

## Update on PIN Polar Targeting

# PIN Polar Targeting<sup>1</sup>

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Cell polarity is one of the fundamental properties of multicellular organisms and is tightly linked with processes such as cell division, differentiation, cellular signaling, and intercellular communication. Polarities of individual cells, transmitted by cell divisions, are reflected at the tissue and organ levels and contribute to the overall shape of the multicellular organism. In addition, in plants, cell polarity provides means by which they maintain developmental continuity and adapt their development to optimally conform to environmental conditions by flexible redefinition of cell polarities. Thus, the mechanisms underlying the establishment and maintenance of cell polarity belong to the important themes of developmental and cell biology.

At the level of the individual cell, polarity is mirrored by the asymmetric distribution of intracellular components, such as organelles, cytoskeletal strands, and single proteins. This asymmetric distribution of intracellular components defines functionally and/or morphologically distinct domains (Bonifacino and Lippincott-Schwartz, 2003). Mechanisms that underlie the sorted delivery of intracellular cargos to these domains and thus contribute to the generation or maintenance of cell polarity have been extensively studied in different model organisms, including mammals, flies, worms, and yeast (Knoblich, 2000; Irazoqui and Lew, 2004; Nance, 2005). Animal epithelial cells are a preferred model system, because their plasma membrane (PM) is divided into two distinct domains, the apical domain facing the lumen and the basolateral domain (Mostov et al., 2003; Janssens and Chavrier, 2004). Selective recruitment of apical and basolateral cargos is achieved by their targeted delivery to these domains and results mainly from three processes: (1) newly synthesized proteins are sorted on their way to the PM (mainly in the trans-Golgi network) into vesicles that discriminately deliver them to the apical or basolateral surface; (2) other proteins are selectively

retained at the PM polar domain; and (3) proteins that are not retained are rapidly endocytosed and either recycled back or, alternatively, delivered to a different polar PM domain by a process called transcytosis (Rodriguez-Boulán et al., 2005).

Comparable knowledge on the cellular mechanisms underlying the polar localization of proteins in plant cells is lacking, but conceptually similar modes of polar delivery can be assumed. Plant and animal cells might differ fundamentally in the manner by which the polar-competent cargos are kept in their polar domains. In animal cells, anchored protein complexes, called tight junctions, form a physical barrier and limit lateral diffusion of proteins between adjacent polar PM domains (Brown and Stow, 1996). So far, no indications for analogical structures in plant cells have been found; therefore, it remains unclear how lateral diffusion of polar cargos is limited in the plant PM. Furthermore, despite significant advances in recent years, the subcellular trafficking pathways in plants are still only rudimentarily sketched; many destinations of secretory and endocytic pathways as well as their interconnections are vaguely defined, and findings are contradictory. These deficiencies in the basic cell biology knowledge of plants restrict our understanding on where and how the main decision on sorting and delivery of polar cargos occurs. The availability of necessary tools and technologies along with enough attention paid to these topics guarantee a rapid advancement in coming years.

## POLARITY IN PLANTS AND AUXIN

For the first time, the term polarity was applied to plants by Hermann von Vöchting in 1878, when he showed that pieces of willow (*Salix* species) stems form roots and shoots at the corresponding ends irrespective of the orientation toward gravity (for review, see Mohr and Schopfer, 1995). Plant polarity is determined at the cellular level and is tightly connected to the polarity of tissues and organs. Better than other multicellular systems, plants can redefine their cellular and tissue polarity based on the influence of many factors. The plant hormone auxin (indole-3-acetic acid) has been identified as an important factor mediating tissue and organ polarity in plants, mainly on account of its strictly directional (polar) flow through plant tissues. The existence of polar auxin transport together with its physiological and develop-

<sup>1</sup> This work was supported by the Odysseus Programme of the Research Foundation-Flanders (grant no. G091608) and the Young Investigator Programme of the European Molecular Biology Organization.

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The author responsible for distribution of materials integral to the findings presented in this article in accordance with the policy described in the Instructions for Authors ([www.plantphysiol.org](http://www.plantphysiol.org)) is: Jiří Friml ([jiri.friml@psb.ugent.be](mailto:jiri.friml@psb.ugent.be)).

[www.plantphysiol.org/cgi/doi/10.1104/pp.108.121756](http://www.plantphysiol.org/cgi/doi/10.1104/pp.108.121756)

mental roles, for example in the growth reorientation in response to environmental stimuli, has led to the hypothesis that a combination of environmental and endogenous factors regulate the auxin flow in different plant tissues and, thus, provide the vectorial information for defining the cellular behavior (Went, 1974; Friml, 2003). The important advancement in auxin biology was the formulation of the chemiosmotic model (Rubery and Sheldrake, 1974; Raven, 1975) that proposed the existence of PM-localized auxin carrier proteins that facilitate auxin uptake and auxin efflux out of the cells. The model also proposed that the asymmetric localization of the efflux carriers at one side of the transporting cells determines the direction of the intercellular auxin movement within the field of cells. This remarkable insight, which was later verified experimentally, connected polarities at the cellular and tissue levels.

### POLAR-COMPETENT PROTEINS IN PLANT CELLS

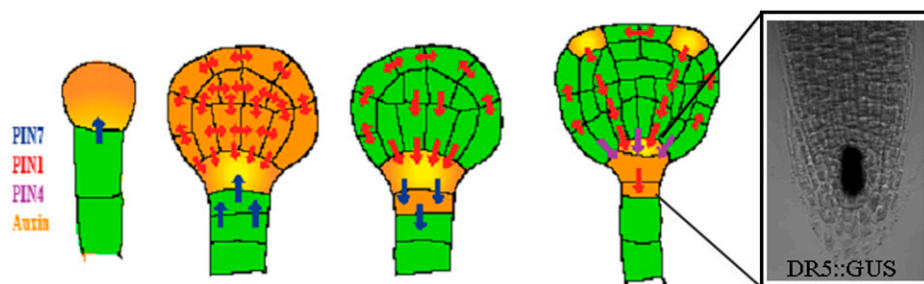
More than 20 years after the chemiosmotic model had been formulated, the hypothetical molecular components were identified. Molecular genetics and physiological studies in *Arabidopsis* (*Arabidopsis thaliana*) led to the discovery of genes coding for auxin influx and efflux carriers.

The PINFORMED (PIN) proteins have been identified and characterized as key regulators of a multitude of auxin-mediated developmental processes, including tropic growth (Chen et al., 1998; Luschnig et al., 1998; Müller et al., 1998; Utsuno et al., 1998; Friml et al., 2002b), axis formation in embryogenesis (Friml et al., 2003), postembryonic organogenesis (Benková et al., 2003; Reinhardt et al., 2003), root meristem maintenance (Friml et al., 2002a; Blilou et al., 2005), and vascular tissue differentiation and regeneration (Sauer et al., 2006; Scarpella et al., 2006). PIN proteins are PM proteins that act as auxin efflux carriers (Petrásek et al., 2006) and have mainly a polar localization that

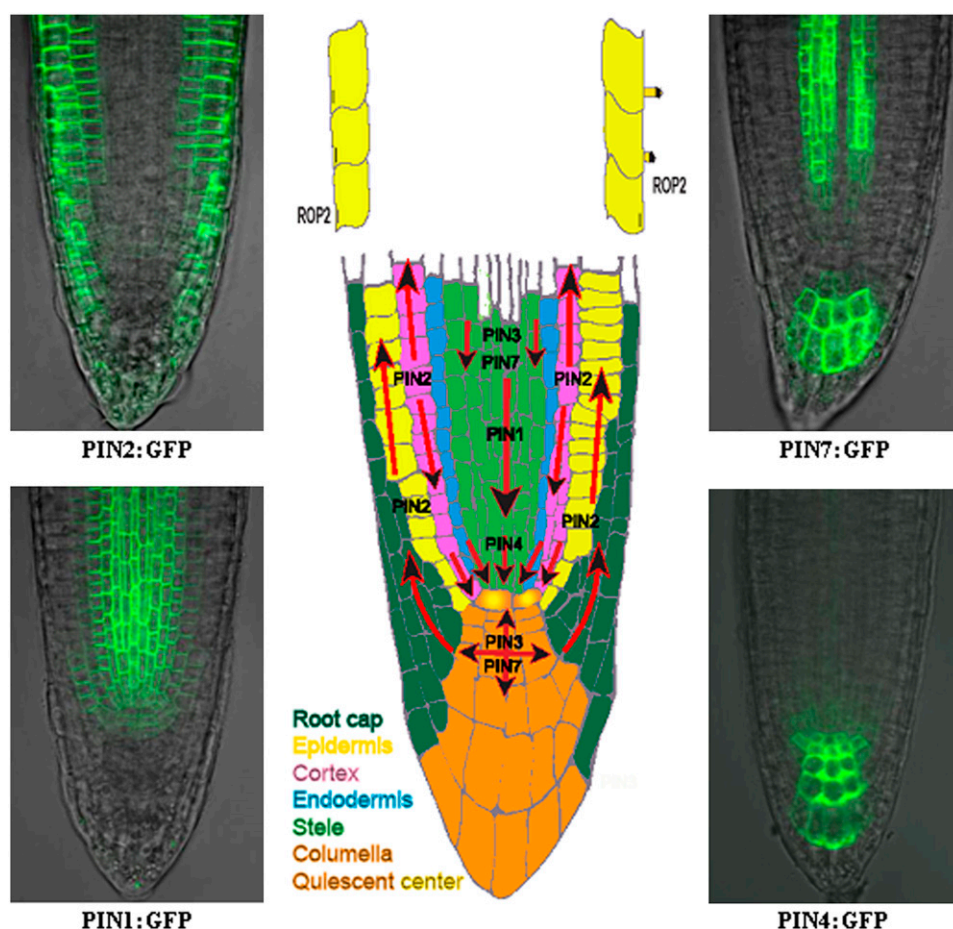
correlates with and is required for the direction of auxin flow (Wiśniewska et al., 2006). The Arabidopsis PIN family consists of eight members, most of which have been functionally characterized and found to be localized polarly at different sides of the various cell types (Vietsen et al., 2007). For example, during embryogenesis, PIN1, PIN4, and PIN7 show polar localizations and act together to specify the apical-basal axis of the embryo (Fig. 1). Postembryonically, PIN proteins have different PM localizations, most being localized at the basal (root apex-facing) side of the vasculature and stele cells, such as PIN1, PIN3, PIN4, and PIN7, whereas some localize also apically (shoot apex-facing side), such as PIN1 in the shoot apex epidermis or PIN2 in the lateral root cap and epidermis cells. In the shoot endodermis and root pericycle cells, PIN3 localizes also at the inner lateral side, whereas it has a symmetric localization in columella cells (Fig. 2). Thus, PIN proteins constitute prominent cell polarity markers in plants. Furthermore, polar targeting of PIN proteins has a clear developmental output, because the polarity of the PIN localization at the single-cell level determines the direction of intercellular auxin transport and the directional signaling to neighboring cells (Wiśniewska et al., 2006).

Besides PIN proteins, other polarly localized components are involved in auxin transport. The AUXIN RESISTANT1/LIKE AUX1 (AUX1/LAX) proteins are PM-localized auxin influx carriers (Bennett et al., 1996; Yang et al., 2006; Swarup et al., 2008). The AUX1 protein has a polar localization in some cells, such as the protophloem and the shoot apical meristem (Swarup et al., 2001; Reinhardt et al., 2003; Kleine-Vehn et al., 2006; Fig. 3). Other auxin transport proteins from the ATP-binding cassette multidrug resistance/P-glycoprotein family are localized mainly symmetrically, but polar localization in some cells has been reported as well (Geisler et al., 2005; Terasaka et al., 2005).

Other plant proteins, whose functions are not directly connected to auxin transport, also have been



**Figure 1.** Establishment of apical-basal polarity during *Arabidopsis* embryogenesis. At early stages, PIN7 localizes to the apical sides of suspensor cells mediating auxin flow into the proembryo, where PIN1 is localized first in a nonpolar manner. At the middle globular stage, PIN1 basal polarity in provascular cells is established, followed by PIN7 relocation to the basal side of suspensor cells. These PIN polarity rearrangements result in redirection of the auxin flow to the basal part of the embryo, where auxin accumulation contributes to root meristem specification. At later stages, PIN4 expression in the central root meristem aids the establishment of local auxin accumulation in the center of developing root meristems. Furthermore, PIN1 relocates at the apical surface of the proembryo to establish two symmetrically positioned auxin accumulation foci marking sites of future embryonic leaves.



**Figure 2.** Polar localization of PIN proteins in the Arabidopsis root tip. The directionality of auxin transport (arrows) is determined by the polar, subcellular localization of PIN proteins. PIN1 is localized at the basal (root apex-facing) side of the root vasculature; PIN2 at the basal side of the cortical cells and at the apical (shoot apex-facing) side of the epidermal and root cap cells; PIN3 in an apolar manner in the columella cells of the root; PIN4 at the basal side of cells in the central root meristem and with less pronounced polarity in the cells of the quiescent center; and PIN7 at the basal side of the stele cells and apolar in the columella cells. ROP2 is also asymmetrically localized, associated with the places of root hair formation. Examples of PIN1:GFP, PIN2:GFP, PIN4:GFP, and PIN7:GFP expression in the root are depicted.

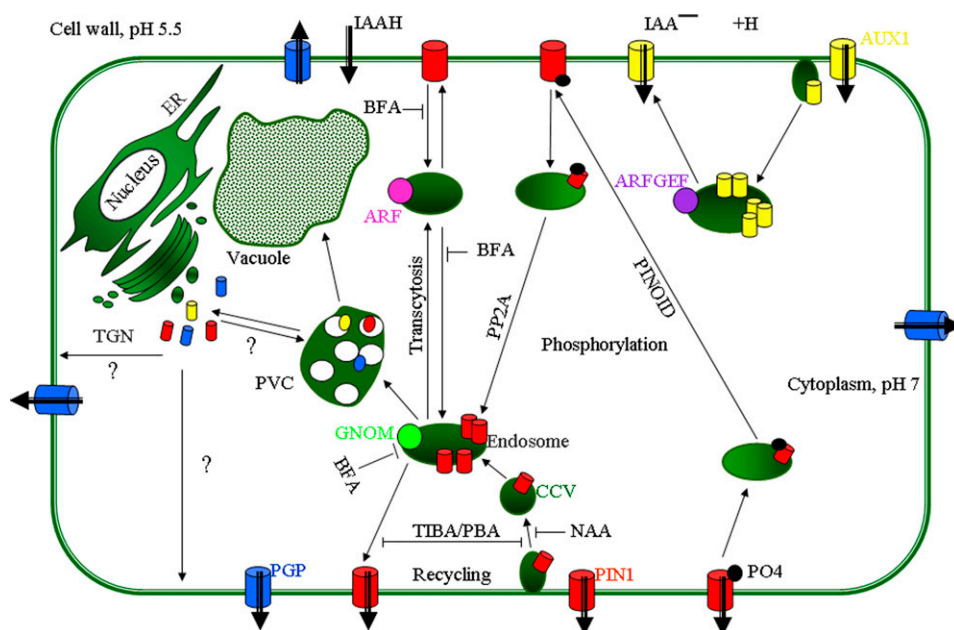
found to be localized asymmetrically in plant cells, namely at the inner lateral side, outer lateral side, and both longitudinal sides of the cells (Roudier et al., 2005; Ma et al., 2006, 2007; Miwa et al., 2007). Examples of laterally localized proteins include transporters for boron (BOR1 and BOR4) and for low silicon (LSI1 and LSI2) in rice (*Oryza sativa*) and the regulator of the anisotropic cell expansion (COBRA). A special asymmetric localization has been detected for a small G protein of the Rho family ROP2 that localizes to a distinct position close to the basal end of the outer membrane of elongating epidermis cells and marks the initiation position of the outgrowing root hair (Molendijk et al., 2001). This unprecedented diversity of polar cargos that localize to different sides of plant cells suggests that plants possess more diverse polar targeting machineries than are known from other eukaryotic systems.

#### ROLE OF PHOSPHORYLATION IN POLAR TARGETING

An important and yet unsolved question concerns how the different polar-competent proteins are recognized and delivered to the correct side of the cell. In

animal systems, polar cargo proteins carry signals that determine their residence at different polar domains (Dugani and Klip, 2005; Rodriguez-Boulán et al., 2005). Similar concepts of polarity determinants in the protein sequence also apply to plants, because different polar cargos, such as PIN1 or PIN2, localize to different polar domains of the same cell, necessitating some identification mechanism. Furthermore, by inserting a GFP tag at a specific position of the PIN1 sequence, the basal targeting of PIN1 is disrupted in epidermal cells, leading to apical localization (Wiśniewska et al., 2006). This observation implies a PIN1 sequence-based signal for decision on the PIN1 subcellular localization. Several findings suggest that the PIN polarity signals are related to the phosphorylation sites found in the PIN sequences. One of the major decision regulators on the PIN polarity are the Ser/Thr protein kinase PINOID (PID; Christensen et al., 2000; Benjamins et al., 2001; Friml et al., 2004) and the protein phosphatase 2A (PP2A; Michniewicz et al., 2007). High levels of PIN phosphorylation as achieved by overexpression of PID or inhibition of PP2A lead to a preferential apical PIN targeting, whereas low phosphorylation levels in the *pid* mutants result in a preferential basal PIN targeting (Friml et al., 2004; Treml et al., 2005). Importantly, PID has been shown to directly phosphorylate





**Figure 3.** Overview of the subcellular trafficking routes in a polarized plant cell. Auxin influx and efflux (black thick arrows) are mediated by AUX1 and PIN proteins, respectively. PGP ATP-binding cassette-type transporters are also involved in auxin efflux. According to the chemiosmotic model, auxin in the protonated form can also enter the cell passively. Basally localized PIN1 recycles in a BFA-sensitive and GNOM-dependent manner between endosomes and the PM. Constitutive cycling of vesicles between these two compartments is disrupted by 2,3,5-triiodobenzoic acid (TIBA) and 1-pyrenoylbenzoic acid (PBA), drugs that inhibit auxin transport and actin cytoskeleton dynamics (Dhonukshe et al., 2008). AUX1 also displays constitutive cycling to and from the apical PM that is dependent of an unknown BFA-insensitive ARF GEF. Auxin, such as  $\alpha$ -naphthaleneacetic acid (NAA), inhibits PIN internalization, increasing the amount and activity of PINs at the PM (Paciorek et al., 2005). PID kinase and PP2A phosphatase mediate reversible PIN phosphorylation, thus contributing to the decision on the apical-basal PIN targeting. Phosphorylated and dephosphorylated PIN proteins are preferentially sorted into the apical and basal targeting pathways, respectively (Michniewicz et al., 2007). PIN1 is able to translocate between the basal and apical cell sides by a combination of constitutive endocytic cycling and alternative recruitment by distinct ARF GEF-dependent apical and basal targeting machineries (Kleine-Vehn et al., 2008).

the hydrophilic loop of PIN proteins in vivo and in vitro and PP2A phosphatase has been shown to antagonize this action (Michniewicz et al., 2007). The available data are thus consistent with the model: when dephosphorylated, PIN is preferentially recruited by the basal targeting machinery, and phosphorylated PIN is trafficked by the apical pathway (Fig. 3). Such a model also has implications for the conditional regulation of PIN polarity and directional auxin fluxes by different signaling pathways that act upstream of the PID-dependent PIN phosphorylation. Other polar-competent proteins probably possess different types of also phosphorylation-unrelated polarity signals that have yet to be identified.

## ROLE OF STEROLS IN POLAR TARGETING

Plant sterols are essential components of plant membranes. Their chemical structure resembles that of animal cholesterol, whose cellular functions are supposed to be similar. Depletion of cholesterol, the main animal sterol, decreases the polar delivery of target proteins (Keller and Simons, 1998), while depletion of

plant sterols leads to cell polarity defects followed by reduced auxin transport and auxin-related developmental defects. Studies carried out on sterol-deficient *orc* and *cyclopropylsterol isomerase1* (*cpi1*) mutants have detected defects in the polar localization of PIN and AUX1 proteins, indicating that the polar delivery of cargos in plants also depends on the sterol composition of the PM (Souter et al., 2002; Grebe et al., 2003; Willemsen et al., 2003; Kleine-Vehn et al., 2006). Recent work has highlighted the essential role of sterols in the reiteration of PIN polarity after the division of polarized cells (Men et al., 2008). PIN proteins have been shown to be targeted to the forming cell plate during cell division (Geldner et al., 2001). This poses a problem that, after the fusion of the cell plate to the PM, PIN will be present at both apical and basal sides of one of the daughter cells. In order to maintain the polarity of the mother cell in both daughter cells, there must be a mechanism that stabilizes the polar cargo at one side and retrieves it from the opposite side of the newly formed cell wall. Little is known of how that is achieved and which cellular and molecular mechanisms are involved, but sterols seem to play a crucial role. The *cpi1* sterol-deficient mutants have impaired

endocytosis and show depositions of PIN2 at both apical and basal PMs in postcytokinetic cells (Men et al., 2008), suggesting that sterol-dependent endocytosis is required to retrieve PINs from the “wrong” side of the cell after cell division. In summary, these observations indicate that sterol-enriched PM microdomains related to the so-called lipid rafts (Martin et al., 2005) are important also in plant cells for different membrane-related trafficking and signaling processes, including the regulation of cell polarity.

## ROLE OF SECRETION AND RECYCLING IN POLAR TARGETING

Direct delivery of secreted proteins and other cargos to distinct polar domains at the PM is one of the basic possibilities for generating an asymmetric distribution at the cell surface. Indeed, in animal epithelial cells, different cargos are secreted directly to the apical and basolateral domains (Mostov et al., 2003). In plants, no data are available on such a mode of polar secretion. On the contrary, PIN proteins, as tested for the basally localized PIN1 or apically localized PIN2, seem to be delivered originally in a nonpolar fashion after the de novo synthesis, and their apical or basal polarity is then established in the next step involving internalization from the PM and polar recycling (Dhonukshe et al., 2008). Thus, the secretion, clathrin-dependent endocytosis (Dhonukshe et al., 2007), and subsequent recycling are important processes in the generation of the PIN polar localization.

The delivery of PIN proteins to the PM is sensitive to brefeldin A (BFA), a known inhibitor of secretion and subcellular trafficking (Steinmann et al., 1999; Geldner et al., 2001). A molecular target of the BFA action is GNOM, an endosomal exchange factor for ARF GTPases (ARF GEF). GNOM functions as the GDP/GTP exchange factor for the small G proteins of the ARF class that mediate vesicle budding processes at different subcellular compartments (Shevell et al., 1994; Geldner et al., 2001). In the presence of BFA, PIN1 largely disappears from the PM and can be found internalized in so-called BFA compartments. The PIN1 internalization is completely reversible and occurs also when de novo protein synthesis is inhibited, indicating constitutive endocytosis and recycling of PIN proteins (Geldner et al., 2001). The repeated cycles of PIN endocytosis and recycling to the PM were also visualized with a green-to-red photoconvertible fluorescent PIN2 version (Dhonukshe et al., 2007). These findings, together with the analysis of the *gnom* mutant (also called *emb30*), have shown that GNOM is a BFA-sensitive regulator of PIN trafficking from the endosomes back to the PM. GNOM seems to be more crucial for basal polar targeting, because the apical PM localization of PIN proteins and AUX1 is not strongly affected when GNOM function is inhibited (Kleine-Vehn et al., 2008). Thus, the apical cargos utilize a different targeting pathway that might require another, possibly BFA-insensitive, ARF GEF. These

observations and, in particular, the PIN polarity defects in the *gnom* loss-of-function mutants tightly link endocytic recycling and polar targeting in plant cells.

The GNOM-dependent, BFA-sensitive recycling pathway applies mainly to the polar PIN targeting in the interphase cells that depend on the actin cytoskeleton. On the contrary, in dividing cells, PIN proteins are delivered to the forming cell plate by the microtubule-dependent pathway (Geldner et al., 2001; Dhonukshe et al., 2006). These observations demonstrate the diversity of different trafficking pathways for the polar delivery of PIN proteins and other polar cargos.

## TRANSCYTOSIS MECHANISM FOR PIN POLARITY SWITCHES

A prominent mechanism for polar delivery of cargos in animal cells is transcytosis, which involves trafficking of polar cargos from one side of the cell to the other. In the animal epithelium, this process is crucial for the polar delivery of multiple cargos (Rodriguez-Boulan et al., 2005; Leibfried and Bellaïche, 2007).

In plants, the directional translocation of PIN1 protein from the basal to the apical PM can be induced by BFA treatment, as has been directly visualized with photoconvertible PIN2 versions. After BFA removal, the basal localization of PIN proteins is restored by translocation in the opposite direction from the apical to the basal cell side (Kleine-Vehn et al., 2008). These results demonstrate that apical and basal targeting pathways in plants are interconnected and can be used by PIN proteins to move between the apical and basal sides of cells. Thus, the transcytosis mechanism in plant cells is realized by a combination of constitutive endocytic recycling and alternative recruitment of cargos by distinct ARF GEF-dependent apical and basal targeting machineries. It remains unclear to what extent the transcytosis mechanism contributes to the establishment of polar PIN localization in different cells and developmental contexts; nonetheless, it is likely that the rapid PIN polarity switches, which can be observed during different developmental processes, are realized by the transcytosis mechanism.

## DEVELOPMENTAL AND ENVIRONMENTAL MODULATION OF PIN POLARITY

The connection between constitutive endocytic recycling and polar PIN localization via transcytosis provides a plausible mechanism for the quick changes in the polarity of the PIN proteins that are utilized to integrate various signals at the single-cell level and translate them into redirecting the intercellular auxin flow through the tissues, ultimately modifying the developmental program of the given tissue or organ.

The earliest known switches in PIN polarity occur during embryogenesis and contribute to the specification of the root pole and the initiation of root meristem

development (Fig. 1). At the early embryogenesis stages, PIN7 is found at the apical side of the suspensor cells, where it mediates the auxin flow toward the small apical cell to specify it as a proembryo. PIN1 is expressed at this stage in a nonpolar manner in the proembryo. At the defined moment of embryo development, PIN1 polarizes toward the basal side of pro-vascular cells and PIN7 changes its localization to the basal side of the suspensor cells. These PIN polarity rearrangements redirect the auxin flow toward the area of the future root meristem, where auxin accumulates and contributes to the specification of the root meristem (Friml et al., 2003; Weijers et al., 2005). Similarly, during postembryonic organogenesis, illustrated by the formation of lateral roots or leaves and flowers at the shoot apical meristem, PIN polarity undergoes rearrangements that are important for determining both the position of the future organ relative to the preexisting ones and the new growth axis of the organ primordium (Benková et al., 2003; Reinhardt et al., 2003; Heisler et al., 2005).

Other examples of PIN polarity changes are related to the canalization hypothesis, which assumes that auxin, by a positive feedback, can induce the capacity and polarity of its own transport within the field of cells that form the auxin channels preceding the flexible formation of the vasculature (Sachs, 1981). Indeed, predicted rearrangements of PIN polarity have been observed during leaf vasculature formation (Scarpella et al., 2006) and during vasculature regeneration after wounding (Sauer et al., 2006). The underlying mechanism is unknown, but feedback regulations of PIN polarity by auxin itself are the indispensable parts of models describing different auxin-dependent patterning processes, such as the phylotactic pattern of organ initiation (Barbier de Reuille et al., 2006; Jönsson et al., 2006; Smith et al., 2006).

Besides the changes in PIN polarity in response to internal signals, PIN polarity switches can occur also in response to environmental stimuli. After gravistimulation, PIN3 has been found to relocate to the lower side of columella cells, thus rerouting the auxin flow to the lower side of the root and triggering root bending (Friml et al., 2002b; Harrison and Masson, 2008). It is possible, although not demonstrated, that other environmental signals, such as light, trigger PIN polarity changes for mediating other developmental responses. These polarity modulations in response to various signals illustrate how the integration of signals at the level of subcellular dynamics of individual cells can be translated into the directional signaling at the tissue level and contribute to the unique plasticity of plant development.

## ACKNOWLEDGMENTS

We apologize to all authors whose important work was not cited due to space constraints. We are grateful to Martine De Cock for help in preparing the manuscript.

Received April 22, 2008; accepted June 24, 2008; published August 6, 2008.

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