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## Covalent Enzyme Coupling on Cellulose Acetate Membranes for Glucose Sensor Development

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**Methods for immobilizing glucose oxidase (GOx) on cellulose acetate (CA) membranes are compared. The optimal method involves covalent coupling of bovine serum albumin (BSA) to CA membrane and a subsequent reaction of the membrane with GOx, which has previously been activated with an excess of *p*-benzoquinone. This coupling procedure is fairly reproducible and allows the preparation of thin membranes (5–20  $\mu\text{m}$ ) showing high surface activities (1–3 U/cm<sup>2</sup>) which are stable over a period of 1–3 months. Electrochemical and radiolabeling experiments show that enzyme inactivation as a result of immobilization is negligible. A good correlation between surface activity of membranes and their GOx load is observed.**

### INTRODUCTION

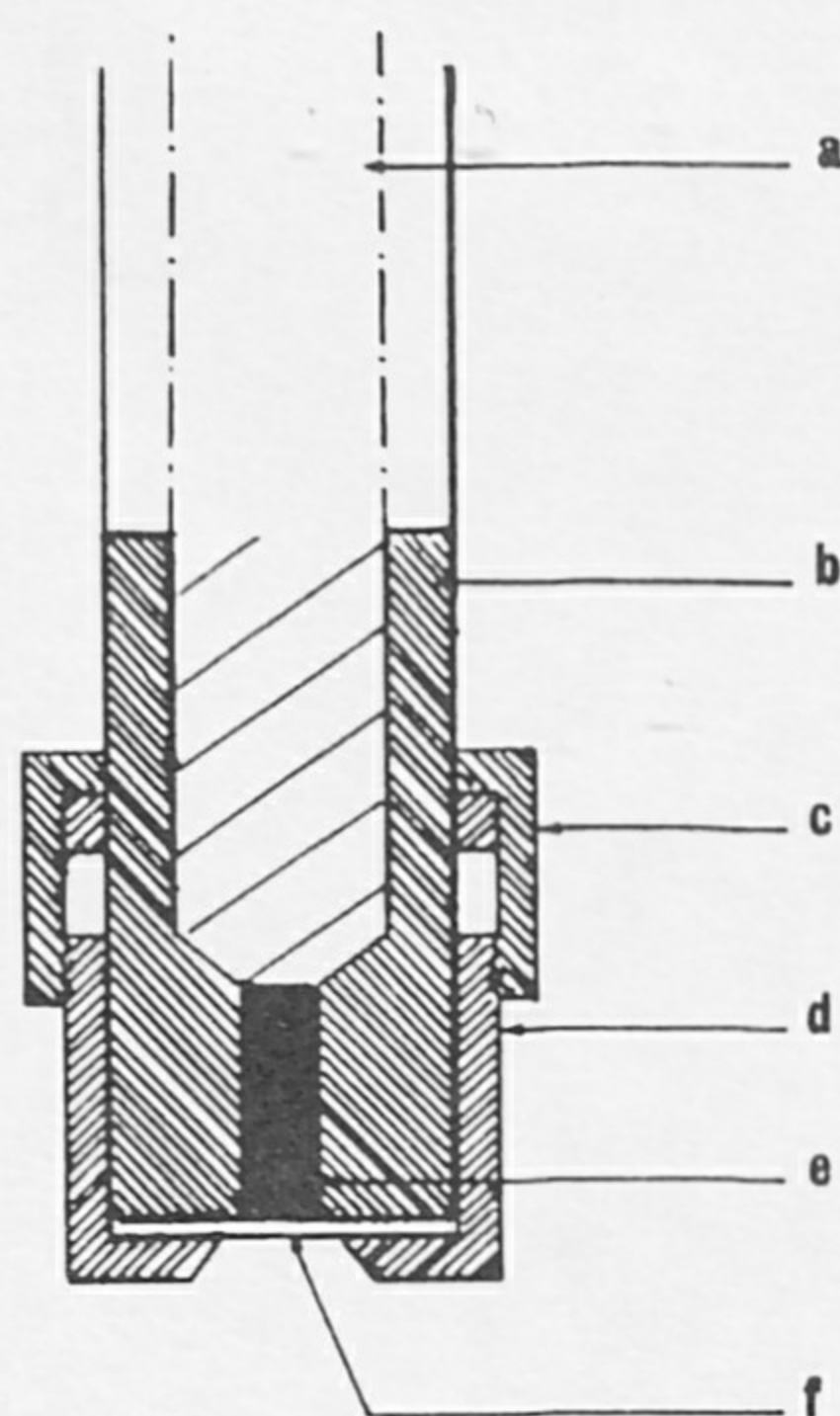
The performance of an enzyme electrode is ultimately dependent on the ability of its enzymatic membrane to sustain

and protect the enzyme. Among the available methods of enzyme immobilization, four methods are currently used: adsorption followed by reticulation with bifunctional reagents such as glutaraldehyde (1), covalent coupling between enzyme and activated support (2–5) or activated enzyme and support (6–8), and reversible immunological coupling (9). Those involving covalent coupling to solid supports are of great interest since they generally yield the best activity stabilities (10). Nevertheless two difficulties may be encountered: low levels of activatable or activated surface groups on the support and denaturation of enzyme if covalent coupling is accomplished through functional groups of the enzyme which are essential to its catalytic activity. Highly active and stable membranes may be prepared by acyl azide activation of reconstituted collagen films (2, 11). However such membranes have been found to be too thick and too fragile, especially at 37 °C, to be recommended for in vivo applications of enzyme electrodes. As cellulose acetate (CA) membranes of different thickness and permeability may easily be prepared by film casting or coating and because they exhibit significant permselectivity toward anions (12), we have studied their ability to support enzyme and be used for an in vivo implantable glucose sensor.

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**Figure 1.** Schematic diagram of rotating membrane electrode: (a) rotating disk electrode shaft and electrical contact, (b) Kel-F body, (c) threaded collar, (d) membrane support cap, (e) platinum electrode, and (f) membrane.

The literature is abundant with reports of glucose sensors and many of these are used routinely for in vitro clinical determinations. Despite a history of more than 25 years (13), no completely reliable implantable sensor has yet been developed (14). For such applications a miniaturized sensor is needed and the fabrication of such devices through multilayer film deposition is a major problem. It is quite evident that miniaturized sensors will yield high sensitivity and stability only if the enzyme layer and the associated polymer layers can be deposited in a reproducible fashion. Commercially available membranes are not suitable for this purpose. The enzyme can be immobilized and protective polymer films deposited by dip coating (15). Because of the low success rate (<40%) associated with such a technique, there exists a need for optimization of procedures for both enzyme and protective layer preparation. Starting with an intact membrane provides a defined system for the optimization process.

This paper proposes and evaluates a method of glucose oxidase (GOx) coupling to CA membranes. These membranes have initially been activated and coupled to bovine serum albumin (BSA) in order to increase the number of functional groups to which GOx may be coupled and to impart a proteic environment to the enzyme. GOx is coupled to CA-BSA membranes by using *p*-benzoquinone (PBQ) as bifunctional reagent (9, 16, 17). Use of radiolabeled GOx, in order to keep a track of enzymatic activity and investigate the reasons for the loss of sensor response with time, is also discussed. This work allows for the correct choice of coupling procedure and support before the whole glucose sensor is assembled.

## EXPERIMENTAL SECTION

**Instrumentation.** GOx activity of membranes was determined by continuous monitoring of enzymatically generated hydrogen peroxide on a combined platinum disk electrode (anode diameter 2 mm) (YSI Model 2150) covered with a general purpose cellulose acetate membrane. This electrode was connected to a YSI Model 25 Oxidase Meter (Yellow Springs Instruments, Yellow Springs, OH) and to a potentiometric recorder (Curken). Determinations of solution and membrane radioactivity were performed on a LKB Wallac 80000 automated gamma counter. GOx membranes were mounted on a rotated platinum disk electrode (anode diameter 3 mm) using a specifically designed membrane holder tip (Figure 1). The rotation speed of the membrane and anode was controlled by a Solea-Tacussel (Lyon, France) EDI motor and Controvit power unit. A Ag/AgCl reference and a platinum wire auxiliary electrode were connected, together with the platinum disk anode, to a BAS LC4A amperometric detector (Bioanalytical Systems, West Lafayette, IN) and potentiometric recorder (Curken). All experiments were performed in thermostated cells at 37 °C.

**Chemicals. Reagents.** Pure *p*-benzoquinone (PBQ) was obtained from Merck and was recrystallized from petroleum ether. Sodium borohydride from Sigma (98%) was used and Cerium(IV) sulfate-2-(sulfuric acid) was obtained from Alfa Division (Ventron). Acetone and ethanol (both AR 100%) were obtained from Mallinckrodt. Cellulose acetate (CA) with 39.8% acetyl content, was purchased from Aldrich Jansen Chimica. Glucose, hydrogen peroxide, and phosphate salts were reagent grade and all solutions were prepared in doubly distilled water.

**Proteins.** Glucose oxidase (GOx) isolated from *Aspergillus niger*, (grade VII, 136 000 U/g) and bovine serum albumin (BSA) fraction V were obtained from Sigma. **Radiochemicals.** Na<sup>125</sup>I (23 mCi/mL in 0.01 N NaOH) was obtained from ICN Radiochemicals (Irvine, CA).

**Glucose Oxidase Membrane Preparation.** *a. Cellulose Acetate Membrane Preparation.* With stirring, 1.8 g of CA was dissolved in 15.8 g of acetone, and 2.8 mL of distilled water was added. After homogenization, this solution was cast on a glass plate and evaporated for 60 s at 22 °C to form a thin membrane (0.015 mm in the wet state and 0.010 mm in the dry state). The CA membrane was removed from the glass plate by immersing the plate in distilled water. The membrane was cut into smaller pieces and stored in distilled water.

*b. Cellulose Acetate Membrane Activation. Procedure A.* Four CA membranes (2.5 cm × 2.5 cm) were suspended for 3 h in 100 mL of 1 mM Ce(IV) solution in 0.1 M nitric acid prepared just prior to use. Then the membranes were washed in distilled water for 5 min and immersed in 10 mL of 10 mg/mL BSA in 0.1 M borate buffer, pH 9.0. The volume of BSA solution was reduced to about 1 mL and three additions of 2 mg of sodium borohydride were made every 20 min at room temperature. After a second washing of 5 min in distilled water, the membranes were stored in 0.1 M phosphate buffered saline solution (PBS), pH 7.4, at room temperature until the GOx coupling reaction was performed.

*Procedure B.* This method is the same as procedure A except that a higher Ce(IV) concentration (100 mM) and a shorter oxidation time (20 min) were employed.

*Procedure C.* In this case 100 mL of 100 mM sodium periodate (20 min) was employed as oxidizing agent. The membranes were washed and BSA added as in procedure A. The Schiff base formed was reduced for 2 h by adding 4 mg of sodium cyanoborohydride to the 1 mL of BSA solution containing the membrane.

*c. Glucose Oxidase Activation.* Freshly prepared PBQ solution 0.1 mL (15 mg in 1 mL ethanol) was introduced in an aluminum-foil-covered tube containing 0.5 mL of GOx solution (20 mg in 0.1 M PBS pH 7.4). After 30 min of incubation at 37 °C, the mixture was filtered through a G-25 Sephadex column (1 × 10 cm) coupled with a peristaltic pump (20 mL/h) and equilibrated with 0.15 M sodium chloride. The first fraction, a pink-brown band of 2–3 mL, was collected and used as the enzyme coupling solution.

*d. Coupling Reaction.* The CA-BSA membranes were suspended in 2–3 mL of activated GOx solution, whose pH was adjusted to 8.0–9.0 with 0.25 mL of 1.0 M sodium carbonate solution. After 38 h of incubation at room temperature, the membranes were removed, washed in stirred 0.15 M potassium chloride solution for 24 h and finally stored in 0.1 M PBS, pH 7.4, containing 1.5 mM sodium azide. The azide was added to all storage and experimental buffers to limit microbial contamination and catalase activity.

**Radiolabeled Glucose Oxidase Preparation.** The iodine monochloride method described by Helmkamp, Contreras, and Bale (18) was chosen to incorporate <sup>125</sup>I into GOx. A 0.5 mM ICl reagent was prepared by adding 4 μL of pure ICl to 100 mL of 2 M NaCl. In a 1-mL conical vial was placed 80 μL of 0.5 mM ICl and the desired amount of Na<sup>125</sup>I solution. Typically, 1.0–2.5 μCi of <sup>125</sup>I was used to iodinate 1.0 mg of protein, depending upon the specific activity needed. The desired amount of glucose oxidase in a volume of 0.20 mL was added to the vessel with the room lights turned off. The contents of the vessel were mixed by tapping and allowed to react for 2 min in the dark. The reaction mixture was applied to the top of a G-25 Sephadex column (25 × 1 cm) that had been equilibrated with PBS, pH 7.4. Elution was continued with PBS at a flow rate of 0.5 mL/min and 1-mL fractions were collected. The effluent was monitored

with a gamma counter and the first fraction, containing labeled protein, was collected. The pooled protein was concentrated to about 1.5 mL by using pressure dialysis. The ratio of protein-bound iodide to total iodide was determined by the trichloroacetic acid (TCA) precipitation method (19). The value, expressed as efficiency of labeling, was usually 95–98%. The same data were used to calculate the specific activity of the iodinated glucose oxidase in microcuries of  $^{125}\text{I}$  attached per milligram of glucose oxidase. The spectrophotometric determination of enzymatic activity of radiolabeled glucose oxidase solution showed retention of 80–85% of the original activity.

**Glucose Oxidase Membrane Characterization.** *a. Enzymatic Surface Activity Determination.* The hydrogen peroxide anodic sensor, covered with a nonenzymatic CA membrane, was immersed into a thermostated cell (37 °C) containing 10 mL of PBS. When the background current was stabilized, i.e. after 10 min of polarization, 0.1 mL of 1 M glucose standard solution was added to the buffer solution. The incremental production of hydrogen peroxide was measured by placing the CA-BSA-PBQ-GOx membrane in the stirred buffer solution and then removing it. This process was repeated several times. The resulting increase in current, after suitable calibration with hydrogen peroxide standards, could be used to estimate the membrane surface enzymatic activity.

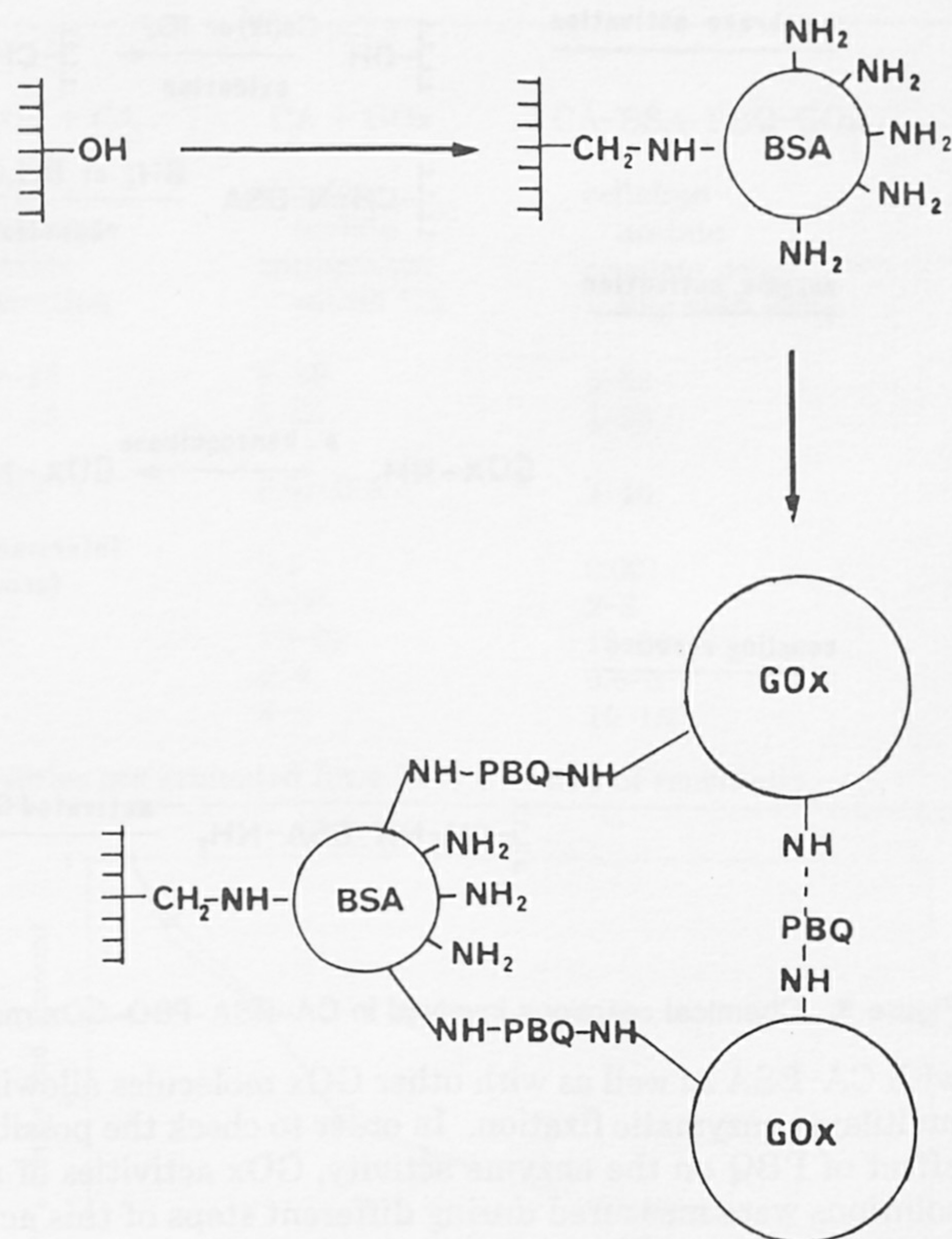
*b. Sensor Response.* The rotating membrane electrode was dipped into a thermostated cell at 37 °C containing 25 mL of PBS. When the background current was stabilized, i.e. after about 20 min of polarization, several 0.025–0.125 mL additions of 1 M glucose standard solution were performed until a 15–20 mM final concentration was obtained. Then one or two 25- $\mu\text{L}$  aliquots of 0.1 M hydrogen peroxide were added to the buffer solution. Response was calculated by comparing steady-state current either to the background current ( $I_b$ ) prior to any glucose addition or to the steady-state current corresponding to the previous addition. Thus either  $I - I_b$  vs  $C$  or  $\Delta I/\Delta C$  vs  $C$  or  $\log C$  curves were plotted.

*c. Immobilized Enzyme Determination.* The mass of immobilized GOx was estimated from the  $\gamma$  activity of the membranes prepared by using radiolabeled GOx and the specific activity of radiolabeled enzyme solution. With the initial enzyme activity of lyophilized powder (U/g) and a 15–20% decrease of this activity due to the radiolabeling procedure taken into account, a further estimation of membrane activity expressed in  $\text{U}/\text{cm}^2$  was accomplished.

## RESULTS AND DISCUSSION

The purpose of this work was to find a suitable method that allows the efficient and reproducible coupling of GOx to a support suitable for an *in vivo* glucose sensor. We were particularly concerned with the methods and support materials that could easily be adaptable to the preparation of needle-type microsensors. Thus all commercially available solid membranes were avoided and we restricted ourselves to cast CA films as support material.

**Cellulose Acetate Membrane Activation.** Bovine serum albumin (BSA) (1, 3, 6, 7), gelatin (20), or collagen (2, 8) are frequently used in enzyme immobilization techniques (21). They bring to the enzyme a proteic environment which seems favorable for its stability (11). As CA membranes usually possess low levels of accessible hydroxyl groups for covalent enzyme coupling, we have decided to increase the number of reacting groups by first covalently coupling BSA to CA (Figure 2). BSA contains more than 57 active amino groups per molecule and may be covalently grafted onto the free aldehyde groups obtained by CA hydroxyl group oxidation (Figure 3). The imine functions are subsequently reduced with borohydride. We have compared three different procedures for CA membrane activation and CA-BSA membranes from each procedure which were reacted with identical activated GOx solutions (*vide supra*). Comparisons of the properties of GOx membranes prepared by these three procedures (Table I) show that the best activities are obtained with procedure C, i.e. periodate oxidation followed by cyanoborohydride reduction. The use of cyanoborohydride, which is a milder reducing agent



**Figure 2.** General scheme of GOx coupling on CA membranes: enhancement of CA surface groups by covalent BSA coupling.

**Table I.** Activation Procedures of CA Membranes and Properties of CA-BSA-PBQ-GOx Membranes Obtained<sup>a</sup>

activation	CA-BSA-PBQ-GOx membrane preparation		
	procedure a	procedure b	procedure c
CA membrane oxidation			
reactant	Ce(IV)	Ce(IV)	$\text{IO}_4^-$
concentration	1	100	100
duration (min)	180	20	20
reaction with BSA			
concentration (g/L)	10	10	10
reduction			
reactant	$\text{BH}_4^-$	$\text{BH}_4^-$	$\text{BH}_3\text{CN}^-$
concentration (g/L)	$3 \times 2$	$3 \times 2$	4
duration (min)	$3 \times 20$	$3 \times 20$	120
radiolabeled GOx attached ( $\mu\text{g}\cdot\text{cm}^{-2}$ )	$3.8 \pm 1.5$ ( $n = 3$ )	$8.0 \pm 2.8$ ( $n = 5$ )	$13 \pm 1.8$ ( $n = 5$ )
GOx surface activity ( $\text{U}\cdot\text{cm}^{-2}$ )	$0.50 \pm 0.10$ ( $n = 3$ )	$1.0 \pm 0.40$ ( $n = 5$ )	$2.8 \pm 0.40$ ( $n = 5$ )

<sup>a</sup> Amounts of GOx and activities are expressed as mean  $\pm$  standard deviation for a set of membranes prepared identically. All GOx masses and activities are referred to membrane surface, i.e.  $0.8 \text{ cm}^2$ .

than borohydride (procedure A and B), probably prevents breaking of disulfide bonds of the BSA protein.

**Glucose Oxidase Activation and Coupling.** Following a procedure described by Avrameas et al. (16), we used *p*-benzoquinone (PBQ) for activating GOx solutions. GOx was treated with an excess of PBQ, reagent known to react with proteins (17), and the excess PBQ was separated by size exclusion chromatography (Figure 3). Most of the above-mentioned coupling reactions take place at low temperature, low ionic strength, and within the physiological pH range. The reason for treating GOx and not the amino groups of the CA-BSA support with PBQ is that it allows GOx-PBQ to react



Table II. Pertinent Parameters for GOx Membranes

parameters	collagen-GOx	Col-GOx + CA	CA + GOx	CA-BSA-PBQ-GOx
membrane material	reconstituted collagen	collagen + cel acetate	cellulose acetate	cellulose acetate
attachment procedure	acyl azide activation	acyl azide activation	entrapment within CA	covalent coupling with PBQ
membrane thickness ( $\mu\text{m}$ )				
dry	100	100 + 15	5-25	5-25
swollen	200-400	300 + 15	5-25	5-25
glucose responses:				
sensitivity <sup>a</sup> ( $\text{mA}\cdot\text{M}^{-1}\cdot\text{cm}^{-2}$ )	8-10	0.01-0.1	0.01-0.1	4-10
linear range (mM)				
lower limit	0.001	0.1	0.1	0.001
upper limit	2-3	40	5-30	2-3
response time transient (s)	20-50	50-90	10-60	10-60
steady state (min)	2-4	3-6	2-4	0.5-3
stability <sup>b</sup> (days)	120-2000		4-8	10-150

<sup>a</sup>All sensor sensitivities are referred to anode area, i.e.  $7.06 \text{ mm}^2$ . <sup>b</sup>Stabilities are evaluated for a 50% decrease of sensitivity.

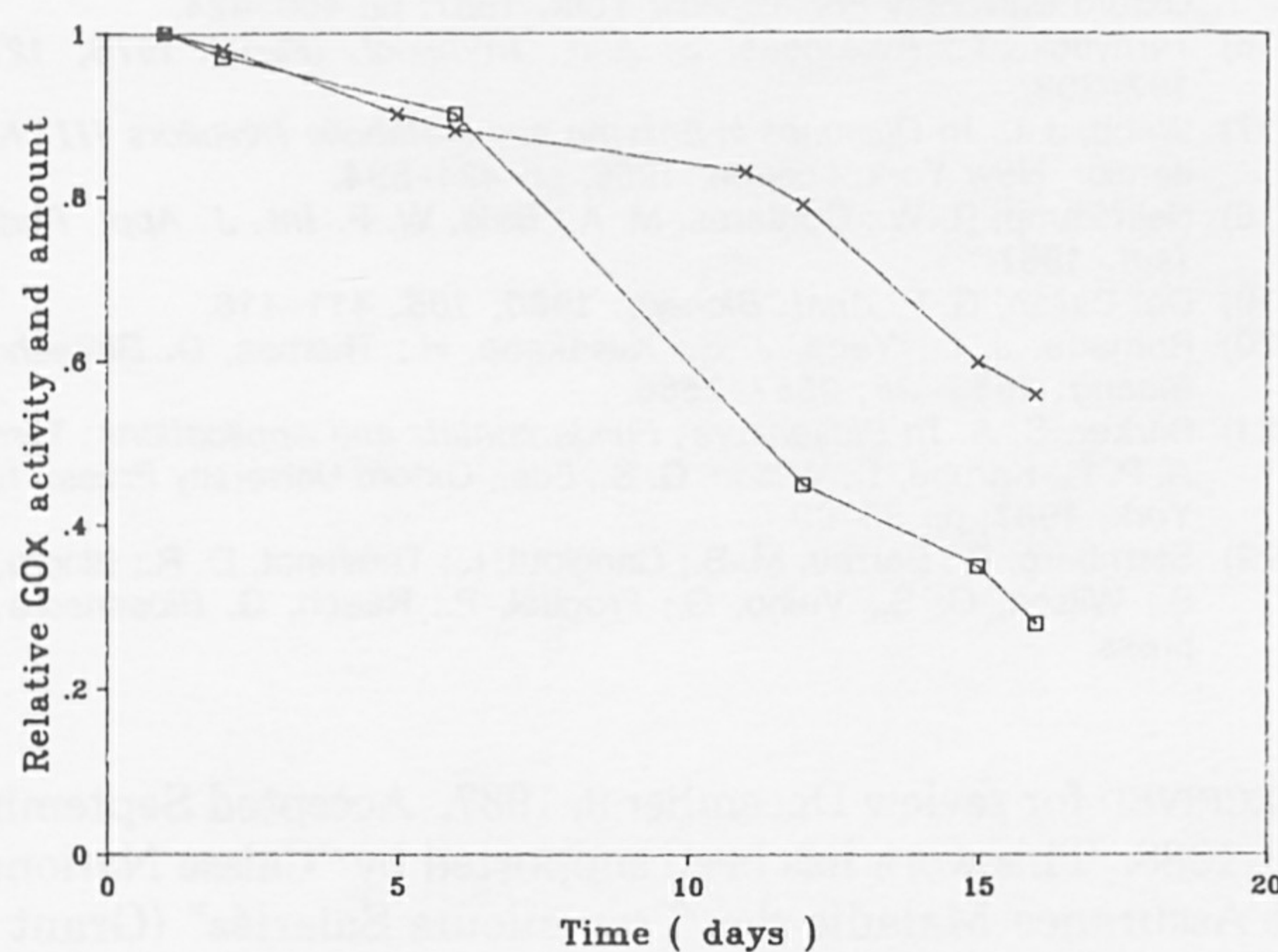


Figure 5. Relative evolution of surface GOx activity (O) and  $^{125}\text{I}$ -GOx (X) in a CA-BSA-PBQ-GOx membrane not treated with lysine after coupling.

membrane, thus preventing further coupling. The effect of this treatment is shown in Figure 6 where it will be noted that the enzymatic activity tracks the mass of immobilized protein. Protein is still lost from the membrane at an approximately comparable rate. However, in the absence of radiochemical measurements, membranes showed significant improvement on these stability values, probably because of decreased handling. A relative activity value as high as 80-85% was frequently observed after passage of 2-3 weeks (Figure 7). As the membranes were very fragile and unsupported, handling them on a routine basis must have caused their partial disintegration. These results demonstrate that enzyme inactivation is negligible after coupling and that any slight surface activity decrease, when observed, is probably related to immobilized enzyme loss from the membrane.

Microbial degradation as a possible explanation for the observed behavior was ruled out on the basis that azide was present as an inhibitor in all storage buffers. The fact that no significant release (<1%) of iodide was observed, as confirmed by the repetitive TCA assays (16) on stored iodinated enzyme solution also supports this belief.

Table II presents pertinent analytical parameters for GOx membranes when mounted on a platinum anode. The sensor sensitivities, as referred to anode area, of the CA-BSA-PBQ-GOx membranes are very close to those for highly active acyl azide activated collagen membranes (2). As CA membranes are significantly thinner than reconstituted collagen, especially when the latter are swollen, both transient and steady-state response times for them are correspondingly much shorter. The stability at  $37^\circ\text{C}$  and anion permselectivity of CA membranes are also preferable to those of collagen for

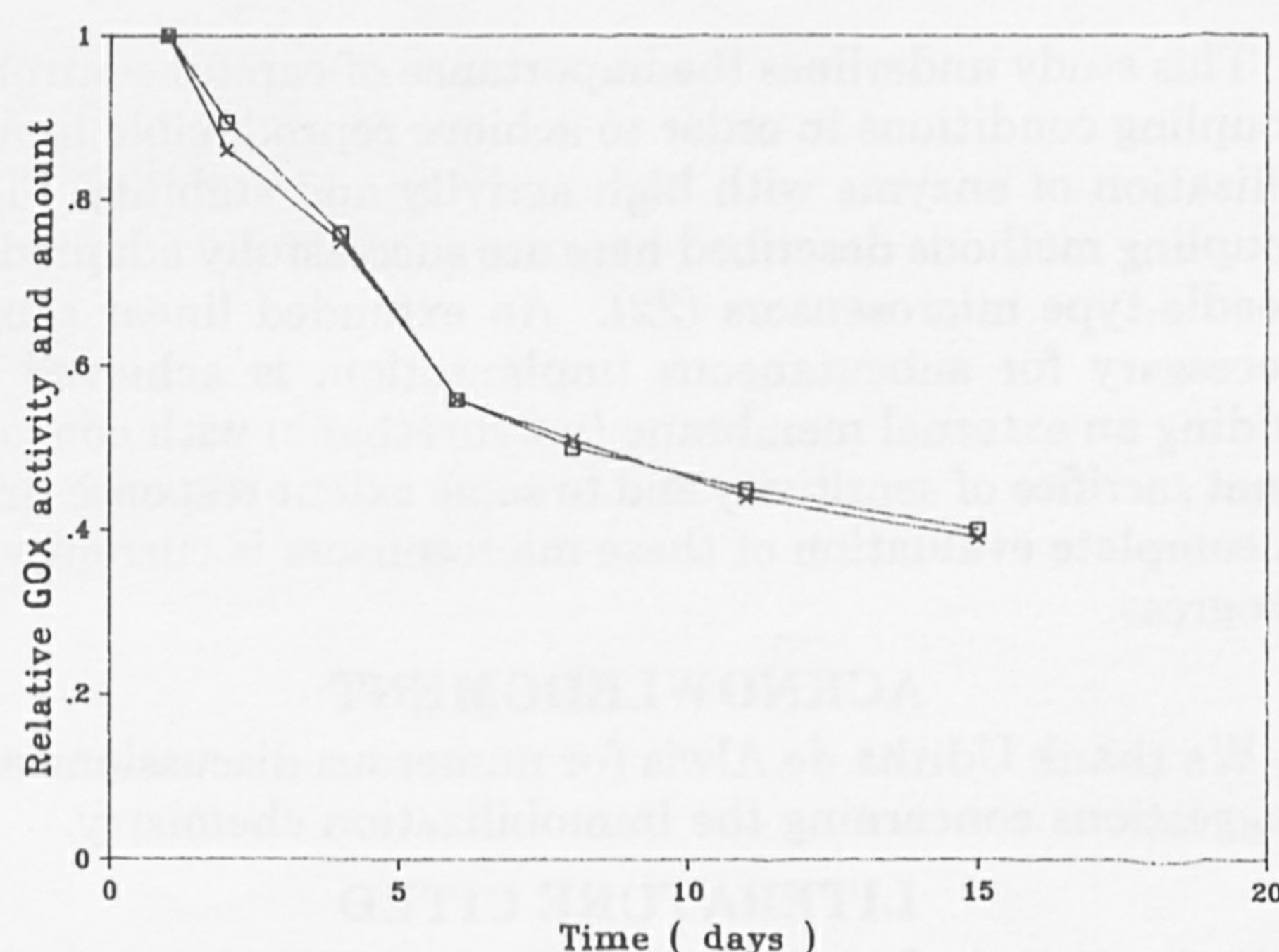


Figure 6. Relative evolution of surface GOx activity (O) and  $^{125}\text{I}$ -GOx amount (X) in a CA-BSA-PBQ-GOx membrane treated with lysine after coupling.

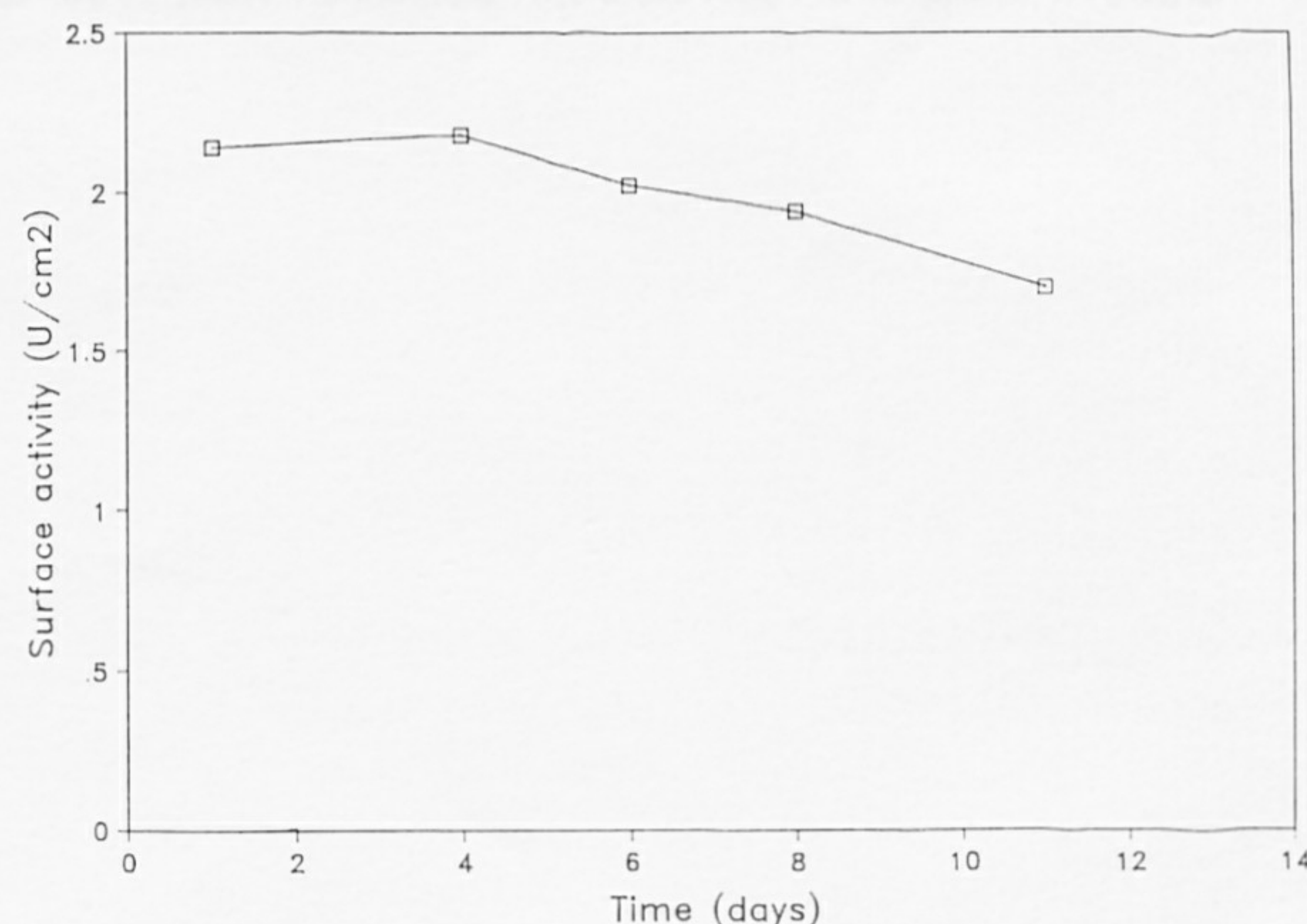
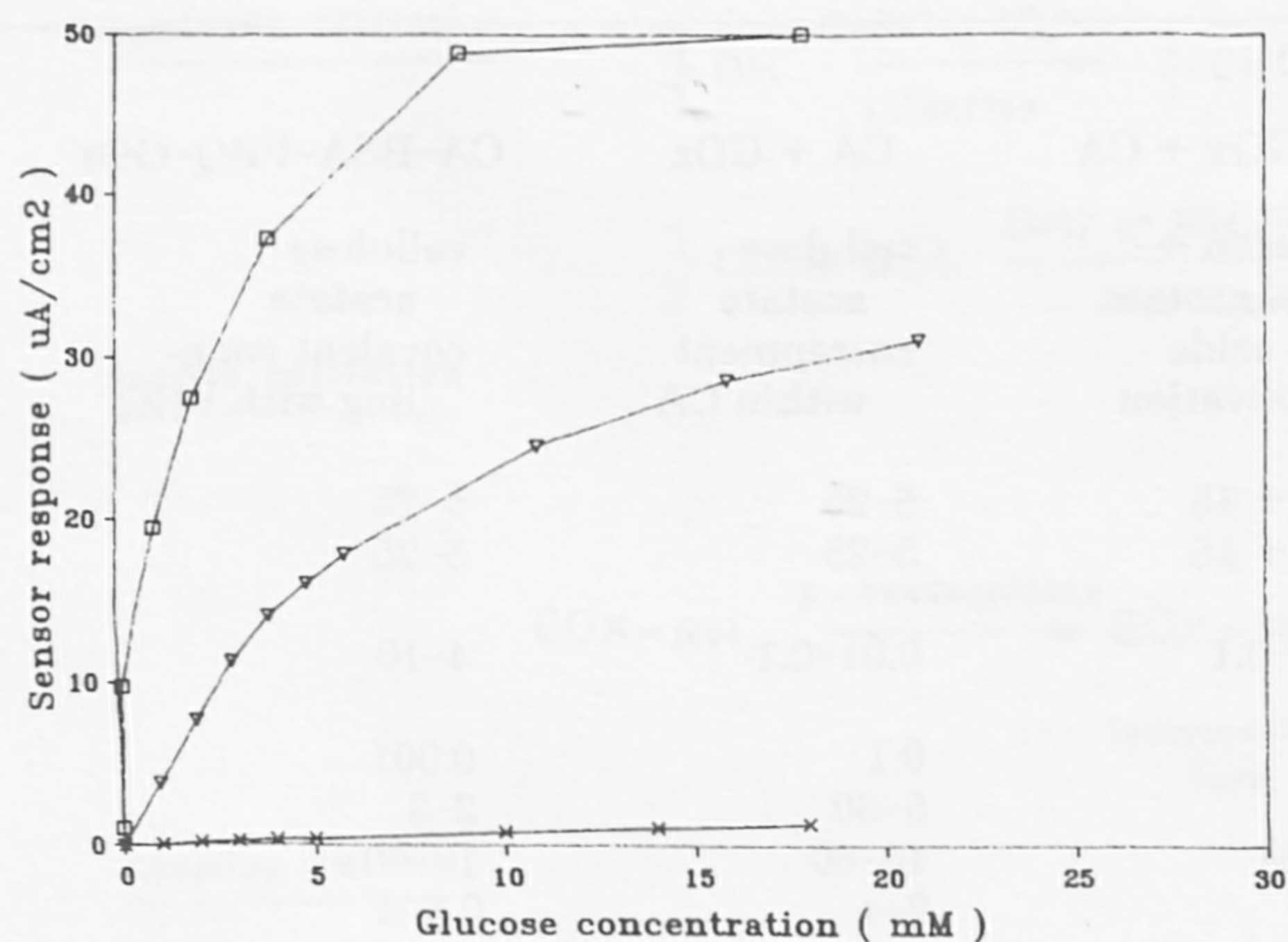


Figure 7. Evolution of surface GOx activity in a CA-BSA-PBQ-GOx membrane treated with lysine after coupling and not routinely handled for radiochemical measurements.

development of in vivo glucose sensors.

Figure 8 presents calibration curves of sensors prepared with such membranes. The linear ranges usually reach 2-3 mM glucose for such active collagen-GOx or CA-BSA-PBQ-GOx membranes indicating that, for higher glucose concentrations, the enzymatic reaction is the rate-limiting step. High linear ranges have been obtained either with less active CA membranes using a GOx entrapment procedure or with collagen-GOx membranes covered with nonenzymatic CA membranes allowing external diffusion restriction (see Table II). Indeed linear ranges as high as 15 mM are needed for potentially implantable glucose sensors (14).



**Figure 8.** Glucose calibration curves of sensors prepared with (O) collagen-GOx, (X) CA with entrapped GOx, and (Δ) CA-BSA-PBQ-GOx membranes.

This study underlines the importance of careful control of coupling conditions in order to achieve reproducible immobilization of enzyme with high activity and stability. The coupling methods described here are successfully adapted to needle-type microsensors (22). An extended linear range, necessary for subcutaneous implantation, is achieved by adding an external membrane (polyurethane) with concomitant sacrifice of sensitivity and to some extent response time. A complete evaluation of these microsensors is currently in progress.

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