

## Common Genetic Variation at *BARD1* Is Not Associated with Breast Cancer Risk in *BRCA1* or *BRCA2* Mutation Carriers

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### Abstract

**Background:** Inherited *BRCA1* and *BRCA2* (*BRCA1/2*) mutations confer elevated breast cancer risk. Knowledge of factors that can improve breast cancer risk assessment in *BRCA1/2* mutation carriers may improve personalized cancer prevention strategies.

**Methods:** A cohort of 5,546 *BRCA1* and 2,865 *BRCA2* mutation carriers was used to evaluate risk of breast cancer associated with *BARD1* Cys557Ser. In a second nonindependent cohort of 1,537 of *BRCA1* and 839 *BRCA2* mutation carriers, *BARD1* haplotypes were also evaluated.

**Results:** The *BARD1* Cys557Ser variant was not significantly associated with risk of breast cancer from single SNP analysis, with a pooled effect estimate of 0.90 (95% CI: 0.71–1.15) in *BRCA1* carriers and 0.87 (95% CI: 0.59–1.29) in *BRCA2* carriers. Further analysis of haplotypes at *BARD1* also revealed no evidence that additional common genetic variation not captured by Cys557Ser was associated with breast cancer risk.

**Conclusion:** Evidence to date does not support a role for *BARD1* variation, including the Cys557Ser variant, as a modifier of risk in *BRCA1/2* mutation carriers.

**Impact:** Interactors of *BRCA1/2* have been implicated as modifiers of *BRCA1/2*-associated cancer risk. Our finding that *BARD1* does not contribute to this risk modification may focus research on other genes that do modify *BRCA1/2*-associated cancer risk. *Cancer Epidemiol Biomarkers Prev*; 20(5): 1032–38.  2011 AACR.

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## Introduction

There is substantial interindividual variability in age at cancer diagnosis in *BRCA1* and *BRCA2* mutation carriers, which persists even among relatives that carry the same *BRCA1* and *BRCA2* mutation (1). Variation in genes that interact with *BRCA1* and *BRCA2* in the recognition and repair of DNA damage are strong candidates for study as genetic modifiers of *BRCA1* and *BRCA2* cancer risk. The *BRCA1*–*BARD1* heterodimer is known to be important for *BRCA1* function, with interaction mediated through the ring finger domains of the 2 proteins (2). In addition, although there is no evidence for a direct interaction between *BARD1* and *BRCA2*, they do operate in the same DNA repair processes, exemplified by the fact that the *BRCA2* partner RAD51, *BARD1*, and *BRCA1* all relocate to proliferating cell nuclear antigen structures after irradiation (3).

The *BARD1* Cys557Ser SNP (rs28997576) was first reported as a germ line alteration in a sporadic breast/uterine tumor (4). This variant lies between the ankyrin repeats and BRCT domains of *BARD1*, and the ectopically expressed Cys557 protein has growth suppression and proapoptotic effects relative to 557Ser (5). This SNP (minor allele frequency in Europeans: 0.025) has been reported to be associated with both breast cancer in the general population and familial breast cancer, but results have not shown consistent across all studies (6–12). Stacey and colleagues (6) initially reported that the Cys557Ser variant was associated with increased breast cancer risk in 756 Icelandic mutation carriers who carry *BRCA2* 999del5 founder mutation [OR = 3.1; 95% CI: 1.2–8.4]. However, subsequent studies reported no elevated risk in

228 Nordic *BRCA1* and *BRCA2* carriers (OR = 0.8, 95% CI: 0.3–2.0; ref. 8), or in 1,207 Polish *BRCA1* mutation carriers (OR = 0.9, 95% CI: 0.4–2.2; ref. 10). There have been no previous haplotype-based studies assessing the role of *BARD1* variation in breast cancer risk in *BRCA1* and *BRCA2* carriers specifically.

To resolve whether *BARD1* is a modifier of *BRCA1* and *BRCA2*-associated breast cancer risk, we undertook a large study to comprehensively assess the association of *BARD1* Cys557Ser as well as haplotypic variation with cancer risk in *BRCA1* and *BRCA2* carriers.

## Materials and Methods

### Study sample

The design for this study has been described in detail previously (13). Briefly, eligible participants included adult women with documented disease-associated inherited mutations in *BRCA1* or *BRCA2*. Mutations were included in the analysis if they were pathogenic according to generally recognized criteria (14, 15). Two overlapping cohorts of women with disease-associated *BRCA1* and *BRCA2* mutations were identified (Table 1). First, a cohort of 5,546 *BRCA1* and 2,865 *BRCA2* mutation carriers from the multicenter CIMBA consortium (13) was used to evaluate risk of breast cancer associated with *BARD1* Cys557Ser. Second, a cohort of 1,537 of *BRCA1* and 839 *BRCA2* mutation carriers participating in the MAGIC consortium was used to further explore the relationship between *BARD1* haplotypes and breast cancer risk. Recruitment and genetic studies were approved by relevant ethics committees at all sites, and informed consent was obtained from each participant.

### Laboratory methods

For analysis of the *BARD1* Cys557Ser SNP, existing genotype data from *BRCA1* and *BRCA2* mutation carriers was requested from members of the CIMBA consortium. The primary methods used for genotyping were Sequenom iPLEX (EMBRACE, -HEBON, kConFab, SWE-BRCA, PISA, Penn, Austria, Mayo, FCCC, GEMO, Georgetown, HEBCS) and by Taqman assays (OUH, Baylor, Beth Israel, City of Hope, Creighton, Dana Farber, NorthShore, IHCC, UCLA, University of Chicago, University of Texas Health Science Center, University of Utah, and Women's College Hospital; ref. 16) Genotypes for the INHERIT samples were typed by direct sequencing using an ABI Prism 3730xl DNA Analyser automated sequencer, with version 3.1 of the Big Dye fluorescent method according to the manufacturer's instructions (Applied Biosystems). Sequence data were analyzed using the Staden preGap4 and Gap4 programs. Samples from IHCC were typed by PCR-RFLP (10). SNP quality control measures included more than 95% success rate, Hardy–Weinberg Equilibrium  $P > 0.005$ . In addition, concordance of more than 98% for duplicate samples was required for studies that had included 2% duplicated samples for quality control purposes (all studies undergoing Sequenom iplex for

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**Table 1.** Sample description and Cys557Ser association

Center	Data in Cys557Ser analysis		Data in haplotype analysis	
	BRCA1	BRCA2	BRCA1	BRCA2
Austria	285	122	196	60
Baylor			14	1
Beth Israel			7	12
City of Hope			68	42
Creighton			180	41
Dana Farber			90	38
EMBRACE	827	638		
NorthShore			31	20
Fox Chase	85	57	40	26
Georgetown	33	16	60	29
GEMO	1,140	559		
HEBON	777	294		
HEBCS	102	104		
IHCC	397			
INHERIT	73	82		
KConFab	506	400	379	302
MAGIC <sup>a</sup>	583	300		
Mayo	218	118	58	31
OUH	33	13		
Penn			202	98
Pisa (PBCS)	73	41		
Swe-BRCA	414	121		
UCLA			32	14
Univ of Chicago			27	11
UT Southwestern			28	19
Utah			99	78
Women's College			26	17
Total	5,546	2,865	1,537	839
Cys557Ser association	0.90 (0.71–1.15)	0.87 (0.59–1.29)	ND	ND

Abbreviation: ND, not done.

<sup>a</sup>Indicates MAGIC data included in single SNP analysis only.

BARD1 Cys557Ser, and all samples included in the haplotype substudy).

For studies of *BARD1* haplotypic variation, 11 haplotype tag SNPs were identified and assayed at the University of Pennsylvania as previously described (16). The rs IDs were as follows: rs6712055, rs16852689, rs280621, rs13021937, rs13423596, rs10190829, rs6751923, rs4234006, rs28997576, rs3768708, rs1374230.

### Statistical methods

To assess the relationship between *BARD1* SNPs and breast cancer risk, proportional hazards models were used as previously described (16, 17). Briefly, participants were followed from the time of genetic testing or study ascertainment until the first diagnosis of breast cancer (the primary event in this analysis) or were censored at ovarian cancer. Participants who developed breast cancer were censored at bilateral prophylactic mastectomy if it

occurred more than a year prior to the cancer diagnosis. This is to avoid censoring at bilateral mastectomies at which occult tumors were detected, but ages are rounded. The remaining participants were censored at the age at last observation. To address the problem of nonrandom sampling of mutation carriers with respect to the disease phenotype, analyses used the weighted Cox regression approach (17), where affected and unaffected individuals were differentially weighted such that observed breast cancer incidence rates in the study sample are consistent with established breast cancer risk estimates for *BRCA1* and *BRCA2* mutation carriers (18). Analyses assessing the association of the *BARD1* Cys557Ser SNP combined heterozygote and homozygote variant carriers under a dominant model because of the rare frequency of this variant. Analyses were assessed separately for *BRCA1* and *BRCA2* mutation carriers, adjusted for Study group, ethnicity (non-Jewish Caucasian, Jewish or other), and

**Table 2.** Analysis of BARD1 haplotype data: failure time analyses stratified on mutation using the MAGIC consortium data<sup>a</sup>

**Haplotypes detected in BRCA1 carriers<sup>b</sup>**

Ht no.	rs280621	rs28997576	rs1374230	rs10190829	rs4234006	rs16852689	rs3768708	rs6712055	rs13021937	Freq	HR	95% CI
B1H1	T	G	C	T	G	C	A	T	T	0.268	1.00	Reference
B1H2	C	<b>G</b>	T	A	G	C	A	T	T	0.011		
B1H3	T	G	T	A	G	C	G	T	T	0.008		
B1H4	T	G	C	A	G	C	A	C	T	0.009		
B1H5	T	G	T	A	A	T	A	T	T	0.047		
B1H6	T	G	T	T	A	C	G	T	C	0.009	1.07 <sup>b</sup>	0.85
B1H7	C	<b>G</b>	T	A	A	C	A	T	C	0.038		1.35
B1H8	C	<b>C</b>	T	A	G	C	A	T	T	0.025		
B1H9	T	G	C	A	G	C	A	T	T	0.005		
B1H10	C	G	T	T	A	C	G	T	C	0.008		
B1H11	T	G	C	T	G	C	A	C	C	0.019		
B1H12	T	G	T	A	G	C	A	C	T	0.056	1.15	0.82
B1H13	T	G	T	A	G	C	A	T	T	0.152	0.97	0.76
B1H14	T	G	T	A	G	C	G	C	C	0.203	1.00	0.80
B1H15	T	G	T	T	A	T	A	T	T	0.062	1.33	0.97
B1H16	T	G	T	T	G	C	A	T	T	0.052	0.99	0.66

**Haplotypes detected in BRCA2 carriers<sup>b</sup>**

Ht#	rs280621	rs28997576	rs1374230	rs6751923	rs16852689	rs13423596	rs3768708	rs6712055	rs13021937	Freq	HR	95% CI
B2H1	T	G	C	T	C	G	A	T	T	0.270	1.00	Reference
B2H2	T	G	T	C	C	G	A	T	T	0.016		
B2H3	C	G	T	C	C	G	G	T	C	0.012		
B2H4	C	G	T	C	C	G	A	T	C	0.043		
B2H5	T	G	T	C	C	G	A	T	C	0.004		
B2H6	T	G	T	C	C	G	A	C	T	0.049		
B2H7	T	G	C	C	C	G	A	C	T	0.020		
B2H8	C	G	T	C	C	G	A	T	T	0.008	0.88 <sup>b</sup>	0.64
B2H9	T	G	C	T	C	C	A	C	C	0.020		1.21
B2H10	C	<b>C</b>	T	T	C	A	A	C	T	0.019		
B2H11	T	G	T	T	C	G	G	T	T	0.007		
B2H12	T	G	T	T	C	G	G	C	T	0.004		
B2H13	T	G	T	T	C	G	G	T	C	0.018		
B2H14	T	G	T	T	C	G	A	T	T	0.172	0.82	0.54
B2H15	T	G	T	T	C	G	G	C	C	0.191	1.10	0.79
B2H16	T	G	T	T	T	A	A	T	T	0.062	0.62	0.37
B2H17	T	G	T	T	T	G	A	T	T	0.064	0.72	0.46

<sup>a</sup>The Cys557Ser variant is highlighted in bold.

<sup>b</sup>Haplotypes with frequency less than 5% were combined into a single "rare haplotype" analysis group.

year of birth cohort (decade of birth, categorized as <1940, 1940–1949, 1950–1959, 1960–1969, 1970–1989). There were 3,047 breast cancer events of 5,546 total for *BRCA1* (55%) and 1,578 breast cancer events of 2,865 total for *BRCA2* (55%) for the Cys557Ser censored analysis datasets. The remainders were censored for analysis. Secondary analyses adjusted for prophylactic oophorectomy, or assessed risk for the subset of carriers with mutations determined to result in unstable transcripts/proteins (class 1 loss of function mutations). R version 2.7.0 was used for single SNP statistical analyses.

To investigate haplotype effects, the Estimation-maximization algorithm (19, 20) was used to estimate haplotype frequencies as implemented in R version 2.1.1 subroutine haplo.em (21) as previously described (16). In this analysis, we included 607 breast cancer cases and 863 censored observations for *BRCA1*, and 813 breast cancer cases and 423 controls for *BRCA2*.

## Results and Discussion

The frequency of the Cys557Ser SNP in the combined dataset (Table 1) was similar to published reports, with 4.4% of individuals carrying at least 1 rare allele (4.5% in *BRCA1* carriers, 4.2% in *BRCA2* carriers). There were no significant associations of Cys557Ser and breast cancer risk for carriers of *BRCA1* mutations (HR = 0.90, 95% CI: 0.71–1.15) or *BRCA2* mutations (HR = 0.87, 95% CI: 0.59–1.29). There was no evidence for heterogeneity by center for either *BRCA1* or *BRCA2* analyses ( $P > 0.5$ ). There was also no evidence for association with additional adjustment for prophylactic oophorectomy, or when analyses were restricted to Class 1 mutations. For example, the HR for the subset of 3,882 individuals with *BRCA1* Class 1 mutations was 0.84 (0.62–1.15), and for the 2,668 individuals with *BRCA2* class 1 mutations was 0.96 (0.64–1.45).

For the haplotype analysis (Table 2), we also observed no overall effect of variation at *BARD1* in either *BRCA1* false discovery rate (FDR-corrected value of  $P = 0.152$ ) or *BRCA2* (FDR-corrected value of  $P = 0.134$ ). No single *BARD1* haplotype was significantly associated with breast cancer risk. Cys557Ser is represented by SNP 16 in Table 2 (*BRCA1* haplotype 8 and *BRCA2* haplotype 10). Since this variant was relatively rare (approximately 2% in both *BRCA1* and *BRCA2* carriers), estimates of its effect were not made in our primary analysis. When we fit a model that allowed the estimation of effects for haplotypes with at least 1% frequency in controls, no single haplotype was significantly associated with risk. The haplotype that contained the 557Ser allele was also not significantly associated with risk in either *BRCA1* (HR = 0.91, 95% CI: 0.45–1.85) or *BRCA2* (HR = 0.69, 95% CI: 0.28–1.72). Indeed, neither of these estimates was associated with increased risk of breast cancer as previously reported.

The data presented here do not provide evidence that neither the *BARD1* Cys557Ser SNP nor additional hap-

lotypic variability not captured by Cys557Ser is associated with breast cancer risk in *BRCA1* and *BRCA2* mutation carriers. Our sample size had more than 99% power to detect the effect size reported by Stacey and colleagues (6) of OR = 3.1. The study had more than 80% power to detect risk ratios of 0.89 (or 1.13) for *BRCA1* carriers and 0.86 (or 1.17) for *BRCA2* carriers. The upper 95% confidence limits on the rate ratio in our analysis exclude any substantial risk.

## Conclusion

Our study found no evidence to support substantial associations of *BARD1* variation with increased breast cancer risk in *BRCA1* and *BRCA2* carriers.

## Disclosure of Potential Conflicts of Interest

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## References

- Goldgar DE, Easton DF, Cannon-Albright LA, Skolnick MH. Systematic population-based assessment of cancer risk in first-degree relatives of cancer probands. *J Natl Cancer Inst* 1994;86:1600–8.
- Wu LC, Wang ZW, Tsan JT, Spillman MA, Phung A, Xu XL, et al. Identification of a RING protein that can interact in vivo with the BRCA1 gene product. *Nat Genet* 1996;14:430–40.
- Irminger-Finger I, Jefford CE. Is there more to BARD1 than BRCA1? *Nat Rev Cancer* 2006;6:382–91.
- Thai TH, Du F, Tsan JT, Jin Y, Phung A, Spillman MA, et al. Mutations in the BRCA1-associated RING domain (BARD1) gene in primary breast, ovarian and uterine cancers. *Hum Mol Genet* 1998;7:195–202.
- Sauer MK, Andrulis IL. Identification and characterization of missense alterations in the BRCA1 associated RING domain (BARD1) gene in breast and ovarian cancer. *J Med Genet* 2005;42:633–8.
- Stacey SN, Sulem P, Johannsson OT, Helgason A, Gudmundsson J, Kostic JP, et al. The BARD1 Cys557Ser variant and breast cancer risk in Iceland. *PLoS Med* 2006;3:e217.
- Vahteristo P, Syrjäkoski K, Heikkinen T, Eerola H, Aittomäki K, von Smitten K, et al. BARD1 variants Cys557Ser and Val507Met in breast cancer predisposition. *Eur J Hum Genet* 2006;14:167–72.
- Karppinen SM, Barkardottir RB, Backenhorst K, Sydenham T, Syrjäkoski K, Schleutker J, et al. Nordic collaborative study of the BARD1 Cys557Ser allele in 3956 patients with cancer: enrichment in familial BRCA1/BRCA2 mutation-negative breast cancer but not in other malignancies. *J Med Genet* 2006;43:856–62.
- Karppinen SM, Heikkinen K, Rapakko K, Winqvist R. Mutation screening of the BARD1 gene: evidence for involvement of the Cys557Ser allele in hereditary susceptibility to breast cancer. *J Med Genet* 2004;41:e114.
- Jakubowska A, Cybulski C, Szymańska A, Huzarski T, Byrski T, Gronwald J, et al. BARD1 and breast cancer in Poland. *Breast Cancer Res Treat* 2008;107:119–22.
- Gorringe KL, Choong DY, Visvader JE, Lindeman GJ, Campbell IG. BARD1 variants are not associated with breast cancer risk in Australian familial breast cancer. *Breast Cancer Res Treat* 2008;111:505–9.
- Johnnatty SE, Beesley J, Chen X, Hopper JL, Southey MC, Giles GG, et al. The BARD1 Cys557Ser polymorphism and breast cancer risk: an Australian case-control and family analysis. *Breast Cancer Res Treat* 2009;115:145–50.
- Chenevix-Trench G, Milne RL, Antoniou AC, Couch FJ, Easton DF, Goldgar DE. An international initiative to identify genetic modifiers of cancer risk in BRCA1 and BRCA2 mutation carriers: the Consortium of Investigators of Modifiers of BRCA1 and BRCA2 (CIMBA). *Breast Cancer Res* 2007;9:104.
- Goldgar DE, Easton DF, Deffenbaugh AM, Monteiro AN, Tavtigian SV, Couch FJ. Integrated evaluation of DNA sequence variants of unknown clinical significance: application to BRCA1 and BRCA2. *Am J Hum Genet* 2004;75:535–44.
- Chenevix-Trench G, Healey S, Lakhani S, Waring P, Cummings M, Brinkworth R, et al. Genetic and histopathologic evaluation of BRCA1 and BRCA2 DNA sequence variants of unknown clinical significance. *Cancer Res* 2006;66:2019–27.
- Rebbeck TR, Mitra N, Domchek SM, Wan F, Chuai S, Friebel TM, et al. Modification of ovarian cancer risk by BRCA1/2-interacting genes in a multicenter cohort of BRCA1/2 mutation carriers. *Cancer Res* 2009;69:5801–10.
- Antoniou AC, Goldgar DE, Andrieu N, Chang-Claude J, Brohet R, Rookus MA, et al. A weighted cohort approach for analysing factors modifying disease risks in carriers of high-risk susceptibility genes. *Genet Epidemiol* 2005;29:1–11.
- Antoniou A, Pharoah PD, Narod S, Risch HA, Eyfjord JE, Hopper JL, et al. Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet* 2003;72:1117–30.
- Excoffier L, Slatkin M. Maximum-likelihood estimation of molecular haplotype frequencies in a diploid population. *Mol Biol Evol* 1995;12:921–7.
- Fallin D, Cohen A, Essioux L, Chumakov I, Blumenfeld M, Cohen D, et al. Genetic analysis of case/control data using estimated haplotype frequencies: application to APOE locus variation and Alzheimer's disease. *Genome Res* 2001;11:143–51.
- Sinnwell JP, Schaid DJ. In: Clinix M, editor. *haplo.stats: Statistical Analysis of Haplotypes with Traits and Covariates when Linkage Phase is Ambiguous*. R Package Version 1.2.2. Rochester, MN: Mayo Clinic; 2005.

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