. *J Dairy Sci* 2014, **97**:2355–2360.
75. Medina-Cruz M, Cruz C, Montaldo HH: Serum protein levels in holstein calves fed pasteurized-frozen-thawed or unpasteurized first-milk colostrum. *Bovine Pr* 2008, **42**:201–205.
76. Urday K, Chigerwe M, Tyler JW: Voluntary colostrum intake in Holstein heifer calves. *Bovine Pr* 2008, **42**:198–200.
77. Russell Sakai R, Coons DM, Chigerwe M: Effect of single oroesophageal feeding of 3 L versus 4 L of colostrum on absorption of colostrum IgG in Holstein bull calves. *Livest Sci* 2012, **148**:296.
78. Kaske M, Werner A, Schubert H-J, Rehage J, Kehler W: Colostrum management in calves: effects of drenching vs. bottle feeding. *J Anim Phys Anim Nutr* 2005, **89**:151–157.
79. Lorenz I: Investigations on the influence of serum D-lactate levels on clinical signs in calves with metabolic acidosis. *Vet J* 2004, **168**:323–327.
80. Nouri M, Constable PD: Comparison of two oral electrolyte solutions and route of administration on the abomasal emptying rate of Holstein-Friesian calves. *J Vet Int Med* 2006, **20**:620–626.
81. Ewaschuk JB, Naylor JM, Palmer R, Whiting SJ, Zello GA: D-lactate production and excretion in diarrheic calves. *J Vet Int Med* 2004, **18**:744–747.
82. Lorenz I: Influence of D-lactate on metabolic acidosis and on prognosis in neonatal calves with diarrhoea. *J Vet Med* 2004, **51**:425–428.
83. Lorenz I, Gentile A, Klee W: Investigations of D-lactate metabolism and the clinical signs of D-lactataemia in calves. *Vet Rec* 2005, **156**:412–415.
84. Trefz FM, Lorch A, Feist M, Sauter-Louis C, Lorenz I: Construction and validation of a decision tree for treating metabolic acidosis in calves with neonatal diarrhea. *Vet Res* 2012, **8**:238–255.
85. Trefz FM, Lorch A, Feist M, Sauter-Louis C, Lorenz I: Metabolic acidosis in neonatal calf diarrhea—Clinical findings and theoretical assessment of a simple treatment protocol. *J Vet Int Med* 2012, **26**:162–170.
86. Lorenz I, Vogt S: Investigations on the association of D-lactate blood concentrations with the outcome of therapy of acidosis, and with posture and demeanour in young calves with diarrhoea. *J Vet Med* 2006, **53**:490–494.
87. Trefz FM, Lorch A, Feist M, Sauter-Louis C, Lorenz I: The prevalence and clinical relevance of hyperkalemia in calves with neonatal diarrhoea. *Vet J* 2013, **195**:350–356.
88. Trefz FM, Constable PD, Sauter-Louis C, Lorch A, Knubben-Schweizer G, Lorenz I: Hyperkalemia in neonatal diarrheic calves depends on the degree of dehydration and the cause of the metabolic acidosis but does not require the presence of acidemia. *J Dairy Sci* 2013, **96**:7234–7244.
89. Sen I, Constable PD, Marshall TS: Effect of suckling isotonic or hypertonic solutions of sodium bicarbonate or glucose on abomasal emptying rate in calves. *Am J Vet Res* 2006, **67**:1377–1384.
90. Constable PD, Grünberg W, Carstensen L: Comparative effects of two oral rehydration solutions on milk clotting, abomasal luminal pH, and abomasal emptying rate in suckling calves. *J Dairy Sci* 2009, **92**:296–312.
91. Bachmann L, Homeier T, Arlt S, Brueckner M, Rawel H, Deiner C, Hartmann H: Influence of different oral rehydration solutions on abomasal conditions and the acid–base status of suckling calves. *J Dairy Sci* 2009, **92**:1649–1659.
92. Reinhold S, Hertsch B-W, Höppner S, Heuwieser W, Hartmann H: Effect of milk and oral electrolyte solutions with and without bicarbonate on abomasal luminal pH and systemic acid–base status in calves. *Tierärztl Prax* 2006, **34**:368–376.
93. Marshall TS, Constable PD, Crochik SS, Wittek T, Freeman DE, Morin DE: Effect of suckling an isotonic solution of sodium acetate, sodium bicarbonate, or sodium chloride on abomasal emptying rate and luminal pH in calves. *Am J Vet Res* 2008, **69**:824–831.
94. Constable PD, Stämpfli HR, Navetat H, Berchtold J, Schelcher F: Use of a quantitative strong ion approach to determine the mechanism for acid–base abnormalities in sick calves with or without diarrhea. *J Vet Int Med* 2005, **19**:581–589.
95. Constable PD, Ahmed AF, Misk NA: Effect of suckling cow's milk or milk replacer on abomasal luminal pH in dairy calves. *J Vet Int Med* 2005, **19**:97–102.
96. Coskun A, Sen I, Guzelbektes H, Ok M, Turgut K, Canikli S: Comparison of the effects of intravenous administration of isotonic and hypertonic sodium bicarbonate solutions on venous acid–base status in dehydrated calves with strong ion acidosis. *J Am Vet Med Assoc* 2010, **236**:1098–1103.
97. Berchtold JF, Constable PD, Smith GW, Mathur SM, Morin DE, Tranquilli WJ: Effects of intravenous hyperosmotic sodium bicarbonate on arterial and cerebrospinal fluid acid–base status and cardiovascular function in calves with experimentally induced respiratory and strong ion acidosis. *J Vet Int Med* 2005, **19**:240–251.
98. Bleul UT, Schwantag SC, Kähn WK: Effects of hypertonic sodium bicarbonate solution on electrolyte concentrations and enzyme activities in newborn calves with respiratory and metabolic acidosis. *Am J Vet Res* 2007, **68**:850–857.
99. Berge ACB, Moore DA, Sischo WM: Field trial evaluating the influence of prophylactic and therapeutic antimicrobial administration on antimicrobial resistance of fecal *Escherichia coli* in dairy calves. *Appl Environ Microbiol* 2006, **72**:3872–3878.
100. Morley PS, Apley MD, Besser TE, Burney DP, Fedorka-Cray PJ, Papich MG, Traub-Dargatz JL, Weese JS: Antimicrobial drug use in veterinary medicine. *J Vet Int Med* 2005, **19**:617–629.
101. Elitok B, Elitok OM, Pulat H: Efficacy of azithromycin dihydrate in treatment of Cryptosporidiosis in naturally infected dairy calves. *J Vet Int Med* 2005, **19**:590–593.
102. Sahal M, Karaer Z, Yasa Duru S, Cizmeci S, Tanyel B: Cryptosporidiosis in newborn calves in Ankara region: clinical, haematological findings and treatment with lasalocid-NA. *Deut Tierärztl Woch* 2005, **112**:203–208.
103. Schnyder M, Kohler L, Hemphill A, Deplazes P: Prophylactic and therapeutic efficacy of nitazoxanide against *Cryptosporidium parvum* in experimentally challenged neonatal calves. *Vet Parasitol* 2009, **160**:149–154.
104. Ewaschuk JB, Zello GA, Naylor JM: *Lactobacillus GG* does not affect D-lactic acidosis in diarrheic calves, in a clinical setting. *J Vet Int Med* 2006, **20**:614–619.
105. Bampidis VA, Christodoulou V, Florou-Paneri P, Christaki E: Effect of dried oregano leaves versus neomycin in treating newborn calves with colibacillosis. *J Vet Med* 2006, **53**:154–156.

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