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Modelling combined effects of temperature and pH on the heat resistance of spores of *Bacillus cereus*.

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Abstract

The development of relationships between the pH of a heating medium and the thermal resistance of contaminant microorganisms is important and of a public health significance. A number of mathematical models have been presented in recent years, including that of Mafart and Leguérinel (1998). However, in this model, the effect of possible interactions between temperature and pH on D-values was not assessed. The consequences of ignoring interaction terms in models has been assessed and a comparison with Mafart's model that includes an interaction term showed that interaction terms can be neglected and that Mafart’s model can be used in thermal process calculations. It appears possible to adopt a standard value of $z_{pH}$, for example the 3.6 value and the conventional concept of biological destruction value $L(T)$ (ratio of the sterilization value and the exposure time at a fixed heating temperature) may then be extended to $L(T,pH)$ (the same ratio at a fixed temperature with a fixed pH of the heating menstruum).

Keywords:
Introduction

Interactions between heating temperature and pH are frequently observed. In most cases, increasing temperature reduces the effect of pH. This type of interaction can be seen in the data of Xezones and Hutchings (1965) for *Clostridium botulinum*. This same behaviour is observed for *Bacillus stearothermophilus* (Rodrigo et al., 1993; Fernandez et al., 1994; Lopez et al., 1996). Rodrigo et al. (1993) observed that pH did not have any effect on the DT values of spores of *Clostridium sporogenes* when they were heated at high temperature. These results do not agree with those of Cameron et al. (1980) from which it can be shown that increasing temperature enhances the effect of pH.

Acidifying the heating medium generally results in a decreased effect of temperature: an increase of the z value is observed when the pH of the medium decreases. This behaviour was showed for *Bacillus stearothermophilus* (Fernandez et al., 1994; Lopez et al., 1996), *Bacillus subtilis* (Condon and Sala, 1992), *Bacillus licheniformis* (Palop et al., 1992) and *Clostridium sporogenes* (Silla Santos et al. 1992). However, Cameron et al. (1980) pointed out an opposite effect for *Clostridium sporogenes* (a decrease of the z value with decreasing pH), that disagrees with results of Silla Santos et al. (1992).

The mathematical relationship between the pH of the heating medium and the thermal resistance of microorganisms has been poorly documented. Jordan and Jacobs (1948) found a linear relationship between pH and the logarithm of the DT values for *E. coli*. Davey et al. (1978) developed the first model for predicting the combined effects of both process temperature and pH on thermal resistance of bacteria. This empirical model, which was developed from data of Xezones and Hutching (1965) for inactivation of *Clostridium botulinum*, shows a satisfactory goodness of fit and is relatively parsimonious (4 parameters for 2 factors). However it can be shown that Davey’s model is still over parameterized (see appendix 1). The four empirical coefficients of the model have no biological significance. As an alternative, Mafart and Leguérinel (1998) proposed a new model with only three parameters, each having a physicochemical significance:

\[
\log D_{(T, pH)} = \log D_{(T*, pH*)} - \frac{T - T^*}{z_T} - \frac{(pH - pH^*)^2}{z_{pH}^2} (1)
\]
Where $T^*$ is the reference temperature (for example 121.1°C), pH* is the pH of maximal heat resistance of spores (generally pH 7), $z_T$ is the conventional thermal z-value, $z_{pH}$ is the distance of pH from pH* which leads to a ten fold reduction of D value. Lastly, $D_{(T^*,pH^*)}$ is the D value at $T^*$ and pH*.

This model was derived from the same data used by Davey et al., 1978 (i.e. Xezones and Hutching, 1965) and from two other sets of data related to \textit{Clostridium sporogenes} (Cameron et al., 1980) and to \textit{Bacillus stearothermophilus} (Lopez et al., 1996).

Neither Davey’s nor Mafart’s models took into account interactions between temperature and pH on thermal resistance of spores. This paper aims to check from our own data related to \textit{Bacillus cereus} that the lack of term accounting for interactions in the Mafart’s model can be justified.

The standard z-value used to compute most heating process is 10°C because it nearly corresponds to the value for \textit{Clostridium botulinum} spores and is closed to $z_T$-value of most other types of bacterial spores. Another aim of this paper is to check that the $z_{pH}$ value of \textit{Bacillus cereus} is in the same range than that of others studied types of spores. If this paper, confirmed by further works, shows that most of $z_{pH}$ values are close to that of \textit{Clostridium botulinum}, it would be possible to adopt a standard $z_{pH}$ for calculations of heat treatements.

**Materials and methods**

**Microorganism and spore production**

The strain of \textit{Bacillus cereus} (CNRZ 110) was obtained from the Institut National de Recherche Agronomie (France). Spores were kept in distilled water at 4°C.

Cells were precultivated at 37°C during 24 h in Brain Heart Infusion (Difco). The preculture was used to inoculate nutritive agar plates (Biokar Diagnostics BK021) added with MnSO$_4$ 40mg l$^{-1}$ and CaCl$_2$ 100 mg.l$^{-1}$ on the surface aera. Plates were incubated at 37°C for 5 days. Spores were then collected by scraping the surface of the agar and suspended in sterile distilled water and washed three times by centrifugation (10000xg for 15 min) (Bioblock Scientific, model Sigma 3K30). The pellet was then resuspended in 5 ml distilled water and 5 ml ethanol. The obtained suspension was then kept at 4°C during 12 hours in order to eliminate vegetative non sporulated bacteria, and washed again three times by centrifugation. The final suspension (about $10^{10}$ spores ml$^{-1}$) was at last distributed in sterile Eppendorfs microtubes and kept at 4°C.
Thermal treatment of spore suspension

$D_{(T,\,pH)}$ values in citrate-phosphate buffers adjusted respectively to 4.1, 4.5, 5.5, 6.5 and 6.9 were determined for temperatures of 86.6°C, 89°C, 95°C, 101°C and 103.4°C with one replicate at each temperature and pH combination. The whole set of data involves a complete 3x3 factorial design from 89°C to 101°C and from pH 4.5 to pH 6.5 and in order to extend the range of validity of the model, the four following combinations were added: pH 5.5 at 86.6°C and 103.4°C, pH 4.1 and 6.9 at 95°C.

First, 30μl of spore suspension was diluted in 3 ml buffer. Capillary tubes of 25 μl (vitrex) were filled with 10μl of sample and submitted to a thermal treatment in a thermostated oil bath. After heating, the tubes were cooled in water/ice bath, washed in a solution of soap and rinsed with sterile distilled water. Finally, ends were flamed with ethanol. The capillary tubes were broken at both ends and their contents poured into a tube containing 9 ml sterile tryptone salt broth (Biokar Diagnostics) by rincing with 1 ml tryptone salt broth contained in a syringe equipped with a needle.

Viable spore count

The viable spores were counted by duplicate plating in nutritive agar (10g tryptone, 5g meat extract, 5g sodium chloride, 15 g agar for 1000ml water)(Biokar Diagnostic) and incubating at 30°C for 48h.

Data analysis

$DT$ values were based on the reciprocal of slopes obtained when the log number of survivors was plotted against time. Multiple linear regressions used to fit the model were carried out with the STAT-ITCF software (Institut Technique du Fourrage France).

The goodness of fit of the model was evaluated by using the per cent variance accounted for (Snedecor and Cochran, 1969) which is given by:

$$R^2 = 1 - \frac{(1 - r^2)(n - 1)}{(n - N - 1)}$$

where $n$ is the number of observations, $N$ the number of terms and $r^2$ is the multiple regression coefficient.
Results

Raw data of the experimental design are shown in Table 1. As the thermal resistance of *Bacillus cereus* is relatively low, we adopted the standard temperature $T^*=100°C$ instead of $121.1°C$. On the other hand, a preliminary experiment showed that the pH of maximal thermal resistance related to the studied strain of *Bacillus cereus* was close to 7.5 (data not shown). The model was then fitted with $pH^*=7.5$. Fitted parameters according to equation 1 were:

$$D_{(T^*,pH^*)} = 60 \text{ seconds}$$
$$z_T = 9.15°C$$
$$z_{pH} = 3.70$$

with a per cent variation accounted for $R^2 = 97.7%$.

Families of graphs of $D_{(T,pH)}$ as a function of temperature at constant values of pH and as a function of pH at constant values of temperature are shown in Figures 1 and 2 respectively. Decreasing pH reduced the effect of temperature: at pH 6.5, the $z_T$ value was 7.97°C whilst it becomes 9.27°C and 10.3°C at pH5.5 and 4.5 respectively. At 95°C and 101°C, the $z_{pH}$ value remains constant and equal to 3.87. On the other hand, a decrease of the $z_{pH}$ value is observed at 89°C ($z_{pH}=3.06$). It is then clear that the heat resistance of spores depends not only on temperature and pH of the medium, but also on interactions between these two factors. In order to take into account interactions, the model (1) was modified by adding an interaction term:

$$\log D_{(T,pH)} = \log D_{(T^*,pH^*)} + C_1(T - T^*) + C_2(pH - pH^*)^2 + C_3(T - T^*)(pH - pH^*)^2$$

(2)

Where $C_1$, $C_2$ and $C_3$ are empirical parameters without known physiological significance. Fitted parameters according to this new model were:

$$D_{(T^*,pH^*)} = 83 \text{ seconds}$$
$$C_1 = -0.2900°C^{-1}$$
$$C_2 = -0.0281$$
$$C_3 = 0.1807°C^{-1}$$

with a per cent variation accounted for $R^2 = 98.5%$.

The new interaction term was highly significant ($P<0.01%$).
The compariasion between goodness of fit of both models (equations 1 and 2) are illustrated by Figures 3 and 4.

4. Discussion

Variation of the thermal resistance of *Bacillus cereus* spores as a function of temperature and pH is similar to those for many bacterial spores. It is confirmed that the pH of the heating medium has a prominent effect on the \(D_{(T, \text{pH})}\) values. For example, at 95°C a decrease of D-value of 5.7 times can be observed when the pH decrease from 6.9 to 4.1. As for most bacterial spores, an increase of the temperature results in a decrease in the effect of the pH while a decrease of the pH medium results in a decrease of the effect of the temperature. This behaviour means that some interactions on \(D_{(T, \text{pH})}\)-values exist between temperature and pH. This is confirmed by the fact that the interactions term of equation (2) is highly significant. However, adding one more parameters to our model in order to account for interactions results in a poor improvement of its goodness of fit: (with a \(R^2\)-value of 0.985 instead of 0.977). Moreover, in Equation 2, only one parameter \(D_{(T^*, \text{pH}^*)}\) keeps a biological significance: because of the occurrence of the interaction term, it is not possible to reparameterize \(C_1\) and \(C_2\) into \(z_T\) and \(z_{\text{pH}}\) respectively. Fernandez et al. (1996) proposed two models for combined effects of temperature and pH on the thermal resistance of *Bacillus stearothermophilus* and *Clostridium sporogenes*: one a polynomial quadratic model including an interaction term (6 parameters), the other a simple linear model without interaction terms (3 parameters). From their data it may be concluded that the slight improvement of goodness of fit obtained by the quadratic model with respect to the linear model does not justify the choice of the 6 parameters model rather than that of 3 parameters model. The linear model of Fernandez et al. is of the form:

\[
\log K = a + bT + c\text{pH} \quad (3)
\]

where \(K\) is the death rate of spores and \(a\), \(b\) and \(c\) empirical coefficients. It can be easily shown (see Appendix 2) that equation (3) can be reparameterized to give:

\[
\log D = \log D^* - \frac{1}{z_T} (T - T^*) - \frac{1}{z_{\text{pH}}} (pH - pH^*) \quad (4)
\]

Disregarding pH terms, both models (1) and (4) are reduced to Bigelow’s equation. However equation (1) where the pH term is squared differs from the
simple linear model (equation (4)) and is the only one to take the sigmoïdal
pattern of D values curves versus pH into account.

The \( z_{pH} \) value (3.7) related to *Bacillus cereus* have the same magnitude
that the observed values for *Clostridium botulinum* (3.56-3.61), *Clostridium
sporogenes* (3.33-4.29) and *Bacillus stearothermophilus* (2.94-3.97) (Mafart
and Leguérinel, 1998). Like conventional \( z_T \) values of spores close to 10°C and
keep generally the range of 8°C to 12°C, it appears that \( z_{pH} \) values are, in most
situations, included in a 3 to 4 range. If this observation is confirmed, it may be
possible to adopt a standard value of \( z_{pH} \) (for example the 3.6 value of
*Clostridium botulinum*) in order to include the pH in thermal process
calculations. The sterilization value could be defined as a heat treatment
equivalent to an exposure time of 1 min at 121.1°C and at pH 7. The
conventional concept of biological destruction value \( L(T) \) could then be
extended to \( L(T,pH) \) with

\[
L(T, pH) = 10 \left( \frac{T - T^*}{z_T} + \left( \frac{pH - pH^*}{z_{pH}} \right)^2 \right)
\]

(5)

This function corresponds to the exposure time, at a T temperature and at a
given pH, which would be necessary in order to obtain one sterilization unit.


Appendix 1: Reparameterization of Davey’s model

Davey et al. (1978) described variation of death rates versus temperature and pH by the following equation:

\[ \ln K = C_0 + \frac{C_1}{T} + C_2 pH + C_3 pH^2 \]  

(1)

where \( K \) is the death rate constant, \( T \) is the absolute temperature and \( C_0, C_1, C_2 \) and \( C_3 \) are empirical coefficients. Let \( pH^* \) be the pH value of maximal thermal resistance of spores (in most cases \( pH^* = 7 \)).

In the situation of maximal thermal resistance,

\[ \frac{d (\ln K)}{dpH} = C_2 + 2C_3 pH = 0 \]  

(2)

\[ pH^* = \frac{-C_2}{2C_3} \]  

(3)

so that Davey’s model can be rewritten with only three parameters:

\[ \ln K = C_0 + \frac{C_1}{T} + C_2 pH \left(1 - \frac{1}{2} \frac{pH}{pH^*}\right) \]  

(4)

For example, in the case of the heat treatment of *Clostridium botulinum* spores in spaghetti tomato sauce, Davey’s coefficient values were \( C_2 = -2.3967; \) \( C_3 = 0.1965 \). In macaroni creole, \( C_2 = -2.6170 \) and \( C_3 = 0.1871 \). Corresponding \( pH^* \) values are then 7.07 and 6.99 respectively.
Appendix 2: Reparameterization of Fernandez’s linear model

The following model was proposed:

\[
\log K = a + bT + cpH \quad (1)
\]

with \( K = \frac{Ln10}{D} \)

so equation (1) is equivalent to

\[
\log D = \log (Ln10) - a - bT - cpH \quad (2)
\]

or

\[
\log D = \log D^* - \frac{1}{z_T} (T - T^*) - \frac{1}{z_pH} (pH - pH^*) \quad (3)
\]

By comparison between equation (2) and (3) new parameters \( D^*, z_T \) and \( z_pH \) can be identified versus Fernandez’s parameters:

\[
\log D^* = \log (Ln10) - a - bT^* - cpH^*
\]

\[
z_T = \frac{1}{b} \quad (5)
\]

\[
z_pH = \frac{-1}{c} \quad (6)
\]

For example, for *Bacillus stearothermophilus* heated in mushroom extract acidified with glucono-δ-lactone, \( a=-12.0495; \ b=0.1156; \ c=0.2990 \). With \( T^*=121.1^\circ C \) and \( pH^*=7 \), new parameters are

\( D^*=3.20 \) minutes

\( z_T=8.65^\circ C \)

\( z_pH=3.34 \)
Table 1

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<th>95°C</th>
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Figure 1
Figure 2
Three parameters model

Figure 3
Erreur! Objet incorporé incorrect.
Legends of figures

Figure 1:
Log D value of *Bacillus cereus* spores versus (T-T*).
Key: □, log D at pH 6.5 ; Δ, log D at pH 5.5 ; ○, log D at 4.5.

Figure 2:
Log D value of *Bacillus cereus* spores versus (pH-pH*)
Key: □, log D at 89°C ; Δ, log D at 95°C ; ○, log D at 101°C.

Figure 3:
Observed log D value compared to log D value calculated according to the three parameters model

Figure 4:
Observed log D value compared to log D value calculated according to the four parameters model
Legends of Table

Table 1

D values (minutes) of experimental design.