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RESEARCH ARTICLE

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A transcriptional-switch model for Slr1738-controlled gene expression in the cyanobacterium *Synechocystis*

Paul Garcin¹, Olivier Delalande¹, Ju-Yuan Zhang¹, Corinne Cassier-Chauvat^{1,2}, Franck Chauvat¹ and Yves Boulard^{1*}

Abstract

Background: Protein-DNA interactions play a crucial role in the life of biological organisms in controlling transcription, regulation, as well as DNA recombination and repair. The deep understanding of these processes, which requires the atomic description of the interactions occurring between the proteins and their DNA partners is often limited by the absence of a 3D structure of such complexes.

Results: In this study, using a method combining sequence homology, structural analogy modeling and biochemical data, we first build the 3D structure of the complex between the poorly-characterized PerR-like regulator Slr1738 and its target DNA, which controls the defences against metal and oxidative stresses in *Synechocystis*. In a second step, we propose an expanded version of the Slr1738-DNA structure, which accommodates the DNA binding of Slr1738 multimers, a feature likely operating in the complex Slr1738-mediated regulation of stress responses. Finally, in agreement with experimental data we present a 3D-structure of the Slr1738-DNA complex resulting from the binding of multimers of the Fur-like regulator onto its target DNA that possesses internal repeats.

Conclusion: Using a combination of different types of data, we build and validate a relevant model of the tridimensional structure of a biologically important protein-DNA complex. Then, based on published observations, we propose more elaborated multimeric models that may be biologically important to understand molecular mechanisms.

Background

DNA-binding proteins play a crucial role in many fundamental biological processes including transcription, regulation, as well as DNA replication and repair. Thus, a better understanding of DNA-protein interactions has both a fundamental research interest and an applied importance in medicine (development of drugs interfering with oncogene expression) and biotechnology (genetic engineering of microbial organisms).

In the past, a lot of effort has been made to understand the basic principles that govern the specificity of protein-DNA interactions. It appeared that there is no simple recognition code linking the DNA interacting

amino acids of a protein with their target DNA nucleotides [1]. Furthermore, there are currently no standard methods to build a 3D-structure model for the representation of a DNA-protein complex, unlike what occurs for protein-protein interactions [2]. All current methods for predicting the structures of protein-DNA complexes use the features of the unbound protein and DNA partners and various algorithms (shape complementarity, surfaces properties, experimental contacts...) to drive the docking, and propose a model for the studied protein-DNA complexes. By contrast, in this study, we used the experimentally-determined structures of protein-DNA complexes that are presumably similar to the one we study, to build a model representation of its possible structure. For this purpose, we selected among the DNA-protein complexes available at the PDB database, those sharing secondary structure motif analogy with our protein of interest, irrespective of the sequence

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homology between these reference proteins and our studied protein. This strategy, aims to preserve the structural conformations required to establish the interactions between amino acids and nucleotides in the model complexes. In order to test the feasibility of this strategy, we applied it to manually build a reliable model of the complex occurring between Slr1738, an important but structurally uncharacterized member of the family of PerR transcription regulators, and its target DNA. The PerR family of regulators belongs to the larger family of bacterial FUR regulators (ferric uptake regulator), which control the responses to iron or zinc availabilities [3], for a review see [4]. PerR was initially characterized as the master regulator of the *Bacillus subtilis* responses to hydrogen peroxide [5], which regulates the anti-oxidant genes encoding the DNA binding protein MgrA, the catalase KatA, the alkyl hydroperoxide reductase AhpCF, PerR itself, and FUR, in accordance with the interplay between iron homeostasis and protection against oxidative stress. PerR is a small dimeric protein that contains two metal ions per monomer and binds to AT-rich DNA motifs of the promoter region of its target genes. One metal-binding site coordinates a zinc ion that plays a structural role, while the second site binds the regulatory metal, Fe²⁺ (PerR-Zn-Fe) or Mn²⁺ (PerR-Zn-Mn). PerR senses H₂O₂ through the Fe-catalyzed oxidation of its H37 or H91 amino acid residues, leading to dissociation of the PerR-DNA complex [6]. The recent crystallographic structures of the PerR protein, though not complexed to its target DNA, suggested possible mechanisms by which PerR undergoes similar conformational changes upon binding either Fe or Mn. The structure of the PerR-Zn protein lacking Fe [7] reveals the two CXXC motifs involved in the tetrahedral coordination of Zn²⁺ that stabilizes the PerR dimer in a flat conformation poorly suited to bind DNA. The structures of the fully metalated proteins PerR-Zn-Mn and PerR-Zn-Fe indicate that the binding of either Mn²⁺ or Fe²⁺ ions, likely to the same pair of regulatory sites on the dimer, lead to a caliper-like close conformation better suited to bind DNA [5,8].

PerR-like regulators occur in a wide variety of prokaryotic organisms, including cyanobacteria, which are important for the Biosphere in producing a large part of the atmospheric oxygen and the biomass for the food chain [9], and have promising biotechnological potentials [10-12]. By their nature, cyanobacteria are frequently challenged by the intrinsically related oxidative and iron stresses, as they perform the two main iron-requiring oxidant-generating processes respiration and photosynthesis [13]. In this study, we pursued the analysis of the PerR-like regulator Slr1738, which controls the responses to oxidative and metal stresses in the widely-used model cyanobacterium *Synechocystis*

PCC6803 [14-16]. We identified the transcription start site and the crucial -10 promoter element for the two oppositely oriented genes *slr1738* and *sll1621* (*sll1621* encodes the antioxidant enzyme AhpC (alkylhydroperoxidase [13])). We also characterized the long (33 bp) AT rich motif involved in the Slr1738-mediated repression of *sll1621*. Also interestingly, we built a 3D structural model of Slr1738 complexed with its AT-rich target DNA. This model will be of great help to decipher the molecular mechanisms operating in the tight interplay between iron homeostasis and tolerance to metal and oxidative stresses. Furthermore, starting from our protein dimer model, we also propose more speculative complex, *i.e.* oligomeric, structures (tetramer, hexamer, etc...) as possible molecular effectors of the numerous regulations controlled by Slr1738 [17].

Methods

Experimental work

Determination of the transcription start sites of the divergently transcribed *slr1738* and *sll1621* genes

Total RNAs were isolated from *Synechocystis* and treated with shrimp alkaline phosphatase (SAP) that does not affect full-length mRNA, which have 5'-triphosphate ends, but dephosphorylates degraded RNA, which have a 5'-monophosphate extremity. Then RNAs were treated with tobacco acid pyrophosphatase (TAP), which converts the 5'-triphosphate of full length mRNA into 5'-monophosphate, but does not modify the 5'-OH of the degraded RNA. Then the 5'-monophosphate extremity of the full length mRNA was ligated to an RNA anchor with the T4 RNA ligase, and the resulting chimeric RNA was reversed transcribed with a gene-specific primer, thus creating the first strand of cDNA. This cDNA strand was amplified by PCR using both the DNA version of the RNA anchor at the 5' extremity, and the gene specific primer at the 3' side. Finally, the PCR-amplified DNA was sequenced to determine the nature of the nucleotide immediately downstream of the DNA anchor oligonucleotide that corresponds to the transcription start site (TSS).

Construction of transcriptional fusions to the *cat* reporter gene and CAT assay

The *slr1738* promoter region and segments thereof were amplified by PCR, using site-specific oligonucleotides that flanked the PCR DNA product with *Sna*BI blunt-ended restriction sites in such a way that all nucleotide substitutions were eliminated upon cleavage with *Sna*BI. The resulting blunt-ended promoter fragments were cloned in the unique *Sna*BI site of the pSB2A promoter probe vector [18], *i.e.* in front of its promoter-less *cat* reporter gene. The sequence of every promoter insert was verified (Big Dye kit; ABI Perkin-Elmer) before and after replication in *Synechocystis*. Then, 1-2 × 10⁹

reporter cells grown on standard plates up to mid-log phase culture were rapidly harvested and disrupted with an Eaton press, prior to CAT assay [19]. CAT activities are the mean value of three measurements performed on two independent cellular extracts; 1 CAT unit = 1 nmol of chloramphenicol acetylated. min^{-1} . mg^{-1} of protein.

Modeling work

Slr1738 monomer construction

The homology model for the monomer of the *Synechocystis* PerR-like regulator Slr1738 was obtained after sequence alignment of the structure of the closely related FUR protein of *Pseudomonas aeruginosa* (PDB ID 1MZB) [20] and built using the modeller program. Completion of the starting structure of the PerR-like model was achieved with the xLeap module of Amber 9 suite, to finally get the correct 139 residues protein sequence (Figure 1A). The final structural model of the PerR-like protein was obtained by short MD relaxation.

Metallic sites parametrization

Considering the nature of the amino acid ligands of the two potential metal-binding sites (using analogy to the family of FUR regulators and its PerR-type sub-family), the Slr1738 protein should likely be a PerR type containing one iron atom and one zinc atom per monomer. Zinc site forcefield parameters have been already proposed for Amber [21,22], so we used a combination of these data and of the tetrahedral geometry of a four-cysteine zinc site in a metallothionein (PDB ID 1JJD) [23] as a structural reference to set the zinc site parameters. The four C95, C98, C134 and C137 cysteines of Slr1738 likely involved in metal coordination (determination by CxxC pattern analogy to PerR regulator) are covalently bonded (*i.e.* explicit bonded terms in the forcefield) to the zinc atom via their sulfur atom.

The iron site is not sufficiently defined for a rigorous semi-empirical treatment at simple atomic resolution. Iron should be in its ferrous state [24] and penta- or hexa-coordinated to histidine (via deprotonated pyrrole

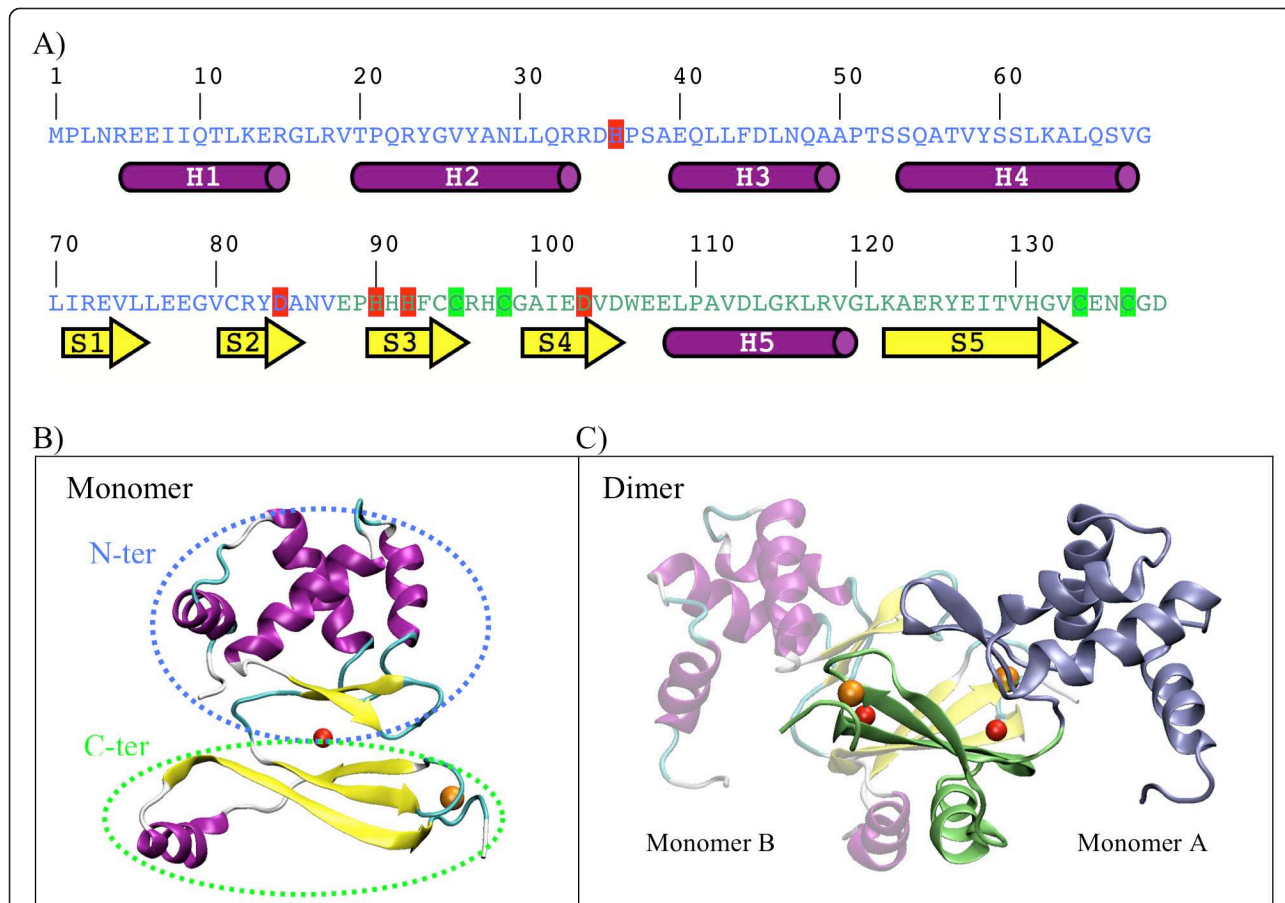


Figure 1 Protein Slr1738 features. (A) Slr1738 primary sequence (N-terminal domain in blue and C-terminal domain in green) and predicted secondary structure elements (α -helix in purple and β -strands in yellow). Metallic ligands are highlighted in red for iron and in green for zinc site. B) 3D monomeric and C) dimeric model of Slr1738 in ribbon representation (iron and zinc ions are respectively represented in red and orange van der Waals spheres).

nitrogen) and aspartic acid residues [8]. We chose to model the metal ion as a hard sphere with a +2.0 raw charge and a 1.2 Å van der Waals radius. The coordination site was set via distances harmonic constraints (100 Kcal.mol⁻¹.Å⁻¹) between iron and its amino acid ligands. Despite inducing potential local distortions, the overestimation of the charge should prevent dissociation of the metal ion from its binding site and preferentially stabilize the folded conformation. In our initial model, iron presents a tetrahedral geometry (H32, E80, H89 and E100 in the *Pseudomonas aeruginosa* *FUR* regulator are respectively H36, D84, H92 and D103 in the *Synechocystis* Slr1738 regulator) resulting from the experimental Fe/Zn-substituted structure. Bipyramidal base-square geometry is reached with histidine H90 residue coordination and the addition of a water molecule that could be crucial for the function of the protein [6].

Molecular mechanics and molecular dynamics (MD) simulations

All simulations have been performed using Parm99 force-field and programs of Amber 9 suite [25]. Molecules were neutralized with Na⁺ ions and placed in TIP3P water boxes for explicit solvation. After energy minimisations, models were used to initiate MD simulations. Short MD simulations (5ns for the production period) were principally used to allow accelerated geometry optimisation after structure modifications. This provided also qualitative information about stability of the models and the flexible regions of a structure. Final models were obtained after geometric average on the whole stabilized trajectories followed by energy minimization in a solvent box.

Energy association and contact surface calculations

Binding free energies between DNA and proteins molecules were estimated using the MM-PBSA method [26]. This method was used with success for several biological protein-ligand complexes [27-29] though the final values need to be interpreted with caution due to approximations in entropic contributions. To avoid such problems, we compared only the relative, not absolute, values between the different complexes. Each complex used was rigorously comparable in terms of number of atoms. Contact surface interactions were computed with the MSMS program [30]. It gives access to the solvent accessible surface area (SASA), which was used to calculate the contact surface values between a receptor and its ligand by using the formula reported below.

$$CS = \frac{SASA_{rec} + SASA_{lig} - SASA_{cplx}}{2}$$

Three dimensional construction of the [(Slr1738-Zn-Fe)₂-DNA] complex

Choice of the template structure We built a Slr1738-DNA complex by structural analogy with the 239 PDB

structures of DNA-protein complexes involving transcription factors. Among those, we considered only the DNA binding proteins harbouring a presumptive helix-turn-helix (HTH) motif predicted by the Pfam database. HTH motifs are known to vary widely in sequence over the whole DNA-binding domain and their relationships can often only be based on structure similarity [31]. Finally, we retained 4 structures [32-35] because they complied with the following qualitative criteria (Additional file 1 Table S1) found in *FUR* proteins [20]: 1) the size of the double stranded DNA target site is ≥ 20 bp; 2) to be complexed with DNA the protein must be homodimeric; 3) the HTH motif (*ca.* 25 residues in length) must be followed by two anti-parallel β-strands. For information, the sequence alignment of the HTH motifs is given in Additional file 2 Figure S1; 4) the resolution of the experimental X-ray structure must be good. Note that as observed for the *FUR* dimer of *Escherichia coli* [36], the two recognition helices H₄ are almost perpendicular and thus should be positioned on both sides of the DNA global axis, and not on the same side as occurs with parallel helices (Additional file 3 Figure S2).

Fitting procedure Using successive energy minimisation steps, the Slr1738 monomer was fitted onto each of the 4 reference complexes by positioning the H₄ recognition helix in the major groove of the target DNA. This global protein-DNA association driven by distance restraints was performed with frozen Slr1738 secondary structure and ended after reproducing the geometry observed in the selected structural patterns. We tested four kinds of superposition motifs to select the best one enabling the largest surface of protein/DNA contacts as shown in Additional file 4 Table S2. We note that the contact surface differences between the template structures reflect differences in the DNA sequences. Then, we replaced the template nucleotides in the model by the Slr1738 DNA-binding sequence while maintaining the phosphodiester backbone of DNA. After geometry optimization, we selected only three structures on the quality of their surface contacts and association energies (Table 1). The final three models were derived from different PDB structures (1SAX, 1U8R and 1Z9C).

Closure of the structure The final step of our protocol was to close the protein-DNA complex by positioning the second monomer of Slr1738 onto its DNA target. This was done using a two-step method. First, we matched the dimer Slr1738 structure to the protein/DNA complex where both Slr1738 monomers were properly positioned. In this way, we conserved the integrity of the Slr1738 protein structure and the recognition helices remained close to their optimal positions. Second, we applied distance restraints derived from structural HTHw patterns on both monomers in order to insert the

Table 1 DNA-protein complexes parameters of the structures built with the target DNA sequence.

PDB name	Superposition type			
	H4S1S2		H3H4S1S2	
	Contact surface (Å ²)	Association energy with DNA (21 bp)	Contact surface (Å ²)	Association energy with DNA (21 bp)
1COW	900	35.4	888	19.03
1SAX	890	23.92	946	13.18
1U8R	949	25.88	885	7.18
1Z9C	1006	-6.82	939	11.14

Association energies and contact surfaces of 3D models with different secondary structure motifs superpositions for the construction of the DNA-(Slr1738)₂ complex. Best values are in bold.

Slr1738 recognition helices into the major groove of DNA. The integrity of the sugar-phosphate DNA backbone, the dimer interface and protein secondary structures was maintained under harmonic restraints throughout the procedure. The internal energies of the complexes were finally minimized with decreasingly harmonic restraints to ensure a smooth transition of the atomic system toward a relaxed configuration. The procedure was completed with short MD simulations in solvated and neutralized conditions. Contact surfaces and the association energies of the three best-predicted models are presented in Table 2. Structures at different steps of our construction strategy are presented in Figure 2.

Results and Discussion

In vivo analysis of the *slr1738* and *sll1621* divergent promoters: evidence for repression by Slr1738 and role of its AT-rich DNA binding motif

We pursued the analysis of the PerR-like regulator Slr1738 that controls the responses to oxidative and metal stresses in the model cyanobacterium *Synechocystis* PCC6803 [14-16]. It was reported [37] that the deletion of *slr1738* increases the expression of the two oppositely-oriented genes <http://genome.kazusa.or.jp/cyanobases> *slr1738* itself and *sll1621*, which encodes the anti-oxidant peroxiredoxin enzyme AhpC. It has also been shown that the Slr1738 protein binds the 300 bp *sll1621-slr1738* intergenic region [14-16], which possesses a long (30 bp) DNA motif containing only A and T nucleotides (the AT-only motif). Together, these

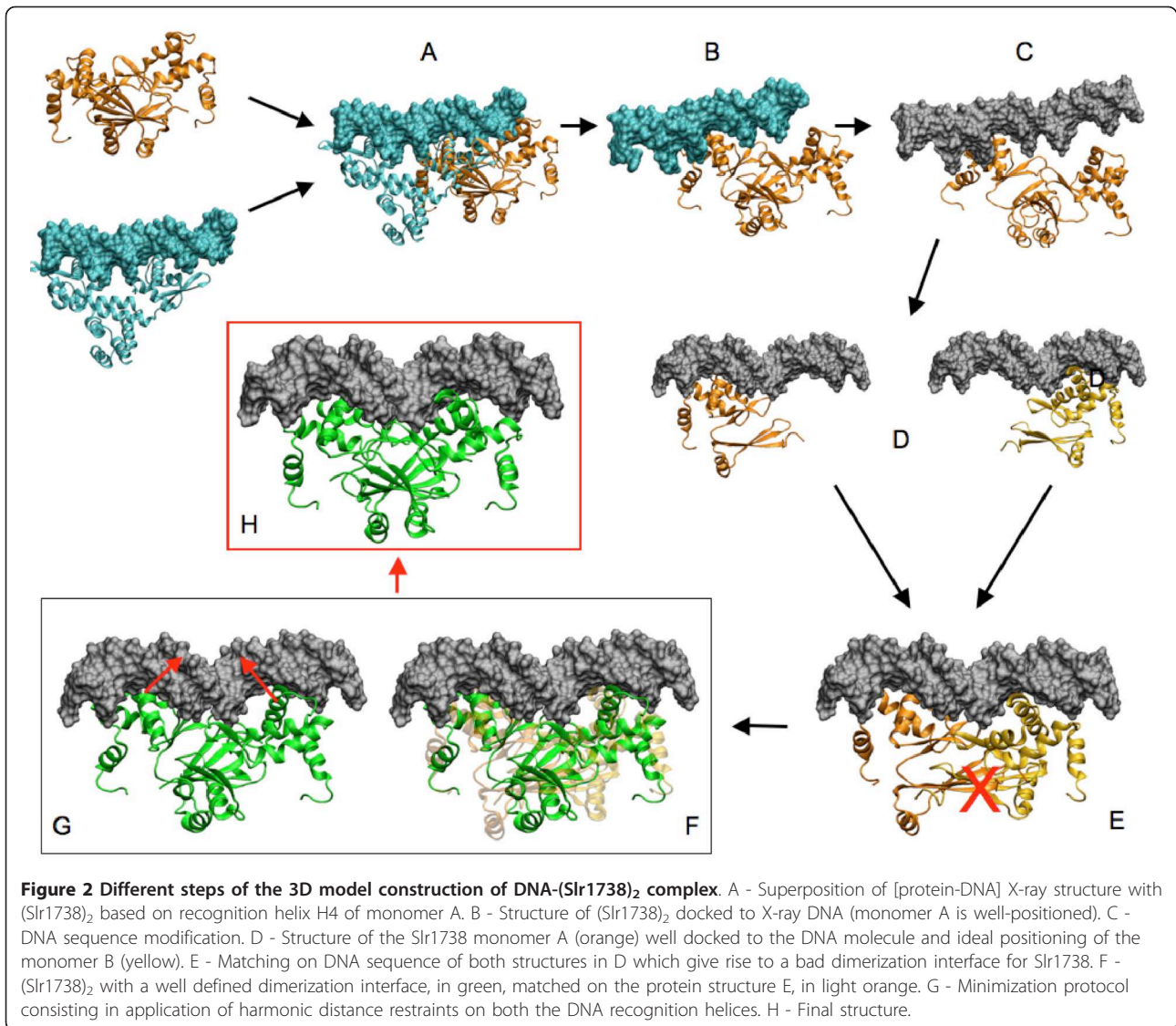
findings suggested that Slr1738 represses both the *slr1738* and *sll1621* promoters. To test this interpretation, and the possible role of the long AT-only DNA motif occurring in the *sll1621-slr1738* promoter region, we performed the following experiments. We cloned the 300 bp *sll1621-slr1738* promoter region, in both orientations relative to the promoter-less *cat* reporter gene of our promoter-probe plasmid vector pSB2A which replicates in *Synechocystis* at about 10 copies per cell, *i.e.* at one copy per copy of the polyploid chromosome [38]. This generated the reporter plasmids psll1621-*cat* and pslr1738-*cat*, which replicated stably in *Synechocystis*, as expected (data not shown), where they directed similar level of *cat* expression (Figure 3). As the usual control, we verified that the empty pSB2A plasmid carrying no promoter insert produced no CAT activity. Collectively, these data showed that the *sll1621* and *slr1738* promoters have similar strengths, which resemble those of other *Synechocystis* genes we previously studied with pSB2A [39] and references therein.

We and others [17] tried to find consensus sequences in the promoter regions of the wealth of genes presumably regulated by Slr1738. However we found none (Additional file 5 Figure S3), as occurred in the case of other PerR-like regulators [4,5,40], suggesting that particular DNA structures rather than defined nucleotide motifs in the target DNA govern PerR regulation of its target genes. This context encouraged us to perform a mutational analysis of the *slr1738* and *sll1621* promoters to identify their *cis*-acting promoter elements. In the *slr1738* promoter, we studied the 5'-TAagcT-3' hexamer that matches the canonical -10 box of σ 70-type *Escherichia coli* promoters [41] in both sequence (5'-TATAAT-3') and position (-12 to -7) from the transcription start site, which we presently mapped with the classical 5' RACE technique [42] that works well in *Synechocystis* [43]. It is the A nucleotide we noted as +1, which is located 131 bp upstream the *slr1838* start codon (see Additional file 6 Figure S4). Consistent with its identification as the -10 promoter box, we found the 5'-TAagcT-3' element to be crucial to *slr1738* transcription. Indeed, the transversion mutagenesis of its proximal T nucleotide

Table 2 DNA-protein complexes parameters of the final structures.

Structure name	Contact surface (Å ²)	Association energy with DNA (kcal/mol)
1SAX (H3H4S1S2)	1852	36
1U8R (H3H4S1S2)	1083	51.23
1Z9C (H4S1S2)	1230	50.45

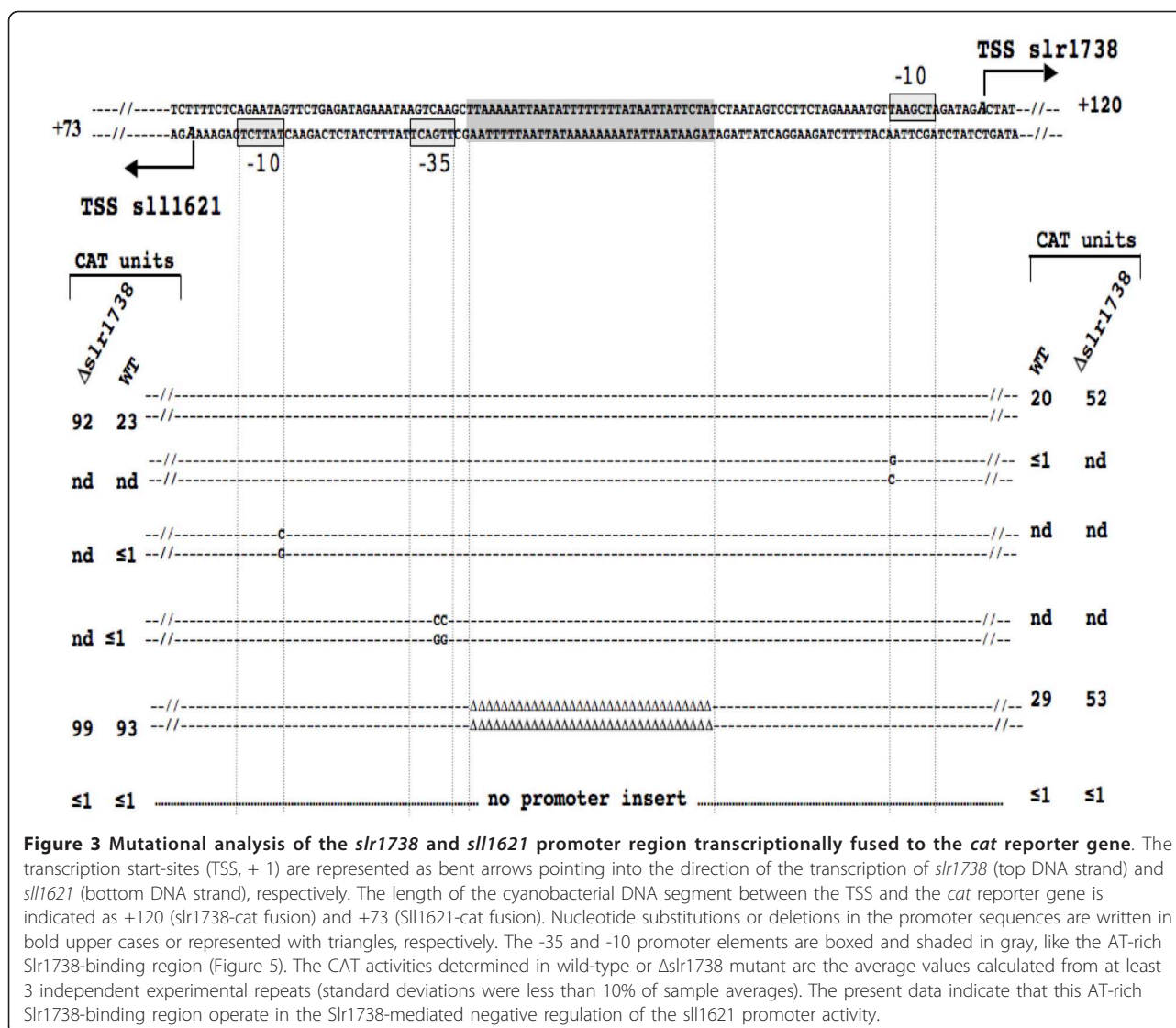
Association energies and contact surfaces of 3D models obtained for the final structures of DNA-(Slr1738)₂ complex. Best values are in bold.



(5'-TAagcT-3' to 5'-GAagcT-3') completely abolished *slr1738* promoter activity (Figure 3), as occurred with the -10 promoter boxes of other *Synechocystis* genes [19,39,44-46]. By contrast, no sequence resembling a -35 promoter box (5'-TTGACA-3') was found upstream of the *slr1738* -10 box, either at 17 bp (*i.e.* the canonical *Escherichia coli* spacing [47]) or at 30 bp (*i.e.* the distance sometimes encountered in *Synechocystis* [19,44,45]). The absence of a -35 promoter box in a *Synechocystis* promoter is not unprecedented as we previously showed that both the *lexA* and *recA* genes are well expressed from -35-less promoters [39]. Furthermore, as σ 70-type promoters lacking a -35 box often possess an "extended -10 box" (5'-TGnTATAAT-3') mediating all contacts with the RNA polymerase σ 70 factor [47,48], it is worth noting that the *slr1738* promoter harbors such an "extended -10 box" (5'-TGnTAagcT-3'), like the *Synechocystis secA*

promoter [19]. In contrast, the *sll1621* promoter appeared to possess the two canonical boxes, *i.e.* a -10 element (5'-TAttcT-3'; Figure 3) located 6 nucleotides upstream of the transcription start site (the A nucleotide noted as +1, we found to be located 73 bp upstream the ATG start codon; see Additional file 6 Figure S4) and a -35 (5'-TTGACT-3') box located 17 nucleotides upstream of the -10 element.

We also introduced and tested the *slr1738*-cat and *sll1621*-cat reporter plasmids in the Δ *slr1738* deletion mutant we previously constructed [14]. We found the *slr1738* promoter to be more active in the absence of the Slr1738 protein, demonstrating that Slr1738 is an autorepressor (Figure 3). Similarly, the *sll1621* promoter appeared to be more active in the absence of the Slr1738 protein (Figure 3), demonstrating that the Slr1738-mediated downregulation of the *sll1621* gene



[14-16] is exerted at the level of its promoter activity. Furthermore, we also found that the above-mentioned long (33 bp) AT-rich motif, which we anticipated to interact with *Slr1738*, truly operates in the *Slr1738*-mediated repression of the *sll1621* promoter (Figure 3).

Structure modelisation of (*Slr1738*-Zn-Fe)₂ protein complexed to DNA

At the beginning of our present study of the PerR-like regulator *Slr1738*, no structure of the metal-containing form of a PerR regulator was available in data banks. Therefore, starting from the *Slr1738* primary sequence, (Figure 1A), we built its tertiary structure using homology modelling methods, energy minimization and short MD simulation. Psi-Blast analysis showed that *Slr1738* exhibits 21% sequence identity (37% for homology) with the *Pseudomonas aeruginosa* FUR protein for which a

crystal structure was available (PDB ID 1MZB) [20]. Based on this and other findings and on our metal titration experiments (data not shown), we completed the two *Slr1738* metal binding sites with both a zinc ion (Zn²⁺) and a ferrous ion (Fe²⁺). These two metal sites were described as essential for the folding and activity of genuine PerR and FUR regulators [4,6-8,49]. In our *Slr1738* model, highly conserved cysteines (C95, C98, C134 and C137) operate in the coordination of the crucial zinc atom (Additional file 7 Figure S5A). The iron-binding site displays a hexa-coordinated geometry (H36, H90, H92, and D84, D103 and a water molecule) and the anchoring of the N-terminal domain of *Slr1738* on its C-terminal domain results essentially from H36-Fe coordination. However, as previously observed [15], we found experimentally that iron-less protein samples of *Slr1738* were still able to bind DNA, unlike what was

observed for the PerR protein where iron is crucial to DNA binding [6,8].

The resulting 3D model of Slr1738 comprises two well-defined domains (Figure 1B). The N-terminal domain from amino acids 1 to 84 contains four α -helices (H_1 to H_4) and a typical helix-turn-helix motif involving helices H_3 and H_4 that is responsible for the binding to DNA [50]. The helix H_4 is the recognition helix that interacts specifically with the major groove of DNA. The two H_3 and H_4 helices are followed by two anti-parallel β -strands (S_1 and S_2) that formed a winged helix-turn-helix motif (wHTH) [51]. The C-terminal domain of Slr1738 from amino acid 85 to 139 comprises three β -sheets (S_3 to S_5) and one α -helix (H_5) involved in the dimerization of Slr1738.

The Slr1738 dimer model we propose (Figure 1C) was first built from the *Pseudomonas aeruginosa* FUR structural information of Pohl et al. [20]. Our model is consistent with previous findings showing that the active form of FUR-type regulators is a dimer [52,53] and that Slr1738 too was proposed to bind to DNA as a dimer [15]. We then refined our model with the structure of the *Bacillus subtilis* PerR protein (PDB ID 2FE3 without Fe and 3F8N with Mn replacing Fe) [7,8]. The final RMSD values for the heavy atoms of the backbone were about 15 Å and 4 Å respectively for the refined structures, demonstrating the great role of iron in modifying the global folding. The zinc metal ion appears to be indispensable for structuring the C-terminal domain to enable dimerization [49]. The interactions involved at the interface of the dimer structure are depicted in Additional file 8 Figure S6.

Currently there is no 3D structure in the PDB database of a FUR/PerR-like regulator complexed with DNA. Therefore, we developed an approach, detailed in the Methods section, based on experimental 3D structures deposited in databases for predicting the structure of protein-DNA complex. Our model is consistent with the above-mentioned findings that Slr1738 binds to the 310 bp-long promoter region of the divergent genes *sll1621* and *slr1738* that contains a long AT-rich motif important for Slr1738-mediated regulation (Figure 3), which harbors a central symmetry for double strands, *i.e.* an ideal feature for binding a dimeric protein acting like pliers. Consequently, we chose a symmetrical 25 bp AT-rich DNA fragment for our modelling purposes, in agreement with the fact that the well-defined FUR-binding DNA sequences usually contain AT-rich palindromes [54]. We obtained the protein-DNA complex by fitting the (Slr1738)₂ model onto well positioned monomers using a strategy based on structural analogy with experimental structures, *i.e.* the recognition helices of the HTH motif being inserted into the major groove of target DNA. The selection of the final model was based

on both structural criteria and energy calculations. Our best complex model is the one based on the structural pattern derived from 1SAX.

Considerations of DNA conformation

In our approach, we did not take into account the DNA structural specificities possibly imposed by the oligonucleotide sequence. Indeed, we kept the DNA backbone structure of the template model to maintain the structural conformations occurring at the protein-DNA interface. Hence, we selected the template structures based on the wHTH motif identification of the Slr1738 protein. This choice was dictated by our long-standing expertise concerning DNA structure [55-59]. DNA molecules are very flexible and they can adopt many local conformations [60] depending on both their nucleotide sequence and the physico-chemical environment, as discussed in a wealth of papers. As discussed above, in absence of a genuine consensus sequence for the binding of Slr1738 onto DNA, we used its AT-rich regulatory element emerging from our work (Figure 3). Such an AT-rich DNA sequence may of course adopt particular local conformations that are not easy to predict and integrate in a 3D structure. Nevertheless, we note that: 1) the final structure was minimized in allowing DNA to relax; 2) MD simulations were performed to allow the structure, and thereby the DNA molecule, to explore 3D space; 3) the four 3D structures retained for the final selection (Table 1) are very different in terms of the AT content of the target DNA sequences, it varies from 48% for 1U8R to 86% for 1Z9C. The template structure we finally retained, *i.e.* 1SAX, is 68% AT-rich. Furthermore, all these DNA structures, analyzed with the 3DNA program [61], were found to adopt a globally B-form without bending in spite of their widely different AT contents; 4) it remains difficult nowadays to predict local DNA conformations from nucleotide sequences, unlike the situation of proteins in which the secondary structure is predictable from their primary sequence. Thus, we can obviously be more confident in selecting a protein template than a DNA template.

Structural analysis of the complex formed by the Slr1738 dimer and its DNA target

Recognition of DNA sequence is mediated by both direct interactions between amino acids and the bases in the major DNA groove (direct readout) and by contacts with DNA backbone (indirect readout). Specific interactions between Slr1738 and its target DNA concerned essentially the residues of the top of the recognition helices and *ca.* ten bases. Even though the dimeric form of Slr1738 docks an oligonucleotide of 25 bp in length, each Slr1738 monomer specifically interacts with only 5 bases in a symmetric way. The interactions are

schematically represented (see Additional file 9 Figure S7) inside a simple 7-1-7 bp DNA motif, the minimal recognition motif for FUR binding [62].

The final model of the complex predicts that there are six contact regions between the Slr1738 dimer and its target DNA, and each monomer operates in three of them via its three sub-regions, namely: (i) the loop between H_1 and H_2 , (ii) the α -helix H_4 and (iii) the loop between S_1 and S_2 (Figure 4A), which are discussed below.

(i) *The loop between H_1 and H_2 .* This region of Slr1738 comprises 11 amino acids including 4 positively-charged residues ($^{13}\text{KERGLRVTPQR}^{23}$). Two of them (R18 and R23) are presumably involved in electrostatic interactions with atoms of the negatively-charged phosphate groups of the DNA backbone in the minor groove. Sequence alignments of FUR-like proteins show that K13, R18 and R23 are highly conserved, especially among cyanobacterial Slr1738 orthologs (Additional file 7 Figure S5). This first region establishes only non-specific interactions with DNA in that there is no contact of protein side-chains with DNA bases. We propose that these long-range electrostatic interactions operate in the pre-orientation of the DNA-binding protein domain to facilitate protein/DNA interactions and in the stabilization of the resulting Slr1738/DNA complex that

precedes the sequence selective interaction mediated by the H_4 protein helix.

(ii) *The H_4 recognition helix.* H_{4A} and H_{4B} , the respective recognition helices of monomers A and B are inserted into the major DNA groove where they likely act as a pair of pliers. A detailed description of the H_4 helix is really informative in this respect. H_4 comprises 14 amino acids (residues 55-68) organized in three different sides (Figure 4B). The first side gathers the hydrophobic residues (V58, L62 & L65) lying in direct contact with numerous hydrophobic residues of the three other helices of the N-terminal domain, thereby forming a compact hydrophobic cluster. The second side, composed of 7 small residues (A56, T57, S60, S61, A64, S67 & V68), is close to the DNA strand 2. The third side comprising bulky and charged residues (Q55, Y59, K63 & Q66) faces the DNA strand 1. The residues in close vicinity of DNA bases are mainly those of the upper part of the helix which are localized in sides two and three: Q55, A56, T57, Y59, S60 and K63. Sequence alignment of FUR proteins shows that the DNA recognition helix always contains a conserved amino acid motif (A)TVY or (A)TIY where the tyrosine is important for DNA binding [36]. For Slr1738 the H_4 helix also has four hydroxyl-containing residues (serine and threonine residues), a feature possibly important because this

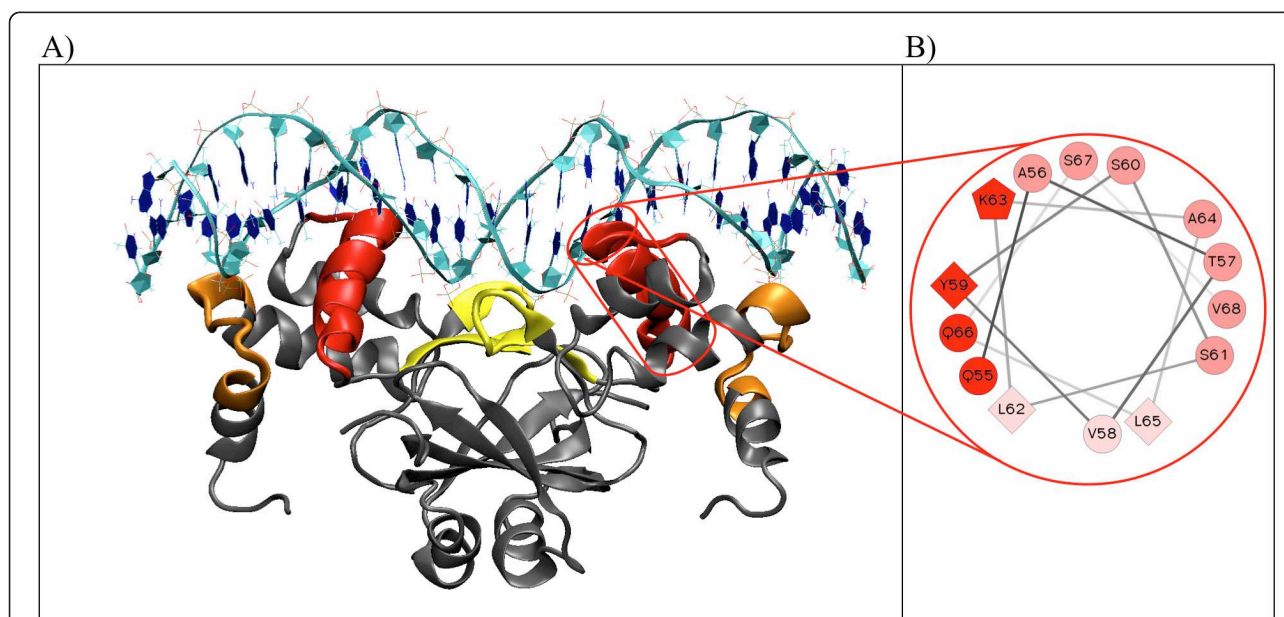


Figure 4 DNA-(Slr1738)₂ complex structure properties. A) 3D structure of DNA-(Slr1738)₂ complex showing six contact regions between the protein (in grey) and the DNA molecule (in cyan). For each monomer, the predicted contact regions are the loop between H_1 and H_2 (in orange), the helix H_4 (in red) and the loop between S_1 and S_2 (in yellow). B) Helical wheel representation of the recognition helix (H_4) of the monomer A. Three faces with different properties are presented. In soft pink are shown the hydrophobic residues turn toward the protein. In dark pink are shown the small residues facing the DNA strand 2 and in red are represented the large and charged residues in interaction with the DNA strand 1. Hydrophilic residues are present as circles, hydrophobic residues as diamonds and potentially positively charged as pentagons <http://r3lab.ucr.edu/scripts/wheel/wheel.cgi>.

DNA contact region seems to be the only one involved in sequence recognition.

(iii) *The loop between S₁ and S₂*. The third region of Slr1738 in contact with DNA contains three negatively-charged glutamic acids (⁷³EVLL^{EE}GC⁸¹) that may interfere with the approach of the S₁ and S₂ anti-parallel strands to the negatively-charged DNA backbone. However, the presence of divalent cations such as Mg²⁺ can bridge interactions between these negatively charged residues (phosphate group and glutamic amino acid) allowing the approach of the S₁ and S₂ anti-parallel strands. In our model, E78 is very well positioned to form such an electrostatic bridge. Furthermore, we know that the iron-binding regulatory site allows proper folding of Slr1738. In particular, its coordination by H36 induces correct contacts between the N-terminal and C-terminal domains. Thus, the modification of this site by oxidation could break the link between the N- and C-terminal domains, thereby preventing the damaged protein from binding DNA [6]. Consequently, we propose that the negatively-charged amino acids of this DNA-binding region participate via strong electrostatic repulsions with the DNA backbone to unlock Slr1738 by inducing a rotation of its N-terminal domain.

During the MD, the three contact zones described above show different behaviours. The first one, the loop between H₁ and H₂, remains stable thanks to the presence of the positively charged residues R18_{AB}, Q22_{AB} and R23_{AB} which establish strong electrostatic interactions with the phosphate atoms of the target DNA that were maintained during the whole simulation. Thus, the dimeric form of Slr1738 makes six non-specific electrostatic interactions that maintain the integrity of the DNA-protein complex. The second region comprising the recognition helix H₄ of each monomer exhibited a non-symmetrical behaviour for each monomer, in agreement with the fact that the H_{4A} and H_{4B} helices interact with non-similar target DNA sequences. In our model structure, only the H_{4A} helix of the monomer A of Slr1738 has its tyrosine residue at a correct distance to the pair of thymines of the target DNA, which are known to interact strongly with tyrosine [33,36]. The alcohol group of the tyrosine is involved in a hydrogen bond with the phosphodiester backbone while its aromatic cycle is facing the methyl groups of the adjacent thymines. Moreover, the recognition helix and the turn preceding it contain five serines and two threonines, the repetition of which might operate in sequence specific recognition. Indeed, we noticed during MD simulations that serine or threonine could contact a thymine residue via a specific dual interaction. First, the close vicinity of the methyl group of thymine and the CH₂ group of serine or the CH₃ group of threonine allows the formation of a long-range hydrophobic interaction. It can also

form a hydrogen bond between the alcohol group of serine or threonine and the ketone group of the thymine. Concerning the third Slr1738-DNA contact region that corresponds to the loop between S₁ and S₂, we observed no significant difference in the mobility of the Slr1738 protein bound or unbound to DNA. The three glutamic acids in this region induce the DNA to move slightly away from the antiparallel strands S₁ and S₂.

Multimer complex hypothesis

The FUR box consensus sequence classically defined as a 19 bp inverted repeat sequence [63] binding a dimeric FUR protein is regarded by some authors as a 15 bp region with a 7-1-7 motif [62] binding a tetrameric (dimer of dimers) FUR. It was also shown that FUR does not bind to PerR boxes though they share with FUR boxes six identical nucleotides within each heptamer. A model of two *Pseudomonas aeruginosa* FUR dimers binding a canonical B-DNA was also proposed by Pohl et al. [20] to take into account that multiple FUR proteins protect a larger DNA region (at least 27-30 bp) than a single FUR dimer (around 20 bp). In this model, the two FUR dimers are located on opposite sides of the DNA molecule. Escolar et al. [64] have reinterpreted the 19 bp consensus FUR binding site as an array of three repeats of the invariable 6 bp GATAAT sequence in *Escherichia coli* while *in vitro* gel shift and DNase footprinting assays led Lavrarr et al. to propose that three FUR dimers (hexamer form) may bind to the 19 bp FUR box [65,66]. The possible occurrence of different forms of FUR and PerR regulators binding with different affinities onto their target genes may explain why some of them are not always co-regulated, depending on the environmental conditions.

Considering these interpretations and our 3D model, we calculated the Slr1738-buried surface of DNA with the NACCESS program [67] and found that the PerR-like regulator Slr1738 likely protects 21 bp of its target DNA, a value close to those mentioned above for FUR/PerR.

Also interestingly, while Slr1738 represses both *slr1738* and *sll1621* in normal conditions (see above), these two genes are not always co-regulated [68] since, for instance, *slr1738* but not *sll1621* is regulated positively by cadmium [14]. Consequently, by analogy with FUR and PerR regulators, we decided to explore the idea that several dimers of Slr1738 could bind to the *sll1621-slr1738* promoter region with different patterns, thereby accounting for the similar or different regulation of the *slr1738* and *sll1621* genes depending on the stress. This idea was reinforced by the observation that Slr1738-like metalloregulators may occur as multimers (dimers, trimers or tetramers) under different redox conditions [69]. Therefore, we built different 3D

structures of the Slr1738/target DNA complex by changing the number of and/or spacing between the Slr1738 dimers bound to the DNA. In practise, we tested all possibilities of DNA-binding of Slr1738 tetramers (dimer of dimers) by fixing the first dimer and moving the second one along the DNA with a one base pair increment so as to steadily increase the spacing between the dimers. This strategy enabled us to (i) structurally validate the different models; (ii) characterize the contact surfaces between each dimer; and (iii) examine more complex multimeric models. As expected, the interaction surface between the DNA and the proteins calculated for our models varies significantly when changing the spacing length between dimers (Additional file 10 Table S3). The DNA overlapped surface globally decreases when the spacing between dimers increases, until it reaches a stabilized value around 3300 Å². Obviously, the hypothetical tetrameric models corresponding to a dimer spacing of 1, 2, 8, 9, 10 or 11 bp are unrealistic because the resulting large surface overlapping between Slr1738 dimers would generate severe steric clashes. Others model combinations without steric clashes can be divided in two groups. The first group comprises the tetramers with no contact surface between its dimers, as occurs in the 5 bp spacer model for the FUR regulator [20], and in models with spacing of 4, 14, 15, 16, 22 bp or more. The second group includes models with a spacing of 3, 6, 7, 12, 13, 18, 19, 20 or 21 bp for which we observe a contact surface between the dimers. With our model of (Slr1738)₂, 21 bp is the maximum offset allowing inter-dimer contacts. Among all these favourable combinations, offsets of 3 and 12 bp show the highest contact surface between dimers, possibly involving a more stable tetrameric complex. However, the 6 bp-spacing model may have biological relevance, though the contact surface between dimers is smaller than in other constructions. This particular model, where the protein tetramer contacts both faces of the DNA helix, is fully compatible with the 7-1-7 inverted repeats of the *Escherichia coli* FUR target DNA, as well as the DNase I footprinting and gel shift evidence of the existence of an overlapping between *Escherichia coli* FUR dimers bound to DNA [65,66].

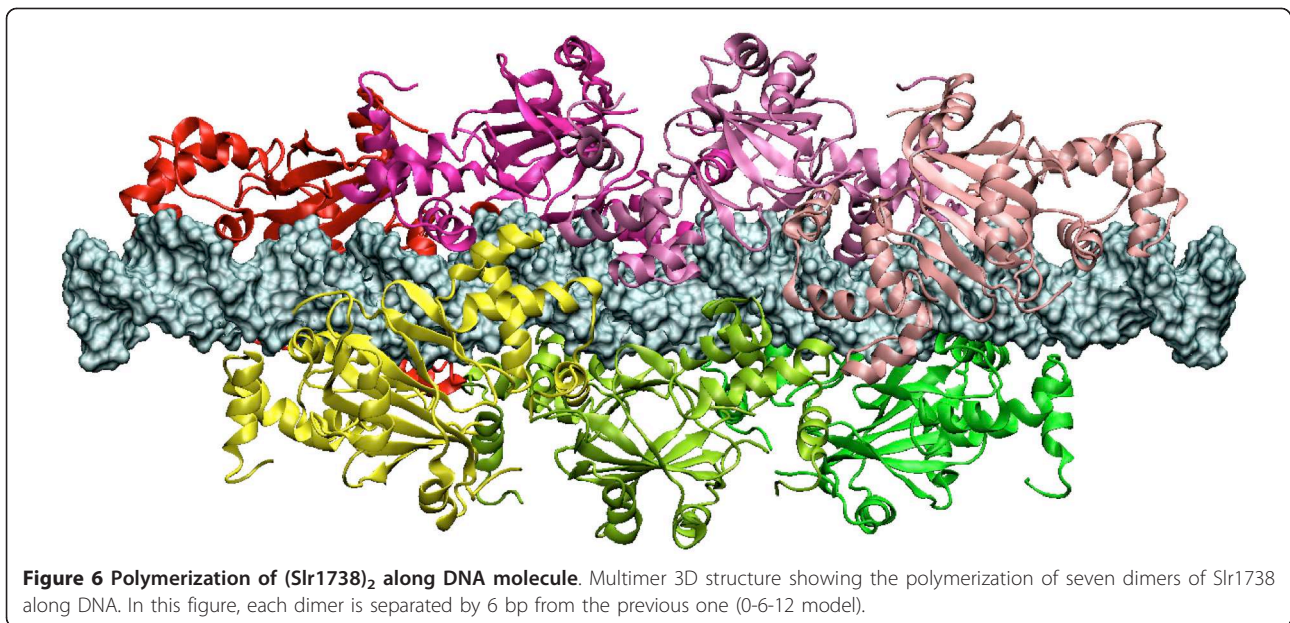
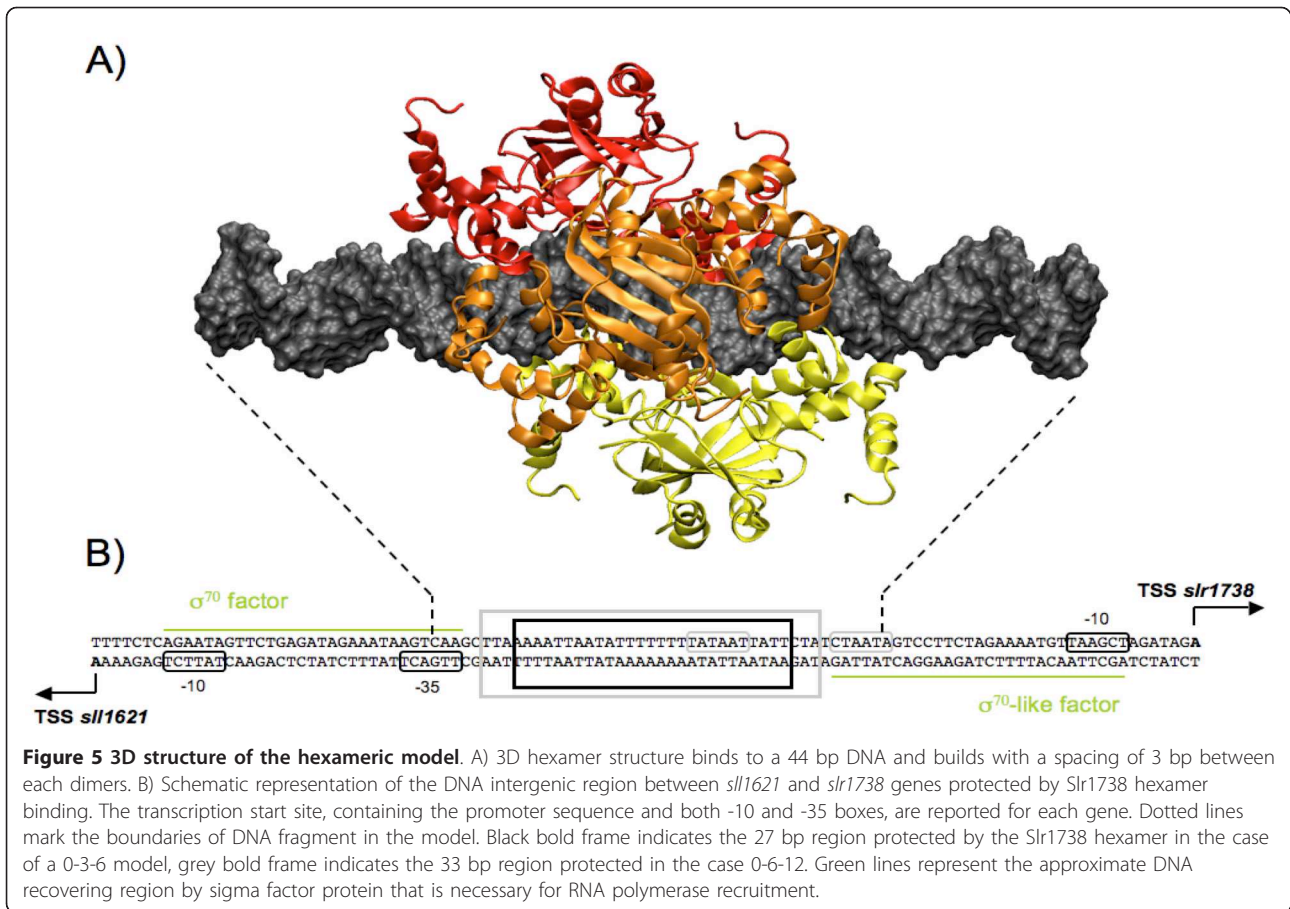
We also constructed more complex structural systems describing higher order multimeric regulators *i.e.* hexamers and more, and found two structures that may account for some biological observations. The first multimer structure is a hexamer composed of three dimers with a spacing of either 3 bp (0-3-6 model) or 6 bp (0-6-12 model). These two models allow the formation of a hexamer-DNA complex with no steric clash, which may operate in the co-regulation of the two genes *slr1738* and *sll1621* as shown for the 3 bp-spacing model in Figure 5. Detailed structural interactions between each

dimer in these hexamer models are shown in Additional file 11 Figure S8. Schematic representation of the *sll1621-slr1738* promoter region in these models predict that the binding of Slr1738 will preclude the binding of sigma factors, thereby impairing the transcription of the *slr1738* and *sll1621* genes. Furthermore, in this switch mechanism the two Slr1738 dimers bordering the hexameric regulator might be alternatively released in some environmental conditions thereby allowing the specific transcription of either *sll1621* or *slr1738*. The second particular multimeric structure, possibly occurring in response to a high concentration of the FUR regulator [65,70], involves polymerization of multiple FUR proteins along their target DNA molecule which is thereafter no longer accessible to other enzymes. Such polymerization is only possible when a 6 bp-spacing (0-6-12 model) occurs between the FUR dimers (Figure 6). It could explain the structural organization of FUR-binding sites with repeated DNA sequence (GATAAT-GATAAT)_n. By contrast, in the 0-3-6 hexameric model, this polymerization is not possible because a spacing of about 21 bp is necessary to allow the binding of the next hexamer onto the DNA helix.

Conclusions

In the current post-genomic era the thousands of 3D protein structures available at the PDB database can be used as template to predict the possible fold of structurally uncharacterized proteins of biological interest. This task is important for mind-oriented analysis of the interactions between these proteins and their DNA and/or protein partners. In this frame, we used a combination of different types of data (structural, biochemical and genetic) to build and validate a relevant model of the tridimensional structure of a biologically important protein-DNA complex. This complex plays a central role in the regulation of cyanobacteria (environmentally crucial organisms) by interdependent oxidative and metal stresses. It is formed between the PerR/FUR-like Slr1738 regulator and its main DNA target, *i.e.* the promoter region of the divergent genes *slr1738* and the peroxiredoxin (anti-oxidant) encoding gene *sll1621*. The detailed analysis of the Slr1738/DNA complex, and the related complex between genuine PerR/FUR-type regulators and their target DNA allowed us to better understand the interactions involved in the protein-DNA recognition and the stability/integrity of the structure.

The method we propose in this work to build the (Slr1738)₂/DNA complex, that is based on fold-level similarity between DNA binding domains to transfer DNA orientation from a co-complex structure to a protein-only model, needs to be formalized and automated to DNA to be applied by other researchers to build other protein/DNA complexes of interest. We also



develop a simple theoretical strategy to predict more complex structures involving the binding of multiple dimers of regulators to the same molecule of DNA. In this strategy, we consider each dimer as a molecular building block that can be moved along the target DNA molecule to hug it, and combine the resulting potential structure with footprinting and gel shift data to propose attractive model structures. One of these, a switch model, may explain a biological mechanism *i.e.* the presence/absence of coordinated expression of the genes co-regulated by the same Slr1738/FUR/PerR-type regulator, depending on the environmental conditions. We believe that our strategy for studying protein/DNA interactions will help to decipher the molecular basis of a wealth of regulatory mechanisms that are crucial for life.

Additional material

Additional file 1: Table S1. Parameters of experimental RX structures used for the construction of the SLR-DNA complex.

Additional file 2: Figure S1. Sequence alignment of the HTHw motif.

Additional file 3: Figure S2. Schematic representation of two different protein-DNA enclosures involving a helix-turn-helix recognition motif.

Additional file 4: Table S2. DNA-protein complexes parameters of the structures built with DNA template sequence.

Additional file 5: Figure S3. Sequence of *ahpC* promoter region in cyanobacteria harbouring a *Synechocystis*-like *ahpC*-Slr1738 intergenic organisation.

Additional file 6: Figure S4. Determination of the transcription start site for the opposite genes *slr1738* and *slr11621* with the 5'-RACE technique.

Additional file 7: Figure S5. Amino acids sequence alignments of the *Synechocystis* regulator Slr1738 with the protein from several cyanobacteria.

Additional file 8: Figure S6. The different interactions that participate to the stabilization of the dimer interface of Slr1738.

Additional file 9: Figure S7. FUR binding site motifs proposed in literature.

Additional file 10: Table S3. Tetramer construction possibilities.

Additional file 11: Figure S8. Interactions description in hexameric models.

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Authors' contributions

YB coordinated the project. YB, OD, FC and CC conceived and designed the experiments. PG and OD performed docking and analyzed the data. JYZ and CC carried out the molecular biology experiments. YB, OD and FC wrote the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Pabo CO, Nekludova L: Geometric analysis and comparison of protein-DNA interfaces: why is there no simple code for recognition? *J Mol Biol* 2000, **301**:597-624.
2. Ritchie DW: Recent progress and future directions in protein-protein docking. *Curr Protein Pept Sci* 2008, **9**:1-15.
3. Hantke K: Iron and metal regulation in bacteria. *Curr Opin Microbiol* 2001, **4**:172-177.
4. Lee J-W, Helmann JD: Functional specialization within the Fur family of metalloregulators. *Biomol* 2007, **20**:485-499.
5. Duarte V, Latour J-M: PerR vs OhrR: selective peroxide sensing in *Bacillus subtilis*. *Mol Biosyst* 2010, **6**:316-323.
6. Lee JW, Helmann JD: The PerR transcription factor senses H₂O₂ by metal-catalysed histidine oxidation. *Nature* 2006, **440**:363-367.
7. Traore DAK, El Ghazouani A, Ilango S, Dupuy J, Jacquamet L, Ferrer J-L, Caux-Thang C, Duarte V, Latour J-M: Crystal structure of the apo-PerR-Zn protein from *Bacillus subtilis*. *Mol Microbiol* 2006, **61**:1211-1219.
8. Jacquamet L, Traore DAK, Ferrer JL, Proux O, Testemale D, Hazemann JL, Nazarenko E, El Ghazouani A, Caux-Thang C, Duarte V, Latour JM: Structural characterization of the active form of PerR: insights into the metal-induced activation of PerR and Fur proteins for DNA binding. *Mol Microbiol* 2009, **73**:20-31.
9. Dismukes GC, Klimov VV, Baranov SV, Kozlov YN, DasGupta J, Tyrshkin A: The origin of atmospheric oxygen on Earth: the innovation of oxygenic photosynthesis. *Proc Natl Acad Sci USA* 2001, **98**:2170-2175.
10. Abed RMM, Dobretsov S, Sudesh K: Applications of cyanobacteria in biotechnology. *J Appl Microbiol* 2009, **106**:1-12.
11. Dismukes GC, Carrieri D, Bennete N, Ananyev GM, Posewitz MC: Aquatic phototrophs: efficient alternatives to land-based crops for biofuels. *Curr Opin Biotechnol* 2008, **19**:235-240.
12. Williams PG: Panning for chemical gold: marine bacteria as a source of new therapeutics. *Trends Biotechnol* 2009, **27**:45-52.
13. Latifi A, Ruiz M, Zhang C-C: Oxidative stress in cyanobacteria. *FEMS Microbiol Rev* 2009, **33**:258-278.
14. Houot L, Floutier M, Marteyn B, Michaut M, Piccicocchi A, Legrain P, Aude J-C, Cassier-Chauvat C, Chauvat F: Cadmium triggers an integrated reprogramming of the metabolism of *Synechocystis* PCC6803, under the control of the Slr1738 regulator. *BMC Genomics* 2007, **8**:350.
15. Kobayashi M, Ishizuka T, Katayama M, Kanehisa M, Bhattacharyya-Pakrasi M, Pakrasi HB, Ikeuchi M: Response to oxidative stress involves a novel peroxiredoxin gene in the unicellular cyanobacterium *Synechocystis* sp. PCC 6803. *Plant Cell Physiol* 2004, **45**:290-299.
16. Li H, Singh AK, McIntyre LM, Sherman LA: Differential gene expression in response to hydrogen peroxide and the putative PerR regulon of *Synechocystis* sp. strain PCC 6803. *J Bacteriol* 2004, **186**:3331-3345.
17. Singh AK, Elvitigala T, Cameron JC, Ghosh BK, Bhattacharyya-Pakrasi M, Pakrasi HB: Integrative analysis of large scale expression profiles reveals core transcriptional response and coordination between multiple cellular processes in a cyanobacterium. *BMC Syst Biol* 2010, **4**:105.
18. Marraccini P, Bulteau S, Cassier-Chauvat C, Mermat-Bouvier P, Chauvat F: A conjugative plasmid vector for promoter analysis in several cyanobacteria of the genera *Synechococcus* and *Synechocystis*. *Plant Mol Biol* 1993, **23**:905-909.
19. Mazouni K, Bulteau S, Cassier-Chauvat C, Chauvat F: Promoter element spacing controls basal expression and light inducibility of the cyanobacterial *secA* gene. *Mol Microbiol* 1998, **30**:1113-1122.
20. Pohl E, Haller JC, Mijovilovich A, Meyer-Klaucke W, Garman E, Vasil ML: Architecture of a protein central to iron homeostasis: crystal structure and spectroscopic analysis of the ferric uptake regulator. *Mol Microbiol* 2003, **47**:903-915.
21. Stote RH, Karplus M: Zinc binding in proteins and solution: a simple but accurate nonbonded representation. *Proteins* 1995, **23**:12-31.
22. Blindauer CA, Harrison MD, Parkinson JA, Robinson AK, Cavet JS, Robinson NJ, Sadler PJ: A metallothionein containing a zinc finger within

- a four-metal cluster protects a bacterium from zinc toxicity. *Proc Natl Acad Sci USA* 2001, **98**:9593-9598.
23. Blindauer CA, Harrison MD, Parkinson JA, Robinson AK, Cavet JS, Robinson NJ, Sadler PJ: A metallothionein containing a zinc finger within a four-metal cluster protects a bacterium from zinc toxicity. *Proc Natl Acad Sci USA* 2001, **98**:9593-9598.
 24. Bagg A, Neilands JB: Ferric uptake regulation protein acts as a repressor, employing iron (II) as a cofactor to bind the operator of an iron transport operon in *Escherichia coli*. *Biochemistry* 1987, **26**:5471-5477.
 25. Case DA, Darden TA, Cheatham TE III, Simmerling CL, Wang J, Duke RE, Luo R, Merz KM, Pearlman DA, Crowley M, Walker RC, Zhang W, Wang B, Hayik S, Roitberg A, Seabra G, Wong KF, Paesani F, Wu X, Brozell S, Tsui V, Gohlke H, Yang L, Tan C, Mongan J, Hornak V, Cui G, Beroza P, Matthews DH, Schafmeister C, Ross WS, Kollman PA: *AMBER 9* University of California, San Francisco; 2006.
 26. Kollman PA, Massova I, Reyes C, Kuhn B, Huo S, Chong L, Lee M, Lee T, Duan Y, Wang W, Donini O, Cieplak P, Srinivasan J, Case DA, Cheatham TE III: Calculating structures and free energies of complex molecules: combining molecular mechanics and continuum models. *Acc Chem Res* 2000, **33**:889-897.
 27. Gohlke H, Case DA, Converging free energy estimates: MM-PB(GB)SA studies on the protein-protein complex Ras-Raf. *J Comput Chem* 2004, **25**:238-250.
 28. Masukawa KM, Kollman PA, Kuntz ID: Investigation of neuraminidase-substrate recognition using molecular dynamics and free energy calculations. *J Med Chem* 2003, **46**:5628-5637.
 29. Wang J, Morin P, Wang W, Kollman PA: Use of MM-PBSA in reproducing the binding free energies to HIV-1 RT of TIBO derivatives and predicting the binding mode to HIV-1 RT of efavirenz by docking and MM-PBSA. *J Am Chem Soc* 2001, **123**:5221-5230.
 30. Sanner MF, Olson AJ, Spehner JC: Reduced surface: an efficient way to compute molecular surfaces. *Biopolymers* 1996, **38**:305-320.
 31. Pellegrini-Calace M, Thornton JM: Detecting DNA-binding helix-turn-helix structural motifs using sequence and structure information. *Nucleic Acids Res* 2005, **33**:2129-2140.
 32. Garcia-Castellanos R, Mallorquí-Fernandez G, Marrero A, Potempa J, Coll M, Gomis-Ruth FX: On the transcriptional regulation of methicillin resistance: Mecl repressor in complex with its operator. *J Biol Chem* 2004, **279**:17888-17896.
 33. Hong M, Fuangthong M, Helmann JD, Brennan RG: Structure of an OhrR-ohrA operator complex reveals the DNA binding mechanism of the MarR family. *Mol Cell* 2005, **20**:131-141.
 34. Pohl E, Holmes RK, Hol WG: Crystal structure of a cobalt-activated diphtheria toxin repressor-DNA complex reveals a metal-binding SH3-like domain. *J Mol Biol* 1999, **292**:653-667.
 35. Wisedchaisri G, Holmes RK, Hol WG: Crystal structure of an IdeR-DNA complex reveals a conformational change in activated IdeR for base-specific interactions. *J Mol Biol* 2004, **342**:1155-1169.
 36. Tiss A, Barre O, Michaud-Soret I, Forest E: Characterization of the DNA-binding site in the ferric uptake regulator protein from *Escherichia coli* by UV crosslinking and mass spectrometry. *FEBS Lett* 2005, **579**:5454-5460.
 37. Kaneko T, Sato S, Kotani H, Tanaka A, Asamizu E, Nakamura Y, Miyajima N, Hirotsawa M, Sugiura M, Sasamoto S, Kimura T, Hosouchi T, Matsuno A, Muraki A, Nakazaki N, Naruo K, Okumura S, Shimpo S, Takeuchi C, Wada T, Watanabe A, Yamada M, Yasuda M, Tabata S: Sequence analysis of the genome of the unicellular cyanobacterium *Synechocystis* sp. strain PCC6803. II. Sequence determination of the entire genome and assignment of potential protein-coding regions. *DNA Res* 1996, **3**:185-209.
 38. Marraccini P, Bulteau S, Cassier-Chauvat C, Mermet-Bouvier P, Chauvat F: A conjugative plasmid vector for promoter analysis in several cyanobacteria of the genera *Synechococcus* and *Synechocystis*. *Plant Mol Biol* 1993, **23**:905-909.
 39. Domain F, Houot L, Chauvat F, Cassier-Chauvat C: Function and regulation of the cyanobacterial genes *lexA*, *recA* and *ruvB*: LexA is critical to the survival of cells facing inorganic carbon starvation. *Mol Microbiol* 2004, **53**:65-80.
 40. Gonzalez A, Bes MT, Peleato ML, Fillat MF: Unravelling the regulatory function of FurA in *Anabaena* sp. PCC 7120 through 2-D DIGE proteomic analysis. *J Proteomics* 2011, **74**:660-671.
 41. Pribnow D: Nucleotide sequence of an RNA polymerase binding site at an early T7 promoter. *Proc Natl Acad Sci USA* 1975, **72**:784-788.
 42. Scotto-Lavino E, Du G, Frohman MA: Amplification of 5' end cDNA with 'new RACE'. *Nat Protoc* 2006, **1**:3056-3061.
 43. Klahn S, Hohne A, Simon E, Hagemann M: The gene *ssl3076* encodes a protein mediating the salt-induced expression of *gppS* for the biosynthesis of the compatible solute glucosylglycerol in *Synechocystis* sp. strain PCC 6803. *J Bacteriol* 2010, **192**:4403-4412.
 44. Figge RM, Cassier-Chauvat C, Chauvat F, Cerff R: The carbon metabolism-controlled *Synechocystis* *gap2* gene harbours a conserved enhancer element and a Gram-positive-like -16 promoter box retained in some chloroplast genes. *Mol Microbiol* 2000, **36**:44-54.
 45. Mazouni K, Domain F, Chauvat F, Cassier-Chauvat C: Expression and regulation of the crucial plant-like ferredoxin of cyanobacteria. *Mol Microbiol* 2003, **49**:1019-1029.
 46. Michel KP, Pistorius EK, Golden SS: Unusual regulatory elements for iron deficiency induction of the *idIA* gene of *Synechococcus elongatus* PCC 7942. *J Bacteriol* 2001, **183**:5015-5024.
 47. Records MT, Reznikoff WS, Craig ML, McQuade KL, Schlx PJ: *Escherichia coli* and *Salmonella*. In *Neidhardt*. Edited by: Ingraham FC, Lin JL, Magasanik ECC, Reznikoff B, Riley WS, Schaechter M, Umberger HE. American Society for Microbiology, Washington DC; 1996:792-820.
 48. Barne KA, Bown JA, Busby SJ, Minchin SD: Region 2.5 of the *Escherichia coli* RNA polymerase sigma70 subunit is responsible for the recognition of the 'extended-10' motif at promoters. *Embo J* 1997, **16**:4034-4040.
 49. Pecqueur L, D'Autreaux B, Dupuy J, Nicolet Y, Jacquamet L, Brutscher B, Michaud-Soret I, Bersch B: Structural changes of *Escherichia coli* ferric uptake regulator during metal-dependent dimerization and activation explored by NMR and X-ray crystallography. *J Biol Chem* 2006, **281**:21286-21295.
 50. Brennan RG, Matthews BW: The helix-turn-helix DNA binding motif. *J Biol Chem* 1989, **264**:1903-1906.
 51. Donaldson LW, Petersen JM, McIntosh LP: Secondary structure of the ETS domain places murine Ets-1 in the superfamily of winged helix-turn-helix DNA-binding proteins. *Biochemistry* 1994, **33**:13509-13516.
 52. Coy M, Neilands JB: Structural dynamics and functional domains of the fur protein. *Biochemistry* 1991, **30**:8201-8210.
 53. Michaud-Soret I, Adrait A, Jaquinod M, Forest E, Touati D, Latour JM: Electrospray ionization mass spectrometry analysis of the apo- and metal-substituted forms of the Fur protein. *FEBS Lett* 1997, **413**:473-476.
 54. Fuangthong M, Helmann JD: Recognition of DNA by three ferric uptake regulator (Fur) homologs in *Bacillus subtilis*. *J Bacteriol* 2003, **185**:6348-6357.
 55. Cognet JAH, Boulard Y, Fazakerley GV: Helical parameters, fluctuations, alternative hydrogen bonding, and bending in oligonucleotides containing a mismatched base-pair by NOESY distance restrained and distance free molecular dynamics. *Journal of Molecular Biology* 1995, **246**:209-226.
 56. Roll C, Ketterlé C, Faibis V, Fazakerley GV, Boulard Y: Conformations of nicked and gapped DNA structures by NMR and molecular dynamic simulations in water. *Biochemistry* 1998, **37**:4059-4070.
 57. Maufrais C, Fazakerley GV, Guy A, Cadet J, Boulard Y: Solution structures of an oligonucleotide containing a N-(2'-deoxy-β-D-erythropentofuranosyl) formamide-guanine base pair by NMR and molecular dynamics studies. *Biochemistry* 2000, **39**:5614-5621.
 58. Maufrais C, Fazakerley GV, Cadet J, Boulard Y: Structural study of DNA duplex containing a N-(2'-deoxy-β-D-erythropentofuranosyl)-formamide frameshifts by NMR and restrained molecular dynamics. *Nucleic Acids Research* 2003, **31**:1-11.
 59. Alvarez Salgado F, Desvaux H, Boulard Y: NMR Assessment of the global shape of a non-labeled DNA dodecamer containing a tandem of G-T mismatches. *Magnetic Resonance in Chemistry* 2006, **44**:1081-1089.
 60. Svozil D, Kalina J, Omelka M, Schneider B: DNA conformations and their sequence preferences. *Nucleic Acids Res* 2008, **36**:3690-3706.
 61. Lu X-J, Olson WK: 3DNA: a software package for the analysis, rebuilding and visualization of the three-dimensional nucleic acid structures. *Nucleic Acids Research* 2003, **31**:5108-5121.
 62. Baichoo N, Helmann JD: Recognition of DNA by Fur: a reinterpretation of the Fur box consensus sequence. *J Bacteriol* 2002, **184**:5826-5832.
 63. de Lorenzo V, Wee S, Herrero M, Neilands JB: Operator sequences of the aerobactin operon of plasmid ColV-K30 binding the ferric uptake regulation (*fur*) repressor. *J Bacteriol* 1987, **169**:2624-2630.

64. Escolar L, Perez-Martin J, de Lorenzo V: **Binding of the fur (ferric uptake regulator) repressor of Escherichia coli to arrays of the GATAAT sequence.** *J Mol Biol* 1998, **283**:537-547.
65. Lavrrar JL, Christoffersen CA, McIntosh MA: **Fur-DNA interactions at the bidirectional *fepDGC-entS* promoter region in Escherichia coli.** *J Mol Biol* 2002, **322**:983-995.
66. Lavrrar JL, McIntosh MA: **Architecture of a fur binding site: a comparative analysis.** *J Bacteriol* 2003, **185**:2194-2202.
67. Hubbard SJ, Thornton JM: *NACCESS* 1993.
68. Singh AK, Elvitigala T, Cameron JC, Ghosh BK, Bhattacharyya-Pakrasi M, Pakrasi HB: **Integrative analysis of large scale expression profiles reveals core transcriptional response and coordination between multiple cellular processes in a cyanobacterium.** *BMC Syst Biol* 2010, **4**:105.
69. Lostao A, Peleato ML, Gomez-Moreno C, Fillat MF: **Oligomerization properties of FurA from cyanobacterium *Anabaena* sp. 7120: Direct visualization by *in situ* atomic force microscopy under different redox conditions.** *Biochim Biophys Acta* 2010, **1804**:1723-1729.
70. Escolar L, Perez-Martin J, de Lorenzo V: **Evidence of an unusually long operator for the fur repressor in the aerobactin promoter of Escherichia coli.** *J Biol Chem* 2000, **275**:24709-24714.

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