



Mutations in ZIC2 in human holoprosencephaly: description of a novel ZIC2 specific phenotype and comprehensive analysis of 157 individuals.

Benjamin Solomon, Felicitas Lacbawan, Sandra Mercier, Nancy Clegg, Mauricio Delgado, Kenneth Rosenbaum, Christèle Dubourg, Véronique David, Ann Haskins Olney, Lars-Erik Wehner, et al.

▶ To cite this version:

Benjamin Solomon, Felicitas Lacbawan, Sandra Mercier, Nancy Clegg, Mauricio Delgado, et al.. Mutations in ZIC2 in human holoprosencephaly: description of a novel ZIC2 specific phenotype and comprehensive analysis of 157 individuals.. Journal of Medical Genetics, BMJ Publishing Group, 2010, 47 (8), pp.513-24. <10.1136/jmg.2009.073049>. <inserm-00439659>

> HAL Id: inserm-00439659 http://www.hal.inserm.fr/inserm-00439659

> > Submitted on 9 Dec 2009

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. Mutations in *ZIC2* in Human Holoprosencephaly: Comprehensive Analysis of 153 Individuals and Description of a Novel *ZIC2*-Specfic Phenotype

Benjamin D. Solomon¹#, Felicitas Lacbawan^{1,2}#, Sandra Mercier^{3,4}, Nancy J. Clegg⁵, Mauricio R. Delgado⁵, Kenneth Rosenbaum⁶, Christèle Dubourg³, Veronique David³, Ann Haskins Olney⁷, Lars-Erik Wehner^{8,9}, Ute Hehr^{8,9}, Sherri Bale¹⁰, Aimee Paulussen¹¹, Hubert J. Smeets¹¹, Emily Hardisty¹², Anna Tylki-Szymanska¹³, Ewa Pronicka¹³, Michelle Clemens¹⁴, Elizabeth McPherson¹⁵, Raoul C.M. Hennekam¹⁶, Jin Hahn¹⁷, Elaine Stashinko¹⁸, Eric Levey¹⁸, Dagmar Wieczorek¹⁹, Elizabeth Roeder²⁰, Kiyoshi Imaizumi²¹, Chayim Can Schell-Apacik^{22,23}, Carol W. Booth²⁴, Ronald L. Thomas²⁵, Sue Kenwrick²⁶, Amelia Keaton¹, Joan Z. Balog¹, Donald Hadley¹, Nan Zhou¹, Robert Long¹, Jorge I. Vélez¹, Daniel E. Pineda-Alvarez¹, Sylvie Odent^{3,4}, Erich Roessler¹, Maximilian Muenke¹* ¹National Human Genome Research Institute, National Institutes of Health, Bethesda, MD 20892, USA; ²Department of Pathology, State University of New York-Downstate Medical Center, Brooklyn, NY 11203, USA; ³CNRS Génétique et Développement, Université de Rennes, 35042 Rennes Cedex, France; ⁴Service de génétique clinique, CHU Hôpital Sud, 35042 Rennes Cedex, France ⁵Department of Neurology, Texas Scottish Rite Hospital for Children, University of Texas Southwestern Medical Center, Dallas, TX 75219, USA; ⁶Department of Genetics, Children's National Medical Center, Washington, DC 20010, USA; ⁷Department of Genetics, Munroe-Meyer Institute for Genetics and Rehabilitation, University of Nebraska Medical Center, Omaha, NE 68109, USA; ⁸Center for Human Genetics Regensburg, Regensburg 93053, Germany, ⁹Department of Human Genetics, University of Regensburg, Regensburg 93053, Germany; ¹⁰GeneDx, Gaithersburg, MD 20877, USA; ¹¹Department of Clinical Genetics, Academic Hospital Maastricht, 6229 GR Maastricht, Netherlands; ¹²Department of Obstetrics and Gynecology, University of Chapel Hill School of Medicine, Chapel Hill, NC 27514, USA; ¹³Clinic of Metabolic Diseases, Endocrinology and Diabetology, The Children's Memorial Health Institute, 02-004 Warsaw, Poland; ¹⁴Department of Genetics, University of Pittsburgh Medical Center, Pittsburgh, PA 15219, USA; ¹⁵Department of Genetics, Marshfield Clinic, Marshfield, WI 54449, USA; ¹⁶Department of Clinical Genetics, Academic Medical Center, 1105 AZ, Amsterdam, Netherlands; ¹⁷Department of Neurology, Stanford University School of Medicine, Palo Alto, CA 94305, USA; ¹⁸Kennedy Krieger Institute, Johns Hopkins University, Baltimore, MD 21218, USA, ¹⁹Institute of Human Genetics, University Duisburg-Essen, 174, 45147 Essen, Germany, ²⁰Department of Pediatrics, Division of Genetics and Metabolic Disorders, University of Texas Health Science Center at San Antonio, San Antonio, TX 78229, USA, ²¹Division of Medical Genetics, Kanagawa Children's Medical Center, Yokohama City, Kanagawa 232-8555, Japan, ²²Institute of Social Pediatric and Adolescent Medicine of the University of Munich, D-81377 Munich, Germany; ²³Practice of Human Genetics, 14050 Berlin, Germany, ²⁴Department of Genetics, Lutheran General Hospital, Park Ridge, IL 60068, USA; ²⁵Department of Obstetrics and Gynecology, Drexel School of Medicine, Philadelphia, PA 19129. USA; ²⁶Cambridge University Hospitals, Cambridge CB2 0QQ, UK.

#These authors contributed equally.
*Corresponding author: National Institutes of Health, Building 35, Room 1B-203 Bethesda, MD 20892 USA Phone: (301)594-7487 Fax: (301)496-7184 e-mail: <u>mamuenke@mail.nih.gov</u>

<u>Abstract</u>

Holoprosencephaly (HPE) is the most common malformation of the human forebrain, and may be due to cytogenetic anomalies, teratogens, occur in the context of a syndrome, or be due to mutations in single genes associated with non-syndromic HPE. Mutations in ZIC2, a transcription factor located on chromosome 13q32, are the secondmost common cause of non-syndromic, non-chromosomal HPE. Blood samples from over 1000 individuals with HPE-spectrum disorders and their relatives were analyzed for sequence variations in ZIC2. We examined clinical details and included all other known previously published and unpublished cases of mutations in ZIC2 through a literature search and collaboration with other centers. We find mutations in ZIC2 in 8% of probands with HPE, and describe 153 individuals from 116 unrelated kindreds, including 137 patients with molecularly-determined mutations in ZIC2 and 16 patients with deletions of the ZIC2 locus. Unlike HPE due to mutations in other genes, the vast majority of cases are sporadic and the proportional distribution of HPE types differs significantly from previously published analyses of non-chromosomal non-syndromic HPE. Furthermore, we describe a novel facial phenotype in patients with mutations in ZIC2 which includes bitemporal narrowing, upsplanting palpebral fissures, a short nose with anteverted nares, and a broad and well-demarcated philtrum, and large ears. This phenotype is distinct from the standard facial dysmorphisms associated with nonchromosomal, non-syndromic HPE. Our findings show that HPE due to mutations in ZIC2 is distinct from that due to mutations in other genes. This may shed light on the

mechanisms that contribute to the formation of the face and the forebrain and may help direct genetic counseling and diagnostic strategies.

<u>Manuscript</u>

Holoprosencephaly (HPE) is the most common malformation of the human forebrain, and results from failed or incomplete forebrain cleavage early in gestation. HPE occurs in 1 in 250 gestations, though the vast majority of conceptions with HPE do not survive to birth^{1,2}. HPE is categorized by the degree of forebrain separation into alobar, in which there is no interhemispheric division, semilobar, and lobar HPE, from the most to least severe type. More recently, middle interhemispheric variant (MIHV) HPE has also been described, which includes failed separation of the posterior frontal and parietal lobes³⁻⁶. The distribution of HPE types in both living patients and deceased fetuses with non-chromosomal, non-syndromic HPE has been estimated to be 22% alobar, 45% semilobar, and 33% lobar HPE⁷.

Common clinical features among patients with HPE include neurological impairment (often severe), seizures, diabetes insipidus, and characteristic dysmorphic facies. Traditionally, it is thought that in HPE "the face predicts the brain": in other words, more severe craniofacial anomalies correlate with more severe neuroanatomic findings.⁴ At the most severe end of the spectrum, facial features in patients with alobar HPE may include cyclopia and a proboscis (a tubular nasal structure located above the fused eyes). Other, more common facial dysmorphisms in less-severely affected patients include microcephaly (though hydrocephalus can lead to macrocephaly), hypotelorism, a flat nasal bridge, and cleft lip and/or palate. At the least severe end of the spectrum, termed microform HPE, patients may have subtle features such as mild microcephaly,

hypotelorism, single maxillary central incisors (SMCI) without appreciable CNS anomalies on conventional neuroimaging. These individuals are often identified due to the presence of a severely affected relative^{6,8-9}.

HPE is etiologically heterogeneous, and may be caused by cytogenetic anomalies, teratogenic influences, occur in the context of a syndrome, or be due to mutations in one of over 10 HPE-associated genes^{6-8,10-12}. In patients with HPE who have a normal chromosome analysis, a typical initial diagnostic strategy is to screen for mutations in four genes: *SHH* [MIM 600725], *ZIC2* [MIM 603073], *SIX3* [MIM 603714], and *TGIF* [MIM 602630]. Mutations in these genes can arise *de novo* or may be found in multiple members of large families segregating HPE-spectrum anomalies. In large kindreds, family studies demonstrate the incomplete penetrance and highly variable expressivity of these mutations^{3-4,6,13}.

ZIC2, located at chromosome 13q32, was first identified as an HPE candidate gene due to individuals with brain anomalies who were found to have deletions involving the long arm of chromosome 13. Subsequent analysis of patients with HPE identified mutations in *ZIC2*¹⁴⁻¹⁶. Mutations in this gene have previously been thought to be the second-most-common identified cause of non-chromosomal non-syndromic HPE (after mutations in *SHH*). In recent estimates, at least 3% of probands with HPE have mutations in *ZIC2*^{6,17}. *ZIC2* codes for a transcription factor which plays several roles in neurological development. Early in development, ZIC2 is predicted to play a role in axial midline establishment; later, ZIC2 appears to affect the development of the dorsal telencephalon^{18,19}. This latter role may explain the occurrence of neural tube defects in individuals with mutations in *ZIC2*, as well as the presence of MIHV-type HPE, though

this type can be seen in HPE due to mutations in other genes as well²⁰. Mouse models show that complete absence of Zic2 activity results in HPE due to mid-gastrulation failure of axial midline development, homozygous hypomorphic alleles result in normal gastrulation but dorsal forebrain malformations at later stages, and heterozygotes for null alleles are phenotypically normal. However, features in homozygous null mice may recapitulate the entire spectrum of HPE severity, suggesting that the phenotypic consequences of mutations depend on the perturbed developmental stage and may be affected by interacting genes^{17,19,21-22}. Of note, it has been suggested that mutations in *ZIC2* may result in HPE, but often do not result in facial features typically seen in human patients with HPE due to mutations in other genes²³.

Here we present clinical and genetic data on all known individuals with mutations in *ZIC2*, over half of whom were identified through our laboratory via direct sequencing. We also present data on individuals with deletions of the *ZIC2* locus ascertained by Multiplex Ligation-dependent Probe Amplification (MLPA) and Fluorescence in Situ Hybridization (FISH), chromosome analysis, or by oligonucleotide array comparative genomic hybridization. Through this comprehensive evaluation, we can identify specific characteristics of these individuals that can differentiate the phenotypic findings in patients with HPE due to *ZIC2* mutations from patients with HPE due to other genetic causes.

Blood samples from approximately 800 individuals with HPE-spectrum disorders and their relatives were collected prospectively over 18 years. These samples were analyzed for potential sequence variations in the *ZIC2* gene under our NHGRI-approved brain research protocol after appropriate consent had been obtained. A strategy for screening the *ZIC2* gene has previously been described¹⁷. Clinical history, photographs, and neuroimaging were reviewed where available, again after appropriate consent was obtained. Three patients were seen at the National Institutes of Health for a comprehensive evaluation. Collaborators sent us de-identified clinical and laboratory data on patients with identified mutations in *ZIC2*.

A Medline search was conducted to find previously reported cases of holoprosencephaly due to mutations in *ZIC2*. The key words and patient terms included "*ZIC2*", "holoprosencephaly", "HPE", "13q", and "13q32". References were also obtained from papers found through the literature search. As loci nearby *ZIC2* may contribute to brain malformations and there have been numerous reported cases of deletions of 13q with unreported clinical and genetic characterizations, only cases which had clear HPE and definitive deletion of the *ZIC2* locus without involvement of other chromosomes were considered. Cases were used from the following papers and abstracts: [Brown et al., 1993]; [Brown et al., 1995]; [Brown et al., 1998]; [Chen et al., 1998]; [Nanni et al., 2000]; [Gutierrez et al., 2001]; [Orioli et al., 2001]; [Brown et al., 2001]; [Marcorelles et al., 2002]; [Dubourg et al., 2004]; [Brown et al., 2005]; [Júnior et al., 2006]; [A. Paulussen et al., 2008, Eur. Soc. of Hum. Genet., abstract.]; [Roessler et al., 2009]; [Quélin et al., 2009]^{14-16,23-32}.

We describe a total of 153 patients, including 137 patients from 100 unrelated kindreds with molecularly-determined mutations in *ZIC2*, 7 patients with deletions of *ZIC2* ascertained by FISH testing and 9 patients with deletions of *ZIC2* ascertained by

chromosome analysis or by oligonucleotide microarray. By direct sequencing of DNA samples of an unselected group of unrelated patients with HPE, 8.25% (99/1200) have mutations in *ZIC2* (NIH: 49/285; Rennes: 41/532; Maastricht: 9/86). Additional cases among the approximately 800 tested in our laboratory were ascertained through screening methodology, including screening methods involving single-strand conformational polymorphism (SSCP) analysis and denaturing high-performance liquid chromatography (dHPLC).

Of note, in the descriptions below, unless otherwise stated, results refer only to individuals with molecularly determined mutations in *ZIC2*. Denominators differ among findings, as the prevalence of each phenotypic manifestation was calculated only where data was available for that specific finding. A summary of all patients is presented in the Supplementary Table.

Inheritance

Among probands in whom parents were available for testing, mutations were found to be *de novo* in 74% (49/66), maternally inherited in 18% (12/66), and paternally-inherited in 8% (5/66) of patients. There were no kindreds in which mutations or affected individuals were identified in more than 2 generations. However, in 4 cases, pedigree analysis showed that a mutation appeared to be inherited from a parent who had multiple affected children but for whom mutation testing was negative, implying either allele drop-out or, more likely, germline mosaicism.

HPE type

Prevalences of HPE types for both all described individuals and probands are presented as tables 1 and 2. Among patients with HPE, the distribution of classic HPE types (not including MIHV-type HPE) among patients with mutations in *ZIC2* differs significantly from a previously published analysis of HPE distribution among patients with nonchromosomal, non-syndromic HPE ($\chi^2 = 16.401$; p = 0.0003)⁷. Patients with mutations in *ZIC2* had a higher prevalence of more severe HPE types. Examples of characteristic findings on neuroimaging are shown in Figure 1.

Table 1.	Preval	lences	of H	PE	types.
----------	--------	--------	------	----	--------

HPE type	Patients with	Patients with		
	mutations in	deletion of		
	ZIC2 (%)	<i>ZIC2</i> (%) (n =		
	(n = 137)	16)		
Alobar	21	38		
Semilobar	32	19		
Lobar	8	7		
MIHV	3	0		
Microform	4	0		
None	5	0		
Unknown	26	38		

Table 2. Prevalences of HPE types among probands with known HPE type.

HPE	Patients with	Patients with
Туре	mutations in	deletion of
	ZIC2 (%)	<i>ZIC2</i> (%) (n =
	(n = 83)	10)

Alobar	33	60
Semilobar	51	30
Lobar	12	10
MIHV	5	0

Clinical Features

Among all individuals with mutations (including both probands and relatives of probands) for whom gender was known, 52% (61/118) were female and 48% (57/118) were male. Among probands for whom gender was known, 51% (43/84) were female and 49% (41/84) were male. There was no statistically significant difference between genders for either all individuals or probands alone.

Patients with recognizable brain anomalies invariably had some degree of neurological impairment. Of 66 families tested, 18 parents were identified as having mutations initially found in their severely-affected children; of those who were subsequently fully examined, only 2 parents were not found to have mild features of microform HPE. The overall penetrance of phenotypic manifestations (including microform HPE) due to mutations in *ZIC2* is estimated to be 96%; the prevalence of brain anomalies is estimated to be 90%.

While many individuals who received a full genetics evaluation had facial dysmorphisms, 67% (39/58) of patients with mutations in *ZIC2* did not display typical HPE facial features such as hypotelorism, flat nasal bridge, cleft lip/palate, or SMCI, features frequently seen in patients with mutations in genes such as *SHH* and *SIX3*²⁰. No patients had facial findings at the most severe end of the spectrum, such as cyclopia or synophthalmia, though one patient with semilobar HPE was described as having a

proboscis. A review of photos (figure 2) of available probands (n = 30) with mutations in *ZIC2* revealed a common phenotype in many patients consisting of bitemporal narrowing, upsplanting palpebral fissures, a short nose with anteverted nares, broad and well-demarcated philtrums, and relatively large ears, even accounting for microcephaly (Table 3). Although additional photos were not available for review, a similar facial phenotype was independently described by collaborators (S.M., S.O., CNRS Génétique et Développement, Université de Rennes/ Service de génétique clinique, CHU Hôpital Sud, Rennes, France). On review, this facial phenotype also occurs in previously published patients with mutations in *ZIC2*^{16,23}. Facial clefts, ranging from cleft lip and palate to a small unilateral nostril cleft, were described in 10% (7/69), while 17% (12/69) did not have clefts, but had high palates.

Patient	HPE type	BN	USPF	FNB	SNAN	BDP	LE	Other	Reference
1	А	+	+	+	+	+		Synophrys	This
									report
2	А		+	+	+				This
									report
3	А	+	+		+	+		Tall	This
								forehead	report
4	А	+	+			+			This
									report
5	А	+	+	+	+	+			[16]
6	А		+				+	Sloping	[16]
								forehead	
7	S		+		+				This
									report
8	S	+	+	+	+	+			This
									report
9	S		+		+	+			This
									report
10	S	+	+	+	+	+	+	Tall, broad	This
								forehead	report
11	S	+				+		Tall,	This
								narrow	report

Table 3. Description of common dysmorphic features in probands shown in Figure 2.

								head	
12	S		+		+	+	+	Slight	This
								synophrys,	report
								epicanthal	
								folds,	
								cupid-bow	
								upper lip	
13	S		+	+				Broad	This
								forehead	report
14	S	+	+		+		+		This
									report
15	S		+		+		+		This
									report
16	S		+		+		+	Synophrys	This
									report
17	S		+				+	Synophrys	This
1.0	~								report
18	S		+	+	+				This
10	~								report
19	S		+		+		+		This
•	~								report
20	S	+	+			+	+	Tall	This
	~							forehead	report
21	S		+						This
	G								report
22	S	+	+		+		+	Tall	[23]
00	C							forehead	[22]
23	S	+	+		+	+		T · 1	[23]
24	3	+	+	+	+			Iriangular	[23]
								mouth,	
								fooiog	
25	T				1			Tacles	[23]
25		+	+	+	+				[23]
20			+		+		+		1 IIIS report
27	MILIV				1				This
21	171111 1		T						report
28	MIHV			+					[23]
20	Unknown		_ _	Т	+		+	Sloping	
27	UIIKIIUWII						–	forehead	report
30	Unknown	+			+			Tall	This
50	UIKIUWII	Т						forehead	report
								TUTUTEau	report

BN: Bitemporal narrowing; USPF: Upslanting palpebral fissures; FNB: Flat nasal bridge; SNAN: Short nose and/or anteverted nares; BDP: Broad or deep philtrum; LE: Large ears

In terms of neurological defects beyond HPE, 12% (11/93) of individuals had hydrocephalus, and 4% (4/93) were reported as having neural tube defects. Finally, in terms of non-neurological manifestations, 14% had skeletal anomalies, 9% had cardiac anomalies, 7% had renal anomalies, 7% had genital anomalies, 4% had gastrointestinal anomalies, and 4% had pulmonary anomalies (n =76). Five percent had more than 3 congenital anomalies in these systems, including complex congenital heart, renal, and skeletal abnormalities.

Genotypic and functional analysis

The molecular findings among patients with mutations in *ZIC2* have been recently and extensively analyzed¹⁷. Among kindreds with molecularly-identified mutations, 84% (84/100) were unique. One mutation, which resulted in an alanine expansion and which has been show to result in greatly reduced function, occurred in 11 apparently unrelated kindreds.

Among the 100 unrelated kindreds with molecularly-demonstrated mutations, 38% (38/100) had frameshift mutations, 21% (21/100) had missense mutations, 16% (16/100) were nonsense mutations, 16% (16/100) were in-frame duplications, 5% (5/100) were predicted to result in alternative splicing, 3% (3/100) were in-frame deletions, and 1% (1/100) was an in-frame insertion. 89% (17/19) of the in-frame deletions and duplications occurred in the poly-alanine segment of the gene.

The vast majority (98%) of family-specific mutations were predicted or proven significant loss-of-function. Interestingly, among the very few patients whose mutations were not predicted null, alobar HPE was not observed and 66% (2/3) were inherited, in contrast to the overall estimation that 69% of mutations were *de novo*. There was no

correlation between the type, location, and functional activity conferred by a mutation with the presence of facial dysmorphisms or with HPE severity.

Mutations in *ZIC2* are one of the two most common single-gene causes of nonsyndromic HPE (with *SHH*). As patients with *ZIC2* mutations may not have facial dysmorphisms typically associated with HPE, the diagnosis of HPE may not be obvious on clinical encounter. Mutations in *ZIC2* may be an underappreciated cause of HPE, especially in the instance of an early fetal demise when high-quality brain imaging or pathologic analysis is not available.

However, our analysis of this large cohort of patients with mutations in *ZIC2* reveals several unique features resulting from mutations in this gene which distinguish the patients described here from patients with mutations in other HPE-associated genes. First, many patients with mutations in *ZIC2* have a subtle but distinct dysmorphic facial phenotype which may help aid diagnosis. This facial appearance is unique among patients with HPE, and has not been seen in patients with HPE due to other genetic etiologies. Second, unlike other genes associated with HPE, the majority of mutations occur *de novo*. Our data suggests the presence of at least 4 families in which germline mosaicism seems to be causative of HPE in a child, which has important implications for genetic counseling. Parents who test negative for *ZIC2* mutations through analysis of peripheral blood may still be at risk for having other affected children.

Third, along these lines, we did not identify any large pedigrees in which numerous individuals from multiple generations were identified, which is not the case for the other common HPE-associated genes such as *SHH* or *SIX3*. This could imply that mutations in *ZIC2* are less likely to result in mildly-affected individuals than mutations in other HPE-associated genes. Since *ZIC2* mutations occur relatively frequently in non-syndromic HPE, this would further imply that the mutation rate for these mutations is higher than, for example, mutations in *SIX3*, which are overall less frequent, but occur more often in large kindreds with multiple affected generations²⁰. However, the high penetrance and relatively severe findings may bring individuals to clinical attention earlier, resulting in ascertainment bias.

Finally, laboratories and clinicians must be aware of the importance of functional data in order to characterize mutations and to inform counseling of affected families; we know specifically of certain repeat variants in *ZIC2* resulting in different numbers of histidine repeats, which have previously been thought to be pathogenic, but on later family analysis, are now thought to be polymorphisms which may be common in ethnicities not originally part of control populations¹⁷.

One shortcoming of this report is that the available retrospective collection of clinical data was not uniform. For this reason, it is likely that we underestimate the prevalence of many of the findings (such as neural tube defects and other congenital anomalies). Despite the challenges synthesizing the data, the availability of a large cohort of patients with mutations affecting the same gene greatly enriches our understanding of HPE in general and *ZIC2* in particular. This study demonstrates the existence of a previously unnoticed *ZIC2*-specific phenotype, and highlights the importance of a comprehensive and collaborative approach to study HPE and other complex genetic disorders.

ACKNOWLEDGEMENTS AND AFFILIATIONS

We would like to express our deep gratitude to the patients and families who participated in these studies. The authors would also like to thank all of the members of the Carter Centers for Brain Research in Holoprosencephaly and Related Malformations.

This research was supported by the Division of Intramural Research, National Human Genome Research Institute, National Institutes of Health, Department of Health and Human Services, United States of America and GIS Maladies Rares GISMR0701/DHOS, France.

There are no competing interests.

References

1. Matsunaga E., Shiota K. (1977). Holoprosencephaly in human embryos: epidemiologic studies of 150 cases. Teratology. *16*, 261-72.

2. Leoncini E., Baranello G., Orioli I.M., Annerén G., Bakker M., Bianchi F., Bower C., Canfield M.A., Castilla E.E., Cocchi G., et al. (2008). Frequency of holoprosencephaly in the International Clearinghouse Birth Defects Surveillance Systems: Searching for population variations. Birth Defects Res A Clin Mol Teratol. *82*, 585-591.

3. Muenke M., Beachy P.A. (2000). Genetics of ventral forebrain development and holoprosencephaly. Curr Opin Genet Dev. *10*, 262-269.

4. Cohen M.M. Jr. (2006). Holoprosencephaly: clinical, anatomic, and molecular dimensions. Birth Defects Res A Clin Mol Teratol. *76*, 658-673.

5. Barkovich A.J., Simon E.M., Clegg N.J., Kinsman S.L., Hahn J.S. (2002). Analysis of the cerebral cortex in holoprosencephaly with attention to the sylvian fissures. AJNR Am J Neuroradiol. *23*, 143-150.

Dubourg C., Bendavid C., Pasquier L., Henry C., Odent S., David V. (2007).
 Holoprosencephaly. Orphanet J Rare Dis. 2, 8.

7. Lazaro L., Dubourg C., Pasquier L., Le Duff F., Blayau M., Durou M.R., de la Pintière A.T., Aguilella C., David V., Odent S. (2004). Phenotypic and molecular variability of the holoprosencephalic spectrum. Am J Med Genet A. *129A*, 21-24.

8. Cohen M.M. Jr. (1989). Perspectives on holoprosencephaly: Part I. Epidemiology, genetics, and syndromology. Teratology. *40*, 211-235.

9. Cohen M.M. Jr, Sulik K.K. (1992). Perspectives on holoprosencephaly: Part II. Central nervous system, craniofacial anatomy, syndrome commentary, diagnostic approach, and experimental studies. J Craniofac Genet Dev Biol. *12*, 196-244.

10. Edison R.J., Muenke M. (2004). Mechanistic and epidemiologic considerations in the evaluation of adverse birth outcomes following gestational exposure to statins. Am J Med Genet. *131*, 287-298.

11. Edison R.J., Muenke M. (2005). Central nervous system and limb anomalies in case reports of first-trimester statin exposure. N Engl J Med. *350*, 1579-1582. Erratum in: N Engl J Med. (2005). *352*, 2759.

12. Croen L.A., Shaw G.M., Lammer E.J. (1996). Holoprosencephaly: epidemiologic and clinical characteristics of a California population. Am J Med Genet. *64*, 465-472.

13. Collins A.L., Lunt P.W., Garrett C., Dennis N.R. (1993). Holoprosencephaly: a family showing dominant inheritance and variable expression. J Med Genet. *30*, 36-40.

14. Brown S., Gersen S., Anyane-Yeboa K., Warburton D. (1993). Preliminary definition of a "critical region" of chromosome 13 in q32: report of 14 cases with 13q deletions and review of the literature. Am J Med Genet. *45*, 52-59.

15. Brown S., Russo J., Chitayat D., Warburton D. (1995). The 13q- syndrome: the molecular definition of a critical deletion region in band 13q32. Am J Hum Genet. *57*, 859-8866.

Brown S.A., Warburton D., Brown L.Y., Yu C.Y., Roeder E.R., Stengel-Rutkowski
 S., Hennekam R.C., Muenke M. (1998). Holoprosencephaly due to mutations in *ZIC2*, a homologue of Drosophila odd-paired. Nat Genet. 20, 180-183.

17. Roessler E., Lacbawan F., Dubourg C., Paulussen A., Herbergs J., Hehr U., Bendavid C., Zhou N., Ouspenskaia M., Bale S., et al. (2009). The full spectrum of holoprosencephaly-associated mutations within the *ZIC2* gene in humans predicts loss-of-function as the predominant disease mechanism. Hum Mutat. *30*, E541-554.
18. Cheng X., Hsu C.M., Currle D.S., Hu J.S., Barkovich A.J., Monuki E.S. (2006). Central roles of the roof plate in telencephalic development and holoprosencephaly. J

Neurosci. 26, 7640-7649.

19. Warr N., Powles-Glover N., Chappell A., Robson J., Norris D., Arkell R.M. (2008). *Zic2*-associated holoprosencephaly is caused by a transient defect in the organizer region during gastrulation. Hum Mol Genet. *17*, 2986-2996.

20. Lacbawan F., Solomon B.D., Roessler E., El-Jaick K., Domené S., Velez J.I., Zhou N., Hadley D., Balog J.Z., Long R., et al. (2009). Clinical Spectrum of *SIX3*-Associated Mutations in Holoprosencephaly: Correlation between Genotype, Phenotype, and Function. J Med Genet. *46*, 389-398.

21. Elms P., Siggers P., Napper D., Greenfield A, Arkell R. (2003). *Zic2* is required for neural crest formation and hindbrain patterning during mouse development. Dev Biol. *264*, 391-406.

22. Nagai T., Aruga J., Minowa O., Sugimoto T., Ohno Y., Noda T., Mikoshiba K. Zic2 regulates the kinetics of neurulation. Proc Natl Acad Sci U S A. *97*, 1618-1623.

23. Brown L.Y., Odent S., David V., Blayau M., Dubourg C., Apacik C., Delgado M.A.,
Hall B.D., Reynolds J.F., Sommer A., et al. (2001). Holoprosencephaly due to mutations
in *ZIC2*: alanine tract expansion mutations may be caused by parental somatic
recombination. Hum Mol Genet. *10*, 791-796.

24. Chen C.P., Chern S.R., Lee C.C., Chen L.F., Chuang C.Y., Chen M.H. (1998). Prenatal diagnosis of de novo isochromosome 13q associated with microcephaly, alobar holoprosencephaly and cebocephaly in a fetus. Prenat Diagn. *18*, 393-398.

25. Nanni L., Croen L.A., Lammer E.J., Muenke M. (2000). Holoprosencephaly: molecular study of a California population. Am J Med Genet. *90*, 315-319.

26. Gutierrez J., Sepulveda W., Saez R., Carstens E., Sanchez J. (2001). Prenatal

diagnosis of 13q- syndrome in a fetus with holoprosencephaly and thumb agenesis.

Ultrasound Obstet Gynecol. 17, 166-168.

27. Orioli I.M., Castilla E.E., Ming J.E., Nazer J., Burle de Aguiar M.J., Llerena J.C.,

Muenke M. (2001). Identification of novel mutations in SHH and ZIC2 in a South

American (ECLAMC) population with holoprosencephaly. Hum Gene. 109, 1-6.

28. Brown L., Paraso M., Arkell R., Brown S. (2005). In vitro analysis of partial loss-offunction *ZIC2* mutations in holoprosencephaly: alanine tract expansion modulates DNA binding and transactivation. Hum Mol Genet. *14*, 411-420. 29. Marcorelles P., Loget P., Fallet-Bianco C., Roume J., Encha-Razavi F., Delezoide A.L. (2002). Unusual variant of holoprosencephaly in monosomy 13q. Pediatr Dev Pathol. *5*, 170-178.

30. Dubourg C., Lazaro L., Pasquier L., Bendavid C., Blayau M., Le Duff F., Durou M.R., Odent S., David V. (2004). Molecular screening of *SHH*, *ZIC2*, *SIX3*, and *TGIF* genes in patients with features of holoprosencephaly spectrum: Mutation review and genotype-phenotype correlations. Hum Mutat. *24*, 43-51.

31. Araujo Júnior E., Filho H.A., Pires C.R., Filho S.M. (2006). Prenatal diagnosis of the 13q- syndrome through three-dimensional ultrasonography: a case report. Arch Gynecol Obstet. *274*, 243-245.

33. Quélin C., Bendavid C., Dubourg C., de la Rochebrochard C., Lucas J., Henry C.,
Jaillard S., Loget P., Loeuillet L., Lacombe D., et al. (2009). Twelve new patients with
13q deletion syndrome: genotype-phenotype analyses in progress. Eur J Med Genet. *52*,
41-46.

Figure Legends

Figure 1. Characteristic findings on neuroimaging. a: Alobar HPE with shunt in place; b,c: semilobar HPE with large dorsal cyst; d: semilobar HPE without dorsal cyst; e,f: MIHV-type HPE

Figure 2. Patients with mutations in *ZIC2*, arranged by HPE type. Note *ZIC2*-specific facial findings, consisting of bitemporal narrowing, upslanting palpebral fissures, flat nasal bridge and a short nose with upturned nares, a broad and/or deep philtrum, and large ears.