



Review

Immunosenescence in multiple sclerosis: the identification of new therapeutic targets

María Dema^{a,b,c}, Herena Eixarch^{a,b,c}, Luisa M. Villar^{c,d}, Xavier Montalban^{a,b,c},
Carmen Espejo^{a,b,c,*}

^a Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya, Vall d'Hebron Institut de Recerca, Hospital Universitari Vall d'Hebron, Vall d'Hebron Barcelona Hospital Campus, 08035, Barcelona, Spain

^b Universitat Autònoma de Barcelona, 08193, Bellaterra, Cerdanyola del Vallès, Spain

^c Red Española de Esclerosis Múltiple (REEM), Spain

^d Servicio de Inmunología, Hospital Universitario Ramón y Cajal de Investigación Sanitaria (IRYCIS), 28034, Madrid, Spain



ARTICLE INFO

Keywords:

Immunosenescence
Ageing
Autoimmunity
Multiple sclerosis

ABSTRACT

The number of elderly multiple sclerosis (MS) patients is growing, mainly due to the increase in the life expectancy of the general population and the availability of effective disease-modifying treatments. However, current treatments reduce the frequency of relapses and slow the progression of the disease, but they cannot stop the disability accumulation associated with disease progression. One possible explanation is the impact of immunosenescence, which is associated with the accumulation of unusual immune cell subsets that are thought to have a role in the development of an early ageing process in autoimmunity. Here, we provide a recent overview of how senescence affects immune cell function and how it is involved in the pathogenesis of autoimmune diseases, particularly MS. Numerous studies have demonstrated age-related immune changes in experimental autoimmune encephalomyelitis models, and the premature onset of immunosenescence has been demonstrated in MS patients. Therefore, potential therapeutic strategies based on rejuvenating the immune system have been proposed. Senolytics and regenerative strategies using haematopoietic stem cells, therapies based on rejuvenating oligodendrocyte precursor cells, microglia and monocytes, thymus cells and senescent B and T cells are capable of reversing the process of immunosenescence and could have a beneficial impact on the progression of MS.

1. Introduction

The world's population is rapidly ageing. By 2050, it is estimated that 22% of people will be over 60 years old [1]. Ageing is broadly defined as a progressive functional decline that leads to impaired biological functions, which are the main cause of multiple human pathologies, such as cancer, diabetes, cardiovascular disorders and neurodegenerative diseases. Several hallmarks define the process of ageing, including genomic instability, telomere attrition, epigenetic alterations, the loss of proteostasis, deregulated nutrient sensing, mitochondrial dysfunction, stem cell exhaustion, altered intercellular communication and cellular senescence [2]. The age-related changes that take place in the immune system, a process known as

immunosenescence, generally result in a higher susceptibility to infections, a reduced response to vaccines [3] and a higher prevalence of autoimmunity [4] and neurodegenerative disorders [5]. The immunosenescence process is the consequence of a series of events that affect the differentiation and maturation processes of different immune cell subtypes as well as their functionality.

Immunosenescence is defined by a set of immune markers known as the immune risk phenotype, which includes the inversion of the CD4:CD8 ratio, the expansion of CD8⁺CD28⁻ T cells, the presence of cytomegalovirus (CMV) seropositivity, poor T cell proliferation and low B cell numbers. In fact, immune risk phenotype features are associated with increased morbidity and mortality in elderly individuals [6]. In addition, recent studies have revealed that the ageing process is also

* Corresponding author at: Servei de Neurologia-Neuroimmunologia, Centre d'Esclerosi Múltiple de Catalunya, Vall d'Hebron Institut de Recerca, Hospital Universitari Vall d'Hebron, Pg. Vall d'Hebron 119-129 (Ed. Collserola, Lab. 149), 08035, Barcelona, Spain

E-mail addresses: maria.dema@vhir.org (M. Dema), herena.eixarch@vhir.org (H. Eixarch), luisamaria.villar@salud.madrid.org (L.M. Villar), xavier.montalban@cem-cat.org (X. Montalban), carmen.espejo@vhir.org (C. Espejo).

<https://doi.org/10.1016/j.autrev.2021.102893>

Received 25 April 2021; Accepted 2 May 2021

Available online 5 July 2021

1568-9972/© 2021 The Author(s).

Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license

(<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

associated with an increased activity of immunosuppressive cells such as regulatory T (Treg) cells, regulatory B (Breg) cells and myeloid-derived suppressor cells (MDSCs) [7]. Numerous changes have already been described to occur in the adaptive immune system during immunosenescence; however, fewer studies have been performed to determine the impact of immunosenescence on innate immunity. Innate immunity seems to be more preserved with age, although recent reports point to age-associated impaired functionality of innate immune cells [8].

Multiple sclerosis (MS) is a chronic neurodegenerative, inflammatory and demyelinating disease of the central nervous system (CNS) characterized by the formation of demyelinating lesions. Although the immunopathological events occurring in the initial stages of the disease are not yet well understood, it is clear that the immune system is involved in the destruction of myelin sheaths that ultimately leads to neurological disability in patients. Taking this into account, many experts point to an autoimmune origin of MS [9]. Several studies suggest that autoimmunity itself as well as MS progression could be associated with premature immunosenescence. Thus, an understanding of the mechanisms underlying immunosenescence is a challenging objective in the field of MS to determine the possible efficacy of already approved therapies that have the capacity to reverse the process of immunosenescence. This review gives an overview of recent research findings on immunosenescence features and links the events that cause immunosenescence with MS immunopathogenesis. Finally, the possible beneficial impact of currently approved therapies aiming to rejuvenate the immune system on MS progression is discussed.

Table 1
Age-related alterations in innate and adaptive immunity.

Immune cell type	Age-related alterations	References
T cells	↓ naïve cells, ↑ memory cells	[14–17]
	↓ TCR repertoire diversity, ↓ TRECs	[11–13]
	↓ CD28 co-stimulatory receptor, ↓ CD40L, ↓ helper ability	[19,23,24]
	↑ TIGIT and PD1 inhibitory receptors	[20]
	↑ CD57 and KLRG-1 senescent markers	[17]
	↑ SASP cytokines	[21]
	↑ NK receptors, ↑ cytotoxic response	[28–30]
B cells	↑ autoreactivity	[22,23]
	↓ naïve cells, ↑ memory cells	[34]
	↓ BCR repertoire diversity	[33]
	↓ class switch, ↓ AID enzyme	[37,38]
	↑ autoantibodies	[40]
Monocytes/ macrophages	↑ ABCs	[39,40]
	↑ inflammation recruitment	[36]
	↑ M2 macrophages	[45]
	↓ MHC-II receptor, ↓ TLR receptors, ↑ TAM receptors	[44,46,47]
DCs	↓ antiviral response, ↓ inflammation recruitment	[43]
	↓ DC responsiveness	[50]
	↑ ROS, ↓ phagocytosis, ↓ cross-presentation	[48]
NK cells	↓ stimulation of CD8 ⁺ T cells, ↓ IFN- γ	[49]
	↓ CD56 ^{bright} immunoregulatory cells, ↑ CD56 ^{dim} cytotoxic cells	[51,52]
	↓ mature NK cells in periphery	[53]
Neutrophils	↓ degranulation capacity	[52]
	↓ ROS, ↓ NETs	[54]
	↓ phagocytosis	[55,56]
	↓ inflammation recruitment	[56]

Abbreviations: T cell receptor (TCR); T cell receptor excision circle (TREC); Tyrosine-based inhibitory motif domain (TIGIT); Senescence-associated secretory profile (SASP); B cell receptor (BCR); Activation-induced cytidine deaminase (AID); Age-associated B cell (ABC); Major histocompatibility complex (MHC); Toll-like receptor (TLR); Tyro3, Axl, and Mer (TAM); Dendritic cell (DC); Reactive oxygen species (ROS); Natural killer (NK); Neutrophil extracellular trap (NET).

2. Ageing of the immune system: immunosenescence

The events that take place in the aged immune system result in poor immune responses in older adults (Table 1). Indeed, these events affect not only the functionality of immune cells but also the number and frequency of certain immune cells due to decreased haematopoiesis and progressive atrophy of the thymus. Immune cells are generated in the bone marrow from haematopoietic stem cells (HSCs), which are able to self-renew and differentiate into any of the lineages of the immune system. The ageing of HSCs is characterized by impaired telomerase activity, which leads to telomere shortening with each division; as a consequence, the pool of HSCs is progressively reduced due to decreased self-renewal capacity. Moreover, immunosenescence bears other consequences involving the commitment of progenitors, since myeloid differentiation is potentiated over lymphoid differentiation [10].

2.1. Adaptive immunity

T cell changes are the most described in the immunosenescence process. One of the main consequences of ageing is the progressive involution of the thymus, which deeply affects the maturation of naïve T cells and the diversity of the T cell receptor (TCR) repertoire [11,12]. These two features bear important consequences in T cell-mediated immunity. Moreover, the involution of the thymus also involves a decrease in the detection of T cell receptor excision circles (TRECs), which are circular DNA products generated from the rearrangement of TCR genes in the thymus. TRECs are detected only in naïve T cells, so their content has been suggested as a biomarker of the naïve T cell pool state and thymic function in immune ageing [13]. Although the overall T cell population is maintained in older adults, the number of naïve T cells in the periphery is reduced (especially the number of CD8⁺ T cells) [14,15]. Terminally differentiated memory T cells (especially CD8⁺ T cells) are expanded and accumulate due to persistent infections, most commonly with CMV [16,17]. As a consequence of the lower number of naïve T cells and their decreased diversity in the TCR repertoire, the normal function of the T cell-mediated immune response is also impaired, presenting a decreased ability to respond properly to infections and vaccination [18]. Senescent cells lack the CD28 co-stimulatory molecule and express CD57 and KLRG1 senescence markers. This phenotype is commonly associated with terminally differentiated memory CD8⁺ T cells in elderly individuals [17]. Terminally differentiated memory CD8⁺ T cells that tend to accumulate with age present an impaired capacity to proliferate, more resistance to undergo apoptosis and shortened telomeres, which are typical features of cellular senescence [19]. There is evidence that senescent CD8⁺ T cells also express inhibitory receptors such as T cell immunoglobulin and immunoreceptor tyrosine-based inhibitory motif domain (TIGIT) and PD1, which are characteristic of not only senescent T cells but also exhausted T cells [20]. Although these terminally differentiated memory CD8⁺ T cells could fight against persistent infections in older adults [17], senescent T cells are also characterized by the secretion of a determinate cytokine profile termed the senescence-associated secretory profile (SASP) [21], which could contribute to tissue damage.

Although major changes are observed in the CD8⁺ T cell compartment, CD4⁺ T cells also experience modifications with age, but to a lesser extent. Terminally differentiated CD4⁺ T cells present senescence features that are similar to those of terminally differentiated CD8⁺ T cells, since they also accumulate with age due to persistent antigen stimulation with pathogens or self-antigens, present shortened telomeres, produce a high number of pro-inflammatory cytokines, express CD57 and KLRG1 senescence markers and lose CD28 co-stimulatory molecule. Particularly, in senescent CD4⁺ T cells, there is an increased number of autoreactive cells [19,22,23]. Furthermore, the loss of CD28 leads to a deficiency in the induced expression of CD40L, impeding CD4⁺ T cells from interacting with CD40 molecules present on the surface of B cells; consequently, CD4⁺ T cells lose their helper ability to

induce B cell proliferation and the production of antibodies. Moreover, senescent CD4⁺ T cells lose their helper ability to induce somatic hypermutation on B cells and the formation of germinal centres, which is a T cell-dependent process that negatively interferes with the humoral immune response in older adults [23,24].

It is well reported that a subset of CD8⁺ T cells constitutively express the NK-associated receptor NKG2D in humans and upon stimulation in mice, as NKG2D serves as an activating receptor in cytotoxic cells [25]. However, CD8⁺ T cells with a senescent-like phenotype, lacking CD28 expression, accumulate NK receptors with age [26]. Despite their inability to respond to TCR-mediated signals, CD8⁺ senescent T cells mediate cytotoxic immune responses through NKG2D and other NK receptors instead. The switch from TCR- to NK-receptor-mediated cytotoxicity probably increases their capacity for immune surveillance and the elimination of senescent cells in aged tissues as well as tumour cells [27]. In contrast, CD4⁺ CD28⁻ T cells express NK receptors *de novo*, and these cells are expanded in certain pathological conditions, including acute coronary syndrome [28] and rheumatoid arthritis [29]. In aged individuals, CD4⁺ CD28⁻ T cells with senescent features that specifically express the NKG2D receptor are expanded and, moreover, have been suggested as a differentiation marker in the CD4⁺ T cell compartment in elderly individuals [30]. Innate-like properties in senescent T cells confer beneficial adaptation mechanisms to mediate faster effector functions, suggesting that the limits between the innate and adaptive immune systems may not be as distinct as was first thought [31].

Numbers of peripheral B cells are not deemed with age, but many changes are observed in the composition of the different compartments and in the functional features of B cell subtypes, mainly due to a deficit in CD4⁺ T helper cells, as mentioned above. The production of B cells in the bone marrow is decreased in elderly individuals [32], as well as the diversity of the B cell receptor (BCR) repertoire in the bone marrow, lymph nodes and peripheral blood [33]. Moreover, a shift from naïve to memory B cells has been described, which is consistent with the decrease in IgD levels observed [34]. Late memory B cell numbers are increased in older people and express markers associated with migration to sites of inflammation [35,36]. Another age-associated feature is the decreased isotype switch from IgM to IgG, IgE or IgA [37], possibly due to a decrease in the expression of activation-induced cytidine deaminase (AID) [38]. Age-associated B cells (ABCs) are antigen-experienced B cells with features of memory B cells that are activated through Toll-like receptor (TLR) 7 and TLR9 and not by BCR stimulation alone [39,40]. ABCs continuously expand with age and are hardly detectable in youth. However, ABCs are particularly expanded at earlier ages in individuals with autoimmune diseases, and ABCs often produce antibodies recognizing self-antigens [41].

2.2. Innate immunity

Although alterations induced by age in the adaptive response are well established, changes in the innate immune response in human ageing are not well understood. Most studies suggest that the total numbers of different innate immune cell populations do not change dramatically; however, recent findings support that many of their functions, such as cytotoxicity, phagocytosis, antigen presentation and the secretion of inflammatory cytokines, are altered.

Monocytes play an important role as starters of the inflammatory response, as they are circulating cells that are recruited to the site of inflammation, where they can differentiate into macrophages or into DCs in certain inflammatory situations [42]. Aged monocytes stimulated with agonists of pattern recognition receptors show differences at the transcriptional and functional levels, resulting in reduced production of IFN- α , IFN- γ , IL-1 β , CCL20 and CCL8, which participate in antiviral responses and in the recruitment of monocytes to the sites of inflammation [43]. Monocytes in elderly individuals also express more TAM (Tyro3, Axl and Mer) tyrosine kinase receptors, a family of receptors that

negatively regulate the immune responses mediated by TLRs and, in turn, inhibit inflammation [44]. Macrophages are considered the main phagocytes of potential pathogens and are capable of initializing and regulating inflammation. Polarization to alternatively activated anti-inflammatory M2 macrophages, rather than pro-inflammatory M1 macrophages, is predominant in the tissues of older mice [45]. Moreover, age negatively impacts the expression of TLRs and major histocompatibility complex II (MHC-II), which probably contributes to impaired antigen presentation and the activation of CD4⁺ T cells [46,47].

DCs are the most important antigen-presenting cells, thereby acting as a link between innate and adaptive immunity. Age-related alterations in DCs, such as mitochondrial dysfunction and increased reactive oxygen species (ROS) production, have specific deleterious effects on the phagocytosis and cross-presenting capacity of DCs [48]. The alteration of the antigen processing and presentation machinery specifically contributes to an impaired stimulation and cytotoxic response of CD8⁺ T cells, together with lower secretion of IFN- γ [49]. Moreover, circulating plasmacytoids (pDCs) and myeloid DCs (mDCs) are less represented and less responsive to TLR stimulation in elderly women [50].

NK cells are the main innate cell subtype responsible for killing cells infected by viruses, which have altered self or missing self-antigens. Unlike T and B cells, the absolute number of NK cells is slightly increased in elderly individuals, which could be associated with a redistribution of NK cell subtypes. There is a decrease in CD56^{bright} immunoregulatory cells and an increase in CD56^{dim} cytotoxic cells; however, older subjects show a defective degranulation capacity [51,52]. In contrast, the maturation of NK cells in the bone marrow of mice was shown to be impaired with age and is associated with reduced proliferation, resulting in reduced numbers of mature circulating cells in peripheral tissues [53]. Taken together, these data highlight the need for more studies on how ageing affects NK cells.

Neutrophils are the first cells recruited against bacterial and fungal infections and they produce many degradative enzymes, antimicrobial peptides and ROS for their activity. Aged neutrophils display a reduced ability to target infected tissue, as these cells present a lower capacity to generate ROS and neutrophil extracellular traps (NETs) [54]. In elderly individuals, alterations in surface molecules of neutrophils, such as CD11b or CD16, have been reported, affecting phagocytosis and intravascular adhesion to the endothelium and, consequently, the recruitment of immune cells to the infected tissue [55,56].

3. Age-related autoimmunity

One of the most important features of immunosenescence is inflammaging, a chronic low-grade inflammation characterized by a gradual increase in pro-inflammatory mediators, including TNF- α , IL-6 and IL-1 β , which results from an imbalance between pro-inflammatory and anti-inflammatory networks during ageing [57]. Pro-inflammatory responses can confer not only a high resistance to infectious diseases but also an increased susceptibility to inflammatory-based diseases. On the other hand, anti-inflammatory responses not only can increase susceptibility to infectious diseases but also can provide a survival advantage in older individuals. In fact, centenarians seem to deal with this chronic low-grade inflammation through an anti-inflammatory response called anti-inflammaging, which may be one of the secrets of longevity [58]. The principal stimulus involved in the progression of inflammaging is the accumulation of cell debris and self-antigens derived from cellular stress or viral infections. The mechanisms devoted to cleaning cell debris and misfolded self-antigens become defective with age, leading to an overexposure of the immune system to these products and sustained low-grade inflammation, which favours autoimmunity [59].

Immune tolerance is a mechanism of the unresponsiveness of the immune system to antigens and is aimed at discriminating self from non-self-antigens. However, some autoreactive lymphocytes escape all tolerance mechanisms, leading to the expansion of autoreactive T cells

and the production of autoantibodies. Most likely related to a higher exposure to exogenous factors, elderly people show an increased prevalence of autoantibodies such as rheumatoid factor, anti-neutrophil cytoplasm and anti-nuclear antibodies, among others [60,61].

Ageing is associated with the emergence of unusual cell subsets that are thought to have a role in the development of autoimmunity. Senescent CD4⁺CD28⁻ T cells were originally isolated in the context of an autoimmune disease and were found to be outgrowths and autoreactive to self-antigens [22]. Their autoreactive properties and inability to serve as helpers in B cell responses are very similar to the deficiencies found in senescent CD4⁺ T cells from aged individuals [23]. Furthermore, senescent CD8⁺CD28⁻ T cells expressing NK receptors can be found not only in elderly individuals but also in patients suffering from autoimmune diseases, who present alterations in the number and function of these cells [26]. ABCs continuously expand with age and are hardly detectable in youth; however, they are particularly expanded at earlier ages in people with autoimmune diseases presenting enriched autoantibody production [40]. There are other factors that might accelerate immunosenescence and contribute to the onset of autoimmune diseases, like genetic factors such as the human leucocyte antigen (HLA) haplotype and environmental factors such as persistent viral infections and high consumption of different medications in elderly individuals [62].

Systemic lupus erythematosus (SLE) has immune features that overlap with features of ageing, suggesting the presence of premature immunosenescence, such as telomere attrition and a generally lower phagocytic capacity [63,64]. The clearance of apoptotic bodies is mediated by macrophages and its deregulation could generate danger signals and the exposure of self-antigens to DCs [65], which present an affected tolerance and a greater autoreactivity with age [66]. In addition, ABCs are significantly increased in these patients and positively correlate with high anti-chromatin antibody production [67]. Moreover, ABCs are B cell activating factor (BAFF)-dependent, and the blockade of this cytokine has been proven to be effective in SLE patients, which confirms the implication of ABCs in the disease [68]. Furthermore, autoreactive B and T cells are thought to be activated after infection by CMV or Epstein-Barr virus (EBV) with pathogen antigens that share structural similarities with self-antigens. This cross-recognition mechanism principally provokes the production of anti-nuclear autoantibodies, which are also found in healthy elderly individuals [69].

As in ageing, in rheumatoid arthritis (RA), it is well established that CD4⁺CD28⁻ T cells accumulate and show signs of senescence; in fact, this population of cells was first identified in RA patients [22]. Senescent T cells no longer require a CD28 co-stimulatory signal for complete activation, and at the same time, they acquire cytotoxic properties [62]. In addition, a novel CD4⁺CD28⁻ T cell subset has been described in RA patients, revealing features of both Treg cells and senescent T cells. These CD4⁺CD28⁻FoxP3⁺ Treg-like cells insufficiently suppress the proliferation of effector T cells and induce a pro-inflammatory cytokine profile [70].

It is known that age-related gene expression profiles overlap with gene expression profiles in tissues affected by autoimmune thyroid disease (AITD), suggesting that there is also a relation between premature immunosenescence and autoimmunity in the thyroid gland. These changes include the downregulation of genes related to mitochondrial and proteasomal functions and the upregulation of genes related to the immune response [71]. AITD patients present a peripheral T cell phenotype reminiscent of findings in elderly patients and patients with other autoimmune diseases. There is a significant increase in CD4⁺CD28⁻ T cells associated with CMV seropositivity in individuals with Hashimoto's disease [72] and an increase in CD4⁺CD28⁻ T cells producing IFN- γ in individuals with Graves' disease [73], which might drive the destruction of thyroid tissue.

The most frequent causes of Sjögren syndrome (SjS) are the chronic use of medications and viral infections [74], which are both common features in elderly individuals, suggesting a relationship between premature ageing of the immune system and the development of the disease.

ABCs are also implicated in the pathogenesis of this syndrome, as they are elevated in the blood of SjS patients. ABCs remain unresponsive in peripheral blood and are probably activated through TLRs, breaking tolerance and leading to B cell lymphoproliferation [75].

4. Immunosenescence in MS

MS is a chronic neurodegenerative, inflammatory and demyelinating disease of the CNS that is probably of autoimmune origin. MS affects approximately 2.5 million people worldwide, and MS is the primary cause of non-traumatic disability in young adults [76]. MS typically initiates between 20 and 40 years of age, with a higher prevalence in women than men (2.3–3.5:1), a ratio that is increasing over time [77]. Clinically isolated syndrome (CIS) is the first clinical acute episode that shows inflammatory demyelination; however, CIS does not fulfil clinical and radiological criteria to be considered definite MS, and not all patients with CIS necessarily develop MS. Most patients present a relapsing-remitting (RR) form of MS, the most frequent form of the disease (80%), characterized by episodes of neurological dysfunction, and the majority of these patients (65%) progressively accumulate disability with time, developing secondary progressive (SP) forms of MS. A minority of patients (20%) present a progressive disease course from the onset, known as primary progressive (PP) MS [78].

Although the aetiology of MS remains unknown, MS is known to result from an interplay between genetic and environmental risk factors. Genome-wide association studies have identified 233 independent associations that are significantly linked to MS susceptibility, the majority of which were found in the autosomal non-MHC genome [79–81]. Nonetheless, the main genetic risk factors are related to HLA genes, with the variants DRB1*15:01, DRB5*01:01 and DQA1*01:02 being associated with an increased risk of developing MS, while HLA-A*02 is associated with protection from the disease [82]. Sex, living in high-latitude areas, smoking, low vitamin D levels caused by insufficient sun exposure or dietary intake, obesity during adolescence, commensal microbiota composition and EBV infection have been identified as possible environmental risk factors related to MS development, whereas the use of oral nicotine or alcohol, seropositivity to CMV and high coffee consumption are related to a reduced risk [83,84]. Among these risk factors, EBV is strongly associated with the development of MS. Several myelin antigens, including myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG) and proteolipid protein (PLP), have been proposed as the principal candidate autoantigens involved in the development of MS [85]; however, other non-myelin antigens have been identified to trigger immune attack on myelin components [86]. The most accepted hypothesis to explain the immunopathogenesis of MS involves an initial immune activation in the periphery of CD4⁺ T cells reactive to myelin antigens. Then, activated autoreactive CD4⁺ T cells infiltrate the CNS, where they are reactivated by the recognition of myelin antigens presented by microglial cells, initiating a series of immune events that contribute to demyelination and tissue damage. These immune events provoke the activation of astrocytes and microglia, the apoptosis of oligodendrocytes and axonal loss in both the white matter and the grey matter of the brain and the spinal cord. In addition, the release of pro-inflammatory mediators facilitates the permeabilization of the blood-brain barrier (BBB), attracting monocytes and additional lymphocytes, thus sustaining the inflammatory process that leads to progressive tissue degeneration [87]. The analysis of the autoimmune response showed structural homology between MBP and EBV peptides presented by HLA-DRB1*15:01, as well as the presence of T cells with specificity for both antigens. Altogether, molecular mimicry with EBV antigens is one of the main mechanisms involved in MS pathogenesis [88].

The prevalence of MS and the age of affected patients are increasing due to the increasing longevity of the general population and the availability of effective disease-modifying treatments. Patients with PPMS normally have a later disease onset than those with RRMS, the majority of whom develop SPMS over time. However, current

treatments reduce the frequency of relapses and slow the progression of the disease, but they cannot stop the disability accumulation associated with disease progression [89]. These facts support the idea that there are differences between the processes driving relapses and those driving chronic progression commonly in elderly individuals, which could be due to the impact of age on the immune system. Indeed, there are numerous studies in the animal model of MS experimental autoimmune encephalomyelitis (EAE) and in MS patients that demonstrate the impact of immunosenescence on the disease [90].

4.1. Age-related immune changes in EAE

EAE is a well-established animal model used for the study of MS that has provided helpful information about various pathological processes, including inflammation, demyelination, axonopathy and neuron loss mediated by immune cells [91]. However, there are some differences in immunopathology between EAE and MS, since CD4⁺ T cells predominate in the inflammatory infiltrate in EAE, while in MS lesions, CD4⁺ T cells are much less frequent, with CD8⁺ T cells and macrophages being the predominant infiltrating cells [92,93].

A key mediator in central tolerance is the autoimmune regulator (Aire), a transcription factor highly expressed in medullary thymic epithelial cells (TECs) that promotes ectopic expression of peripheral-tissue-specific self-antigens, leading to central tolerance. Some autoreactive T cells escape this process, but they are regulated by peripheral tolerance mechanisms, including immune regulation by Treg cells. There is an age-related association between Aire and Treg cells in susceptibility to EAE. The resistance of young Aire-knockout (KO) mice to EAE correlated with an elevated percentage of peripheral Treg cells, whereas older Aire-KO mice presented more severe disease and no differences in the frequency of Treg cells in the periphery, demonstrating age-related impaired peripheral tolerance [94]. A spontaneous EAE mouse model also showed a decrease in intrinsic regulatory mechanisms involved in the maintenance of self-tolerance and Treg cell suppressive function with age [95]. EAE rats show an age-associated alteration in the autoimmune response, while young rats show an anti-MBP antibody response during the acute period, no epitope dominance was detected in the older rats [96].

Little is known about the age effect on BBB disruption in EAE, a critical step in the development of the disease, as myelin-specific T cells infiltrate the CNS by crossing the BBB. Several molecules are altered in response to BBB disruption, such as nicotinamide adenine dinucleotide phosphate oxidase, matrix metalloproteinases and cell adhesion molecules. These molecules were elevated with age in a non-relapse EAE mouse model, suggesting that they play a role in BBB disruption and the subsequent autoreactive T cell infiltration that leads to neurodegeneration [97]. However, age not only is a risk factor for neurodegeneration but also influences neuroregeneration. Using a focal EAE rat model, immunization with MOG followed by localized injections of cytokines in the spinal cord to generate focal demyelinating injury increased vulnerability to axonal injury and reduced the efficiency of remyelination in older animals compared with young animals. Moreover, remyelination in aged rats was mediated by Schwann cells, while in younger rats, oligodendrocytes mediated the regeneration process [98].

4.2. Age-related immune changes in MS

Age, sex and viral infections are known to influence the phenotype and function of the immune system. The frequencies of immune cells depend on lifetime exposure to pathogens such as CMV, which leads to late differentiation and the accumulation of senescent immune cells with age [99]. Indeed, senescent neurons and glial cells also accumulate in the nervous system with age, predisposing individuals to the appearance and aggravation of neurodegenerative events [100]. Furthermore, the evolution from RRMS to SPMS normally occurs over time at older ages,

and this transition is dependent on age rather than disease duration [101]. Neuroinflammation mechanisms such as synaptopathy and synaptic plasticity impairments are exacerbated and accelerate with age in MS patients [102]. In particular, disease progression is characterized by a marked decrease with age in the CD8⁺ T cell response to EBV-infected B cells, which may result from the deletion of EBV-specific T cells through exhaustion mechanisms that occur during chronic viral infections [103].

4.3. Premature ageing of the immune system in MS

Patients suffering from MS are also characterized by premature immunosenescence [104] and reduced immune function that resembles that of healthy elderly individuals. The reduced immune function can be a consequence of the diminished proliferative capacity of bone-marrow-derived cells and the shortening of telomeres, which are both observed in MS patients [105]. Telomere length reduction and increased oxidative stress have been suggested as premature ageing markers in many autoimmune diseases, including MS. The shortened telomeres and oxidative stress found in PPMS patients may reflect the most severe state of the disease and may also be associated with greater disability and brain atrophy [106,107]. Furthermore, RRMS and PPMS patients also present prematurely reduced immune functions related to thymic dysfunction, including a contraction of naïve T cells, Treg cell dysfunction and decreased signal joint TRECs [108]. Similar alterations in T cell homeostasis are observed in paediatric MS patients, confirming the presence of an early thymic involution [109]. The main common characteristic between ageing and MS is the accumulation of CD4⁺CD28⁻ T cells. These effector memory T cells result from repeated viral antigenic stimulation, typically CMV and EBV, before migrating to the CNS, which contributes to tissue damage, thereby causing the release of self-antigens. IL-15 produced by peripheral B cells, astrocytes and infiltrating macrophages in CNS inflammatory lesions is known to enhance the cytotoxic properties of CD4⁺CD28⁻ T cells [110]. Alterations in B cell subsets are also found in the cerebrospinal fluid of MS patients with acute relapse, as there is a higher frequency of non-switched memory B cells in paediatric patients and class-switched memory B cells in adults [111]. However, B cell participation in MS goes beyond the production of antibodies. ABCs are increased in MS patients as well, corroborating the presence of early ageing [112].

MS is primarily mediated by adaptive immune responses, while chronic stages are believed to be mediated by compartmentalized innate immune responses in the CNS [9]. Therefore, a complete understanding of the impact of age on the immune system and disease progression, as well as the premature immune ageing observed in MS patients, needs to be achieved to develop effective immunotherapies against immunosenescence in MS.

5. Rejuvenation of the immune system as a therapy for MS

Currently, there is no cure for MS and no effective therapy options available that can stop disability accumulation in individuals with progressive forms of the disease, who represent the majority of patients over 65 years of age. Food and Drug Administration (FDA)-approved therapies for RRMS have failed to demonstrate efficacy as a treatment for PPMS, and there are few disease-modifying therapy options for SPMS [113,114]. In addition, emerging data suggest that premature immunosenescence strongly influences MS progression; thus, several therapeutic strategies based on rejuvenating the immune system have been proposed to reverse this phenomenon (Fig. 1).

5.1. Senolytics and HSC regenerative strategies

Senescent cells accumulate with age in the CNS and contribute to disease progression. Senolytic drugs have been developed to selectively remove senescent cells by targeting anti-apoptotic pathways, and

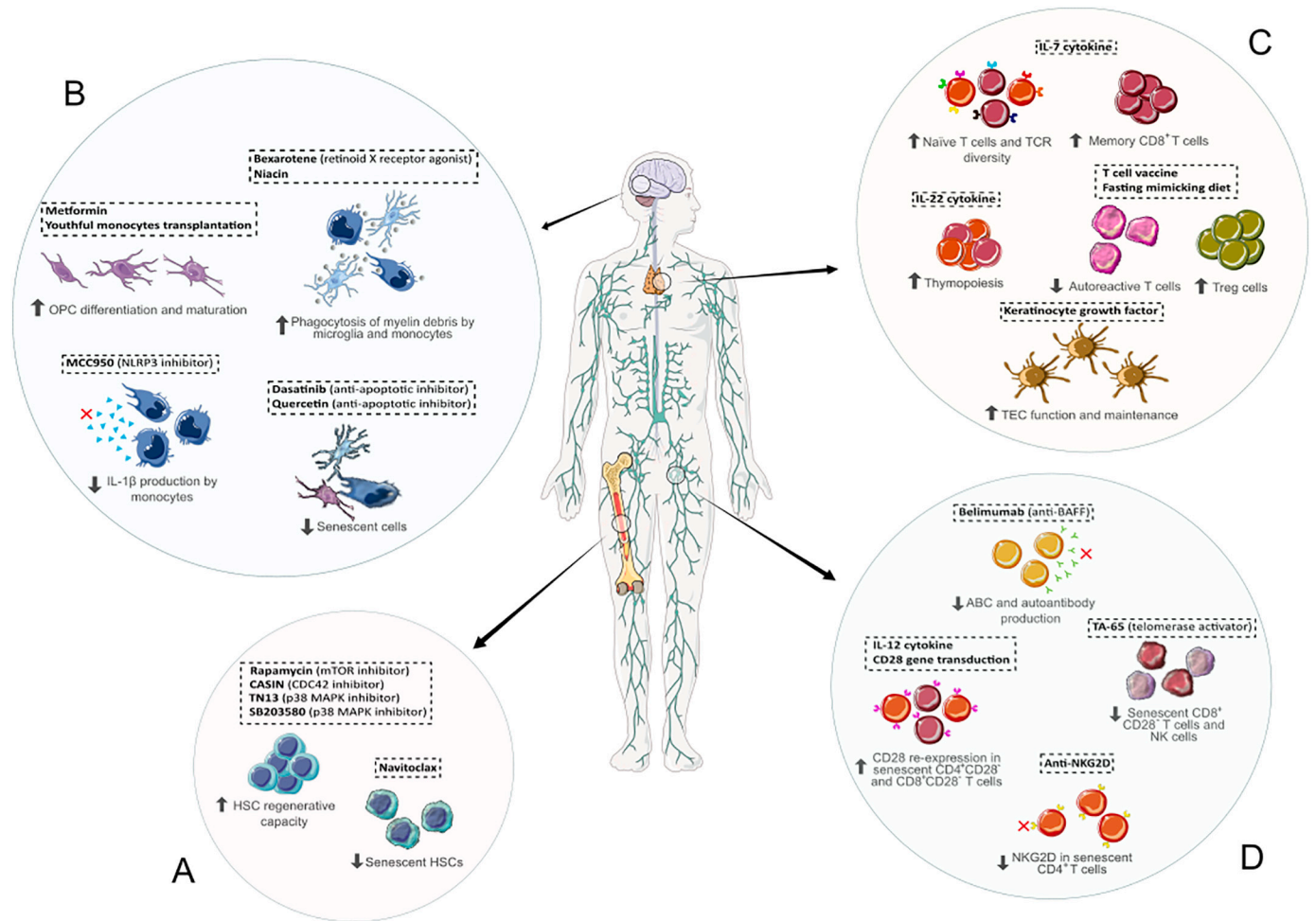


Fig. 1. Senolytics and regenerative strategies using haematopoietic stem cells (A), therapies based on rejuvenating oligodendrocyte precursor cells, microglia and monocytes (B), thymus cells (C) and senescent B and T cells (D) are capable of reversing the process of immunosenescence and could have a beneficial impact on the progression of MS. *Abbreviations:* Oligodendrocyte precursor cell (OPC); Haematopoietic stem cell (HSC); T cell receptor (TCR); Regulatory T cell (Treg); Thymic epithelial cell (TEC); Age-associated B cell (ABC); Natural killer (NK).

senolytics have recently been suggested as potential rejuvenation strategies against progressive MS [115]. The use of the tyrosine kinase inhibitor **dasatinib** and **quercetin** in combination decreased the accumulation of senescent cells and increased survival in older mice. Moreover, these drugs showed immunomodulatory effects on MS when used separately [116–118]. Another important senolytic drug is **navitoclax**, or ABT263, which is able to induce apoptosis in senescent bone marrow HSCs, reduce myeloid bias and improve haematopoietic functionality [119]. In fact, HSC ageing is related to reduced self-renewal capacity, which leads to altered immune cell numbers and functions. The ageing of the HSC compartment was initially considered to be irreversible; however, recent findings show rejuvenating strategies that are capable of reversing HSC dysfunctions related to ageing, including prolonged fasting and pharmacological targeting to inhibit molecules such as mTOR (**rapamycin**) [120], CDC42 (**CASIN**) [121] and p38 MAPK (**TN13** and **SB203580**) [122,123]. These strategies are capable of enhancing the regenerative capacity of HSCs, reducing ROS levels and re-establishing an immune system similar to that of young organisms (Fig. 1A).

5.2. Rejuvenation of oligodendrocyte precursor cells (OPCs), microglia and monocytes

MS progressive forms are characterized by accumulative neurodegeneration due to persistent demyelination of axons. Although the

CNS has the capacity to remyelinate axons and, thus, prevent neurodegeneration, the efficiency of CNS remyelination declines with age, and age-related failure of OPCs to differentiate into myelinating oligodendrocytes is particularly involved. The fasting mimetic drug **metformin** reverses the poor differentiation and maturation of OPCs and restores remyelination and neuroregeneration in aged rodents [124]. Furthermore, older mice with a focal demyelination exposed to a youthful systemic milieu through heterochronic parabiosis (a young mouse surgically joined to an aged partner) could recruit young macrophages and rejuvenate OPCs, suggesting that **youthful monocyte transplantation** could be considered a remyelination therapy [125]. Microglia and macrophages are essential for remyelination, as both are responsible for the phagocytosis of myelin debris that inhibits remyelination and for the release of growth factors necessary for OPC maturation. With age, a decline in the phagocytic clearance of myelin debris occurs, a phenomenon that has also been observed in MS patients. Given that retinoid X receptor activation in monocytes enhances myelin debris clearance in MS, the agonist **bexarotene** was found to reverse phagocytosis defects and lead to a younger state of these cells [126]. In addition, with age, a decrease in the expression of CD36 by microglia occurs, which is also involved in myelin debris clearance by phagocytosis. Daily treatment of old microglia with **niacin** or vitamin B3 upregulates CD36 expression and enhances phagocytosis, which leads to remyelination. Thus, niacin can stimulate innate immunity and represents a regenerative therapy for chronic demyelination, which is an interesting therapeutic strategy for

MS [127]. **MCC950** regulates innate immunity by specifically inhibiting the NLRP3 inflammasome and subsequently reducing IL-1 β production. MCC950 attenuated the severity of EAE, improved the histopathology of the CNS and protected against axonal damage in organotypic cerebellar cultures. Therefore, MCC950 could be considered an anti-inflammaging therapy for MS [128,129] (Fig. 1B).

5.3. Rejuvenation of the thymus

Recently, many potential rejuvenation therapeutic strategies have been developed to restore aged thymic function and, consequently, defects in negative selection and in the generation of Treg cells. As the FOXN1 transcription factor is strongly implicated in the differentiation of TECs cells and is responsible for the ectopic expression of peripheral-tissue-specific self-antigens that lead to tolerance, it has been proposed as a target for several rejuvenation strategies against thymic involution [130]. Because thymic atrophy is related to a failure in the thymic microenvironment and consequently in thymopoiesis, targeting this process could also reverse the observed thymus dysfunction in MS patients. There is evidence that an age-related decrease in the expression of IL-7 is involved in thymus involution. Treatment with IL-7 promotes the maturation of thymocytes to naïve T cells, enhances TCR diversity and results in higher numbers of memory CD8⁺ T cells [131]. Another cytokine involved in thymopoiesis is IL-22, which was shown to enhance thymic recovery after the depletion of CD4⁺CD8⁺ double-positive thymocytes [132]. **Keratinocyte growth factor** is important for the maintenance of TECs and has been proven to increase thymopoietic capacity in aged mice by restoring the function of TECs [133]. However, thymic involution results in not only decreased numbers of naïve T cells but also increased output of potential autoreactive T cells to the periphery and consequently to higher susceptibility to developing autoimmune diseases such as MS. The most promising rejuvenation immunotherapies focused on targeting these autoreactive T cells are therapeutic vaccinations that include TCR peptides, MBP-based DNA, altered peptide ligands and attenuated autoreactive T cells [134]. Among them, **T cell vaccines** have shown clinical efficacy in clinical trials, as they induce an anti-idiotypic response against MBP-reactive T cells and restore Treg cell function in MS patients, making them a potential strategy against MS [135]. Additionally, a **fasting-mimicking diet**, a very low-calorie and low-protein diet, prevents autoimmunity in an EAE model by reducing the levels of autoreactive T cells and increasing the production of Treg cells, which are both cell subsets that are altered with immunosenescence [136]. Another strategy to counteract immunosenescence is to reduce as much as possible the antigenic load represented by common infectious agents such as influenza virus and CMV to avoid inflammaging. Strategies of specific vaccination should be applied to prevent not only morbidity and mortality but also any additional persistent stimulation of the immune system and, thus, reduce the impact of inflammaging in elderly individuals and in the context of autoimmune diseases (Fig. 1C).

5.4. Rejuvenation of senescent B and T cells

Different cell types that become senescent during MS could be interesting targets of rejuvenation. Anti-BAFF treatment with **belimumab** decreases the proliferation of ABCs and, consequently, the production of autoantibodies [68]. As in MS, chronic viral infections such as CMV promote telomere shortening and premature ageing of the immune system, particularly of cytotoxic T cells. These T cells also lack the expression of the co-stimulatory receptor CD28, and hence, their capacity to respond to infections is diminished. **TA-65** is a purified telomerase activator that, when given orally to aged subjects, can lengthen short leucocyte telomeres and decrease the percentage of senescent cytotoxic CD8⁺CD28⁻ T cells as well as NK cells. This effect has been observed mostly in CMV-seropositive subjects, whose leukocytes switch towards a younger profile [137]. The re-expression of CD28 and the

restoration of helper function in senescent CD4⁺ T cells can be partially achieved when CD4⁺ T cells are exposed to the cytokine IL-12 [138]. A similar functional restoration and production of IL-2 was shown in senescent CD8⁺ T cells by **CD28 gene transduction** [139]. Both strategies can delay or reverse the ageing effects of the loss of the CD28 co-stimulatory molecule. In addition, the expression of innate receptors such as NKG2D on CD4⁺ T cells is characteristic of ageing and regulates the cytotoxic functions of these cells by promoting pro-inflammatory cytokine production. Treatment with a blocking **anti-NKG2D** antibody ameliorates EAE inflammation, making this receptor a promising therapeutic target for MS treatment [140] (Fig. 1D).

6. Conclusions

Overall, the evidence presented here demonstrates that age-related immune changes are observed in EAE and MS patients, as well as premature ageing of the immune system in MS patients. Therefore, we propose different therapeutic strategies based on rejuvenating the immune system, which could reverse the process of immunosenescence and have a beneficial impact on the treatment of MS, as potential add-on treatments to current FDA-approved therapies. Moreover, therapies focused on the rejuvenation of the immune system could provide new therapeutic opportunities for the PP form of the disease. A broad knowledge of the impact of age on the immune system and disease progression in MS patients would facilitate the development of new effective immunotherapies against immunosenescence in MS.

Funding

This work was supported by the Fondo de Investigación Sanitaria, Instituto de Salud Carlos III, and co-funded by the European Union (European Regional Development Fund/European Social Fund), “A way to build Europe”, under Grant PI18/01146 and RD16/0015/004 and the “Agència de Gestió d’Ajuts Universitaris i de Recerca” (AGAUR; Generalitat de Catalunya) under Grant 2017SGR527.

Declaration of Competing Interest

MD, HE, LMV and CE: None.

XM has received speaking honoraria and travel expenses for participation in scientific meetings and has been a steering committee member of clinical trials or has participated in advisory boards of clinical trials in the past years with Actelion, Alexion, Bayer, Biogen, Celgene, EMD Serono, Genzyme, Immunic, MedDay, Merck KGaA, Darmstadt Germany, Mylan, Nervgen, Novartis, Roche, Sanofi-Genzyme, Teva Pharmaceutical, TG Therapeutics, Excemed, MSIF and NMSS.

References

- [1] World Health Organisation (WHO). Ageing and Health. <https://www.who.int/news-room/fact-sheets/detail/ageing-and-health>; 2018.
- [2] López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013;153:1194–217. <https://doi.org/10.1016/j.cell.2013.05.039>.
- [3] Pera A, Campos C, López N, Hassouneh F, Alonso C, Tarazona R, et al. Immunosenescence: implications for response to infection and vaccination in older people. *Maturitas* 2015;82:50–5. <https://doi.org/10.1016/j.maturitas.2015.05.004>.
- [4] Goronzy J. Immune aging and autoimmunity. *Cell Mol Life Sci* 2012;69:1615–23. <https://doi.org/10.1007/s00018-012-0970-0>.
- [5] Costantini E, D’Angelo C, Reale M. The role of immunosenescence in neurodegenerative diseases. *Mediat Inflamm* 2018;1–12. <https://doi.org/10.1155/2018/6039171>.
- [6] Wikby A, Ferguson F, Forsey R, Thompson J, Strindhall J, Löfgren S, et al. An immune risk phenotype, cognitive impairment, and survival in very late life: impact of allostatic load in Swedish octogenarian and nonagenarian humans. *J Gerontol - Ser A Biol Sci Med Sci* 2005;60:556–65. <https://doi.org/10.1093/gerona/60.5.556>.
- [7] Salminen A. Activation of immunosuppressive network in the aging process. *Ageing Res Rev* 2020;57:1–15. <https://doi.org/10.1016/j.arr.2019.100998>.
- [8] Solana R, Pawelec G, Tarazona R. Aging and innate immunity. *Immunity* 2006;24:491–4. <https://doi.org/10.1016/j.immuni.2006.05.003>.

- [9] Sospedra M, Martin R. Immunology of multiple sclerosis. *Semin Neurol* 2016;36:115–27. <https://doi.org/10.1055/s-0036-1579739>.
- [10] Rossi DJ, Bryder D, Zahn JM, Ahlenius H, Sonu R, Wagers AJ, et al. Cell intrinsic alterations underlie hematopoietic stem cell aging. *Proc Natl Acad Sci U S A* 2005;102:9194–9. <https://doi.org/10.1073/pnas.0503280102>.
- [11] Naylor K, Li G, Vallejo AN, Lee W-W, Koetz K, Bryl E, et al. The influence of age on T cell generation and TCR diversity. *J Immunol* 2005;174:7446–52. <https://doi.org/10.4049/jimmunol.174.11.7446>.
- [12] Qi Q, Liu Y, Cheng Y, Glanville J, Zhang D, Lee JY, et al. Diversity and clonal selection in the human T-cell repertoire. *Proc Natl Acad Sci U S A* 2014;111:13139–44. <https://doi.org/10.1073/pnas.1409155111>.
- [13] Lang PO, Govind S, Aspinall R. Reversing T cell immunosenescence: why, who, and how. *Age (Omaha)* 2013;35:609–20. <https://doi.org/10.1007/s11357-012-9393-y>.
- [14] Fagnoni FF, Vescovini R, Passeri G, Bologna G, Pedrazzoni M, Lavagetto G, et al. Shortage of circulating naive CD8+ T cells provides new insights on immunodeficiency in aging. *Blood* 2000;95:2860–8. <https://doi.org/10.1182/blood.v95.9.2860.009k35.2860.2868>.
- [15] Ferrando-Martínez S, Ruiz-Mateos E, Hernández A, Gutiérrez E, Rodríguez-Méndez MDM, Ordoñez A, et al. Age-related deregulation of naive T cell homeostasis in elderly humans. *Age (Omaha)* 2011;33:197–207. <https://doi.org/10.1007/s11357-010-9170-8>.
- [16] Almanzar G, Schwaiger S, Jenewein B, Keller M, Herndler-Brandstetter D, Würzner R, et al. Long-term cytomegalovirus infection leads to significant changes in the composition of the CD8+ T-cell repertoire, which may be the basis for an imbalance in the cytokine production profile in elderly persons. *J Virol* 2005;79:3675–83. <https://doi.org/10.1128/jvi.79.6.3675-3683.2005>.
- [17] Derhovanessian E, Maier AB, Hähnel KH, Beck R, de Craen AJM, Slagboom EP, et al. Accumulation with cytomegalovirus but not herpes simplex virus induces the accumulation of latently differentiated CD4+ and CD8+ T-cells in humans. *J Gen Virol* 2011;92:2746–56. <https://doi.org/10.1099/vir.0.036004.0>.
- [18] Lang PO, Govind S, Mitchell WA, Siegrist CA, Aspinall R. Vaccine effectiveness in older individuals: what has been learned from the influenza-vaccine experience. *Ageing Res Rev* 2011;10:389–95. <https://doi.org/10.1016/j.arr.2010.09.005>.
- [19] Vallejo AN. CD28 extinction in human T cells: altered functions and the program of T-cell senescence. *Immunol Rev* 2005;205:158–69. <https://doi.org/10.1111/j.0105-2896.2005.00256.x>.
- [20] Song Y, Wang B, Song R, Hao Y, Wang D, Li Y, et al. T-cell Immunoglobulin and ITIM domain contributes to CD8+ T-cell immunosenescence. *Aging Cell* 2018;17:1–12. <https://doi.org/10.1111/acel.12716>.
- [21] Callender LA, Carroll EC, Beal RWJ, Chambers ES, Nourshargh S, Akbar AN, et al. Human CD8+ EMRA T cells display a senescence-associated secretory phenotype regulated by p38 MAPK. *Aging Cell* 2018;17:1–9. <https://doi.org/10.1111/acel.12675>.
- [22] Schmidt D, Goronzy JJ, Weyand CM. CD4+ CD7- CD28- T cells are expanded in rheumatoid arthritis and are characterized by autoreactivity. *J Clin Invest* 1996;97:2027–37. <https://doi.org/10.1172/JCI118638>.
- [23] Weyand CM, Brandes JC, Schmidt D, Fulbright JW, Goronzy JJ. Functional properties of CD4+ CD28- T cells in the aging immune system. *Mech Ageing Dev* 1998;102:131–47. [https://doi.org/10.1016/S0047-6374\(97\)00161-9](https://doi.org/10.1016/S0047-6374(97)00161-9).
- [24] Haynes L, Eaton SM. The effect of age on the cognate function of CD4+ T cells. *Immunol Rev* 2005;205:220–8. <https://doi.org/10.1111/j.0105-2896.2005.00255.x>.
- [25] Bauer S, Groh V, Wu J, Steinle A, Phillips JH, Lanier LL, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. *Science* 1999;285:727–9. <https://doi.org/10.1126/science.285.5428.727>.
- [26] Tarazona R, DelaRosa O, Alonso C, Ostos B, Espejo J, Pea J, et al. Increased expression of NK cell markers on T lymphocytes in aging and chronic activation of the immune system reflects the accumulation of effector/senescent T cells. *Mech Ageing Dev* 2001;121:77–88. [https://doi.org/10.1016/S0047-6374\(00\)00199-8](https://doi.org/10.1016/S0047-6374(00)00199-8).
- [27] Pereira BI, De Maeyer RPH, Covre LP, Nehar-Belaid D, Lanna A, Ward S, et al. Sestrins induce natural killer function in senescent-like CD8+ T cells. *Nat Immunol* 2020;21:684–94. <https://doi.org/10.1038/s41590-020-0643-3>.
- [28] Nakajima T, Goek Ö, Zhang X, Kopecky SL, Frye RL, Goronzy JJ, et al. De novo expression of killer immunoglobulin-like receptors and signaling proteins regulates the cytotoxic function of CD4 T cells in acute coronary syndromes. *Circ Res* 2003;93:106–13. <https://doi.org/10.1161/01.RES.0000082333.58263.58>.
- [29] Warrington KJ, Takemura S, Goronzy JJ, Weyand CM. CD4+, CD28- T cells in rheumatoid arthritis patients combine features of the innate and adaptive immune systems. *Arthritis Rheum* 2001;44:13–20. [https://doi.org/10.1002/1529-0131\(200101\)44:1<13::AID-ANR3>3.0.CO;2-6](https://doi.org/10.1002/1529-0131(200101)44:1<13::AID-ANR3>3.0.CO;2-6).
- [30] Alonso-Arias R, Moro-García MA, López-Vázquez A, Rodrigo L, Baltar J, García FMS, et al. NKG2D expression in CD4+ T lymphocytes as a marker of senescence in the aged immune system. *Age (Omaha)* 2011;33:591–605. <https://doi.org/10.1007/s11357-010-9200-6>.
- [31] Pereira BI, Akbar AN. Convergence of innate and adaptive immunity during human aging. *Front Immunol* 2016;7:1–9. <https://doi.org/10.3389/fimmu.2016.00445>.
- [32] Riley RL. Impaired B lymphopoiesis in old age: a role for inflammatory B cells? *Immunol Rev* 2013;257:361–9. <https://doi.org/10.1007/s12026-013-8444-5>.
- [33] Tabibian-Keissar H, Hazanov L, Schiby G, Rosenthal N, Rakovsky A, Michaeli M, et al. Aging affects B-cell antigen receptor repertoire diversity in primary and secondary lymphoid tissues. *Eur J Immunol* 2016;46:480–92. <https://doi.org/10.1002/eji.201545586>.
- [34] Colonna-Romano G, Aquino A, Bulati M, Di Lorenzo G, Listi F, Vitello S, et al. Memory B cell subpopulations in the aged. *Rejuvenation Res* 2006;9:149–52. <https://doi.org/10.1089/rej.2006.9.149>.
- [35] Frasca D, Diaz A, Romero M, Blomberg BB. Human peripheral late/exhausted memory B cells express a senescent-associated secretory phenotype and preferentially utilize metabolic signaling pathways. *Exp Gerontol* 2017;87:113–20. <https://doi.org/10.1016/j.exger.2016.12.001>.
- [36] Frasca D, Diaz A, Romero M, D'Ermo F, Blomberg BB. Aging effects on T-bet expression in human B cell subsets. *Cell Immunol* 2017;321:68–73. <https://doi.org/10.1016/j.cellimm.2017.04.007>.
- [37] Bagnara D, Squillario M, Kipling D, Mora T, Walczak AM, Da Silva L, et al. A reassessment of IgM memory subsets in humans. *J Immunol* 2015;195:3716–24. <https://doi.org/10.4049/jimmunol.1500753.A>.
- [38] Frasca D, Diaz A, Romero M, Ferracci F, Blomberg BB. Micro-RNAs miR-155 and miR-16 decrease AID and E47 in B cells from elderly individuals. *J Immunol* 2015;195:2134–40. <https://doi.org/10.1016/j.physbeh.2017.03.040>.
- [39] Hao Y, O'Neill P, Naradikian MS, Scholz JL, Cancro MP. A B-cell subset uniquely responsive to innate stimuli accumulates in aged mice. *Blood* 2011;118:1294–304. <https://doi.org/10.1182/blood-2011-01-330530>.
- [40] Rubtsov AV, Rubtsova K, Fischer A, Meehan RT, Gillis JZ, Kappler JW, et al. Toll-like receptor 7 (TLR7)-driven accumulation of a novel CD11c+ B-cell population is important for the development of autoimmunity. *Blood* 2011;118:1305–15. <https://doi.org/10.1182/blood-2011-01-331462>.
- [41] Cancro MP. Age-associated B cells. *Annu Rev Immunol* 2020;38:315–40. <https://doi.org/10.1146/annurev-immunol-092419-031130>.
- [42] Collin M, Mcgovern N, Haniffa M. Human dendritic cell subsets. *Immunology* 2013;140:22–30. <https://doi.org/10.1111/imm.12117>.
- [43] Metcalf TU, Wilkinson PA, Cameron MJ, Ghneim K, Chiang C, Wertheimer AM, et al. Human monocyte subsets are transcriptionally and functionally altered in aging in response to pattern recognition receptor agonists. *J Immunol* 2017;199:1405–17. <https://doi.org/10.4049/jimmunol.1700148>.
- [44] Wang X, Malawista A, Qian F, Ramsey C, Allore HG, Montgomery RR. Age-related changes in expression and signaling of TAM receptor inflammatory regulators in monocytes. *Oncotarget* 2018;9:9572–80. <https://doi.org/10.18632/oncotarget.23851>.
- [45] Jackaman C, Radley-Crabb HG, Soffe Z, Shavlakadze T, Grounds MD, Nelson DJ. Targeting macrophages rescues age-related immune deficiencies in C57BL/6J geriatric mice. *Aging Cell* 2013;12:345–57. <https://doi.org/10.1111/acel.12062>.
- [46] Herrero C, Marqués L, Lloberas J, Celada A. IFN-γ dependent transcription of MHC class II IA is impaired in macrophages from aged mice. *J Clin Invest* 2001;107:485–93. <https://doi.org/10.1172/JCI11696>.
- [47] Renshaw M, Rockwell J, Engleman C, Gewirtz A, Katz J, Sambhara S. Cutting edge: impaired toll-like receptor expression and function in aging. *Mech Ageing Dev* 2002;169:4697–701. <https://doi.org/10.4049/jimmunol.169.9.4697>.
- [48] Choungnet CA, Thacker RI, Shehata HM, Hennies CM, Lehn MA, Lages CS, et al. Loss of phagocytosis and antigen cross-presenting capacity in aging DCs is associated with mitochondrial dysfunction. *J Immunol* 2015;195:2624–32. <https://doi.org/10.4049/jimmunol.1501006>.
- [49] Zacca ER, Crespo MI, Acland RP, Roselli E, Núñez NG, Maccioni M, et al. Aging impairs the ability of conventional dendritic cells to cross-prime CD8+ T cells upon stimulation with a TLR7 ligand. *PLoS One* 2015;10:1–20. <https://doi.org/10.1371/journal.pone.0140672>.
- [50] van Splunter M, Perdijk O, Fick-Brinkhof H, Floris-Vollenbroek EG, Meijer B, Brugman S, et al. Plasmacytoid dendritic cell and myeloid dendritic cell function in ageing: a comparison between elderly and young adult women. *PLoS One* 2019;14:1–19. <https://doi.org/10.1371/journal.pone.0225825>.
- [51] Borrego F, Alonso MC, Galiani MD, Carracedo J, Ramirez R, Ostos B, et al. NK phenotypic markers and IL2 response in NK cells from elderly people. *Exp Gerontol* 1999;34:253–65. [https://doi.org/10.1016/S0531-5565\(98\)00076-X](https://doi.org/10.1016/S0531-5565(98)00076-X).
- [52] Le Garff-Tavernier M, Béziat V, Decocq J, Siguret V, Gandjbakch F, Pautas E, et al. Human NK cells display major phenotypic and functional changes over the life span. *Aging Cell* 2010;9:527–35. <https://doi.org/10.1111/j.1474-9726.2010.00584.x>.
- [53] Beli E, Duriancik DM, Clinthorne JF, Lee T, Kim S, Gardner EM. Natural killer cell development and maturation in aged mice. *Mech Ageing Dev* 2014;135:33–40. <https://doi.org/10.1016/j.mad.2013.11.007.NATURAL>.
- [54] Hazeldine J, Harris P, Chapple IL, Grant M, Greenwood H, Livesey A, et al. Impaired neutrophil extracellular trap formation: a novel defect in the innate immune system of aged individuals. *Aging Cell* 2014;13:690–8. <https://doi.org/10.1111/acel.12222>.
- [55] Butcher SK, Chahal H, Nayak L, Sinclair A, Henriquez NV, Sapey E, et al. Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. *J Leukoc Biol* 2001;70:881–6. <https://doi.org/10.1189/jlb.70.6.881>.
- [56] Lopes AB, Lopes LB, da Silveira Antunes RN, Fukasawa JT, de A. Cavaretto D, Calamita Z. Effects of immunosenescence on the lower expression of surface molecules in neutrophils and lymphocytes. *Curr Aging Sci* 2018;11:118–25. <https://doi.org/10.2174/1874609811666180605092234>.
- [57] Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000;908:244–54. <https://doi.org/10.1111/j.1749-6632.2000.tb06651.x>.
- [58] Franceschi C, Capri M, Monti D, Giunta S, Olivieri F, Sevini F, et al. Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. *Mech Ageing Dev* 2007;128:92–105. <https://doi.org/10.1016/j.mad.2006.11.016>.

- [59] Franceschi C, Garagnani P, Vitale G, Capri M, Salvioli S. Inflammaging and 'Garbaging'. *Trends Endocrinol Metab* 2017;28:199–212. <https://doi.org/10.1016/j.tem.2016.09.005>.
- [60] Ramos-Casals M, García-Carrasco M, Brito MP, López-Soto A, Font J. Autoimmunity and geriatrics: clinical significance of autoimmune manifestations in the elderly. *Lupus* 2003;12:341–55. <https://doi.org/10.1191/0961203303lu383ed>.
- [61] Rosato E, Salsano F. Immunity, autoimmunity and autoimmune diseases in older people. *J Biol Regul Homeost Agents* 2008;22:217–24.
- [62] Thewissen M, Somers V, Venken K, Linsen L, Van Paassen P, Geusens P, et al. Analyses of immunosenescence markers in patients with autoimmune disease. *Clin Immunol* 2007;123:209–18. <https://doi.org/10.1016/j.clim.2007.01.005>.
- [63] Haque AS, Rakić H, Marriage F, Ho P. Shortened telomere length in patients with systemic lupus erythematosus. *Arthritis Rheum* 2013;65:1319–23. <https://doi.org/10.1002/art>.
- [64] Herrmann M, Voll RE, Zoller OM, Hagenhofer M, Ponner BB, Kalden JR. Impaired phagocytosis of apoptotic cell material by monocyte-derived macrophages from patients with systemic lupus erythematosus. *Arthritis Rheum* 1998;41:1241–50. [https://doi.org/10.1002/1529-0131\(199807\)41:7<1241::AID-ART15>3.0.CO;2-H](https://doi.org/10.1002/1529-0131(199807)41:7<1241::AID-ART15>3.0.CO;2-H).
- [65] Baumann I, Kolowos W, Voll RE, Manger B, Gaipal U, Neuhuber WL, et al. Impaired uptake of apoptotic cells into tingible body macrophages in germinal centers of patients with systemic lupus erythematosus. *Arthritis Rheum* 2002;46:191–201. [https://doi.org/10.1002/1529-0131\(200201\)46:1<191::AID-ART10027>3.0.CO;2-K](https://doi.org/10.1002/1529-0131(200201)46:1<191::AID-ART10027>3.0.CO;2-K).
- [66] Agrawal A, Tay J, Ton S, Agrawal S, Gupta S. Increased reactivity of dendritic cells from aged subjects to selfantigen, the human DNA. *J Immunol* 2009;182:1138–45. <https://doi.org/10.4049/jimmunol.182.2.1138>.
- [67] Liu Y, Zhou S, Qian J, Wang Y, Yu X, Dai D, et al. T-bet+CD11c+ B cells are critical for antichromatin immunoglobulin G production in the development of lupus. *Arthritis Res Ther* 2017;19:1–11. <https://doi.org/10.1186/s13075-017-1438-2>.
- [68] Ramsköld D, Parodis I, Lakshminanth T, Sippl N, Khademi M, Chen Y, et al. B cell alterations during BAFF inhibition with belimumab in SLE. *EBioMedicine* 2019;40:517–27. <https://doi.org/10.1016/j.ebiom.2018.12.035>.
- [69] Xavier RM, Yamauchi Y, Nakamura M, Tanigawa Y, Ishikura H, Tsunematsu T, et al. Antinuclear antibodies in healthy aging people: a prospective study. *Mech Ageing Dev* 1995;78:145–54. [https://doi.org/10.1016/0047-6374\(94\)01532-Q](https://doi.org/10.1016/0047-6374(94)01532-Q).
- [70] Fessler J, Raicht A, Husic R, Ficjan A, Schwarz C, Duftner C, et al. Novel senescent regulatory T-cell subset with impaired suppressive function in rheumatoid arthritis. *Front Immunol* 2017;8:1–11. <https://doi.org/10.3389/fimmu.2017.00300>.
- [71] Cho BA, Yoo SK, Song YS, Kim SJ, Lee KE, Shong M, et al. Transcriptome network analysis reveals aging-related mitochondrial and proteasomal dysfunction and immune activation in human thyroid. *Thyroid* 2018;28:656–66. <https://doi.org/10.1089/thy.2017.0359>.
- [72] Prelog M, Schönlaub J, Würzner R, Koppelstaetter C, Almanzar G, Brunner A, et al. Lower CD28+ T cell proportions were associated with CMV-seropositivity in patients with Hashimoto's thyroiditis. *BMC Endocr Disord* 2013;13:1–11. <https://doi.org/10.1186/1472-6823-13-34>.
- [73] Sun Z, Zhong W, Lu X, Shi B, Zhu Y, Chen L, et al. Association of Graves' disease and prevalence of circulating IFN- γ -producing CD28- T cells. *J Clin Immunol* 2008;28:464–72. <https://doi.org/10.1007/s10875-008-9213-4>.
- [74] Ship JA, Pillemer SR, Baum BJ. Xerostomia and the geriatric patient. *J Am Geriatr Soc* 2002;50:535–43. <https://doi.org/10.1046/j.1532-5415.2002.50123.x>.
- [75] Saadoun D, Terrier B, Bannock J, Vazquez T, Massad C, Kang I, et al. Expansion of autoreactive unresponsive CD21⁻/low B cells in sjögren's syndrome-associated lymphoproliferation. *Arthritis Rheum* 2013;65:1085–96. <https://doi.org/10.1002/art.37828>.
- [76] Filippi M, Bar-Or A, Piehl F, Preziosa P, Solari A, Vukusic S, et al. Multiple sclerosis. *Nat Rev Dis Prim* 2018;4:1–27. <https://doi.org/10.1038/s41572-018-0041-4>.
- [77] Harbo HF, Gold R, Tintora M. Sex and gender issues in multiple sclerosis. *Ther Adv Neurol Disord* 2013;6:237–48. <https://doi.org/10.1177/1756285613488434>.
- [78] Lublin FD, Reingold SC, Cohen JA, Cutter GR, Sorensen PS, Thompson AJ, et al. Defining the clinical course of multiple sclerosis: the 2013 revisions. *Neurology* 2014;83:278–86. <https://doi.org/10.1212/WNL.0000000000000560>.
- [79] International Multiple Sclerosis Genetics.. Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. *Nat Genet* 2013;45:1353–62. <https://doi.org/10.1038/ng.2770>.
- [80] International Multiple Sclerosis Genetics.. A systems biology approach uncovers cell-specific gene regulatory effects of genetic associations in multiple sclerosis. *Nat Commun* 2019;10:2236. <https://doi.org/10.1038/s41467-019-09773-y>.
- [81] Patsopoulos NA, Baranzini SE, Santaniello A, Shoostari P, Cotsapas C, Wong G, et al. Multiple sclerosis genomic map implicates peripheral immune cells and microglia in susceptibility. *Science* 2019;1:25. <https://doi.org/10.1126/science.aav7188>.
- [82] International Multiple Sclerosis Genetics.. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature* 2011;476:214–9. <https://doi.org/10.1038/nature10251>.
- [83] Calvo-Barreiro L, Eixarch H, Montalban X, Espejo C. Combined therapies to treat complex diseases: the role of the gut microbiota in multiple sclerosis. *Autoimmun Rev* 2018;17:165–74. <https://doi.org/10.1016/j.autrev.2017.11.019>.
- [84] Olsson T, Barcellos LF, Alfredsson L. Interactions between genetic, lifestyle and environmental risk factors for multiple sclerosis. *Nat Rev Neurol* 2016;13:25–36. <https://doi.org/10.1038/nrneurol.2016.187>.
- [85] Bielekova B, Sung M-H, Kadom N, Simon R, McFarland H, Martin R. Expansion and functional relevance of high-avidity myelin-specific CD4+ T cells in multiple sclerosis. *J Immunol* 2004;172:3893–904. <https://doi.org/10.4049/jimmunol.172.6.3893>.
- [86] Planas R, Santos R, Tomas-Ojer P, Cruciani C, Lutterotti A, Faigle W, et al. GDP-fucose synthase is a CD4+ T cell-specific autoantigen in DRB3*02:02 patients with multiple sclerosis. *Sci Transl Med* 2018;10:1–16. <https://doi.org/10.1126/scitranslmed.aat4301>.
- [87] Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, et al. The relation between inflammation and neurodegeneration in multiple sclerosis brains. *Brain* 2009;132:1175–89. <https://doi.org/10.1093/brain/awp070>.
- [88] Lünemann JD, Jelčić I, Roberts S, Lutterotti A, Tackenberg B, Martin R, et al. EBNA1-specific T cells from patients with multiple sclerosis cross react with myelin antigens and co-produce IFN- γ and IL-2. *J Exp Med* 2008;205:1763–73. <https://doi.org/10.1084/jem.20072397>.
- [89] Vaughn CB, Jakimovski D, Kavak KS, Ramanathan M, Benedict RHB, Zivadinov R, et al. Epidemiology and treatment of multiple sclerosis in elderly populations. *Nat Rev Neurol* 2019;15:329–42. <https://doi.org/10.1038/s41582-019-0183-3>.
- [90] Bolton C, Smith PA. The influence and impact of ageing and immunosenescence (ISC) on adaptive immunity during multiple sclerosis (MS) and the animal counterpart experimental autoimmune encephalomyelitis (EAE). *Ageing Res Rev* 2018;41:64–81. <https://doi.org/10.1016/j.arr.2017.10.005>.
- [91] Bjelobaba I, Begovic-Kupresanin V, Pekovic S, Lavrnja I. Animal models of multiple sclerosis: focus on experimental autoimmune encephalomyelitis. *J Neurosci Res* 2018;96:1021–42. <https://doi.org/10.1002/jnr.24224>.
- [92] Denic A, Johnson AJ, Bieber AJ, Warrington AE, Rodriguez M, Pirko I. The relevance of animal models in multiple sclerosis research. *Pathophysiology* 2011;18:21–9. <https://doi.org/10.1038/jid.2014.371>.
- [93] Hauser SL, Bhan AK, Gilles F, Kemp M, Kerr C, Weiner HL. Immunohistochemical analysis of the cellular infiltrate in multiple sclerosis lesions. *Ann Neurol* 1986;19:578–87. <https://doi.org/10.1002/ana.410190610>.
- [94] Aharoni R, Aricha R, Eilam R, From I, Mizrahi K, Arnon R, et al. Age dependent course of EAE in Aire^{-/-} mice. *J Neuroimmunol* 2013;262:27–34. <https://doi.org/10.1016/j.jneuroim.2013.06.001>.
- [95] Zhang H, Podojil JR, Luo X, Miller SD. Intrinsic and induced regulation of the age-associated onset of spontaneous experimental autoimmune encephalomyelitis. *J Immunol* 2008;181:4638–47. <https://doi.org/10.4049/jimmunol.181.7.4638>.
- [96] Ditamo Y, Degano AL, Maccio DR, Pistoiresi-Palencia MC, Roth GA. Age-related changes in the development of experimental autoimmune encephalomyelitis. *Immunol Cell Biol* 2005;83:75–82. <https://doi.org/10.1111/j.1440-1711.2004.01294.x>.
- [97] Seo JE, Hasan M, Han JS, Kang MJ, Jung BH, Kwok SK, et al. Experimental autoimmune encephalomyelitis and age-related correlations of NADPH oxidase, MMP-9, and cell adhesion molecules: the increased disease severity and blood-brain barrier permeability in middle-aged mice. *J Neuroimmunol* 2015;287:43–53. <https://doi.org/10.1016/j.jneuroim.2015.08.005>.
- [98] Hampton DW, Innes N, Merkler D, Zhao C, Franklin RJM, Chandran S. Focal immune-mediated white matter demyelination reveals an age-associated increase in axonal vulnerability and decreased remyelination efficiency. *Am J Pathol* 2012;180:1897–905. <https://doi.org/10.1016/j.ajpath.2012.01.018>.
- [99] Di Benedetto S, Derhovanessian E, Steinhagen-Thiessen E, Goldeck D, Müller L, Pawelec G. Impact of age, sex and CMV-infection on peripheral T cell phenotypes: results from the Berlin BASE-II Study. *Biogerontology* 2015;16:631–43. <https://doi.org/10.1007/s10522-015-9563-2>.
- [100] Kritsilis M, Rizou SV, Koutsoudaki PN, Evangelou K, Gorgoulis VG, Papadopoulos D. Ageing, cellular senescence and neurodegenerative disease. *Int J Mol Sci* 2018;19:1–37. <https://doi.org/10.3390/ijms19102937>.
- [101] Tutuncu M, Tang J, Zeid NA, Kale N, Crusan DJ, Atkinson EJ, et al. Onset of progressive phase is an age-dependent clinical milestone in multiple sclerosis. *Mult Scler* 2013;19:188–98. <https://doi.org/10.1177/1352458512451510.Onset>.
- [102] Musella A, Gentile A, Rizzo FR, De Vito F, Fresogna D, Bullitta S, et al. Interplay between age and neuroinflammation in multiple sclerosis: effects on motor and cognitive functions. *Front Aging Neurosci* 2018;10:1–13. <https://doi.org/10.3389/fnagi.2018.00238>.
- [103] Pender MP, Csurhes PA, Pfluger CM, Burrows SR. CD8 T cell deficiency impairs control of Epstein-Barr virus and worsens with age in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 2012;83:353–4. <https://doi.org/10.1136/jnnp.2011.241349>.
- [104] Thewissen M, Linsen L, Somers V, Geusens P, Raus J, Stinissen P. Premature immunosenescence in rheumatoid arthritis and multiple sclerosis patients. *Ann N Y Acad Sci* 2005;1051:255–62. <https://doi.org/10.1196/annals.1361.066>.
- [105] Redondo J, Sarkar P, Kemp K, Virgo PF, Pawade J, Norton A, et al. Reduced cellularity of bone marrow in multiple sclerosis with decreased MSC expansion potential and premature ageing in vitro. *Mult Scler* 2018;24:919–31. <https://doi.org/10.1177/1352458517711276>.
- [106] Guan JZ, Guan WP, Maeda T, Guoqing X, GuangZhi W, Makino N. Patients with multiple sclerosis show increased oxidative stress markers and somatic telomere length shortening. *Mol Cell Biochem* 2014;400:183–7. <https://doi.org/10.1007/s11010-014-2274-1>.
- [107] Habib R, Ocklenburg S, Hoffjan S, Haghikia A, Epplen JT, Arning L. Association between shorter leukocyte telomeres and multiple sclerosis. *J Neuroimmunol* 2020;341:1–5. <https://doi.org/10.1016/j.jneuroim.2020.577187>.

- [108] Haegert DG, Hackenbroch JD, Duszczyszyn D, Fitz-Gerald L, Zastepa E, Mason H, et al. Reduced thymic output and peripheral naïve CD4 T-cell alterations in primary progressive multiple sclerosis (PPMS). *J Neuroimmunol* 2011;233:233–9. <https://doi.org/10.1016/j.jneuroim.2010.12.007>.
- [109] Balint B, Haas J, Schwarz A, Jarius S, Fürwentsches A, Engelhardt K, et al. T-cell homeostasis in pediatric multiple sclerosis: old cells in young patients. *Neurology* 2013;81:784–92. <https://doi.org/10.1212/WNL.0b013e3182a2ce0e>.
- [110] Broux B, Mizze MR, Vanheusden M, van der Pol S, van Horsen J, Van Wijmeersch B, et al. IL-15 amplifies the pathogenic properties of CD4 + CD28 – T cells in multiple sclerosis. *J Immunol* 2015;194:2099–109. <https://doi.org/10.4049/jimmunol.1401547>.
- [111] Schwarz A, Balint B, Korporal-Kuhnke M, Jarius S, Von Engelhardt K, Fürwentsches A, et al. B-cell populations discriminate between pediatric- and adult-onset multiple sclerosis. *Neurol Neuroimmunol NeuroInflamm* 2017;4:1–9. <https://doi.org/10.1212/NXI.0000000000000309>.
- [112] Claes N, Fraussen J, Vanheusden M, Hellings N, Stinissen P, Van Wijmeersch B, et al. Age-associated B cells with proinflammatory characteristics are expanded in a proportion of multiple sclerosis patients. *J Immunol* 2016;197:4576–83. <https://doi.org/10.4049/jimmunol.1502448>.
- [113] Frau J, Coghe G, Lorefice L, Fenu G, Cocco E. New horizons for multiple sclerosis therapeutics: milestones in the development of ocrelizumab. *Neuropsychiatr Dis Treat* 2018;14:1093–9. <https://doi.org/10.2147/NDT.S147874>.
- [114] Hartung H, Gonsette R, König N, Kwieciński H, Guseo A, Morrissey SP, et al. Mitoxantrone in progressive multiple sclerosis: a placebo-controlled, double-blind, randomised, multicentre trial. *Lancet* 2002;360:2018–25. [https://doi.org/10.1016/S0140-6736\(02\)12023-X](https://doi.org/10.1016/S0140-6736(02)12023-X).
- [115] Oost W, Talma N, Meilof JF, Laman JD. Targeting senescence to delay progression of multiple sclerosis. *J Mol Med* 2018;96:1153–66. <https://doi.org/10.1007/s00109-018-1686-x>.
- [116] Azizi G, Goudarzvand M, Afræi S, Sedaghat R, Mirshafiey A. Therapeutic effects of dasatinib in mouse model of multiple sclerosis. *Immunopharmacol Immunotoxicol* 2015;37:287–94. <https://doi.org/10.3109/08923973.2015.1028074>.
- [117] Sternberg Z, Chadha K, Lieberman A, Hojnacki D, Drake A, Zamboni P, et al. Quercetin and interferon- β modulate immune response(s) in peripheral blood mononuclear cells isolated from multiple sclerosis patients. *J Neuroimmunol* 2008;205:142–7. <https://doi.org/10.1016/j.jneuroim.2008.09.008>.
- [118] Xu M, Pirtskhalava T, Farr JN, Weigand BM, Palmer AK, Weivoda MM, et al. Senolytics improve physical function and increase lifespan in old age. *Nat Med* 2018;24:1246–56. <https://doi.org/10.1038/s41591-018-0092-9>.
- [119] Chang J, Wang Y, Shao L, Laberge RM, Demaria M, Campisi J, et al. Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat Med* 2016;22:78–83. <https://doi.org/10.1038/nm.4010>.
- [120] Chen C, Liu Y, Liu Y, Zheng P. MTOR regulation and therapeutic rejuvenation of aging hematopoietic stem cells. *Sci Signal* 2009;2:1–8. <https://doi.org/10.1126/scisignal.2000559>.
- [121] Florian MC, Dörr K, Niebel A, Daria D, Schrezenmeier H, Rojewski M, et al. Cdc42 activity regulates hematopoietic stem cell aging and rejuvenation. *Cell Stem Cell* 2012;10:520–30. <https://doi.org/10.1016/j.stem.2012.04.007>.
- [122] Ito K, Hirao A, Arai F, Takubo K, Matsuoka S, Miyamoto K, et al. Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat Med* 2006;12:446–51. <https://doi.org/10.1038/nm1388>.
- [123] Jung H, Kim DO, Byun JE, Kim WS, Kim MJ, Song HY, et al. Thioredoxin-interacting protein regulates haematopoietic stem cell ageing and rejuvenation by inhibiting p38 kinase activity. *Nat Commun* 2016;7:1–12. <https://doi.org/10.1038/ncomms13674>.
- [124] Neumann B, Baror R, Zhao C, Segel M, Dietmann S, Rawji KS, et al. Metformin restores CNS remyelination capacity by rejuvenating aged stem cells. *Cell Stem Cell* 2019;25:473–85. <https://doi.org/10.1016/j.stem.2019.08.015>.
- [125] Ruckh JM, Zhao JW, Shadrach JL, Van Wijngaarden P, Rao TN, Wagers AJ, et al. Rejuvenation of regeneration in the aging central nervous system. *Cell Stem Cell* 2012;10:96–103. <https://doi.org/10.1016/j.stem.2011.11.019>.
- [126] Natrajan MS, De La Fuente AG, Crawford AH, Linehan E, Nuñez V, Johnson KR, et al. Retinoid X receptor activation reverses age-related deficiencies in myelin debris phagocytosis and remyelination. *Brain* 2015;138:3581–97. <https://doi.org/10.1093/brain/awv289>.
- [127] Rawji KS, Young AMH, Ghosh T, Michaels NJ, Mirzaei R, Kappen J, et al. Niacin-mediated rejuvenation of macrophage/microglia enhances remyelination of the aging central nervous system. *Acta Neuropathol* 2020;139:893–909. <https://doi.org/10.1007/s00401-020-02129-7>.
- [128] Coll RC, Robertson AAB, Chae JJ, Higgins SC, Muñoz-Planillo R, Insera MC, et al. A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. *Nat Med* 2015;21:248–57. <https://doi.org/10.1038/nm.3806>.
- [129] Malhotra S, Costa C, Eixarch H, Keller CW, Amman L, Martínez-Banaclocha H, et al. NLRP3 inflammasome as prognostic factor and therapeutic target in primary progressive multiple sclerosis patients. *Brain* 2020;143:1414–30. <https://doi.org/10.1093/brain/awaa084>.
- [130] Thomas R, Wang W, Su DM. Contributions of age-related thymic involution to immunosenescence and inflammaging. *Immun Ageing* 2020;17:1–17. <https://doi.org/10.1186/s12979-020-0173-8>.
- [131] Aspinall R, Pido-Lopez J, Imami N, Henson SM, Ngom PT, Morre M, et al. Old rhesus macaques treated with interleukin-7 show increased TREC levels and respond well to influenza vaccination. *Rejuvenation Res* 2007;10:5–17. <https://doi.org/10.1089/rej.2006.9098>.
- [132] Dudakov JA, Hanash AM, Jenq RR, Young LF, Ghosh A, Singer NV, et al. Interleukin-22 drives endogenous thymic regeneration in mice. *Science* 2012;335:91–5. <https://doi.org/10.1126/science.1218004>.
- [133] Min D, Panoskaltis-Mortari A, Kuro-o M, Holländer GA, Blazar BR, Weinberg KI. Sustained thymopoiesis and improvement in functional immunity induced by exogenous KGF administration in murine models of aging. *Blood* 2007;109:2529–37. <https://doi.org/10.1182/blood-2006-08-043794>.
- [134] Coder B, Wang W, Wang L, Wu Z, Zhuge Q, Su DM. Friend or foe: the dichotomous impact of T cells on neuro-de/re-generation during aging. *Oncotarget* 2017;8:7116–37. <https://doi.org/10.18632/oncotarget.12572>.
- [135] Zhang J, Medaer R, Stinissen P, Hafler D, Raus J. MHC-restricted depletion of human myelin basic protein-reactive T cells by T cell vaccination. *Science* 1993;261:1451–4. <https://doi.org/10.1126/science.7690157>.
- [136] Choi IY, Piccio L, Childress P, Bollman B, Ghosh A, Brandhorst S, et al. A diet mimicking fasting promotes regeneration and reduces autoimmunity and multiple sclerosis symptoms. *Cell Rep* 2016;15:2136–46. <https://doi.org/10.1016/j.celrep.2016.05.009>.
- [137] Harley CB, Liu W, Blasco M, Vera E, Andrews WH, Briggs LA, et al. A natural product telomerase activator as part of a health maintenance program. *Rejuvenation Res* 2011;14:45–56. <https://doi.org/10.1089/rej.2010.1085>.
- [138] Warrington KJ, Vallejo AN, Weyand CM, Goronzy JJ. CD28 loss in senescent CD4 + T cells: reversal by interleukin-12 stimulation. *Blood* 2003;101:3543–9. <https://doi.org/10.1182/blood-2002-08-2574>.
- [139] Topp MS, Riddell SR, Akatsuka Y, Jensen MC, Blattman JN, Greenberg PD. Restoration of CD28 expression in CD28- CD8+ memory effector T cells reconstitutes antigen-induced IL-2 production. *J Exp Med* 2003;198:947–55. <https://doi.org/10.1084/jem.20021288>.
- [140] Babic M, Dimitropoulos C, Hammer Q, Stehle C, Heinrich F, Sarsenbayeva A, et al. NK cell receptor NKG2D enforces proinflammatory features and pathogenicity of Th1 and Th17 cells. *J Exp Med* 2020;217:1–16. <https://doi.org/10.1084/jem.20190133>.