Alcoholism and alcohol drinking habits predicted from alcohol dehydrogenase genes

Janne Schurmann Tolstrup^{1,2} Børge Grønne Nordestgaard^{2,3}, Søren Rasmussen¹, Anne Tybjærg-Hansen⁴ and Morten Grønbæk^{1,3}

¹Center for Alcohol Research, National Institute of Public Health, Copenhagen, Denmark; ²Department of Clinical Biochemistry, Herlev University Hospital, Herlev, Denmark; ³The Copenhagen City Heart Study, Bispebjerg University Hospital, Copenhagen, Denmark and ⁴Department of Clinical Biochemistry, Copenhagen University Hospital, Copenhagen, Denmark

Correspondence:

Dr JS Tolstrup, Center for Alcohol Research, National Institute for Public Health, Oster Farimagsgade 5a, Copenhagen, 1399 Denmark. E-mail: jst@niph.dk

> published online 9 October 2007 dehydrogenase; acetaldehyde Introduction

> > Alcoholism and alcohol drinking in general represent huge public health problems in most countries worldwide, preventing many individuals from successfully holding a job or looking after a family. In addition, excessive alcohol use leads to diseases such as liver cirrhosis, chronic pancreatitis, upper gastrointestinal cancers, cardiomyopathy, polyneuropathy and dementia. It has been shown in twin studies that heritability explains approximately 50% of alcoholism and problem drinking in the white population.^{1,2}

> > The region surrounding the alcohol dehydrogenase (ADH) gene cluster is known to be associated with alcoholism from whole-genome scans.³ Well-known

Received 18 March 2007; revised 4 June 2007; accepted 13 June 2007; published online 9 October 2007

Alcohol drinking habits and alcoholism are partly genetically determined. Alcohol is degraded primarily by alcohol dehydrogenase (ADH) wherein genetic variation that affects the rate of alcohol degradation is found in ADH1B and ADH1C. It is biologically plausible that these variations may be associated with alcohol drinking habits and alcoholism. By genotyping 9080 white men and women from the general population, we found that men and women with ADH1B slow vs fast alcohol degradation drank more alcohol and had a higher risk of everyday drinking, heavy drinking, excessive drinking and of alcoholism. For example, the weekly alcohol intake was 9.8 drinks (95% confidence interval (CI): 9.1–11) among men with the ADH1B \cdot 1/1 genotype compared to 7.5 drinks (95% CI: 6.4-8.7) among men with the ADH1B · 1/2 genotype, and the odds ratio (OR) for heavy drinking was 3.1 (95% CI: 1.7-5.7) among men with the $ADH1B \cdot 1/1$ genotype compared to men with the ADH1B · 1/2 genotype. Furthermore, individuals with ADH1C slow vs fast alcohol degradation had a higher risk of heavy and excessive drinking. For example, the OR for heavy drinking was 1.4 (95% CI: 1.1–1.8) among men with the *ADH1C* · 1/2 genotype and 1.4 (95% CI: 1.0–1.9) among men with the $ADH1B \cdot 2/2$ genotype, compared with men with the $ADH1C \cdot 1/1$ genotype. Results for ADH1B and ADH1C genotypes among men and women were similar. Finally, because slow ADH1B alcohol degradation is found in more than 90% of the white population compared to less than 10% of East Asians, the population attributable risk of heavy drinking and alcoholism by ADH1B · 1/1 genotype was 67 and 62% among the white population compared with 9 and 24% among the East Asian population. The Pharmacogenomics Journal (2008) 8, 220–227; doi:10.1038/sj.tpj.6500471;

Keywords: genetic association study; alcoholism; population-based study; alcohol; alcohol

1000 221

functional polymorphisms of *ADH1B* and *ADH1C* may explain this finding because the *ADH1B* · 2 vs the *ADH1B* · 1 allele confer a 38-fold increase in *in vitro* alcohol degradation rate (that is, the conversion of ethanol to acetaldehyde) and the *ADH1C* · 1 vs the *ADH1C* · 2 allele confer a 2.5-fold increase in *in vitro* alcohol degradation rate.⁴

During alcohol degradation, acetaldehyde is only found in low concentrations. If concentrations become high, for example, during treatment with disulfiram (used in some countries to prevent alcohol intake among alcoholics) or in individuals with a defective acetaldehyde dehydrogenase (found among Asians), individuals experience severe nausea and flushing and automatically abstain from drinking alcohol. It is possible that similar but less pronounced responses to alcohol are more likely to be produced among individuals carrying the fast alcohol degradation $ADH1B \cdot 2$ and $ADH1C \cdot 1$ alleles compared to individuals carrying the slow alcohol degradation ADH1B · 1 and ADH1C · 2 alleles. If so, individuals with slow alcohol degradation may be able to drink larger quantities of alcohol without experiencing discomfort due to elevated acetaldehyde levels, and consequently are more likely to use alcohol excessively and to develop alcoholism. This issue has been addressed in different populations in case-control settings where allele frequencies of ADH1B and ADH1C are compared between alcoholics and nonalcoholics.⁵⁻²⁸ To our knowledge, this has not been studied in a prospective setting in the general white population and it is unknown if the ADH1B and ADH1C polymorphisms are associated with alcohol drinking habits, such as amount of usual intake.

In the present study, we have genotyped a sample of 9080 men and women from the general white population to test the hypotheses that slow alcohol degradation $ADH1B \cdot 1$ and $ADH1C \cdot 2$ alleles are associated with alcohol drinking habits and with increased risk of alcoholism.

Results

The frequencies of the *ADH1B* and *ADH1C* alleles coding for slow alcohol degradation was 0.98 (*ADH1B* · 1) and 0.42 (*ADH1C* · 2) (Table 1). Genotypes were in Hardy–Weinberg equilibrium (P = 0.8 for *ADH1B* genotypes and P = 0.7 for *ADH1C* genotypes by χ^2 -test). *ADH1B* · 2 were associated with *ADH1C* · 1 (linkage disequilibrium coefficients D' = 0.90 and $r^2 = 0.012$).

Table 1	Distribution	of ADH1B and	ADH1C genotypes
---------	--------------	--------------	-----------------

For *ADH1B*, we found that men and women who were homozygous for the slow alcohol degrading *ADH1B* · 1 allele had a higher alcohol intake than men and women who were fast alcohol degrading *ADH1B* · 2 heterozygotes or homozygotes. For example, men with the *ADH1B* · 1/1 genotype drank on average 9.8 drinks per week (95% confidence interval (CI): 9.1–11) and men with the *ADH1B* · 1/2 genotype drank on average 7.5 drinks per week (95% CI: 6.4–8.7) (Table 2). Furthermore, odds for any, daily, heavy and excessive alcohol drinking were 2–4 times higher among men and women who were *ADH1B* · 1 homozygotes than among men and women who were *ADH1B* · 2 heteroor homozygotes.

Using the brief Michigan Alcoholism Screening Test (brief MAST) we found that men with the slow alcohol degradation $ADH1B \cdot 1/1$ genotype had a two- to fourfold risk of alcoholism compared to men with the fast alcohol degradation $ADH1B \cdot 2/1$ or $ADH1B \cdot 2/2$ genotypes (Table 2). The hazard ratio of hospitalization for alcoholism in men and women with the slow alcohol degradation $ADH1B \cdot 1/1$ genotype was 3.9 (95% CI: 1.0–16) and 2.7 (95% CI: 0.4–20).

For *ADH1C*, odds for heavy and excessive alcohol drinking were 40–70% higher among men who were hetero- or homozygous for the slow alcohol degrading $ADH1C \cdot 2$ allele than among men who were homozygous for the fast alcohol degrading $ADH1C \cdot 1$ allele (Table 3). Similar results were found in women; however, effect sizes were slightly smaller and only statistically significant for heavy drinking. ADH1C genotype was not associated with alcoholism (Table 3).

Analyses on daily, heavy and excessive drinking and on alcoholism (MAST score and hospitalizations) were repeated excluding consistent nondrinkers, that is, participants who reported no alcohol intake at every examination in which they participated (7.5% of the total study population). This restriction did not affect any of our results (data not shown).

Because of linkage disequilibrium between the $ADH1B \cdot 2$ and $ADH1C \cdot 1$ alleles, and the relatively large effect on enzyme activity of the ADH1B polymorphism, our results for ADH1C could be influenced by ADH1B genotype. Therefore, ADH1C analyses were repeated solely on individuals who were $ADH1B \cdot 1$ homozygotes (95% of the study cohort): we found similar results, indicating that the effect of ADH1Cgenotype was independent of ADH1B genotype (data not shown).

We also performed analyses on genotype combinations, ranking genotypes in order of expected total enzyme

ADH1B	ADH1C						
	Men (n = 4039)			<i>Women</i> (n = 5041)			
	1/1 (fast)	1/2 (intermediate)	2/2 (slow)	1/1 (fast)	1/2 (intermediate)	2/2 (slow)	
1/1 (slow)	1272	1882	697	1570	2347	907	
1/2 (intermediate)	110	70	5	130	81	3	
2/2 (fast)	2	1	0	3	0	0	

npg	
222	

Women (n = 5041)ADH1B alcohol degradation *Men* (n = 4039) 1/2+2/2 (n = 188) (fast) 1/1 (n = 3851) (slow) 1/2+2/2 (n = 217) (fast) 1/1 (n = 4824) (slow) Alcohol drinking habits Weekly alcohol intake $(n = 3784 \text{ M}, 4052 \text{ W})^{a}$ 7.5 (6.4-8.7) 9.8 (9.1-11) 3.0 (2.6-3.5) 4.0 (3.7-4.3) Any alcohol intake $(n = 3784 \text{ M}, 4052 \text{ W})^{\text{b}}$ 1.0 (reference) 2.1 (1.0-4.5) 1.0 (reference) 3.1 (1.8-5.3) Daily drinking $(n = 2015 \text{ M}, 1259 \text{ W})^{\text{b}}$ 1.0 (reference) 2.5 (1.5-4.1) 1.0 (reference) 1.9(1.1 - 3.5)Heavy drinking (n = 1262 M, 814 W)^b 1.0 (reference) 3.1 (1.7-5.7) 1.0 (reference) 3.0 (1.4-6.4) Excessive drinking $(n = 507 \text{ M}, 353 \text{ W})^{\text{b}}$ 1.0 (reference) 2.7 (1.1-6.5) 1.0 (reference) 4.2 (1.3–13) Alcoholism bMAST score ≥ 5 (n = 351 M, 105 W)^b 1.0 (reference) 2.9 (1.3-6.6) 1.0 (reference) 1.7(0.5-5.2)bMAST score ≥ 10 (n = 217 M, 55 W)^b 1.0 (reference) 2.6 (1.0-7.1) 1.0 (reference) 2.8 (0.4-20) Hospitalization $(n = 167 \text{ M}, 74 \text{ W})^{c}$ 1.0 (reference) 3.9 (1.0-16) 1.0 (reference) 2.7 (0.4-20)

Table 2 Associations between ADH1B genotype and alcohol drinking habits and alcoholism

Abbreviations: bMAST, the brief Michigan Alcoholism Screening Test; M, men; W, women.

Heavy drinking was defined as drinking more than 21 drinks per week for men and 14 drinks per week for women, and excessive drinking as drinking more than 35 drinks per week for men and 21 drinks per week for women. *n* indicates the number of men and women who are cases in respective analyses. Adjustment was made for *ADH1C* genotype, age, years of school education and examination year.

^aShown numbers are mean number of drinks per week (95% CI), which is estimated among those who report any alcohol intake.

^bShown numbers are OR (95% CI).

^cShown numbers are hazard ratios (95% Cl).

ADH1C alcohol degradation		<i>Men (</i> n = 4039)			<i>Women (</i> n = 5041)	
	1/1 (n = 1384) (fast)	1/2 (n = 1953) (intermediate)	2/2 (n = 702) (slow)	1/1 (n = 1703) (fast)	1/2 (n = 2428) (intermediate)	2/2 (n = 910) (slow)
Alcohol drinking habits						
Weekly alcohol intake $(n = 3784 \text{ M}, 4052 \text{ W})^{a}$	9.8 (9.1–11)	10.4 (9.4–12)	10.5 (9.4–12)	4.0 (3.7–4.3)	4.1 (3.7–4.5)	4.1 (3.7–4.6)
Any alcohol intake $(n = 3784 \text{ M}, 4032 \text{ W})^{\text{b}}$	1.0 (reference)	1.3 (0.9–1.8)	0.7 (0.5–1.2)	1.0 (reference)	1.1 (0.9–1.4)	1.0 (0.7–1.4)
Daily drinking $(n = 2015 \text{ M}, 1259 \text{ W})^{\text{b}}$	1.0 (reference)	1.1 (0.9–1.4)	1.1 (0.8–1.5)	1.0 (reference)	1.2 (0.9–1.5)	1.2 (0.9–1.7)
Heavy drinking $(n=1262 \text{ M}, 814 \text{ W})^{\text{b}}$	1.0 (reference)	1.4 (1.1–1.8)	1.4 (1.0–1.9)	1.0 (reference)	1.3 (1.0–1.7)	1.2 (0.8–1.7)
Excessive drinking $(n = 507 \text{ M}, 353 \text{ W})^{\text{b}}$	1.0 (reference)	1.6 (1.1–2.3)	1.7 (1.1–2.6)	1.0 (reference)	1.4 (0.9–2.1)	1.0 (0.6–1.6)
Alcoholism						
bMAST score \geq 5 (n=1076 M, 597 W) ^b	1.0 (reference)	1.0 (0.8–1.3)	1.0 (0.7–1.4)	1.0 (reference)	0.8 (0.5–1.2)	0.8 (0.5–1.5)
bMAST score ≥ 10 (n = 290 M, 65 W) ^b	1.0 (reference)	1.0 (0.7–1.3)	1.0 (0.7–1.6)	1.0 (reference)	0.6 (0.3–1.0)	0.7 (0.3–1.5)
Hospitalization (n=209 M, 90 W) ^c	1.0 (reference)	1.2 (0.9–1.7)	0.9 (0.6–1.5)	1.0 (reference)	1.4 (0.8–2.6)	2.2 (1.2–4.2)

Table 3 Associations between ADH1C genotype and alcohol drinking habits and alcoholism

Abbreviations: bMAST, the brief Michigan Alcoholism Screening Test; M, men; W, women.

Heavy drinking was defined as drinking more than 21 drinks per week for men and 14 drinks per week for women, and excessive drinking as drinking more than 35 drinks per week for men and 21 drinks per week for women. *n* indicates the number of men and women who are cases in the respective analyses. Adjustment was made for *ADH1B* genotype, age, years of school education and examination year.

^aShown numbers are mean number of drinks per week (95% CI), which is estimated among those who report any alcohol intake.

^bShown numbers are OR (95% CI).

^cShown numbers are hazard ratios (95% CI).

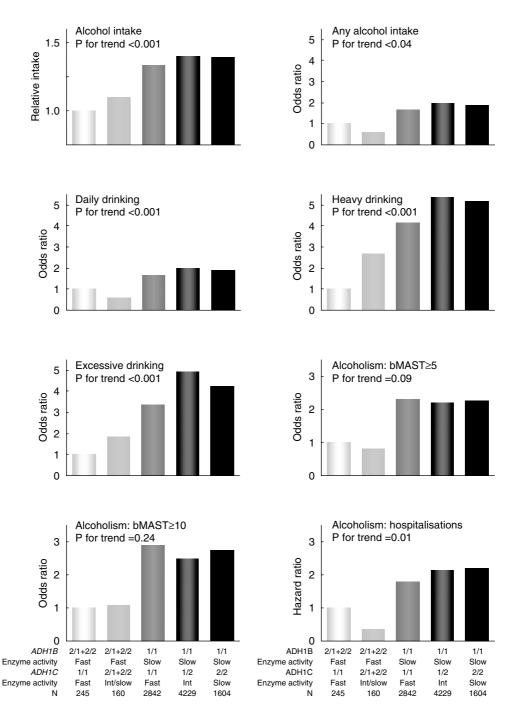


Figure 1 Association between the combined *ADH1B* and *ADH1C* genotypes and alcohol drinking habits and alcoholism. The combined *ADH1B* and *ADH1C* genotypes are ranked according to expected enzyme activity (($ADH1B \cdot 2/1 + ADH1B \cdot 2/2$, $ADH1C \cdot 1/1$) > ($ADH1B \cdot 2/1 + ADH1B \cdot 2/2$, $ADH1C \cdot 2/2$) > ($ADH1B \cdot 1/1$, $ADH1C \cdot 1/1$) > ($ADH1B \cdot 1/1$, $ADH1C \cdot 1/2$) > ($ADH1B \cdot 1/1$, $ADH1C \cdot 2/2$) ($ADH1B \cdot 1/1$, $ADH1C \cdot 1/2$) > ($ADH1B \cdot 1/1$, $ADH1C \cdot 1/2$) > ($ADH1B \cdot 1/1$, $ADH1C \cdot 2/2$)). $ADH1C \cdot 1/2$ and $ADH1C \cdot 2/2$ were combined because of few subjects in these categories among the $ADH1B \cdot 2$ hetero- and homozygotes. Analyses are for men and women combined and *P*-values are for linear trend tests.

activity, and tested for linear trend in each of the variables for alcohol drinking habits and alcoholism (Figure 1). For most end points, there was a statistically significant trend test in the expected direction: individuals with slow vs fast alcohol degradation drank more alcohol and more often, and had a higher risk of alcoholism. *ADH1B* genotypes are very differently distributed among whites and East Asian populations (Figure 2). Among the white population, more than 90% carry the *ADH1B* \cdot *1/1* genotype, coding for slow alcohol degradation, whereas among the East Asian population less than 10% carry this genotype. Hence, the population attributable risk of heavy

npg 223

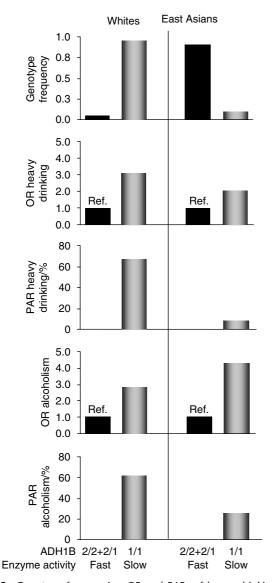


Figure 2 Genotype frequencies, OR and PARs of heavy drinking and alcoholism according to *ADH1B* genotype among white and East Asian populations. Slow alcohol degradation $ADH1B \cdot 1/1$ genotype is predominant among Caucasians, but rare among East Asian population. Risks of heavy drinking and alcoholism according to $ADH1B \cdot 1/1$ among white and East Asian populations are comparable, but because of the different genotype distributions in the two populations, PARs are much higher among white population. For East Asian population, genotype frequencies, OR and PARs are calculated from previous studies.^{29,30} Ref, reference group; OR, odds ratio; PAR, population attributable risk.

drinking and alcoholism according to the $ADH1B \cdot 1/1$ genotype is 67 and 62% among white population compared with 9 and 24% among the East Asian population.

Discussion

224

Our results suggest that *ADH1B* and *ADH1C* genotypes are associated with alcoholism and alcohol drinking habits.

Men and women with ADH1B slow vs fast alcohol degradation drank more alcohol, were more often daily, heavy and excessive drinkers and had higher risks of alcoholism. Men and women with ADH1C intermediate and slow vs fast alcohol degradation were more often heavy and excessive drinkers. As expected, effect sizes were smaller for ADH1C than for ADH1B, but given the high frequency of the $ADH1C \cdot 2$ allele, it is nevertheless a very interesting finding. Moreover, the impact of the $ADH1C \cdot 2$ allele on cumulative lifetime alcohol intake may be significant. Our results also suggest that ADH1B and ADH1C genotypes may partly explain why white people generally drink more alcohol than East Asian. The population risk of heavy drinking and alcoholism attributable to the ADH1B · 1/1 genotype was 67 and 62% among white population and only 9 and 24% among the East Asian population.

Among men, we found relative estimates for alcoholism from two to three among $ADH1B \cdot 1$ homozygotes, which is comparable to results from a recent meta-analysis consisting predominantly of East Asian studies.²⁹ Also, the odds ratio (OR) for heavy drinking for men was 3.1 which agrees with what is previously found among Asian men.³⁰ Separate estimates for women were not available for any of the end points in previous studies. Our results were remarkably similar for men and women indicating that the investigated associations are not sex-specific.

The various end points were probably subject to some misclassification, that is, sensitivity and/or specificity less than 100%. Measures of heavy and excessive drinking will be misclassified if amount of alcohol intake is under- or overreported, which it in many instances probably is. The MAST score is not a perfect screenings tool for alcoholism either. However, these errors occur most likely independently of genotype, and because end points are binary, will lead to bias toward the null.³¹ Hence, we do not consider misclassification of end points to have caused our results. For alcoholism defined by hospital registry information, sensitivity is likely considerably less than 100% because many alcoholics are untreated or treated at private clinics that are not included in the national registers. However, specificity could be close to perfect; few nonalcoholics are presumably diagnosed as alcoholics. In this scenario, nondifferential misclassification is not affecting the hazard ratio.32

Never-drinkers have not been exposed to alcohol and hence, their drinking status cannot have been affected by *ADH1B* and *ADH1C* genotypes. It was not possible to separate never-drinkers from nondrinkers in this study, so performing analyses without never-drinkers was not an option. Potentially, this could have caused bias, but since excluding consistent nondrinkers from analyses had virtually no impact on our results, we do not consider this a major limitation.

We modeled the amount of alcohol intake in five different ways and alcoholism in three different ways. Hence, several statistical tests have been performed which in some instances call for caution. However, outcomes in this study were not independent but merely represent similar outcomes modeled differently and we chose not to adjust for multiple comparisons.

A likely explanation for our findings is that differences in enzyme activity from the ADH1B and ADH1C polymorphisms result in intraindividual differences in alcohol degradation rate and that, for a given level of alcohol intake, individuals with fast alcohol degradation have higher levels of acetaldehyde and thus more unpleasant symptoms compared with individuals with slow alcohol degradation. However, an effect of the polymorphisms on alcohol degradation rate in vivo has been difficult to demonstrate,33 which may have been due to insufficiently sensitive laboratory methods. In a more recent study which applied a more refined method for measuring the rate of alcohol degradation, results showed a significant difference in degradation rate according to the ADH1B genotype.³⁴ In further support of an *in vivo* effect of the ADH1B genotype is that individuals with the most active enzymes are consistently reported to experience more unpleasant symptoms such as flushing when drinking alcohol compared to individuals with the less active enzymatic forms.35-38

Our study had several strengths. First of all, sample size was large and provided adequate power to study associations between the relatively rare $ADH1B \cdot 1/2$ genotype and several end points, and to detect the small effect sizes associated with the ADH1C genotypes. Furthermore, participants were men and women from the general population, all of Danish descent. Hence, population stratification is unlikely to have affected our results. Alcohol drinking habits were described in several dimensions and information on alcoholism was obtained from two independent sources (questionnaire and hospital registry information). All end points were assessed independently from genotyping and participants were unaware of the purpose of this study when enrolled.

In conclusion, our data suggest that alcoholism and alcohol drinking habits are partly predictable from *ADH1B* and *ADH1C* genotypes. Results for men and women were comparable and, as expected, effects of *ADH1B* were larger than effects of *ADH1C*.

Methods

Study population

Our data originate from The Copenhagen City Heart Study, which is a series of studies conducted in the Danish general population. Examinations consisted of interview and physical examination, and more especially, blood was given for DNA purification at the examination that was performed during 1991–1994. All participants gave written consent and the ethics committee for Copenhagen and Frederiksberg approved the study (no. 100.2039/91). Enrolment and examination procedures have been described in more detail elsewhere.^{39,40} Of the 17 180 individuals who were invited to the 1991–1994 examination, 10 135 participated, 9259 gave blood and 9222 were successfully genotyped. Eligibility criterion for participation was Danish citizenship and therefore, the Copenhagen City Heart Study does not reflect

the ethnic admixture of Copenhagen (the proportion of inhabitants with foreign citizenship was 8% in 1994). However, even a few participants of foreign ethnicity could potentially confound our results since the fast alcohol degradation $ADH1B \cdot 2$ allele is rare among white population and quite frequent in other populations. Information on ethnicity was not assessed at the examinations, and hence information on birthplace was obtained from the Civil Registration System. Participants born in Asia, Africa, the Middle East, South America or Greenland were excluded from further study (n=211). In all, 9080 individuals were eligible for analyses, some of whom also participated in the examinations during 1981–1983 (n=6615) and during 2001–2003 (n=4684).

Genotyping procedures

The $ADH1B \cdot 2$ allele (rs1229984, Arg47His in exon 3) and $ADH1C \cdot 2$ allele (rs698, Ile3490Val in exon 8) were identified by means of duplex polymerase chain reaction followed by Nanogen microelectronic chip technology (Nanogen NMW 1000 Nanochip Molecular Biology Workstation)⁴¹ using standard conditions (details available from authors). In a validation study, the accuracy of the Nanogen method was found to be comparable to restriction fragment length polymorphism.⁴²

End points

Questions on drinking habits were included in the questionnaire at the examinations during 1981–1983, 1991– 1994 and 2001–2003. Amount of alcohol intake was reported as usual intake of weekly beers, wine and spirits. Assuming one drink to be equal to 12 g of pure alcohol, a measure of total weekly intake was calculated. We defined heavy drinking as drinking more than 21 drinks per week for men and 14 drinks per week for women, and excessive drinking as drinking more than 35 drinks per week for men and 21 drinks per week for women.⁴³ Participants were defined as daily drinkers if they reported to drink alcohol every day.

We defined alcoholism from questionnaire as well as from hospital discharge information. The former definition was taken from the 1991–1994 questionnaire, which included a screening test for alcoholism (10 question version of the brief MAST⁴⁴). The test included questions such as 'Do you feel you are a normal drinker?' and 'Have you ever gone to anyone for help about your drinking?' Test scores of \geq 5 and of \geq 10 were used as dichotomized end points. Information on hospitalizations for alcoholism was obtained from the Danish Hospital Discharge Register where all hospitalizations in Denmark, classified according to the World Health Organization's International Classification of Diseases (ICD) are registered.⁴⁵ The following diagnoses indicative of hospitalization due to alcoholism were obtained: ICD-8 codes 303.09–303.99 and ICD-10 codes F10.1–F10.4.

Statistical analyses

All statistical models included *ADH1B* and *ADH1C* genotypes, age and years of school education using the SAS/Stat 226

software (version 8.02). $ADH1B \cdot 2$ heterozygotes were combined with $ADH1B \cdot 2$ homozygotes (n = 6). Estimated haplotype frequencies were calculated by Hplus.^{46,47} Linkage disequilibrium was expressed as r^2 and D'.^{48,49}

To study the association between *ADH1B* and *ADH1C* genotypes and amount of alcohol intake, the correlated mixed distribution model was applied (Mixcorr macro⁵⁰). This model handles data with clumping at zero and a lognormal distribution for nonzero values, and contains components to model the probability of a nonzero value and the mean of nonzero value, allowing for repeated measurements using random effects.⁵⁰ This means that if a variable affects the mean amount by affecting both the probability of occurrence of a nonzero value and also the mean of a nonzero value, these effects can be separated and quantified. Hence, two estimates are produced from this model: the OR for having a nonzero alcohol intake (that is, for not being a non-drinker) and the mean amount of alcohol intake among those with a nonzero intake.

To study the association between *ADH1B* and *ADH1C* genotypes and daily, heavy and excessive drinking, we applied logistic regression allowing for repeated measurements using random effects (proc nlmixed). We applied unconditional logistic regression to study associations between *ADH1B* and *ADH1C* genotypes and dichotomized brief MAST score (proc genmod).

Risk estimates for alcoholism defined from hospitalizations were computed by means of Cox proportional hazard regression (proc phreg). Age was used as the time axis and analyses were corrected for delayed entry. Vital status of the participants was obtained from the National Central Person Register. The observation time for each participant was the period from participation in the Copenhagen City Heart Study, until date of alcoholism, death, emigration outside Denmark or January 1, 2004, whichever came first. We had 100% follow-up.

Population attributable risk was calculated as (proportion of exposed in the population (OR-1))/(proportion of exposed in the population (OR-1)+1).⁵¹

Abbreviations				
ADH bMAST OR PAR Ref	alcohol dehydrogenase brief Michigan Alcoholism Screening Test odds ratio population attributable risk Reference group			

Acknowledgments

This work was supported by grants from the Danish Graduate School of Public Health, the Health Insurance Foundation, the Ministry of the Interior and Health and the Danish National Board of Health and The Danish Heart Foundation, The Danish Medical Research Council, The Copenhagen County Research Foundation and Chief Physician Johan Boserup and Lise Boserups Foundation. We thank the participants of the Copenhagen City Heart Studies. None declared.

References

- 1 Han C, McGue MK, Iacono WG. Lifetime tobacco, alcohol and other substance use in adolescent Minnesota twins: univariate and multi-variate behavioral genetic analyses. *Addiction* 1999; **94**: 981–993.
- 2 Kendler KS, Neale MC, Heath AC, Kessler RC, Eaves LJ. A twin-family study of alcoholism in women. Am J Psychiatry 1994; 151: 707–715.
- 3 Saccone NL, Kwon JM, Corbett J, Goate A, Rochberg N, Edenberg HJ et al. A genome screen of maximum number of drinks as an alcoholism phenotype. Am J Med Genet 2000; 96: 632–637.
- 4 Bosron WF, Li TK. Genetic polymorphism of human liver alcohol and aldehyde dehydrogenases, and their relationship to alcohol metabolism and alcoholism. *Hepatology* 1986; **6**: 502–510.
- 5 Vidal F, Lorenzo A, Auguet T, Olona M, Broch M, Gutierrez C et al. Genetic polymorphisms of ADH2, ADH3, CYP4502E1 Dra-I and Pst-I, and ALDH2 in Spanish men: lack of association with alcoholism and alcoholic liver disease. J Hepatol 2004; 41: 744–750.
- 6 Konishi T, Luo HR, Calvillo M, Mayo MS, Lin KM, Wan YJ. ADH1B*1, ADH1C*2, DRD2 (-141C Ins), and 5-HTTLPR are associated with alcoholism in Mexican American men living in Los Angeles. *Alcohol Clin Exp Res* 2004; **28**: 1145–1152.
- 7 Wall TL, Carr LG, Ehlers CL. Protective association of genetic variation in alcohol dehydrogenase with alcohol dependence in native American Mission Indians. *Am J Psychiatry* 2003; **160**: 41–46.
- 8 Osaka R, Nanakorn S, Sakata R, Nishiyori A, Shibata A, Nakamura J *et al.* Alcohol dehydrogenase-2 and aldehyde dehydrogenase-2 genotypes and male alcohol use disorders in Khon Kaen, north-east Thailand. *Psychiatry Clin Neurosci* 2003; **57**: 37–45.
- 9 Chao YC, Wang SJ, Chu HC, Chang WK, Hsieh TY. Investigation of alcohol metabolizing enzyme genes in Chinese alcoholics with avascular necrosis of hip joint, pancreatitis and cirrhosis of the liver. *Alcohol Alcohol* 2003; **38**: 431–436.
- 10 Frenzer A, Butler WJ, Norton ID, Wilson JS, Apte MV, Pirola RC *et al.* Polymorphism in alcohol-metabolizing enzymes, glutathione S-transferases and apolipoprotein E and susceptibility to alcohol-induced cirrhosis and chronic pancreatitis. *J Gastroenterol Hepatol* 2002; **17**: 177–182.
- 11 Chambers GK, Marshall SJ, Robinson GM, Maguire S, Newton-Howes J, Chong NL. The genetics of alcoholism in Polynesians: alcohol and aldehyde dehydrogenase genotypes in young men. *Alcohol Clin Exp Res* 2002; 26: 949–955.
- 12 Ogurtsov PP, Garmash IV, Miandina GI, Guschin AE, Itkes AV, Moiseev VS. Alcohol dehydrogenase ADH2-1 and ADH2-2 allelic isoforms in the Russian population correlate with type of alcoholic disease. *Addict Biol* 2001; 6: 377–383.
- 13 Lee HC, Lee HS, Jung SH, Yi SY, Jung HK, Yoon JH *et al.* Association between polymorphisms of ethanol-metabolizing enzymes and susceptibility to alcoholic cirrhosis in a Korean male population. *J Korean Med Sci* 2001; **16**: 745–750.
- 14 Borras E, Coutelle C, Rosell A, Fernandez-Muixi F, Broch M, Crosas B et al. Genetic polymorphism of alcohol dehydrogenase in Europeans: the ADH2*2 allele decreases the risk for alcoholism and is associated with ADH3*1. Hepatology 2000; 31: 984–989.
- 15 Amadéo S, Noble EP, Fourcade-Amadéo ML, Tetaria C, Brugiroux MF, Nicolas L *et al.* Association of D2 dopamine receptor and alcohol dehydrogenase 2 genes with polynesian alcoholics. *Eur Psychiatry* 2000; 15: 97–102.
- 16 Chao YC, Wang LS, Hsieh TY, Chu CW, Chang FY, Chu HC. Chinese alcoholic patients with esophageal cancer are genetically different from alcoholics with acute pancreatitis and liver cirrhosis. *Am J Gastroenterol* 2000; **95**: 2958–2964.
- 17 Rodrigo L, Alvarez V, Rodriguez M, Perez R, Alvarez R, Coto E. N-acetyltransferase-2, glutathione S-transferase M1, alcohol dehydrogenase, and cytochrome P450IIE1 genotypes in alcoholic liver cirrhosis: a case–control study. Scand J Gastroenterol 1999; 34: 303–307.

(1)pg

- 18 Osier M, Pakstis AJ, Kidd JR, Lee JF, Yin SJ, Ko HC et al. Linkage disequilibrium at the ADH2 and ADH3 loci and risk of alcoholism. Am J Hum Genet 1999; 64: 1147–1157.
- 19 Chen CC, Lu RB, Chen YC, Wang MF, Chang YC, Li TK *et al.* Interaction between the functional polymorphisms of the alcohol-metabolism genes in protection against alcoholism. *Am J Hum Genet* 1999; **65**: 795–807.
- 20 Whitfield JB, Nightingale BN, Bucholz KK, Madden PA, Heath AC, Martin NG. ADH genotypes and alcohol use and dependence in Europeans. Alcohol Clin Exp Res 1998; 22: 1463–1469.
- 21 Shen YC, Fan JH, Edenberg HJ, Li TK, Cui YH, Wang YF *et al.* Polymorphism of ADH and ALDH genes among four ethnic groups in China and effects upon the risk for alcoholism. *Alcohol Clin Exp Res* 1997; 21: 1272–1277.
- 22 Espinos C, Sanchez F, Ramirez C, Juan F, Najera C. Polymorphism of alcohol dehydrogenase genes in alcoholic and nonalcoholic individuals from Valencia (Spain). *Hereditas* 1997; **126**: 247–253.
- 23 Chen WJ, Loh EW, Hsu YP, Cheng AT. Alcohol dehydrogenase and aldehyde dehydrogenase genotypes and alcoholism among Taiwanese aborigines. *Biol Psychiatry* 1997; **41**: 703–709.
- 24 Nakamura K, Iwahashi K, Matsuo Y, Miyatake R, Ichikawa Y, Suwaki H. Characteristics of Japanese alcoholics with the atypical aldehyde dehydrogenase 2*2. I. A comparison of the genotypes of ALDH2, ADH2, ADH3, and cytochrome P-4502E1 between alcoholics and nonalcoholics. *Alcohol Clin Exp Res* 1996; 20: 52–55.
- 25 Tanaka F, Shiratori Y, Yokosuka O, Imazeki F, Tsukada Y, Omata M. High incidence of ADH2*1/ALDH2*1 genes among Japanese alcohol dependents and patients with alcoholic liver disease. *Hepatology* 1996; 23: 234–239.
- 26 Higuchi S, Muramatsu T, Matsushita S, Murayama M, Hayashida M. Polymorphisms of ethanol-oxidizing enzymes in alcoholics with inactive ALDH2. *Hum Genet* 1996; **97**: 431–434.
- 27 Chen WJ, Loh EW, Hsu YP, Chen CC, Yu JM, Cheng AT. Alcoholmetabolising genes and alcoholism among Taiwanese Han men: independent effect of ADH2, ADH3 and ALDH2. *Br J Psychiatry* 1996; 168: 762–767.
- 28 Maezawa Y, Yamauchi M, Toda G, Suzuki H, Sakurai S. Alcoholmetabolizing enzyme polymorphisms and alcoholism in Japan. Alcohol Clin Exp Res 1995; 19: 951–954.
- 29 Zintzaras E, Stefanidis I, Santos M, Vidal F. Do alcohol-metabolizing enzyme gene polymorphisms increase the risk of alcoholism and alcoholic liver disease? *Hepatology* 2006; **43**: 352–361.
- 30 Sun F, Tsuritani I, Yamada Y. Contribution of genetic polymorphisms in ethanol-metabolizing enzymes to problem drinking behavior in middleaged Japanese men. *Behav Genet* 2002; 32: 229–236.
- 31 Rothman K. Biases in Study Design. Epidemiology—An Introduction. Oxford University Press: New York, 2002: 94–112.
- 32 Rothman K, Greenland S. *Precision and Validity in Epidemiological Studies Modern Epidemiology*.2nd edn, Lippincott, Williams and Wilkins, Philadelphia, USA, 1998, pp 115–134.
- 33 Mizoi Y, Yamamoto K, Ueno Y, Fukunaga T, Harada S. Involvement of genetic polymorphism of alcohol and aldehyde dehydrogenases in individual variation of alcohol metabolism. *Alcohol Alcohol* 1994; 29: 707–710.

- 34 Neumark YD, Friedlander Y, Durst R, Leitersdorf E, Jaffe D, Ramchandani VA *et al.* Alcohol dehydrogenase polymorphisms influence alcoholelimination rates in a male Jewish population. *Alcohol Clin Exp Res* 2004; 28: 10–14.
- 35 Yokoyama T, Yokoyama A, Kato H, Tsujinaka T, Muto M, Omori T *et al.* Alcohol flushing, alcohol and aldehyde dehydrogenase genotypes, and risk for esophageal squamous cell carcinoma in Japanese men. *Cancer Epidemiol Biomarkers Prev* 2003; **12**: 1227–1233.
- 36 Carr LG, Foroud T, Stewart T, Castelluccio P, Edenberg HJ, Li TK. Influence of ADH1B polymorphism on alcohol use and its subjective effects in a Jewish population. *Am J Med Genet* 2002; **112**: 138–143.
- 37 Takeshita T, Yang X, Morimoto K. Association of the ADH2 genotypes with skin responses after ethanol exposure in Japanese male university students. *Alcohol Clin Exp Res* 2001; **25**: 1264–1269.
- 38 Takeshita T, Mao XQ, Morimoto K. The contribution of polymorphism in the alcohol dehydrogenase beta subunit to alcohol sensitivity in a Japanese population. *Hum Genet* 1996; **97**: 409–413.
- 39 The Copenhagen City Heart Study. Osterbroundersogelsen. A book of tables with data from the first examination (1976–78) and a five year follow-up (1981–83). Scand J Soc Med (Suppl) 1989; 41: 1–160.
- 40 Schnohr P, Jensen G, Lange P, Scharling H, Appleyard M. The Copenhagen City Heart Study—Østerbroundersøgelsen. Tables with data from the third examination 1991–94. *Eur Heart J (Suppl)* 2001; **3**: H1–H83.
- 41 Sosnowski RG, Tu E, Butler WF, O'Connell JP, Heller MJ. Rapid determination of single base mismatch mutations in DNA hybrids by direct electric field control. *Proc Natl Acad Sci USA* 1997; **94**: 1119–1123.
- 42 Sethi AA, Tybjaerg-Hansen A, Andersen RV, Nordestgaard BG. Nanogen microelectronic chip for large-scale genotyping. *Clin Chem* 2004; **50**: 443–446.
- 43 Jensen MK, Sorensen TI, Andersen AT, Thorsen T, Tolstrup JS, Godtfredsen NS *et al.* A prospective study of the association between smoking and later alcohol drinking in the general population. *Addiction* 2003; **98**: 355–363.
- 44 Pokorny AD, Miller BA, Kaplan HB. The brief MAST: a shortened version of the Michigan Alcoholism Screening Test. *Am J Psychiatry* 1972; **129**: 342–345.
- 45 Jürgensen HJ, Frølund C, Gustafsen J, Mosbech H, Guldhammer B, Mosbech J. Registration of diagnoses in the Danish National Registry of patients. *Methods Inf Med* 1986; **25**: 158–164.
- 46 Hplus. Fred Hutchinson Cancer Research Center: Seattle, WA, 2003.
- 47 Li SS, Khalid N, Carlson C, Zhao LP. Estimating haplotype frequencies and standard errors for multiple single nucleotide polymorphisms. *Biostatistics* 2003; 4: 513–522.
- 48 Lewontin RC. The interaction of selection and linkage. I. General considerations: heterotic models. *Genetics* 1964; 49-67.
- 49 Pritchard JK, Przeworski M. Linkage disequilibrium in humans: models and data. *Am J Hum Genet* 2001; **69**: 1–14.
- 50 Tooze JA, Grunwald GK, Jones RH. Analysis of repeated measures data with clumping at zero. *Stat Methods Med Res* 2002; **11**: 341–355.
- 51 Walter SD. Calculation of attributable risks from epidemiological data. Int J Epidemiol 1978; 7: 175–182.