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## Review Article

# The Bone Niche of Chondrosarcoma: A Sanctuary for Drug Resistance, Tumour Growth and also a Source of New Therapeutic Targets

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Chondrosarcomas are malignant cartilage-forming tumours representing around 20% of malignant primary tumours of bone and affect mainly adults in the third to sixth decade of life. Unfortunately, the molecular pathways controlling the genesis and the growth of chondrosarcoma cells are still not fully defined. It is well admitted that the invasion of bone by tumour cells affects the balance between early bone resorption and formation and induces an “inflammatory-like” environment which establishes a dialogue between tumour cells and their environment. The bone tumour microenvironment is then described as a sanctuary that contributes to the drug resistance patterns and may control at least in part the tumour growth. The concept of “niche” defined as a specialized microenvironment that can promote the emergence of tumour stem cells and provide all the required factors for their development recently emerges in the literature. The present paper aims to summarize the main evidence sustaining the existence of a specific bone niche in the pathogenesis of chondrosarcomas.

## 1. Introduction

Most chondrosarcomas (90%) are conventional chondrosarcomas which occur in the medullar cavity or at the bone surface. The fact that cartilaginous tumours are mainly observed in bones formed from endochondral ossification strengthens the relationship between the differentiation of normal chondrocytes and these neoplastic cells. Chondrosarcoma cells are cytologically and phenotypically related to the different chondrocyte subtypes observed in the growth plate, and all cell shapes can be observed in the tumour mass [1–4]. Thus, these similarities are in favour of a mesenchymal stem-cell origin for chondrosarcoma cells [1, 5]. The development of cancer cells in bone site responds to several biological mechanisms potentially applicable to numerous other entities. For

instance, invasion of bone by a primary or metastatic tumour cell affects the balance between early bone resorption and bone formation. This dysregulation of osteoblast-osteoclast coupling induces the release of factors initially trapped in the bone matrix, which in turn promote tumour cell proliferation [6]. Thus, the bone tumour microenvironment controls the tumour growth and is also described as a sanctuary that contributes to drug resistance patterns [7]. The specific and different bone sites in which the various sarcomas are able to grow reinforce the prominence of the tumour microenvironment. Chondrosarcomas are also characterized by their chemo- and radioresistance leading to a therapeutic surgical approach which remains the only available treatment with a 10-year survival between 30% and 80% depending on the grade [8, 9]. Currently, surgical excision is the main

treatment for all chondrosarcoma subtypes [10], and non-surgical treatments of their microenvironment are under investigation. In this context, a better understanding of the bone niche which interacts with chondrosarcoma is one of the future therapeutic options. The present paper aims to describe the bone niche of chondrosarcoma, its role in tumour growth and drug resistance, and its clinical interest as a therapeutic target.

## 2. The Bone Niche Is Composed of Heterogeneous Cell Types with Coupled Activities

In 2003, two research laboratories demonstrated that osteoblasts formed an osteoblastic niche to sustain hemopoiesis [11, 12]. Osteoblasts establish an “epithelial-like” tissue which physically interacts with hemopoietic stem cells and contributes to their maintenance in a quiescent stage through the interaction between Tie-2 and angiopoietin-1 [13]. Nilsson et al. showed that primitive hematopoietic cells resided close to the bone surface [14]. From these observations, the concept of bone niche has strongly evolved and has been applied to cancer stem cells [15]. Indeed, the “niche” is a functional microenvironment able to promote the emergence of cancer stem cells and to provide all factors required for their development. Naturally, this concept is well recognized in the context of hematologic malignancies such as multiple myeloma [16] or leukemia [17], and these diseases appear as a stem-cell disease with a hierarchy analogous to normal hematopoietic development. However, the bone niche is not limited to osteoblasts and during skeletal remodelling, numerous cell types (preosteoclasts, preosteoblasts, endothelial cells, macrophages, etc.) are closely located in the bone matrix and their functional coordination is a prerequisite to maintain the bone and the bone niche microarchitecture. Using three-dimensional visualizations, Andersen et al. clearly demonstrated the functional relevance of these cellular interactions in the bone niche [18]. In physiological conditions in which bone resorption and bone formation are coupled, the bone surface is always covered by canopy composed by flat cells expressing osteoblastic markers and associated with sinusoidal vessels [18]. Disruption of this canopy results in the dysregulation of the coupled bone-formation bone-resorption process and leads to a bone deficiency [18]. These very elegant observations revealed that the bone niche is composed of multiple cell entities. Macrophages also contribute to the bone niche as shown by Chang et al. [19]. Indeed, a discrete population of resident macrophages has been identified between bone lining cells within endosteum and periosteum. These osteal tissue macrophages are involved in bone dynamics by controlling osteoblast functions and, more specifically, are required for efficient osteoblast mineralization [19]. Into the bone niche, self-renewal and differentiation activity are clearly balanced as shown for hemopoietic stem cells [16, 17], and this balance is being controlled by the level of hypoxia, which modulates the interactions between tumour cells and the components of bone niche. The proliferation stage of stem

cells is predominant with increased levels of oxygen and hypoxia resulting in opposite effects [20, 21].

The concept of bone niche is also currently discussed for solid tumours and strengthens the very modern theory of “seed and soils” proposed by Paget in 1887 in which tumour cells (“seeds”) would colonize receptive foci (“soils”) [22]. This data is supported by the fact that specific molecules (e.g., cadherin and osteopontin) contribute to the stabilization of cancer cells in bone niches mimicking the cell interactions which take place during hemopoiesis [23, 24]. Such interactions have been identified in the premetastatic niche of breast carcinoma, where carcinoma cells grow avidly in bone which stores a variety of cytokines and growth factors and thus provide an extremely fertile environment for growing cells [25, 26]. The seed and soil theory can be also envisaged for the primary bone tumours. In a recent study, we reported an unexpected local osteosarcoma relapse which occurred at the exact site of autologous fat grafts in a patient who did not present any predictive factor of local recurrence [27]. Moreover, we showed that tumour growth was promoted by fat injection using a human osteosarcoma model induced in athymic nude mice. We then demonstrated that the mesenchymal stem cells isolated from adipose tissue induced exactly the same effect, probably reactivating quiescent tumour cells locally deposited into the bone tissue [27]. A recent study reinforces this theory by presenting 8 cases of osteosarcoma development several years after benign bone tumour treatment by curettage associated with bone graft. To explain the development of “de novo” sarcomas in these patients, an attraction mechanism of mesenchymal stem cells by the scaffold has been hypothesized [28]. Although mechanisms by which cancer stem cells could drive the tumour growth are still unknown, modulation of the microenvironment by mesenchymal stem cells may interfere with the biological behavior of this cell subpopulation. Similarly, inflammatory process associated with surgery may be also responsible for the reactivation of dormant tumour cells [29, 30]. Thus, a disturbance of the microenvironment and the bone niche modifies the proliferation/differentiation program of the tumour cells.

## 3. The Bone Niche of Chondrosarcoma

The key role of bone microenvironment in chondrosarcoma development has been suspected many years ago. Indeed, a rat intraosseous model simulating the progression of human chondrosarcoma has been set up to assess the interactions between bone environment and chondrosarcoma [31]. Transplantation of swarm rat chondrosarcoma within bone marrow or in close contact to the bone with induced periosteal lesions led to extensive bone remodelling with trabecular bone rarefaction and periosteal apposition associated with tumour growth. In contrast with these results, transplantation in close contact to the bone but without any periosteal lesion had no effect on bone, suggesting that bone healing factors interact with tumour development. The tumours which developed in intramedullary environment presented different foci with various gradings confirming

that bone environment is an important factor in the pathogenesis of chondrosarcoma [31]. Histological examination of conventional chondrosarcoma reveals the presence of numerous cells types in close contact to the cartilaginous tumour cells (Figures 1(a) and 2). The morphology of cartilaginous tumour cells depends on the grading of the tumour and associated cartilage-like tissue composed by tumour chondrocytes with heterogeneous shapes (Figures 1(b)–1(e)) and tumour cell types with mesenchymal aspect (Figure 1(e)). The tumour mass is characterized by lobular foci separated by vascularized soft tissue, which establishes a continuum with bone marrow or with the surrounding tissues (Figure 1). When chondrosarcoma develops in the medullary space (central or primary chondrosarcoma), the tumour cells induce the dysregulation of the balance between osteoblasts and osteoclasts, degrading the trabecular bone, perturbing the bone marrow environment. When chondrosarcoma develops from the bone surface (peripheral or secondary chondrosarcoma), tumour mass exhibits a similar lobular morphology associated with a periosteal reaction [31]. These peripheral chondrosarcoma develop on preexisting osteochondroma defined as the most common benign bone tumours and characterized by a cartilage-capped exophytic lesion that arises from the bone cortex. Nevertheless, the limit between osteochondroma and chondrosarcoma is still unclear, especially with low-grade chondrosarcoma that is closely related to osteochondroma. These tumours interact with periosteum mimicking the “bone niche”. Periosteum is a continuous membrane intimately linked covering the bone, well vascularized and containing osteoprogenitor cells including mesenchymal stem cells [32–34]. Thereby, peripheral and central chondrosarcoma can interact with the same kind of bone microenvironment. The permeation of tumour cells into the bone tissue is associated with the activation of bone resorption through the induction of osteoclast formation (Figures 1(f) and 2). In fact, the bone niche of chondrosarcoma includes all cell types described in the other neoplastic bone diseases. The narrow relationship between chondrosarcoma cells, soft tissue, vessels, and bone cells strengthens the relevance of a specific bone niche able to sustain tumour growth.

Can we suspect the existence of cancer stem cells in this bone niche which could be at the origin of chondrosarcoma and become quiescent in specific circumstances? Expression of SOX9 in human chondrosarcomas suggests that chondrosarcomas originate from a multipotent stem cell committed to differentiation along the chondrogenic pathway [35]. Moreover, the results of the cDNA array analyses emphasize the heterogeneous nature of chondrosarcoma. Using similar approaches, Boeuf et al. [36] proposed a new classification of chondrosarcoma in two clusters: a prechondrogenic phenotype with immature cells and a chondrogenic phenotype composed of more mature cells. Primary conventional central chondrosarcoma cells could be then grouped into two main clusters with distinctive marker expression signatures: one group clustering together with mesenchymal stem cells (CD49b-high/CD10-low/CD221-high) and a second group clustering close to fibroblasts (CD49b-low/CD10-high/CD221-low) [37]. These data strongly suggest the existence

of cancer stem cells possibly with mesenchymal stem cells or fibroblast markers. Although most of the literature on chondrosarcoma has confirmed that adequate surgery is the mainstay of treatment for local tumour control, which itself constitutes a risk factor for survival, an additional feature of chondrosarcoma is also the high level of local recidive even in case of adequate surgery [38–40]. This feature is also in favour of the existence of cancer stem cells in the bone marrow which may remain dormant until some yet unknown signals promote their growth or/and metastasis formation in bone tissue.

Hypoxia is a signal resulting in a large number of adaptive changes aimed at surviving in the hypoxic environment as well as correcting the oxygen deficit. Hypoxia inducing factor (HIF)-1 is a dimeric transcription factor composed of HIF-1 alpha and beta subunits. HIF-1 protein levels increase as a result of decreased degradation of the oxygen sensitive subunit HIF-1 $\alpha$ . HIF-1 modulates changes in gene expression during hypoxia. Although the angiogenesis compound of cartilage tumours is heterogeneous [41], hypoxia modulates the proliferation of chondrosarcoma cells similarly to the other solid tumour types and hemopoietic neoplasia. Thus, there is a significant relationship between the expression of HIF-1 $\alpha$ , the microvessel density and the proliferating cell nuclear antigen [42]. Several authors demonstrated that malignant chondrocytes increased HIF-1 $\alpha$  expression in an oxygen concentration-dependent manner and increased VEGF expression in response to hypoxia [43–46] which is closely related to the potential malignancy of chondrosarcoma [47, 48]. Hypoxia is also known to increase chemokine receptor expression such as CXCR4 in numerous cell types [49] and CXCR4/SDF1 also indirectly promotes the proliferation and migration of tumour cells and enhances tumour-associated angiogenesis [50]. CXCR4 expressed by tumour cells contributes to their migration into the premetastatic niche [51]. Interestingly, chondrosarcoma cell invasion is increased by hypoxia-induced expression of CXCR4 and MMP1, a process mediated by HIF1 $\alpha$  and ERK [52], and CXCL12, also called SDF-1, increases the invasiveness of chondrosarcoma cells [53]. Other chemokine/chemokine receptors couples are also involved in chondrosarcoma progression. Thus, the interaction of CCL5 (RANTES), a product of activated T cells present in bone environment during the tumour process with CCR5 expressed on the cell membrane enhances the migration of chondrosarcoma cells through the increase of MMP-3 production [54]. Overall, these data point out the similarities between the behaviour of chondrosarcoma cells and the invasion of leukaemia cells in the bone niche [51]. Osteopontin is also a typical example of these similarities. Indeed, osteopontin could mediate the anchoring of cancer cells in osteoblastic niches in a manner that mimics the mechanisms used by osteoblast to retain hematopoietic stem cells in these niches and to negatively regulate stem-cell pool size [55]. Osteopontin also influence the behaviour of carcinoma cells (proliferation, invasiveness, etc.) [56]. Similarly, osteopontin located in the bone matrix increases the migration and MMP expression in human chondrosarcoma and contributes to the pathogenesis of chondrosarcoma in its bone niche [57]. More



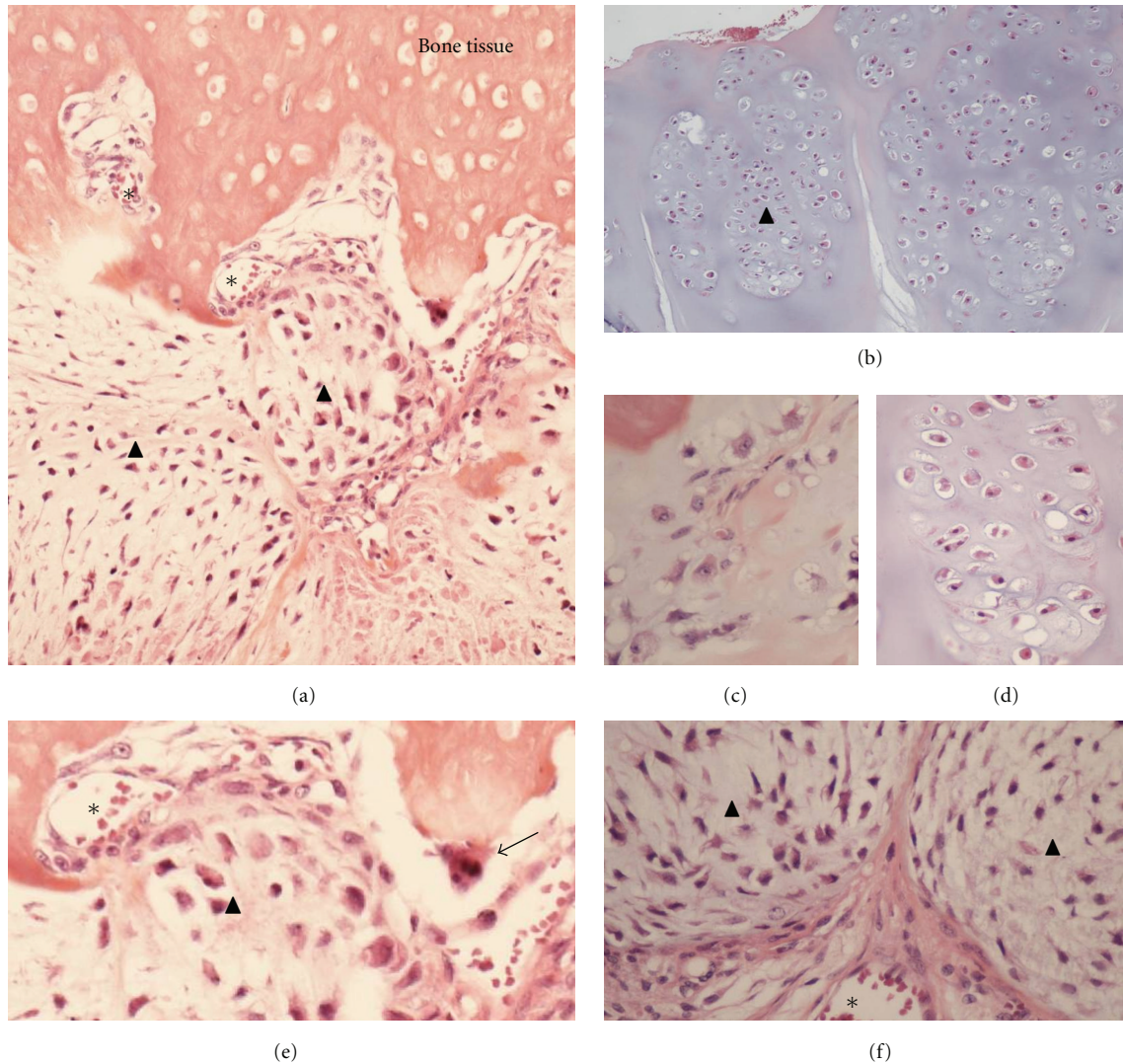


FIGURE 1: The bone niche of chondrosarcoma is composed by various cellular entities. Chondrosarcoma tissue shows heterogeneous cell morphology (a–d) with chondrocyte-like (b–d) and mesenchymal features (f). Chondrosarcoma bone niche is associated with several cell types including osteoclasts (e), endothelial cells vascularized soft tissue (f). HES staining, original magnification ( $\times 20$ , a and b;  $\times 40$ : c–e). Tumour cells: arrow head, asterix: blood vessels, and arrow: osteoclast.

recently, Vincourt et al. [58] demonstrated not only that the respective levels of C-propeptides of procollagens I and II in chondrogenic tumours but also that the interactions of chondrosarcoma cells with the surrounding extracellular matrix may modulate tumour progression, angiogenesis, and metastasis. C-propeptides of procollagen I favor angiogenesis and tumour progression, whereas C-propeptides of procollagen II exert antitumour and antiangiogenic properties through apoptosis induction when they are immobilized, and progression and metastasis when they are soluble [58]. Endostatin derived from collagen XVIII, a potent endogenous antiangiogenic factor that induces regression of various tumours of epithelial origin, prevents the chondrosarcoma growth via its potential activity on endothelial cells [59]. These results demonstrate that bone microenvironment and extracellular matrix establish a very complex bone niche adapted to the tumour progression.

The interactions between the extracellular matrix of bone niche and chondrosarcoma cells are tightly controlled by cytokines and growth factors produced by the environmental cells (osteoblasts, endothelial cells, macrophages, lymphocytes, etc.) and also by tumour cells themselves [60]. Proinflammatory cytokines are particularly associated with the pathogenesis of chondrosarcoma. Interleukin (IL)-1 regulates the expression of a disintegrin and metalloproteinase with thrombospondin motifs 1 (ADAMTS1) and VEGF by chondrosarcoma cells, then contributing to a strong positive impact of IL-1 on vascularization and tumour progression [61]. TNF- $\alpha$ , another proinflammatory cytokine, induced MMP-12 expression in chondrosarcoma cells when chondrocytes undergo malignant transformation [62] and increased also MMP-13 [63]. Members of TGF- $\beta$  superfamily play also a crucial role in migration and metastasis of human chondrosarcoma. For instance, TGF- $\beta$ 1 and BMP-2 increase

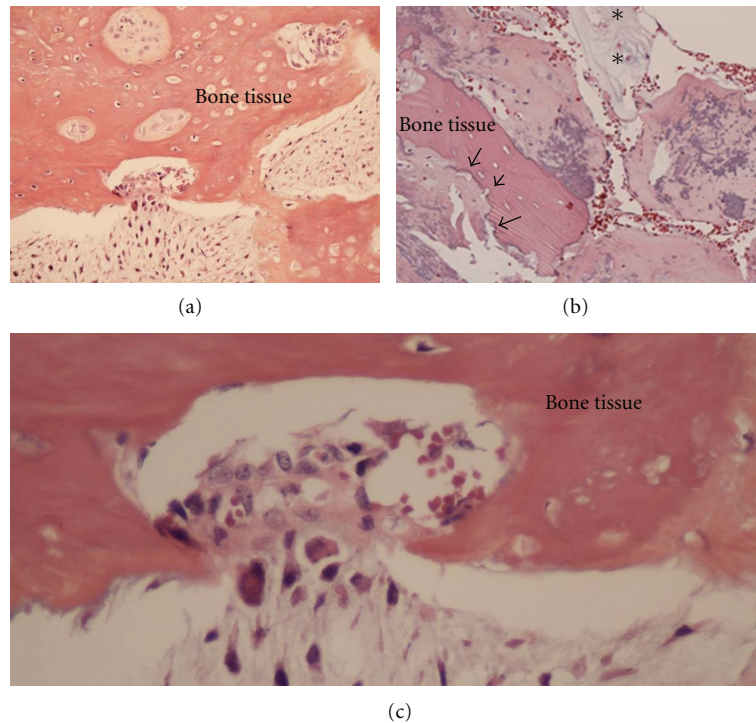


FIGURE 2: Chondrosarcoma growth is strongly linked to the bone tissue. Relationship between bone tissue and chondrosarcoma cells (a–c). Infiltration of chondrosarcoma cells into the bone tissue (permeation) (a–c). Chondrosarcoma development is associated with bone resorption foci (b). HES staining, original magnification ( $\times 20$ , a and b;  $\times 40$ : c). Arrow: bone resorption area, arrow head: necrosis of chondrosarcoma tissue, and \*: viable tumour component.

motility of human chondrosarcoma via the PI3K/Akt pathway [64, 65]. Oncostatin (OSM), a member of IL-6 cytokine family, induces a hypertrophic differentiation, with reduced SOX9 and induced Cbfa1, Coll10, MMP13, VEGF, and RANKL expression in chondrosarcoma cells. RANKL being a pro-osteoclastogenesis factor and then a proresorptive factor, OSM enhances osteoclast formation at the tumour/bone interface and reduces the ectopic bone neoformation [66].

#### 4. The Bone Niche: A Sanctuary for the Drug Resistance and a Source of New Therapeutic Targets

Although bone niche represents an adequate microenvironment for the survival/proliferation of cancer stem cells and has been identified as a major parameter regulating the metastatic process [67], recent studies also described the tumour microenvironment as a sanctuary contributing to the phenomenon of drug resistance [68]. The process of drug resistance has been shown to be mediated through (i) soluble factors such as cytokines or adhesion molecules constituting *de novo* drug resistances or (ii) acquired drug resistance linked to resistance mechanisms caused by selective pressure of chemotherapy or other therapeutic drugs [68]. Chondrosarcomas are poorly vascularized in correlation with resistance to systemic chemotherapy and exhibit poor metastatic potential. However, although this poor vascularization represents a first explanation for the drug

resistance, the bone niche also contributes to this resistance as observed for other tumour entities. In this context, a better definition of bone niche leads to the identification of relevant drug targets to improve the efficiency of the current treatment. This concept has been already validated in leukemia [69]. In sarcomas, similar approaches have been also envisaged [70]. Targeting of angiogenesis has been assessed in combination of chemotherapy and induced tumour necrosis [71]. Cyclooxygenase-2 (COX-2), a mediator of angiogenesis, is expressed in malignant cartilaginous tumours [72]. In chondrosarcoma, the use of celecoxib, a COX-2 inhibitor, first results in a decrease in tumour volume followed unfortunately by a relapsed tumour growth after 6 weeks [73]. Higher doses of COX-2 may be used, or a combinatory therapy based on this concept may be designed. HDAC4 represses VEGF expression and associated angiogenesis in chondrosarcoma [74]. Similarly, a therapeutic approach of chondrosarcoma based on HDAC inhibitor administration may be interesting [75, 76]. Bisphosphonates and rapamycin and its derivatives have been originally developed, respectively, as antiresorptive and antifungal agents [77, 78]. However, *in vitro* and *in vivo* experiments demonstrated that these compounds are multifunctional molecules exerting their effects not only on bone remodelling but also on tumour cell growth. mTOR targeting has been envisaged for numerous cancer types including malignant primary bone tumours [78–80], and a very impressive response of myxoid chondrosarcoma has been obtained



in combination with cyclophosphamide [81]. The main targets of bisphosphonates are bone-resorbing osteoclasts [82] which contribute to the hemopoietic and tumour bone niche [82]. Bisphosphonates also reduce the proliferation and invasion of chondrosarcoma [83, 84]. In preclinical model of chondrosarcoma, zoledronic acid slows down rat primary development and recurrent tumour progression after intralesional curettage and increases overall survival [85]. Thus, osteoclasts targeting may be used in prevention of chondrosarcoma recurrence. Cytokinetic treatment represents another relevant therapeutic approach of chondrosarcoma [2]. Oncostatin M, a member of the IL-6 cytokine family mainly produced by macrophages, neutrophils, and T lymphocytes, is a cytostatic factor for chondrosarcomas *in vitro* and *in vivo* [66]. This growth inhibitory effect is also observed with two other cytokines of the same family able to reduce chondrosarcoma expansion but with a lower efficiency: IL-6 in association with its soluble receptor and IL-27 [66]. This list is not exhaustive but gives some evidence of the interest to target or to modulate the bone niche components to improve chondrosarcoma treatment.

## 5. Conclusion

The treatment of chondrosarcoma is currently based on surgery, radiotherapy, and chemotherapy being occasionally used for metastatic tumours. However, a recent concept has emerged based on the key role played by the tumour micro-environment in the tumour invasiveness and in the drug-resistance phenomenon. This bone niche allows to identify new therapeutic targets for chondrosarcoma, and it appears clearly that a better understanding of the chondrosarcoma bone niche will open nonsurgical therapeutic options for chondrosarcoma which could also be combined with surgery.

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## References

- [1] S. Boeuf, P. Kunz, T. Hennig et al., "A chondrogenic gene expression signature in mesenchymal stem cells is a classifier of conventional central chondrosarcoma," *Journal of Pathology*, vol. 216, no. 2, pp. 158–166, 2008.
- [2] J. V. M. G. Bovée, A. M. Cleton-Jansen, A. H. M. Taminiau, and P. C. W. Hogendoorn, "Emerging pathways in the development of chondrosarcoma of bone and implications for targeted treatment," *Lancet Oncology*, vol. 6, no. 8, pp. 599–607, 2005.
- [3] T. Aigner, "Towards a new understanding and classification of chondrogenic neoplasias of the skeleton—biochemistry and cell biology of chondrosarcoma and its variants," *Virchows Archiv*, vol. 441, no. 3, pp. 219–230, 2002.
- [4] H. Gelderblom, P. C. W. Hogendoorn, S. D. Dijkstra et al., "The clinical approach towards chondrosarcoma," *Oncologist*, vol. 13, no. 3, pp. 320–329, 2008.
- [5] G. De Pinieux and C. Bouvier, "Recent advances in the biology of bone tumors and new diagnostic tools," in *Bone Cancer*, D. Heymann, Ed., chapter 19, pp. 225–234, Academic Press, New York, NY, USA, 2010.
- [6] Y. Wittrant, S. Théoleyre, C. Chipoy et al., "RANKL/RANK/OPG: new therapeutic targets in bone tumours and associated osteolysis," *Biochimica et Biophysica Acta*, vol. 1704, no. 2, pp. 49–57, 2004.
- [7] M. B. Meads, L. A. Hazlehurst, and W. S. Dalton, "The bone marrow microenvironment as a tumor sanctuary and contributor to drug resistance," *Clinical Cancer Research*, vol. 14, no. 9, pp. 2519–2526, 2008.
- [8] M. Campanacci, *Bone and Soft Tissue Tumors*, Springer, New York, NY, USA, 2nd edition, 1999.
- [9] F. Y. Lee, H. J. Mankin, G. Fondren et al., "Chondrosarcoma of bone: an assessment of outcome," *Journal of Bone and Joint Surgery. Series A*, vol. 81, no. 3, pp. 326–338, 1999.
- [10] E. L. Staals, E. Palmerini, S. Ferrari, and M. Mercuri, "Non-surgical treatment of chondrosarcoma: current concepts and future perspectives," in *Bone Cancer*, D. Heymann, Ed., chapter 31, pp. 375–383, Academic Press, New York, NY, USA, 2010.
- [11] J. Zhang, C. Niu, L. Ye et al., "Identification of the haematopoietic stem cell niche and control of the niche size," *Nature*, vol. 425, no. 6960, pp. 836–841, 2003.
- [12] L. M. Calvi, G. B. Adams, K. W. Weibrecht et al., "Osteoblastic cells regulate the haematopoietic stem cell niche," *Nature*, vol. 425, no. 6960, pp. 841–846, 2003.
- [13] F. Arai, A. Hirao, M. Ohmura et al., "Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche," *Cell*, vol. 118, no. 2, pp. 149–161, 2004.
- [14] S. K. Nilsson, H. M. Johnston, and J. A. Coverdale, "Spatial localization of transplanted hemopoietic stem cells: inferences for the localization of stem cell niches," *Blood*, vol. 97, no. 8, pp. 2293–2299, 2001.
- [15] H. Iwasaki and T. Suda, "Cancer stem cells and their niche," *Cancer Science*, vol. 100, no. 7, pp. 1166–1172, 2009.
- [16] G. W. Basak, A. S. Srivastava, R. Malhotra, and E. Carrier, "Multiple myeloma bone marrow niche," *Current Pharmaceutical Biotechnology*, vol. 10, no. 3, pp. 335–346, 2009.
- [17] S. W. Lane, D. T. Scadden, and D. G. Gilliland, "The leukemic stem cell niche: current concepts and therapeutic opportunities," *Blood*, vol. 114, no. 6, pp. 1150–1157, 2009.
- [18] T. L. Andersen, T. E. Sondergaard, K. E. Skorzynska et al., "A physical mechanism for coupling bone resorption and formation in adult human bone," *American Journal of Pathology*, vol. 174, no. 1, pp. 239–247, 2009.
- [19] M. K. Chang, L. J. Raggatt, K. A. Alexander et al., "Osteal tissue macrophages are intercalated throughout human and mouse bone lining tissues and regulate osteoblast function *in vitro* and *in vivo*," *Journal of Immunology*, vol. 181, no. 2, pp. 1232–1244, 2008.
- [20] K. Ito, A. Hirao, F. Arai et al., "Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells," *Nature Medicine*, vol. 12, no. 4, pp. 446–451, 2006.
- [21] K. Hosokawa, F. Arai, H. Yoshihara et al., "Function of oxidative stress in the regulation of hematopoietic stem cell-niche interaction," *Biochemical and Biophysical Research Communications*, vol. 363, no. 3, pp. 578–583, 2007.
- [22] S. Paget, "The distribution of secondary growths in cancer of the breast," *The Lancet*, vol. 133, no. 3421, pp. 571–573, 1889.
- [23] T. Yin and L. Li, "The stem cell niches in bone," *Journal of Clinical Investigation*, vol. 116, no. 5, pp. 1195–1201, 2006.



- [24] K. Hosokawa, F. Arai, H. Yoshihara et al., "Knockdown of N-cadherin suppresses the long-term engraftment of hematopoietic stem cells," *Blood*, vol. 116, no. 4, pp. 554–563, 2010.
- [25] P. Clézardin, "Early bone metastasis-associated molecular and cellular events," in *Bone Cancer*, D. Heymann, Ed., chapter 3, pp. 41–45, Academic Press, New York, NY, USA, 2010.
- [26] C. Coghin and G. I. Murray, "Current and emerging concepts in tumour metastasis," *Journal of Pathology*, vol. 222, no. 1, pp. 1–15, 2010.
- [27] P. Perrot, J. Rousseau, A. -L. Bouffaut et al., "Safety concern between autologous fat graft, mesenchymal stem cell and osteosarcoma recurrence," *PLoS One*, vol. 5, no. 6, Article ID e10999, 2010.
- [28] P. Picci, G. Sieberova, M. Alberghini et al., "Late sarcoma development after curettage and bone grafting of benign bone tumors," *European Journal of Radiology*, vol. 77, no. 1, pp. 19–25, 2011.
- [29] R. Demicheli, M. W. Retsky, W. J. M. Hrushesky, and M. Baum, "Tumor dormancy and surgery-driven interruption of dormancy in breast cancer: learning from failures," *Nature Clinical Practice Oncology*, vol. 4, no. 12, pp. 699–710, 2007.
- [30] R. Demicheli, M. W. Retsky, W. J. M. Hrushesky, M. Baum, and I. D. Gukas, "The effects of surgery on tumor growth: a century of investigations," *Annals of Oncology*, vol. 19, no. 11, pp. 1821–1828, 2008.
- [31] E. Grimaud, C. Damiens, A. V. Rousselle, N. Passuti, D. Heymann, and F. Gouin, "Bone remodelling and tumour grade modifications induced by interactions between bone and Swarm rat chondrosarcoma," *Histology and Histopathology*, vol. 17, no. 4, pp. 1103–1111, 2002.
- [32] M. Chanavaz, "The periosteum: the "umbilical cord" of bone. Quantification of the blood supply of cortical bone of periosteal origin," *Revue de Stomatologie et de Chirurgie Maxillo-Faciale*, vol. 96, no. 4, pp. 262–267, 1995.
- [33] C. A. Squier, S. Ghoneim, and C. R. Kremenak, "Ultrastructure of the periosteum from membrane bone," *Journal of Anatomy*, vol. 171, pp. 233–239, 1990.
- [34] X. Zhang, H. A. Awad, R. J. O'Keefe, R. E. Guldberg, and E. M. Schwarz, "A perspective: engineering periosteum for structural bone graft healing," *Clinical Orthopaedics and Related Research*, vol. 466, no. 8, pp. 1777–1787, 2008.
- [35] M. Söderström, T. Böhling, T. Ekfors, L. Nelimarkka, H. T. Aro, and E. Vuorio, "Molecular profiling of human chondrosarcomas for matrix production and cancer markers," *International Journal of Cancer*, vol. 100, no. 2, pp. 144–151, 2002.
- [36] S. Boeuf, P. Kunz, T. Hennig et al., "A chondrogenic gene expression signature in mesenchymal stem cells is a classifier of conventional central chondrosarcoma," *Journal of Pathology*, vol. 216, no. 2, pp. 158–166, 2008.
- [37] J. Diaz-Romero, S. Romeo, J. V. M. G. Bovée, P. C. W. Hogendoorn, P. F. Heini, and P. Mainil-Varlet, "Hierarchical clustering of flow cytometry data for the study of conventional central chondrosarcoma," *Journal of Cellular Physiology*, vol. 225, no. 2, pp. 601–611, 2010.
- [38] F. Fiorenza, A. Abudu, R. J. Grimer et al., "Risk factors for survival and local control in chondrosarcoma of bone," *Journal of Bone and Joint Surgery. Series B*, vol. 84, no. 1, pp. 93–99, 2002.
- [39] F. Y. Lee, H. J. Mankin, G. Fondren et al., "Chondrosarcoma of bone: An assessment of outcome," *Journal of Bone and Joint Surgery. Series A*, vol. 81, no. 3, pp. 326–338, 1999.
- [40] S. Gitelis, F. Bertoni, P. Picci, and M. Campanacci, "Chondrosarcoma of bone. The experience at the Istituto Ortopedico Rizzoli," *Journal of Bone and Joint Surgery. Series A*, vol. 63, no. 8, pp. 1248–1257, 1981.
- [41] T. Kalinski, S. Sel, I. Kouznetsova, M. Röpke, and A. Roessner, "Heterogeneity of angiogenesis and blood vessel maturation in cartilage tumors," *Pathology Research and Practice*, vol. 205, no. 5, pp. 339–345, 2009.
- [42] T. Kubo, T. Sugita, S. Shimose, T. Matsuo, K. Arihiro, and M. Ochi, "Expression of hypoxia-inducible factor-1 $\alpha$  and its relationship to tumour angiogenesis and cell proliferation in cartilage tumours," *Journal of Bone and Joint Surgery. Series B*, vol. 90, no. 3, pp. 364–370, 2008.
- [43] C. Lin, R. McGough, B. Aswad, J. A. Block, and R. Terek, "Hypoxia induces HIF-1 $\alpha$  and VEGF expression in chondrosarcoma cells and chondrocytes," *Journal of Orthopaedic Research*, vol. 22, no. 6, pp. 1175–1181, 2004.
- [44] G. Ayala, C. Liu, R. Nicosia, S. Horowitz, and R. Lackman, "Microvasculature and VEGF expression in cartilaginous tumors," *Human Pathology*, vol. 31, no. 3, pp. 341–346, 2000.
- [45] R. L. McGough, C. Lin, P. Meitner, B. I. Aswad, and R. M. Terek, "Angiogenic cytokines in cartilage tumors," *Clinical Orthopaedics and Related Research*, no. 397, pp. 62–69, 2002.
- [46] T. Furumatsu, K. Nishida, A. Kawai, M. Namba, H. Inoue, and Y. Ninomiya, "Human chondrosarcoma secretes vascular endothelial growth factor to induce tumor angiogenesis and stores basic fibroblast growth factor for regulation of its own growth," *International Journal of Cancer*, vol. 97, no. 3, pp. 313–322, 2002.
- [47] S. A. Nakagawa, A. Lopes, A. L. De Carvalho et al., "Nitric oxide synthases, cyclooxygenase-2, nitrotyrosine, and angiogenesis in chondrosarcoma and their relation to prognosis," *Journal of Bone and Joint Surgery. Series A*, vol. 92, no. 8, pp. 1738–1746, 2010.
- [48] S. Boeuf, J. V. M. G. Bovée, B. Lehner, P. C. W. Hogendoorn, and W. Richter, "Correlation of hypoxic signalling to histological grade and outcome in cartilage tumours," *Histopathology*, vol. 56, no. 5, pp. 641–651, 2010.
- [49] T. Schioppa, B. Uranchimeg, A. Saccani et al., "Regulation of the chemokine receptor CXCR4 by hypoxia," *Journal of Experimental Medicine*, vol. 198, no. 9, pp. 1391–1402, 2003.
- [50] J. Wang, J. Wang, Y. Sun et al., "Diverse signaling pathways through the SDF-1/CXCR4 chemokine axis in prostate cancer cell lines leads to altered patterns of cytokine secretion and angiogenesis," *Cellular Signalling*, vol. 17, no. 12, pp. 1578–1592, 2005.
- [51] R. N. Kaplan, R. D. Riba, S. Zacharoulis et al., "VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche," *Nature*, vol. 438, no. 7069, pp. 820–827, 2005.
- [52] X. Sun, L. Wei, Q. Chen, and R. M. Terek, "CXCR4/SDF1 mediate hypoxia induced chondrosarcoma cell invasion through ERK signaling and increased MMP1 expression," *Molecular Cancer*, vol. 9, article no. 17, 2010.
- [53] T. H. Lai, Y. C. Fong, W. M. Fu, R. S. Yang, and C. H. Tang, "Stromal cell-derived factor-1 increase  $\alpha\beta 3$  integrin expression and invasion in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 218, no. 2, pp. 334–342, 2009.
- [54] C. H. Tang, A. Yamamoto, Y. T. Lin, Y. C. Fong, and T. W. Tan, "Involvement of matrix metalloproteinase-3 in CCL5/CCR5 pathway of chondrosarcomas metastasis," *Biochemical Pharmacology*, vol. 79, no. 2, pp. 209–217, 2010.
- [55] S. Stier, Y. Ko, R. Forkert et al., "Osteopontin is a hematopoietic stem cell niche component that negatively regulates stem

- cell pool size," *Journal of Experimental Medicine*, vol. 201, no. 11, pp. 1781–1791, 2005.
- [56] J. C. Reichert, V. M. C. Quent, L. J. Burke, S. H. Stansfield, J. A. Clements, and D. W. Huttmacher, "Mineralized human primary osteoblast matrices as a model system to analyse interactions of prostate cancer cells with the bone microenvironment," *Biomaterials*, vol. 31, no. 31, pp. 7928–7936, 2010.
  - [57] Y. J. Chen, Y. Y. Wei, H. T. Chen et al., "Osteopontin increases migration and MMP-9 up-regulation via  $\alpha\beta 3$  integrin, FAK, ERK, and NF- $\kappa$ B-dependent pathway in human chondrosarcoma cells," *Journal of Cellular Physiology*, vol. 221, no. 1, pp. 98–108, 2009.
  - [58] J. B. Vincourt, S. Etienne, J. Cottet et al., "C-propeptides of procollagens Ia1 and II that differentially accumulate in enchondromas versus chondrosarcomas regulate tumor cell survival and migration," *Cancer Research*, vol. 70, no. 11, pp. 4739–4748, 2010.
  - [59] T. Furumatsu, N. Yamaguchi, K. Nishida et al., "Endostatin inhibits adhesion of endothelial cells to collagen I via  $\alpha\beta$  integrin, a possible cause of prevention of chondrosarcoma growth," *Journal of Biochemistry*, vol. 131, no. 4, pp. 619–626, 2002.
  - [60] P. Rutkowski, J. Kamińska, M. Kowalska, W. Ruka, and J. Steffen, "Cytokine and cytokine receptor serum levels in adult bone sarcoma patients: correlations with local tumor extent and prognosis," *Journal of Surgical Oncology*, vol. 84, no. 3, pp. 151–159, 2003.
  - [61] T. Kalinski, S. Krueger, S. Sel, K. Werner, M. Röpke, and A. Roessner, "ADAMTS1 is regulated by interleukin-1 $\beta$ , not by hypoxia, in chondrosarcoma," *Human Pathology*, vol. 38, no. 1, pp. 86–94, 2007.
  - [62] E. Kerkelä, T. Böhling, R. Herva, J. A. Uria, and U. Saarialho-Kere, "Human macrophage metalloelastase (MMP-12) expression is induced in chondrocytes during fetal development and malignant transformation," *Bone*, vol. 29, no. 5, pp. 487–493, 2001.
  - [63] S. W. Yoon, J. S. Chun, M. H. Sung, J. Y. Kim, and H. Poo, " $\alpha$ -MSH inhibits TNF- $\alpha$ -induced matrix metalloproteinase-13 expression by modulating p38 kinase and nuclear factor  $\kappa$ B signaling in human chondrosarcoma HTB-94 cells," *Osteoarthritis and Cartilage*, vol. 16, no. 1, pp. 115–124, 2008.
  - [64] Y. Y. Yeh, C. C. Chiao, W. Y. Kuo et al., "TGF- $\beta$ 1 increases motility and  $\alpha\beta 3$  integrin up-regulation via PI3K, Akt and NF- $\kappa$ B-dependent pathway in human chondrosarcoma cells," *Biochemical Pharmacology*, vol. 75, no. 6, pp. 1292–1301, 2008.
  - [65] Y. C. Fong, T. M. Li, C. M. Wu et al., "BMP-2 increases migration of human chondrosarcoma cells via PI3K/Akt pathway," *Journal of Cellular Physiology*, vol. 217, no. 3, pp. 846–855, 2008.
  - [66] E. David, P. Guihard, B. Brounais et al., "Direct anti-cancer effect of oncostatin M on chondrosarcoma," *International Journal of Cancer*, vol. 128, no. 8, pp. 1822–1835, 2011.
  - [67] J. A. Joyce and J. W. Pollard, "Microenvironmental regulation of metastasis," *Nature Reviews Cancer*, vol. 9, no. 4, pp. 239–252, 2009.
  - [68] M. B. Meads, L. A. Hazlehurst, and W. S. Dalton, "The bone marrow microenvironment as a tumor sanctuary and contributor to drug resistance," *Clinical Cancer Research*, vol. 14, no. 9, pp. 2519–2526, 2008.
  - [69] M. Konopleva, Y. Tabe, Z. Zeng, and M. Andreeff, "Therapeutic targeting of microenvironmental interactions in leukemia: mechanisms and approaches," *Drug Resistance Updates*, vol. 12, no. 4-5, pp. 103–113, 2009.
  - [70] K. Scotlandi, P. Picci, and H. Kovar, "Targeted therapies in bone sarcomas," *Current Cancer Drug Targets*, vol. 9, no. 7, pp. 843–853, 2009.
  - [71] H. Morioka, L. Weissbach, T. Vogel et al., "Antiangiogenesis treatment combined with chemotherapy produces chondrosarcoma necrosis," *Clinical Cancer Research*, vol. 9, no. 3, pp. 1211–1217, 2003.
  - [72] K. M. Sutton, M. Wright, G. Fondren, C. A. Towle, and H. J. Mankin, "Cyclooxygenase-2 expression in chondrosarcoma," *Oncology*, vol. 66, no. 4, pp. 275–280, 2004.
  - [73] Y. M. Schrage, I. Machado, D. Meijer et al., "COX-2 expression in chondrosarcoma: a role for celecoxib treatment?" *European Journal of Cancer*, vol. 46, no. 3, pp. 616–624, 2010.
  - [74] X. Sun, L. Wei, Q. Chen, and R. M. Terek, "HDAC4 represses vascular endothelial growth factor expression in chondrosarcoma by modulating RUNX2 activity," *Journal of Biological Chemistry*, vol. 284, no. 33, pp. 21881–21890, 2009.
  - [75] F. Blanchard and C. Chipoy, "Histone deacetylase inhibitors: new drugs for the treatment of inflammatory diseases?" *Drug Discovery Today*, vol. 10, no. 3, pp. 197–204, 2005.
  - [76] R. Sakimura, K. Tanaka, S. Yamamoto et al., "The effects of histone deacetylase inhibitors on the induction of differentiation in chondrosarcoma cells," *Clinical Cancer Research*, vol. 13, no. 1, pp. 275–282, 2007.
  - [77] B. Ory, G. Moriceau, F. Redini, and D. Heymann, "mTOR inhibitors (Rapamycin and its derivatives) and nitrogen containing bisphosphonates: bi-functional compounds for the treatment of bone tumours," *Current Medicinal Chemistry*, vol. 14, no. 13, pp. 1381–1387, 2007.
  - [78] D. Heymann, B. Ory, F. Gouin, J. R. Green, and F. Rédini, "Bisphosphonates: new therapeutic agents for the treatment of bone tumors," *Trends in Molecular Medicine*, vol. 10, no. 7, pp. 337–343, 2004.
  - [79] R. E. Brown, "Brief communication: morphoproteomic portrait of the mTOR pathway in mesenchymal chondrosarcoma," *Annals of Clinical and Laboratory Science*, vol. 34, no. 4, pp. 397–399, 2004.
  - [80] G. Moriceau, B. Ory, L. Mitrofan et al., "Zoledronic acid potentiates mTOR inhibition and abolishes the resistance of osteosarcoma cells to RAD001 (everolimus): pivotal role of the prenylation process," *Cancer Research*, vol. 70, no. 24, pp. 10329–10339, 2010.
  - [81] O. Merimsky, R. Bernstein-Molho, and R. Sagi-Eisenberg, "Targeting the mammalian target of rapamycin in myxoid chondrosarcoma," *Anti-Cancer Drugs*, vol. 19, no. 10, pp. 1019–1021, 2008.
  - [82] O. Kollet, A. Dar, S. Shviti et al., "Osteoclasts degrade endosteal components and promote mobilization of hematopoietic progenitor cells," *Nature Medicine*, vol. 12, no. 6, pp. 657–664, 2006.
  - [83] T. J. Lai, S. F. Hsu, T. M. Li et al., "Alendronate inhibits cell invasion and MMP-2 secretion in human chondrosarcoma cell line," *Acta Pharmacologica Sinica*, vol. 28, no. 8, pp. 1231–1235, 2007.
  - [84] T. Kubo, S. Shimose, T. Matsuo et al., "Inhibitory effects of a new bisphosphonate, minodronate, on proliferation and invasion of a variety of malignant bone tumor cells," *Journal of Orthopaedic Research*, vol. 24, no. 6, pp. 1138–1144, 2006.
  - [85] F. Gouin, B. Ory, F. Rédini, and D. Heymann, "Zoledronic acid slows down rat primary chondrosarcoma development, recurrent tumor progression after intralesional curettage and increases overall survival," *International Journal of Cancer*, vol. 119, no. 5, pp. 980–984, 2006.