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# Oligonucleotides containing a piperazino-modified 2'-amino-LNA monomer exhibit very high duplex stability and remarkable nuclease resistance†

Chenguang Lou,<sup>a</sup> Birte Vester<sup>b</sup> and Jesper Wengel<sup>\*a</sup>

**Incorporation of a piperazino-modified 2'-amino-LNA monomer (PipLNA-T) into oligonucleotides conferred very high affinity and base-pairing selectivity towards complementary DNA and RNA strands. Furthermore, one PipLNA-T modification provided a robust nuclease resistance that safeguarded three neighbouring natural nucleosides from 3'-exonucleolytic degradation. These favourable properties render PipLNA-T a promising oligonucleotide modification for various biological applications.**

DNA encodes the genetic information in living organisms and has attracted extensive and continuous interest in the areas of biochemistry, biology and nanotechnology within the past few decades.<sup>1–4</sup> Many modified nucleotide monomers have been incorporated into oligonucleotides (ONs) in order to optimise affinity and sequence selectivity towards complementary nucleic acids and to obtain nuclease resistance.<sup>5–9</sup> Locked nucleic acid (LNA) monomers represent a class of nucleotides containing ribofuranose units locked into a C3'-endo (*North-type*) conformation,<sup>10,11</sup> and incorporation of LNA-T and 2'-amino-LNA-T monomers (Fig. 1) into ONs provides increased duplex stability, favorable mismatch discrimination and improved resistance to nucleolytic digestion.<sup>12–16</sup>

Besides LNA, the introduction of alkylamine chains into ONs is recognised as a successful strategy to improve binding affinity, probably due to electrostatic interactions between the protonated amine functionalities and the negatively charged phosphodiester backbone.<sup>17–20</sup> Modification of ONs with a piperazino group introduces a cationic center under physiological conditions,

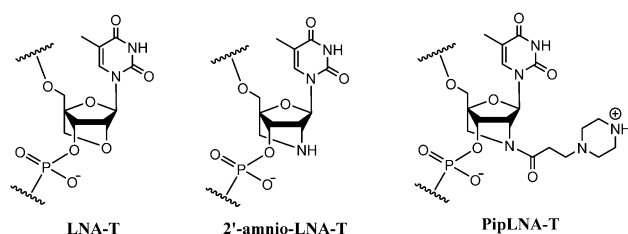


Fig. 1 Chemical structures of LNA-T, 2'-amino-LNA-T and PipLNA-T monomers.

and provides a possibility for further functionalisation.<sup>17,19,21</sup> Therefore, we decided to investigate a piperazino-modified 2'-amino-LNA monomer (PipLNA-T; Fig. 1) by combining the advantageous characteristics of the 2'-amino-LNA and piperazino scaffolds.

Synthesis of the PipLNA-T phosphoramidite monomer **5** was accomplished in three steps from 3-piperazinopropionic acid (**1**) and a DMTr-protected 2'-amino-LNA-T precursor **3**<sup>22</sup> (Scheme 1).

The secondary amine of derivative **1** was reacted with *N*-(9-fluorenylmethoxycarbonyloxy)succinimide (Fmoc-OSu) in pyridine to produce **2** in 88% yield. The carboxylic group of **2** was subsequently reacted with 2'-amino-LNA nucleoside **3** in anhydrous DMF in the presence of 1-[bis(dimethylamino)methylene]-1*H*-1,2,3-triazolo-[4,5-*b*]pyridinium 3-oxid hexafluorophosphate (HATU) and *N,N*-diisopropylethylamine (DIPEA) to give nucleoside **4** in 84% yield, which was finally phosphitylated by reaction with bis(diisopropylamino)(2-cyanoethoxy)phosphine to provide the desired phosphoramidite monomer **5** in 67% yield.

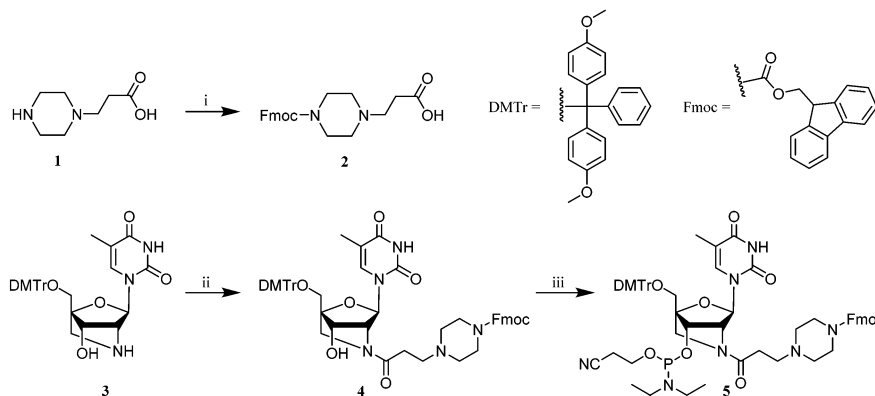
ON synthesis was carried out using standard solid-phase phosphoramidite chemistry on an automated DNA synthesiser where the PipLNA-T phosphoramidite monomer **5** was coupled using a hand-coupling process.<sup>23</sup> Fmoc protection of the secondary amine of PipLNA-T not only prevents it from being irreversibly acetylated in the capping step of the standard synthesis cycle, but also allows ON deprotection under mild conditions after completion of ON synthesis.<sup>24,25</sup> The stepwise coupling yield of 2'-amino-LNA-T and PipLNA-T were 96% and 80%, respectively. The imperfect

<sup>a</sup> Biomolecular Nanoscale Engineering Center, Department of Physics, Chemistry and Pharmacy, University of Southern Denmark, Campusvej 55, 5230 Odense M, Denmark. E-mail: jwe@sdu.dk; Tel: +45 65502510

<sup>b</sup> Department of Biochemistry and Molecular Biology, University of Southern Denmark, Campusvej 55, 5230 Odense M, Denmark

† Electronic supplementary information (ESI) available: Details on synthesis of PipLNA-T monomer, oligonucleotide synthesis, purification and analysis (IE-HPLC and MAIDI-TOF MS), ultraviolet duplex melting studies, ultraviolet triplex melting studies, nuclease resistance assay, molecular modelling; copies of representative IE-HPLC curves, MAIDI-TOF mass spectra, and UV-melting curves. See DOI: 10.1039/c5cc00322a





**Scheme 1** Reagents and conditions: (i) Fmoc-OSu, DIPEA, pyridine, RT, 12 h, 88%; (ii) **2**, HATU, DIPEA, DMF, RT, 1 h, 84%; (iii) bis(diisopropylamino)(2-cyanoethoxy)phosphine, diisopropylammonium tetrazolide, DCM, 18 h, RT, 67%.

coupling efficiency on PipLNA-T monomer may result from a close proximity of the bulky Fmoc-protected piperazino moiety and the diisopropylamino leaving group.

The synthesised ONs were purified by DMTr-ON reversed-phase HPLC (and/or ion-exchange HPLC) and their composition and purity (>90%) characterised by MAIDI-TOF MS and ion-exchange HPLC analysis, respectively.

A 9-mer sequence<sup>12,17</sup> was chosen for evaluation of the binding affinity of PipLNA-containing ONs towards complementary DNA and RNA strands (Table 1). In comparison with all DNA (**ON1**) and 2'-amino-LNA-T-containing controls (**ON2** and **ON4**), ONs containing one and three PipLNA-T modifications were studied (**ON3** and **ON5**). The data show that a PipLNA-T monomer confers substantially higher duplex stability than thymidine (T) and 2'-amino-LNA-T in DNA-DNA contexts. The same tendency was observed for the corresponding DNA-RNA hybrids, but with even greater enhancement of stability relative to T and less dramatic difference between the PipLNA-T and the 2'-amino-LNA-T monomers. These hybridisation data are consistent with previous results showing that ONs containing 2'-amino-LNA-T exhibit higher affinity towards complementary RNA than DNA,<sup>12,26</sup> but show a remarkable additional affinity-enhancing effect due to the incorporation of the piperazino group in monomer PipLNA-T.

The effect of PipLNA-T monomers on the thermal stability of duplexes is cumulative. This is demonstrated by the results that three PipLNA-T monomers provide far greater duplex stability

than a single modification irrespective of either a DNA or RNA complementary strand.

In order to determine the importance of protonation of the distal nitrogen atom of the piperazino moiety ( $pK_a \approx 9$  (ref. 27)), thermal stability of six duplexes formed by **ON1**, **ON4** and **ON5** with complementary DNA and RNA were evaluated at low salt buffer condition (6.7 mM  $\text{NaH}_2\text{PO}_4$ - $\text{Na}_2\text{HPO}_4$  buffer and 0.10 mM EDTA, Table 1). Smaller decreases in melting temperature were found for the duplexes involving **ON5** with three PipLNA-T nucleotides than for the two control duplexes involving **ON1** and **ON4**, suggesting that decreased duplex stability under low salt conditions can be alleviated by a stabilising effect from the positively-charged piperazino moieties of PipLNA-T. A similar effect was not observed for **ON4** containing three 2'-amino-LNA-T monomers, which is consistent with the reported  $pK_a$  value of 6.17 for the N2'-protonated derivative of the 2'-amino-LNA-T monomer.<sup>28</sup>

The mismatch discrimination ability of the novel PipLNA-T monomer was assessed for the central position in **ON3** and **ON5** in comparison to all-DNA control **ON1** (Table S3, ESI<sup>†</sup>). Singly-modified **ON3** showed higher base-pairing specificity for its correctly matched complementary strands than **ON1**, except for slightly reduced discrimination in the case of the mismatched RNA target having a cytidine monomer in the central position. For **ON5** (three substitutions with PipLNA-T), the efficient mismatch discrimination ability of PipLNA-T was similar to that of DNA-T.

**Table 1** Thermal denaturation temperatures of duplexes<sup>a</sup>

Sequence	X		Complementary DNA		Complementary RNA	
			Medium salt	Low salt	Medium salt	Low salt
5'-GTGATATGC	—	<b>ON1</b>	32.0	18.0	29.5	16.0
5'-GTGAXATGC	2'-Amino-LNA-T	<b>ON2</b>	35.5 (+3.5)		37.0 (+7.5)	
	PipLNA-T	<b>ON3</b>	39.0 (+7.0)		38.5 (+9.0)	
5'-GXGAXXGC	2'-Amino-LNA-T	<b>ON4</b>	41.0 (+9.0)	27.5	49.0 (+19.5)	36.0
	PipLNA-T	<b>ON5</b>	49.5 (+17.5)	39.0	54.0 (+24.5)	42.5

<sup>a</sup>  $T_m$  values (°C) of unmodified and modified (X = 2'-amino-LNA-T or PipLNA-T) DNA-DNA and DNA-RNA duplexes measured as an average of two independent melting temperature determinations with a deviation <0.5 °C. Numbers in brackets are  $\Delta T_m$  measured as the difference in  $T_m$  values between modified and unmodified duplexes. The experiments were carried out at pH 7.0 in medium salt buffer (5.8 mM  $\text{NaH}_2\text{PO}_4$ - $\text{Na}_2\text{HPO}_4$  buffer, containing 100 mM NaCl and 0.10 mM EDTA) and low salt buffer (6.7 mM  $\text{NaH}_2\text{PO}_4$ - $\text{Na}_2\text{HPO}_4$  buffer, containing 0.10 mM EDTA). The increase in melting temperature of 2'-amino-LNA-T is in a general agreement with previous results reported for **ON2** and **ON4**.<sup>12</sup>



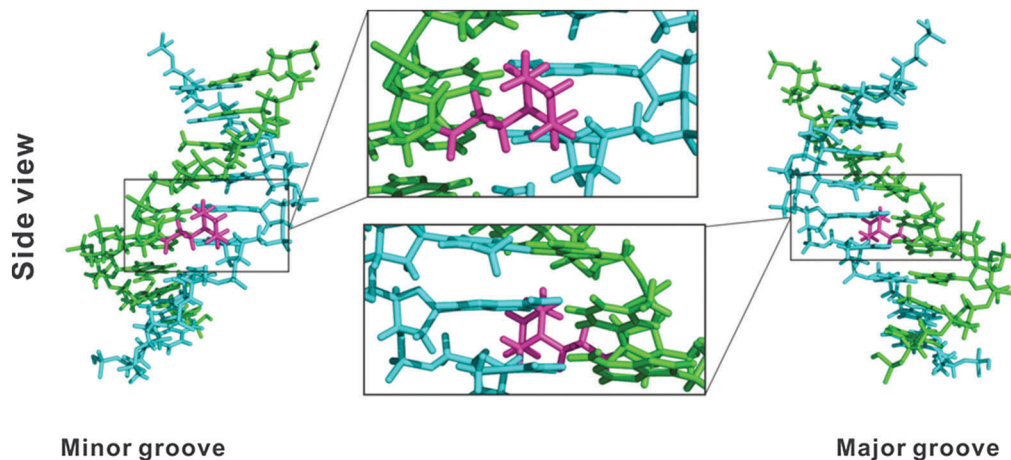


Fig. 2 Modelling structure of a PipLNA-T-modified duplex formed between 5'-GTGAXATGC (**ON3**; green) and its complementary DNA strand (cyan). The 3-(1-piperazino)propanoyl moiety is marked in magenta. Side views are given from the minor and major grooves.

The ability of the PipLNA-T monomer to partake in triplex formation was evaluated using a 29-mer dsDNA target containing the wild-type HIV-1 polypurine tract sequence<sup>29</sup> (Table S2, ESI<sup>†</sup>). Compared to a wild-type all-DNA TFO (Table S5, ESI<sup>†</sup>), incorporation of PipLNA-T greatly improved the triplex stability at both pH 6.0 and 7.0. Interestingly, no obvious improvement in triplex stability was observed when LNA-T was replaced by PipLNA-T, possibly due to the short propanoyl tether between the moderately bulky piperazino group and the nucleoside itself, restricting the conformational flexibility which could prevent the piperazino group from finding an optimal position for triplex stabilisation.

Molecular modelling<sup>30,31</sup> (Fig. 2) on the duplex **ON3**:DNA with one PipLNA-T substitution indicated that the 3-(1-piperazino)propanoyl group of PipLNA-T is oriented in the minor groove of the double helix. The distances measured in the model structure between the protonated amino group of the piperazino ring and the four nearest negatively-charged oxygen anions on the phosphate backbone are ~8–10 Å, which implies that any favourable electrostatic interaction has to be mediated through water molecule(s)<sup>32,33</sup> in the minor groove.

It is known that 2'-modified nucleotides usually increase nuclease resistance of ONs.<sup>34–36</sup> In order to evaluate their resistance towards nucleolytic degradation, we studied the stability of 5'-<sup>32</sup>P labeled **ON1–ON3** upon incubation with snake venom phosphodiesterase (SVPDE; 3'-exonuclease). As shown in Fig. 3, the unmodified oligonucleotide (X = T, **ON1**) was rapidly degraded, with no trace of full-length **ON1** left after 10 min incubation. The degradation progressed over time with bands corresponding to 3- and 4-mers and <sup>32</sup>P-labelled guanosine monophosphate remaining after 60 min. Also **ON2** with the central thymidine substituted by a 2'-amino-LNA-T monomer, showed no full-length product after 15 min incubation. Intriguingly, the degradation was severely inhibited once the TGC-3' triad was digested and the resulting 5'-GTGAXA exhibited very strong nuclease resistance with only little further digestion after 60 min incubation. This shows that a 2'-amino-LNA-T monomer is much more nuclease-resistant than its natural counterpart and that it protects the 3'-neighbouring nucleotide from 3'-exonucleolytic cleavage.

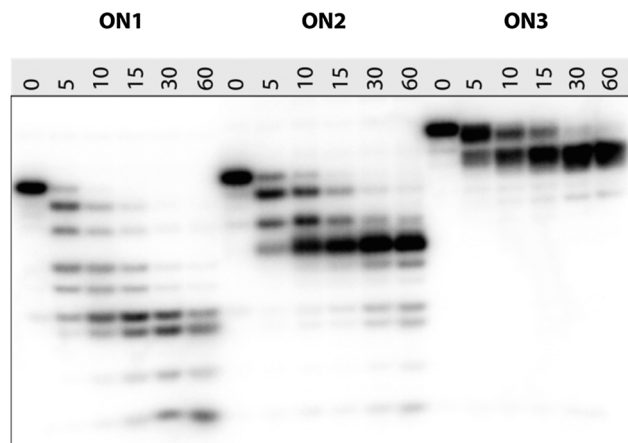


Fig. 3 20% PAGE denaturing gel showing the time-course of SVPDE-mediated degradation of 5'-<sup>32</sup>P-GTGAXATGC **ON1** (X = T), **ON2** (X = 2'-amino-LNA-T) and **ON3** (X = PipLNA-T) under pH 8.0 at 21 °C. Samples were incubated for 5, 10, 15, 30 and 60 min. The negative control (0 min) was taken before the enzyme was added. The gel was visualized by autoradiography.

**ON3** (X = PipLNA-T) showed retarded mobility in the gel compared to **ON1** and **ON2**, probably due to a combination of reduced overall negative charge because of protonation of the piperazino moiety and increased bulkiness. Fig. 3 demonstrates that a single, centrally positioned PipLNA-T monomer when incorporated into the 9-mer ON induces significantly improved resistance to SVPDE when compared to both 2'-amino-LNA-T and DNA-T monomers. This counts with respect to full length product of which a trace remained after 15 min incubation, but more pronouncedly also by the remarkable stability of the 8-mer product resulting from cleavage of the 3'-nucleotide. This product is well preserved even after 60 min incubation. This implies that a PipLNA-T modification protects its neighbourhood up to three natural nucleosides towards its 3'-end from 3'-exonucleolytic degradation. A similar mechanism has also been proposed for C5-substituted LNA pyrimidines.<sup>20</sup> This behaviour strongly contrasts that of a single LNA nucleotide which has been



reported to confer no or only little resistance towards exonucleolytic degradation,<sup>37,38</sup> and suggests that the PipLNA-modification may prove uniquely suited for increasing the biostability of antisense ONs.

In conclusion, a novel PipLNA-T phosphoramidite monomer has been synthesised from a known 2'-amino-LNA-T precursor and has been incorporated into oligonucleotides. The PipLNA-T monomer obeys the Watson–Crick base pairing rules, and has a pronounced stabilising effect on nucleic acid duplexes and triplexes and a strong stabilising effect against 3'-exonucleolytic degradation. These properties make PipLNA-modified oligonucleotides attractive in the context of various biomedical and nanotechnological applications.

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