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## Protective effect of resveratrol against caspase 3 activation in primary mouse fibroblasts

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**Aim** To study the effect of resveratrol on survival and caspase 3 activation in non-transformed cells after serum deprivation.

**Methods** Apoptosis was induced by serum deprivation in primary mouse embryonic fibroblasts. Caspase 3 activation and lactate dehydrogenase release were assayed as cell viability measure by using their fluorogenic substrates. The involvement of PI3K, ERK, JNK, p38, and SIRT1 signaling pathways was also examined.

**Results** Serum deprivation of primary fibroblasts induced significant activation of caspase 3 within 3 hours and reduced cell viability after 24 hours. Resveratrol dose-dependently prevented caspase activation and improved cell viability with 50% inhibitory concentration ( $IC_{50}$ ) =  $66.3 \pm 13.81$   $\mu$ M. It also reduced the already up-regulated caspase 3 activity when it was added to the cell culture medium after 3 hour serum deprivation, suggesting its rescue effect. Among the major signaling pathways, p38 kinase was critical for the protective effect of resveratrol which was abolished completely in the presence of p38 inhibitor.

**Conclusion** Resveratrol showed protective effect against cell death in a rather high dose. Involvement of p38 kinase in this effect suggests the role of mild stress in its cytoprotective action. Furthermore due to its rescue effect, resveratrol may be used not only for prevention, but also treatment of age-related degenerative diseases, but in the higher dose than consumed in conventional diet.

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Age-related degenerative diseases pose enormous challenges both for individuals and society in terms of life quality and economic burden. Since age-related neurodegenerative and cardiovascular diseases develop mainly as a result of cell impairments, it is crucial to find agents that prevent and abolish cell damage and death. Resveratrol (3,5,4'-trihydroxy-trans-stilbene) is a widely investigated phytoalexin compound, which can be found in numerous plants, mainly in the skin and seeds of red grapes (1). It was reported to possess multiple pharmacological properties including antiaging (2), antioxidative, anti-inflammatory (3), anticarcinogenic (4), and neuro- and cardioprotective effects (5). However, in the literature its rather contradictory properties, ie, cytoprotective and proapoptotic, were reported (6). The cause of opposite effects may lie in different cell types, cell states, and the duration or dosage of treatment used in the various models (7). Characteristically, resveratrol has an opposite impact on apoptosis in non-transformed and transformed cells (8,9). The targets of resveratrol and the mechanisms governing its effects are currently unclear. It was reported to affect different metabolic and signaling pathways, exhibit pro- or antioxidative activities, and modify the functions of several transcription factors and cofactors (10).

Since resveratrol might differently affect apoptotic process of tumorigenic and non-transformed normal cells, the aim of this study was to investigate its effect on the death of non-transformed cells as a potential lead compound for research of cytoprotective medications. We used primary mouse embryonic fibroblasts as an easily available non-transformed cell culture model. In order to evaluate its cytoprotective effect, caspase 3 activation was examined following serum deprivation as a model of insufficient availability of trophic factors. The specific background mechanisms, involvement of the PI3K, ERK, JNK, p38, and SIRT1 signaling pathways were also determined.

## MATERIALS AND METHODS

### Reagents and animals

Resveratrol, the inhibitors of kinases (SB202190 for p38 MAPK, SP600125 for JNK, PD184352 for ERK, wortmannin for PI3K) and SIRT1 (EX-527), caspase 3 activity assay kit using fluorogenic caspase 3 substrate (Ac-DEVD-AMC), buffer components and N-acetylcysteine were purchased from Sigma-Aldrich (St. Louis, MO, USA). Non-selective caspase inhibitor (Ac-VAD-CMK) was obtained from AnaSpec (Fermont, CA, USA) and CytoxOne lactate dehydrogenase release kit from Promega (Fitchburg, WI, USA). Cell cul-

ture mediums and fetal bovine serum were supplied by GE Healthcare (Little Chalfont, UK) and Life Technologies (Carlsbad, CA, USA), respectively. Test compounds were dissolved in DMSO and used in cell culture medium to provide 0.5% final DMSO concentration. Control cells were treated with the same concentration of DMSO.

Pregnant NMRI mice were supplied by Toxicoop, Gödöllő, Hungary. All animal procedures were approved by the ethics committee of the Semmelweis University (22.1/606/001/2010, February 5, 2010) and were in accordance with the EU Council directives on laboratory animals (86/609/EEC).

### Cell culture conditions and assay for caspase 3 activity and lactate dehydrogenase release

Mouse embryonic fibroblast culture was established according to CSH protocol (11). Cells were maintained in DMEM supplemented with 10% fetal bovine serum and used between passage 3 and 7. One day before the experiment cells were seeded to 6 cm Petri dishes ( $3 \times 10^5$  cells/dish). Twenty-four hours later fetal bovine serum was withdrawn from the cell culture medium to induce cell death. Resveratrol treatment was initiated simultaneously with serum deprivation. When the rescue effect of resveratrol was investigated, resveratrol was added to the cell culture medium after 3-hour serum deprivation. Inhibitors of various signaling pathways were applied simultaneously with serum deprivation and/or resveratrol treatment.

For caspase activity assay after specified treatment periods (3, 4.5, 6 hours), cells were rinsed with PBS and harvested by trypsin-EDTA, and cytosol extract was prepared by hypotonic lysis with 0.6% Nonidet P40 according to Andrews and Faller (12). In order to evaluate direct caspase inhibitory effect of resveratrol, resveratrol was added directly to cytosol extract of serum-deprived fibroblasts immediately before measuring caspase 3 activity. Ac-VAD-CMK, a non-selective direct caspase inhibitor, was used in 20  $\mu$ M concentration as positive control. Caspase 3 activity and lactate dehydrogenase release were measured by commercially available kits according to the manufacturer instructions. Caspase 3 activity is shown as nanomol substrate cleaved by milligram protein in 3 hours.

### Statistical analysis

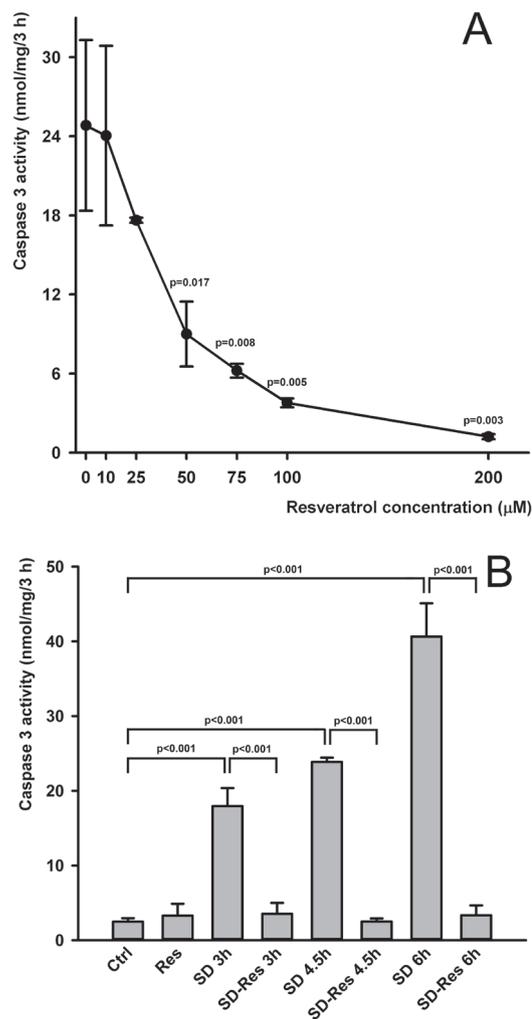
Data were expressed as mean  $\pm$  standard deviation. Comparisons were made by paired *t* test.  $P < 0.05$

was considered statistically significant. Data were analyzed by Microsoft Excel 2010 (Redmond, WA, USA).

## RESULTS

### Resveratrol dose-dependently prevented serum deprivation-induced caspase 3 activation in primary mouse embryonic fibroblasts

Primary mouse fibroblasts were exposed to serum deprivation, which after 3-6 hours induced significant caspase

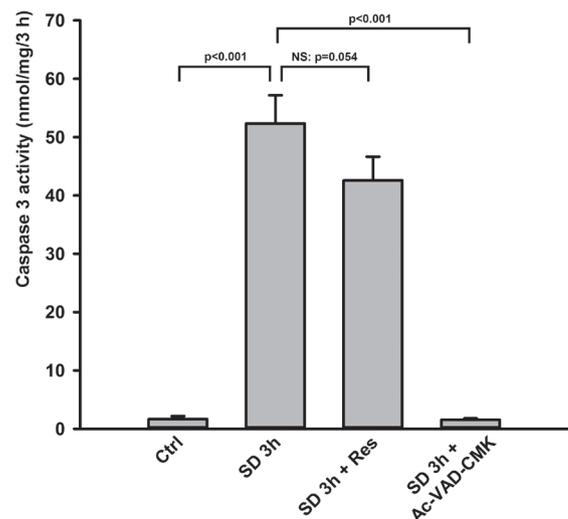


**FIGURE 1.** Resveratrol dose-dependently prevented caspase 3 activation after 3 h serum deprivation. Control value of caspase 3 activity in serum supported cells:  $1.76 \pm 0.097$  nmol/mg/3 h (A). 200 µM of resveratrol prevented caspase 3 activation after 3, 4.5, and 6 h serum deprivation (B).

3 activation ( $P < 0.001$ ). In order to evaluate the protective effect of resveratrol, the cells were treated with several concentrations (10, 25, 50, 75, 100, 200 µM) of resveratrol simultaneously with serum deprivation. Resveratrol prevented caspase 3 activation in a dose-dependent manner, with 50% inhibitory concentration ( $IC_{50}$ ) =  $66.3 \pm 13.81$  µM. Caspase 3 activation following 3 hour serum deprivation was completely inhibited at 200 µM resveratrol concentration (Figure 1A), and thus this level was used in the further experiments. This protective effect was also obtained after up to 6 hours of serum deprivation (Figure 1B). To verify whether resveratrol regulates the cellular response or directly interacts with caspase 3, resveratrol was added directly to the cytosol extract rather than to cell culture medium. Resveratrol showed no direct caspase inhibitory effect, although the known direct inhibitor Ac-VAD-CMK, used as positive control, completely blocked caspase 3 activity (Figure 2).

### Resveratrol exhibited rescue effect on serum deprivation-induced caspase 3 activation

We further investigated whether resveratrol reduced the already up-regulated caspase 3 activity. Primary fibroblasts were exposed to serum deprivation for 3 hours, after which



**FIGURE 2.** Resveratrol showed no direct caspase 3 inhibitory effect. When 200 µM of resveratrol was added to cytosol extract of serum-deprived fibroblast during caspase 3 activity measurement, it did not significantly reduce caspase 3 activity. A known direct caspase inhibitor, Ac-VAD-CMK, was used as positive control in 20 µM concentration.

the culture medium was supplemented with 200  $\mu$ M resveratrol for an additional 2 hours. Resveratrol significantly reduced the already activated caspase 3. It prevented not only its further increase but also reduced it to a level below that observed after 3-hour serum deprivation. These experiments indicate that resveratrol may have both protective and rescue effect on cells (Figure 3).

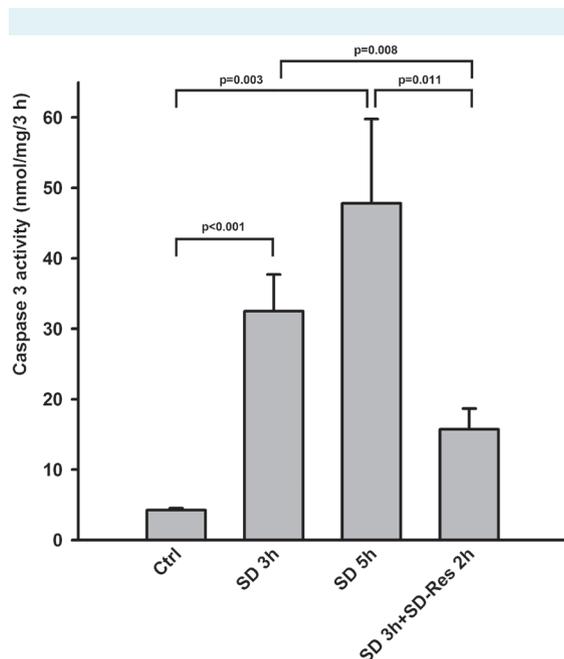
### Resveratrol reduced lactate dehydrogenase release induced by serum deprivation

Lactate dehydrogenase release was measured to evaluate whether the inhibition of caspase 3 activation by resveratrol was accompanied by increased cell viability. Cell viability decreased by 24 hour serum deprivation was significantly improved by 200  $\mu$ M resveratrol treatment (Figure 4).

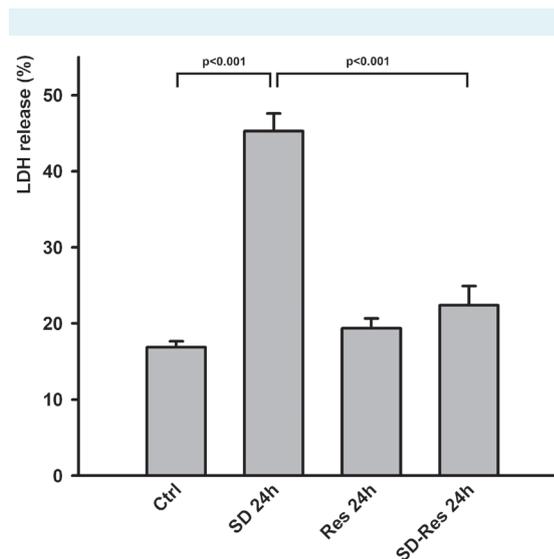
### The effect of resveratrol on caspase 3 activity involves p38 kinase pathway

In order to investigate the signaling cascades involved in the protective effects of resveratrol, we carried out experiments in the presence of specific inhibitors of p38 (50  $\mu$ M SB202190), JNK (50  $\mu$ M SP600125), ERK (50  $\mu$ M PD184352), PI3K (10  $\mu$ M wortmannin) kinase pathways, and SIRT1 (5

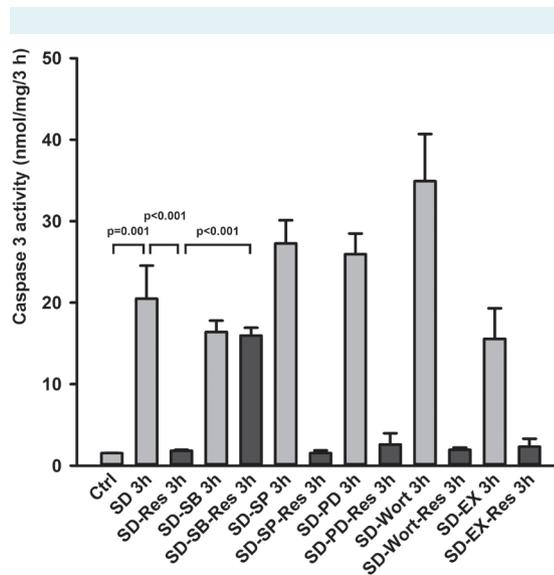
$\mu$ M EX-527). Among them, only p38 MAPK inhibitor SB202190 decreased the protective effect of resveratrol on caspase 3 activation (Figure 5).



**FIGURE 3.** Resveratrol showed rescue effect on caspase 3 activation. Following 3 h of serum deprivation, 200  $\mu$ M resveratrol was supplemented for an additional 2 h.



**FIGURE 4.** 200  $\mu$ M of resveratrol reduced lactate dehydrogenase release after 24 h serum deprivation.



**FIGURE 5.** The effect of 50  $\mu$ M SB202190 (p38 MAPK inhibitor), 50  $\mu$ M SP600125 (JNK inhibitor), 50  $\mu$ M PD184352 (ERK inhibitor), 10  $\mu$ M wortmannin (PI3 kinase inhibitor), and 5  $\mu$ M EX-527 (SIRT-1 inhibitor) on 3-h serum deprivation-induced caspase 3 activation and the protective action of 200  $\mu$ M resveratrol. Only p38 MAPK inhibitor SB202190 abolished the effect of resveratrol on caspase activation.

### The role of oxidative stress in the effect of resveratrol

Considering that p38 kinase pathway is activated by mild intracellular stress (13) and pro- and antioxidant properties of resveratrol had been previously described (14), we hypothesized that reactive oxygen species generation could be involved in caspase 3 activation induced by serum deprivation and/or the protective effect of resveratrol. To clarify if the antioxidant property of resveratrol may play a key role in its cytoprotective effect, we investigated the effect of 5 mM N-acetylcysteine, a well-known antioxidant agent, on caspase 3 activation. Contrary to our expectations, it did not prevent caspase 3 activation but exacerbated it. However, 200  $\mu$ M resveratrol abolished the combined effect of serum deprivation and N-acetylcysteine on caspase 3 activation (Figure 6).

## DISCUSSION

### Cytoprotective effect of resveratrol

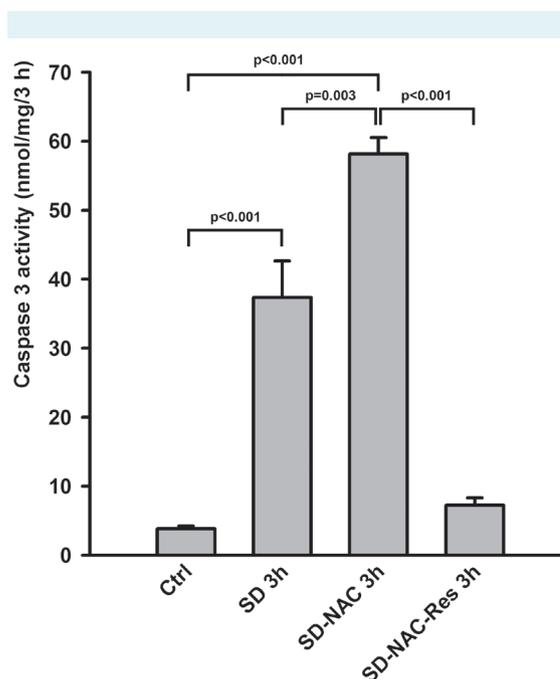
Resveratrol prevented serum deprivation-induced caspase 3 activation in primary fibroblasts and increased their viability. These results are in line with those of previous studies

performed on non-transformed cells using various toxic insults (15,16). In this study, cytoprotective effect of resveratrol was considerable, in 100-200  $\mu$ M concentration range, which is similar to another study (17). However, some recent studies observed lower concentrations, in the 10-20  $\mu$ M range to be efficient as well (9,16). The effective dose probably depends on the cell type and the intensity of the damaging insult used. The concentration found to be effective in the present study is considerably higher than the concentration that can be obtained from dietary sources, suggesting the need for resveratrol supplementation. Furthermore, resveratrol can serve as a lead compound for research of more potent cytoprotective medications.

To the best of our knowledge this is the first report demonstrating that resveratrol abolishes the already elevated caspase 3 activity induced by serum deprivation, suggesting its rescue effect. Resveratrol was found to prevent and improve cardiac function in cardiac fibroblasts (18,19) and to play a neuroprotective role in neurotoxic injury (20). However, our results showed that it is a promising cytoprotective agent which should be explored not only for prevention of age-related degenerative disorders, but also in the early treatment of degeneration following an acute insult.

### Probable mechanism of resveratrol action

It has already been suggested that several kinase pathways have a role in the cytoprotective effects of resveratrol. Cytoprotective functions of resveratrol were associated with the activation of PI3-kinase/Akt (21,22), p38 MAPK/JNK/ERK (23,24) signaling, and molecular pathways involving SIRT1 (9), an NAD dependent histone deacetylase. Our present findings indicate that the most critical signaling pathway in the protective effect of resveratrol against serum deprivation-induced caspase 3 activation is the activation of p38. The reports about the effects of resveratrol on p38 kinase pathway are rather contradictory. It was shown that through inhibition of p38 pathway resveratrol suppresses macrophage and vascular smooth muscle cell apoptosis (17,23). On the other hand, it exerted protective effect in H9c2 embryonic rat heart derived cells by up-regulating the p38 MAPK signaling (25). It was also shown to inhibit the proliferation of human primary fibroblasts and enhance their entry to senescence in p38 dependent manner (26). Therefore, p38 kinase seems to have a dual role as a regulator of cell fate, mediating either survival or death. Adams et al (27) reported that the specific function of p38 MAPKs in apoptosis depended on the cell type, stimuli, and/or p38 isoform. In accordance with their findings, we



**FIGURE 6.** Five mM of N-acetylcysteine (NAC) exacerbated serum deprivation-induced caspase 3 activation, but 200  $\mu$ M of resveratrol prevented their combined effect.

showed that p38 MAPK had a cytoprotective rather than proapoptotic role.

Several articles discuss antioxidant properties of resveratrol as the cause of its cytoprotective effect (14,16). Considering that N-acetylcysteine exacerbated rather than prevented serum deprivation-induced caspase activation and resveratrol abolished their combined effect, antioxidant properties cannot explain its protective action. Previous articles reported similar effect of N-acetylcysteine, concluding that the elevated glutathione level can inhibit NF- $\kappa$ B induced transcription of inhibitor of apoptosis protein, which can explain its potentiating effect on caspase activation (28,29). Since several previous reports demonstrated not only antioxidant but prooxidant characteristics of resveratrol (14,30), the latter might be involved in the activation of p38 MAPK and reduction of caspase 3 activation. A previous study suggested that the prooxidant activity of resveratrol was responsible for its inhibitory effect on apoptosis by creating an intracellular milieu non-permissive for caspase activation (30). These findings are in line with our results, which also indicate the role of p38 kinase in the protective effect of resveratrol and activation of this pathway by mild intracellular stress (13). However, the effect of resveratrol on oxidative state of cells requires further research.

Activation of p38 was also connected to increase in autophagic flux. This process is involved in the degradation of misfolded proteins or damaged organelles, such as depolarized mitochondria, which can prevent the release of proapoptotic mediators and the consequent caspase activation (25). Accordingly, two recent papers reported that resveratrol improved autophagic flux and prevented caspase cleavage in H9c2 rat cardiomyoblast cells (25,31). Similarly to our results, the protective effect of resveratrol depended on p38 MAPK activity (25). Based on these data, we can hypothesize that the effect of resveratrol on caspase 3 activation and cell survival might be connected with its prooxidant property, which may enhance autophagic flux via p38 activation.

A major limitation of this study is its *in vitro* nature, which is why further translational experiments are required to analyze the cytoprotective effect of resveratrol. In conclusion, we demonstrated the p38 MAPK signaling pathway-dependent cytoprotective effect of resveratrol against serum deprivation induced caspase 3 activation in primary fibroblasts. Also, resveratrol exhibited a rescue effect and reduced the already up-regulated caspase 3 activity. This

finding may contribute to the research of drugs used for prevention and treatment of age-related disorders.

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**Ethical approval** received from the ethics committee of the Semmelweis University (22.1/606/001/2010, February 5, 2010)

**Declaration of authorship** ZU, FB, and IV performed the experiments and evaluated the results, and ZU, ES, and TT participated in the planning of experiments, evaluation of results, and manuscript preparation.

**Competing interests** All authors have completed the Unified Competing Interest form at [www.icmje.org/coi\\_disclosure.pdf](http://www.icmje.org/coi_disclosure.pdf) (available on request from the corresponding author) and declare: no support from any organization for the submitted work; no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

## References

- 1 Fonseca-Kelly Z, Nassrallah M, Uribe J, Khan RS, Dine K, Dutt M, et al. Resveratrol neuroprotection in a chronic mouse model of multiple sclerosis. *Frontiers in Neurology*. 2012;3:84. [Medline:22654783](#) [doi:10.3389/fneur.2012.00084](#)
- 2 Darzynkiewicz Z, Zhao H, Halicka HD, Li J, Lee YS, Hsieh TC, et al. In search of antiaging modalities: evaluation of mTOR- and ROS/DNA damage-signaling by cytometry. *Cytometry A*. 2014;85:386-99. [Medline:24677687](#) [doi:10.1002/cyto.a.22452](#)
- 3 Jang JH, Surh YJ. Protective effect of resveratrol on beta-amyloid-induced oxidative PC12 cell death. *Free Radic Biol Med*. 2003;34:1100-10. [Medline:12684095](#) [doi:10.1016/S0891-5849\(03\)00062-5](#)
- 4 Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science*. 1997;275:218-20. [Medline:8985016](#) [doi:10.1126/science.275.5297.218](#)
- 5 Rege SD, Geetha T, Griffin GD, Broderick TL, Babu JR. Neuroprotective effects of resveratrol in Alzheimer disease pathology. *Frontiers in Aging Neuroscience*. 2014;6:218. [Medline:25309423](#) [doi:10.3389/fnagi.2014.00218](#)
- 6 Li Q, Huyen T, Ye LJ, Li J, Shi JL, Huang QS. Concentration-dependent biphasic effects of resveratrol on human natural killer cells *in vitro*. *J Agric Food Chem*. 2014;62:10928-35. [Medline:25360711](#) [doi:10.1021/jf502950u](#)
- 7 Barger JL, Kayo T, Vann JM, Arias EB, Wang J, Hacker TA, et al. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS ONE*. 2008;3:e2264. [Medline:18523577](#) [doi:10.1371/journal.pone.0002264](#)
- 8 Trinchieri NF, Nicotra G, Follo C, Castino R, Isidoro C. Resveratrol induces cell death in colorectal cancer cells by a novel pathway involving lysosomal cathepsin D. *Carcinogenesis*. 2007;28:922-31. [Medline:17116725](#) [doi:10.1093/carcin/bgl123](#)
- 9 Zhang L, Guo X, Xie W, Li Y, Ma M, Yuan T, et al. Resveratrol exerts an anti-apoptotic effect on human bronchial epithelial cells undergoing cigarette smoke exposure. *Molecular Medicine*

- Reports. 2015;11:1752-8. [Medline:25385506](#)
- 10 Khan RS, Fonseca-Kelly Z, Callinan C, Zuo L, Sachdeva MM, Shindler KS. SIRT1 activating compounds reduce oxidative stress and prevent cell death in neuronal cells. *Front Cell Neurosci.* 2012;6:63. [Medline:23293585](#) [doi:10.3389/fncel.2012.00063](#)
  - 11 Nagy A, Gertsenstein M, Vintersten K, Behringer R. Preparing mouse embryo fibroblasts. *CSH Protocols.* 2006;2006(1).
  - 12 Andrews NC, Faller DV. A rapid micropreparation technique for extraction of DNA-binding proteins from limiting numbers of mammalian cells. *Nucleic Acids Res.* 1991;19:2499. [Medline:2041787](#) [doi:10.1093/nar/19.9.2499](#)
  - 13 Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. *Cell Res.* 2005;15:11-8. [Medline:15686620](#) [doi:10.1038/sj.cr.7290257](#)
  - 14 de la Lastra CA, Villegas I. Resveratrol as an antioxidant and pro-oxidant agent: mechanisms and clinical implications. *Biochem Soc Trans.* 2007;35:1156-60. [Medline:17956300](#) [doi:10.1042/BST0351156](#)
  - 15 Liang Q, Wang XP, Chen TS. Resveratrol protects rabbit articular chondrocyte against sodium nitroprusside-induced apoptosis via scavenging ROS. *Apoptosis.* 2014;19:1354-63. [Medline:25001340](#) [doi:10.1007/s10495-014-1012-1](#)
  - 16 Zhou X, Chen M, Zeng X, Yang J, Deng H, Yi L, et al. Resveratrol regulates mitochondrial reactive oxygen species homeostasis through Sirt3 signaling pathway in human vascular endothelial cells. *Cell Death and Disease.* 2014;5:e1576. [Medline:25522270](#) [doi:10.1038/cddis.2014.530](#)
  - 17 Guo R, Su Y, Liu B, Li S, Zhou S, Xu Y. Resveratrol suppresses oxidised low-density lipoprotein-induced macrophage apoptosis through inhibition of intracellular reactive oxygen species generation, LOX-1, and the p38 MAPK pathway. *Cell Physiol Biochem.* 2014;34:603-16. [Medline:25116358](#) [doi:10.1159/000363026](#)
  - 18 Liu J, Zhuo X, Liu W, Wan Z, Liang X, Gao S, et al. Resveratrol inhibits high glucose induced collagen upregulation in cardiac fibroblasts through regulating TGF-beta1-Smad3 signaling pathway. *Chem Biol Interact.* 2015;227:45-52. [Medline:25559857](#) [doi:10.1016/j.cbi.2014.12.031](#)
  - 19 Chen T, Li J, Liu J, Li N, Wang S, Liu H, et al. Activation of SIRT3 by resveratrol ameliorates cardiac fibrosis and improves cardiac function via the TGF-beta/Smad3 pathway. *Am J Physiol Heart Circ Physiol.* 2015;308:H424-34. [Medline:25527776](#) [doi:10.1152/ajpheart.00454.2014](#)
  - 20 Ai Z, Li C, Li L, He G. Resveratrol inhibits beta-amyloid-induced neuronal apoptosis via regulation of p53 acetylation in PC12 cells. *Molecular Medicine Reports.* 2015;11:2429-34. [Medline:25483559](#)
  - 21 Liu MH, Yuan C, He J, Tan TP, Wu SJ, Fu HY, et al. Resveratrol protects PC12 cells from high glucose-induced neurotoxicity via PI3K/Akt/FoxO3a pathway. *Cell Mol Neurobiol.* 2015. [Medline:25471227](#) [doi:10.1007/s10571-014-0147-5](#)
  - 22 Simao F, Matte A, Pagnussat AS, Netto CA, Salbego CG. Resveratrol prevents CA1 neurons against ischemic injury by parallel modulation of both GSK-3beta and CREB through PI3-K/Akt pathways. *Eur J Neurosci.* 2012;36:2899-905. [Medline:22817531](#) [doi:10.1111/j.1460-9568.2012.08229.x](#)
  - 23 Guo R, Li W, Liu B, Li S, Zhang B, Xu Y. Resveratrol protects vascular smooth muscle cells against high glucose-induced oxidative stress and cell proliferation in vitro. *Medical Science Monitor Basic Research.* 2014;20:82-92. [Medline:24971582](#) [doi:10.12659/MSMBR.890858](#)
  - 24 Kutuk O, Poli G, Basaga H. Resveratrol protects against 4-hydroxynonenal-induced apoptosis by blocking JNK and c-JUN/AP-1 signaling. *Toxicological Sciences.* 2006;90:120-32. [Medline:16322078](#) [doi:10.1093/toxsci/kfj055](#)
  - 25 Lv XC, Zhou HY. Resveratrol protects H9c2 embryonic rat heart derived cells from oxidative stress by inducing autophagy: role of p38 mitogen-activated protein kinase. *Can J Physiol Pharmacol.* 2012;90:655-62. [Medline:22537597](#) [doi:10.1139/y2012-051](#)
  - 26 Faragher RG, Burton DG, Majecha P, Fong NS, Davis T, Sheerin A, et al. Resveratrol, but not dihydroresveratrol, induces premature senescence in primary human fibroblasts. *Age (Dordr).* 2011;33:555-64. [Medline:21318333](#) [doi:10.1007/s11357-010-9201-5](#)
  - 27 Adams RH, Porras A, Alonso G, Jones M, Vintersten K, Panelli S, et al. Essential role of p38alpha MAP kinase in placental but not embryonic cardiovascular development. *Mol Cell.* 2000;6:109-16. [Medline:10949032](#) [doi:10.1016/S1097-2765\(05\)00014-6](#)
  - 28 Zhang H, Limphong P, Pieper J, Liu Q, Rodesch CK, Christians E, et al. Glutathione-dependent reductive stress triggers mitochondrial oxidation and cytotoxicity. *FASEB J.* 2012;26:1442-51.
  - 29 Qanungo S, Wang M, Nieminen AL. N-Acetyl-L-cysteine enhances apoptosis through inhibition of nuclear factor-kappaB in hypoxic murine embryonic fibroblasts. *J Biol Chem.* 2004;279:50455-64. [Medline:15375156](#) [doi:10.1074/jbc.M406749200](#)
  - 30 Ahmad KA, Clement MV, Pervaiz S. Pro-oxidant activity of low doses of resveratrol inhibits hydrogen peroxide-induced apoptosis. *Ann NY Acad Sci.* 2003;1010:365-73. [Medline:15033754](#) [doi:10.1196/annals.1299.067](#)
  - 31 Wang B, Yang Q, Sun YY, Xing YF, Wang YB, Lu XT, et al. Resveratrol-enhanced autophagic flux ameliorates myocardial oxidative stress injury in diabetic mice. *J Cell Mol Med.* 2014;18:1599-611. [Medline:24889822](#) [doi:10.1111/jcmm.12312](#)