## RESEARCH



# Incident disease associations with mosaic chromosomal alterations on autosomes, X and Y chromosomes: insights from a phenome-wide association study in the UK Biobank

Shu-Hong Lin<sup>1</sup>, Derek W. Brown<sup>1</sup>, Brandon Rose<sup>2</sup>, Felix Day<sup>3</sup>, Olivia W. Lee<sup>1</sup>, Sairah M. Khan<sup>1</sup>, Jada Hislop<sup>1</sup>, Stephen J. Chanock<sup>1</sup>, John R. B. Perry<sup>3</sup> and Mitchell J. Machiela<sup>1\*</sup>

## Abstract

**Background:** Mosaic chromosomal alterations (mCAs) are large chromosomal gains, losses and copy-neutral losses of heterozygosity (LOH) in peripheral leukocytes. While many individuals with detectable mCAs have no notable adverse outcomes, mCA-associated gene dosage alterations as well as clonal expansion of mutated leukocyte clones could increase susceptibility to disease.

**Results:** We performed a phenome-wide association study (PheWAS) using existing data from 482,396 UK Biobank (UKBB) participants to investigate potential associations between mCAs and incident disease. Of the 1290 ICD codes we examined, our adjusted analysis identified a total of 50 incident disease outcomes associated with mCAs at PheWAS significance levels. We observed striking differences in the diseases associated with each type of alteration, with autosomal mCAs most associated with increased hematologic malignancies, incident infections and possibly cancer therapy-related conditions. Alterations of chromosome X were associated with increased lymphoid leukemia risk and, mCAs of chromosome Y were linked to potential reduced metabolic disease risk.

**Conclusions:** Our findings demonstrate that a wide range of diseases are potential sequelae of mCAs and highlight the critical importance of careful covariate adjustment in mCA disease association studies.

Keywords: Mosaic loss of Y, Mosaic chromosomal alterations, Phenome-wide association study, PheWAS, UK Biobank

## Introduction

Cells accumulate somatic mutations during normal growth and cellular division [36, 48], despite the presence of cellular mechanisms to prevent and repair genomic damage [19]. In many cases, acquired somatic mutations are not compatible with cellular survival resulting in apoptosis [9]. However, some mutations can evade repair mechanisms and are tolerated by cells [40, 48]. These

<sup>1</sup> Division of Cancer Epidemiology and Genetics, National Cancer Institute, 9609 Medical Center Drive, Rockville, MD 20892, USA surviving mutations could provide competitive advantage over normal cells lacking these somatic mutations. Clonal expansion of these cells result in genetic mosaicism, the presence of a clonal subset of cells with a mutation that differs from normal germline DNA [31, 43]. As mCAs generally cover an expansive genomic footprint (often more than 50 kilobases), these events offer opportunities to investigate future health outcomes that could arise due to a non-trivial fraction of the genome impacted by a somatic alteration [53].

Peripheral leukocytes are perhaps the most wellstudied tissue for somatic mutations as blood-derived



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

<sup>\*</sup>Correspondence: mitchell.machiela@nih.gov

Full list of author information is available at the end of the article

DNA is easy to obtain for large populations of individuals [14, 20, 23, 28, 29, 34, 42, 46]. Clonal hematopoiesis (CH) is an age-related process in which a hematopoietic stem cell which has acquired a somatic mutation, passes that mutation onto daughter cells. The chance of observing these subclones carrying genomic alterations increases with increasing age. These daughter cells eventually form a clonal subpopulation of cells with the somatic mutations offering potential cellular survival advantages. Genomic alterations can range in size from a single base pair change (e.g., somatic SNVs) [16, 21, 22], to very large structural mosaic chromosomal alterations (mCAs) [20, 23, 28, 29, 34, 37, 42, 46]. mCAs can be divided into different categories according to the location and span of these events, i.e. telomeric, centromeric, interstitial, and whole chromosome. Additionally, copy number changes can be used to classify these alterations as mosaic chromosomal gains, mosaic chromosomal losses, and mosaic chromosomal losses of heterozygosity (LOH) [34].

Mosaic loss of the Y chromosome (mLOY) is the most frequently occurring mCA in males [12, 20, 46, 50, 56]. The frequency of mLOY increases with age, with < 2.5% of men below the age of 40 having detectable mLOY and estimated frequencies rapidly increasing in excess of 40% by 70 years of age [46, 56]. A number of studies have investigated the impact of mLOY on future health outcomes and noted possible associations with risk of solid tumors [13, 25, 26, 56], Alzheimer's disease [7], cardiovascular disease [18] and other chronic health outcomes [11, 17]. However, the level of evidence for these associations varies by study and some lack adjustment for key confounders like smoking patterns. Likewise, some studies of autosomal mosaicism have reported associations with risk of cancer; particularly hematologic cancers [20, 28], diabetes [1], and infectious disease [54]. Current investigation of phenotypes associated with mosaic loss of the X chromosome (mLOX) in females is limited [33]. mLOX is not observed in males since it is critical male cells carry at least one copy of the X chromosome [55].

Herein, we perform a phenome-wide association study (PheWAS) of 1290 first occurrence of incident health conditions (both self-reported and a subset clinically verified) among 482,396 UK Biobank (UKBB) participants [44]. Our aim is to systematically scan for associations between risk of disease and presence of mCAs. Our approach has been to perform robust statistical adjustment, require stringent significance levels and carry out additional sensitivity analyses to identify a high-confidence set of incident disease associations to be prioritized in future studies of mCAs.

## Methods

## Study population

UK Biobank sample collection and processing protocols are previously described [8]. In brief, UK Biobank enrolled approximately half a million participants, aged 37 to 73, from 22 assessment centers across the UK from 2006 to 2010. Participants provided informed consent for collection of medical histories using touch-screen questionnaires, participation in nurse-led interviews, and blood sample collection at enrollment. 4.5 mL of blood samples were collected in anticoagulant tubes containing acid citrate dextrose, EDTA, or lithium-heparin as well as silica-containing tubes for fast clotting. Upon arrival at the central processing laboratory, blood was aliquoted and cryopreserved at - 80 °C or in liquid nitrogen within 24 h.

### Genotyping and detection of mosaicism

DNA extracted from the blood samples was genotyped on the Applied Biosystems UK BiLEVE or UK Biobank Axiom array. After removing individuals with sex discordance or whose DNA failed QC during genotyping, 482,396 individuals with genotype and phenotype data were included for analyses. We used previously generated data on copy number variation and corresponding cellular fraction [28, 29]. Additional steps designed for detecting mosaic Y loss events were performed as described previously [46]. In brief, log transformed intensities from UK Biobank genotyping arrays were used to yield log<sub>2</sub> R ratio and B-allele frequency for each SNP, and the Eagle2 software [27, 30] was employed to phase SNPs. After phasing, a hidden Markov model was employed to detect allelic imbalances using additional information from long-range haplotype data. Chromosome Y events were detected using B-allele frequency in two pseudo-autosomal regions (PAR) shared between the Y and X chromosomes, and further examination of  $\log_2 R$  ratio across the whole chromosome was employed to determine copy number changes [46].

### Phenotypes

First occurrence of 1764 ICD-10-coded diseases (UK Biobank category 1712) were derived from linkage to primary care, hospital admission, cancer registry, death register data as well as self-report disease history obtained at enrollment. We further combined these data with the most up-to-date inpatient data (UK Biobank fields 41270 and 41280) as well as cancer registry data (UK Biobank fields 40005 and 40006). For the main PheWAS [2, 6] analysis, we first investigated incident cases defined by diagnoses occurring after enrollment; for simplicity in performing the PheWAS, individuals with diseases diagnosed before enrollment were coded as controls. To ensure that this control coding scheme did not bias reported associations, sensitivity analyses by cancer status and additional adjustment for ancestry as well as Mendelian randomization were performed. Further exploratory analyses focused on prevalent disease and medication codes to more comprehensively examine associations with mCAs.

### Medications

The UK Biobank has 6745 unique medication codes (UK Biobank category 20003) derived from the Read Codes, Clinical Terms Version 3 (CTV3). As prior publications curating UK Biobank medications were incomplete [51] or based on text interpreters (Categorising UK Biobank Self-Reported Medication Data using Text Matching [3]), we traced the origins of each UK Biobank medication code back to the original CTV3 Read Codes to assign standardized active ingredients to each code. All UK Biobank medication codes were converted with high fidelity (100%) to 6197 unique CTV3 Read Codes along with hierarchy-path information to 1073 single ingredients and assigned Unified Medical Language System (UMLS) RxNorm codes using the "RxNormR" package in R. RxNorm codes provided connections to other medication classification systems including ATC (WHO), DrugBank (University of Alberta and the Metabolomics Innovation Centre) and MeSH (NLM thesaurus). In some cases, manual edits were required to provide missing ATC codes and exclude codes that did not apply based on CTV3 path information. Finally, PheWAS analysis of mCAs was performed on the reported medications using each of the five levels of ATC codes: chemical substance, chemical subgroup, pharmacological subgroup, therapeutic subgroup, and anatomical main group to identify certain components of medications that may be associated with mCAs.

### Statistical analyses

All statistical analyses were performed in R version 4.0.3 [41]. PheWAS was performed using the "PheWAS" package [2]. Variables adjusted in the PheWAS models include age, age<sup>2</sup>, sex (for analyses other than loss of X or Y chromosomes), and a detailed 25-level smoking variable. The detailed 25-level smoking variable was created as previously described [25] and includes information on smoking intensity, duration and tobacco type. Genetic ancestry proportions were calculated using SNPweights [4], which utilizes SNP weights computed from large reference panels to infer genetic ancestry. The percentage of European, African, and Asian ancestry were computed for each participant, and these percentages were used in adjusted analyses. Phenome-wide significance levels were calculated by Bonferroni correction (0.05/number of diseases tested), in which the number of diseases tested vary by the category of genomic alteration investigated. We set a minimum number of cases for inclusion in analyses of 20 to perform reliable asymptotic association tests. In total, autosomal mCA models with participants of both sexes had 1290 incident diseases with sufficient case numbers, mLOY models with males had 1140 incident diseases, and mLOX models with females included 1128 incident diseases. Manhattan plots for the PheWAS were created with the "ggplot2" package [49].

We utilized the "GLIDE" package to test potential pleiotropy [5] and applied the Steiger filter to remove mLOYrelated SNPs which could be subject to reverse causation from either body mass index or high blood sugar. With a false discovery rate threshold of 0.05, 127 out of 156 previously published mLOY-related SNPs [46] were chosen as the mLOY instrument. Mendelian randomization analyses were conducted with the "MendelianRandomization" package [52].

### Results

### Detectable mCAs in the UK Biobank population

A total of 482,396 individuals were examined for mCAs, with a total of 17,113 (3.5%) having at least one detectable autosomal event and a total of 19,632 mosaic chromosomal alterations detected on the autosomes. Of the detected autosomal events, 8185 (41.7%) were copy number neutral, 3718 (18.9%) were losses, 2389 (12.2%) were gains, and 5341 (27.2%) had undetermined copy number state due to challenges with assigning copy number status to events with a low cellular fraction of affected cells (Additional file 1: Fig. S1). In addition to mosaic events of the autosomes, we also identified 43,297 males (19.6%) with mosaic chromosome Y loss (mLOY) and 12,550 females (4.8%) with chromosome X loss (mLOX). There were no cases of mLOX detected in males.

## Observed associations between mosaic chromosomal alterations and incident disease

We first performed a PheWAS for incident diseases that were captured in the UKBiobank resource (Table 1, Fig. 1a). We observed the strongest association between autosomal mCAs and risk of C91 lymphoid leukemia (OR 23.74,  $p < 5 \times 10^{-324}$ ). In addition to lymphoid leukemia, we also observed a strong association with C83 diffuse non-Hodgkin's lymphoma (OR 4.62,  $p=9.64 \times 10^{-66}$ ). However, lymphoid lineage malignancies were not the only hematologic malignancy associated with autosomal mCAs; diseases of the myeloid lineage were also found to be associated, including D45 polycythaemia vera (OR 12.26,  $p=3.37 \times 10^{-61}$ ), D46 myelodysplastic syndromes (OR 6.19,  $p=1.65 \times 10^{-39}$ ), and C92 myeloid leukemia (both acute C92.0 and chronic C92.1 myeloid

## Table 1 Statistically significant associations for mCAs and incident disease associations.

mCA	Disease	Ν	Percent with preceding cancer	Odds ratio [95% Confidence Interval]	P-value
Autosome	C91 Lymphoid leukaemia	616	35.47	23.74 [20.21–27.88]	<5 × 10 <sup>-324</sup>
Autosome	D47 Other neoplasms of uncertain or unknown behaviour of lymphoid, haemat- opoietic and related tissue	729	37.5	5.29 [4.38–6.39]	1.97 × 10 <sup>-67</sup>
Autosome	C83 Diffuse non-Hodgkin's lymphoma	903	50.67	4.62 [3.88–5.5]	9.64 × 10 <sup>-66</sup>
Autosome	D45 Polycythaemia vera	201	21.47	12.26 [9.1–16.51]	$3.37 \times 10^{-61}$
Autosome	C85 Other and unspecified types of non-Hodgkin's lymphoma	859	59.02	4.28 [3.56–5.15]	$1.27 \times 10^{-53}$
Autosome	D70 Agranulocytosis	4637	72.65	2.24 [2.02–2.49]	9.73 × 10 <sup>-51</sup>
Autosome	D46 Myelodysplastic syndromes	302	47.86	6.19 [4.72-8.13]	$1.65 \times 10^{-39}$
Autosome	C92 Myeloid leukaemia	381	47.22	5.19 [4-6.73]	1.66 x 10 <sup>-35</sup>
Autosome	R16 Hepatomegaly and splenomegaly, not elsewhere classified	611	36.33	3.78 [2.99–4.77]	9.44 × 10 <sup>-29</sup>
Autosome	D69 Purpura and other haemorrhagic conditions	3090	38.04	2.01 [1.76–2.29]	$2.02 \times 10^{-25}$
Autosome	A41 Other septicaemia	10,144	52.41	1.54 [1.42–1.67]	$2.43 \times 10^{-25}$
Autosome	D80 Immunodeficiency with predominantly antibody defects	248	42.63	5.44 [3.91–7.56]	$7.27 \times 10^{-24}$
Autosome	D75 Other diseases of blood and blood-forming organs	1059	25.52	2.77 [2.27–3.38]	$9.33 \times 10^{-24}$
Autosome	C95 Leukaemia of unspecified cell type	50	76.92	13.37 [7.43–24.06]	$4.92 \times 10^{-18}$
Autosome	D72 Other disorders of white blood cells	794	27.25	2.66 [2.1–3.36]	$2.97 \times 10^{-16}$
Autosome	J18 Pneumonia, organism unspecified	17,638	29.62	1.32 [1.24–1.41]	3.19 × 10 <sup>-16</sup>
Autosome	C94 Other leukaemias of specified cell type	32	70.59	14.93 [7.2–30.96]	3.73 × 10 <sup>-13</sup>
Autosome	D61 Other aplastic anaemias	879	46.57	2.24 [1.78–2.82]	$6.94 \times 10^{-12}$
Autosome	Z51 Other medical care	23,428	63.39	1.24 [1.16–1.32]	1.38 × 10 <sup>-11</sup>
Autosome	C84 Peripheral and cutaneous T-cell lymphomas	154	41.08	4.37 [2.83–6.72]	2.27 × 10 <sup>-11</sup>
Autosome	C93 Monocytic leukaemia	51	80.77	6.61 [3.5–12.47]	5.89 × 10 <sup>-9</sup>
Autosome	J90 Pleural effusion, not elsewhere classified	10,097	42.9	1.29 [1.18–1.41]	$1.06 \times 10^{-8}$
Autosome	C88 Malignant immunoproliferative diseases	121	69.7	4.17 [2.55–6.83]	$1.43 \times 10^{-8}$
Autosome	C82 Follicular [nodular] non-Hodgkin's lymphoma	347	37.96	2.71 [1.91–3.83]	1.94 × 10 <sup>-8</sup>
Autosome	Z85 Personal history of malignant neoplasm	20,384	88.65	1.21 [1.13–1.29]	1.98 × 10 <sup>-8</sup>
Autosome	J98 Other respiratory disorders	12,465	29.57	1.26 [1.16–1.37]	$4.06 \times 10^{-8}$
Autosome	Y43 Primarily systemic agents	2494	93.78	1.58 [1.34–1.86]	5.24 × 10 <sup>-8</sup>
Autosome	B25 Cytomegaloviral disease	171	51.13	3.63 [2.27–5.81]	$7.78 \times 10^{-8}$
Autosome	N17 Acute renal failure	14,465	42.7	1.22 [1.13–1.32]	1.61 × 10 <sup>-7</sup>
Autosome	E87 Other disorders of fluid, electrolyte and acid–base balance	15,028	38.77	1.22 [1.13–1.31]	$2.40 \times 10^{-7}$
Autosome	D64 Other anaemias	16,970	30.46	1.2 [1.11–1.29]	$9.06 \times 10^{-7}$
Autosome	Z94 Transplanted organ and tissue status	724	44.28	1.99 [1.5–2.65]	1.94 × 10 <sup>-6</sup>
Autosome	D59 Acquired haemolytic anaemia	177	38.52	2.96 [1.85–4.73]	5.70 × 10 <sup>-6</sup>
Autosome	R50 Fever of unknown origin	3504	48.62	1.4 [1.21–1.63]	6.79 × 10 <sup>-6</sup>
Autosome	B96 Other bacterial agents as the cause of diseases classified to other chapters	9917	33.45	1.23 [1.12–1.35]	8.21 × 10 <sup>-6</sup>
Autosome	R60 Oedema, not elsewhere classified	2075	42.78	1.49 [1.25–1.78]	1.11 × 10 <sup>-5</sup>
Autosome	E83 Disorders of mineral metabolism	5293	43.15	1.3 [1.15–1.47]	2.23 × 10 <sup>-5</sup>
Autosome	R22 Localised swelling, mass and lump of skin and subcutaneous tissue	1175	28.16	1.66 [1.31–2.11]	2.82 × 10 <sup>-5</sup>
Autosome	G72 Other myopathies	258	28.99	2.45 [1.6–3.74]	3.29 × 10 <sup>-5</sup>
Autosome	T86 Failure and rejection of transplanted organs and tissues	232	49.76	2.6 [1.65–4.08]	3.39 × 10 <sup>-5</sup>
mLOX	C91 Lymphoid leukaemia	236	35.47	2.45 [1.7–3.53]	1.40 × 10 <sup>-6</sup>
mLOX	J03 Acute tonsillitis	1167	4.97	1.78 [1.39–2.27]	4.85 × 10 <sup>-6</sup>
mLOY	E11 Non-insulin-dependent diabetes mellitus	14,110	21.37	0.8 [0.77–0.84]	3.67 × 10 <sup>-22</sup>
mLOY	110 Essential (primary) hypertension	46,694	20.37	0.91 [0.89–0.94]	2.91 × 10 <sup>-11</sup>
mLOY	H36 Retinal disorders in diseases classified elsewhere	1359	20.69	0.65 [0.56–0.75]	1.46 × 10 <sup>-9</sup>
mLOY	E66 Obesity	12,647	22.40	0.87 [0.83–0.91]	1.12 × 10 <sup>-8</sup>
mLOY	E14 Unspecified diabetes mellitus	1642	11.07	0.7 [0.61–0.8]	9.34 × 10 <sup>-8</sup>
mLOY	MI0 Gout	5601	14.55	0.8/[0.81-0.93]	3./4 × 10 <sup>-5</sup>
mlOY mlOY	E 16 Other disorders of pancreatic internal secretion G63 Polyneuropathy in diseases classified elsewhere	1641 646	30.03 29.78	0.78 [0.69–0.88] 0.65 [0.53–0.8]	4.28 × 10 <sup>-5</sup> 4.29 × 10 <sup>-5</sup>

### Table 1 (continued)

Only PheWAS significant associations are reported ( $P < 3.88 \times 10^{-5}$  for autosomal,  $P < 4.43 \times 10^{-5}$  for mLOX, and  $P < 4.39 \times 10^{-5}$  for mLOY). All PheWAS associations are adjusted for age, age<sup>2</sup>, smoking, and sex (only for autosomal mCA analyses). We define percent with preceding cancer as the percentage of individuals who have been diagnosed with any cancer prior to the diagnosis of that disease



leukemia; OR 5.19,  $p = 1.66 \times 10^{-35}$ ). This suggests that the association between mCAs and myeloid and lymphoid lineage diseases points towards an early progenitor cell, leading to abnormal growth of distinct sets of blood cells. Additionally, we detected associations which could be partially attributed to cancer progression as well as treatment evidenced by simultaneous or prior diagnoses of neoplasm. Diagnoses included D70 agranulocytosis (OR 2.24,  $p = 9.73 \times 10^{-51}$ ), A41 other septicaemia (OR 1.54,  $p = 2.43 \times 10^{-25}$ ), D80 immunodeficiency with predominantly antibody defects (OR 5.44,  $p = 7.27 \times 10^{-24}$ ), D61 other aplastic anemia (OR 2.24,  $p = 6.94 \times 10^{-12}$ ), J18 pneumonia of unspecified organism (OR 1.32,  $3.19 \times 10^{-16}$ ), J90 pleural effusion (OR 1.29,  $p = 1.06 \times 10^{-8}$ ), B25 cytomegaloviral disease (OR 3.63,  $p = 7.78 \times 10^{-8}$ ), and T86 failure and rejection of transplanted organs and tissues (OR 2.60,  $p = 3.39 \times 10^{-5}$ ). When stratifying by cancer diagnoses prior to these



	Observation	MR IVW		
Trait	Odds ratio [95% confidence interval]	P-value	Odds ratio [95% confidence interval]	P-value
Retinal diseases	0.67 [0.58–0.77]	$6.06 \times 10^{-8}$	0.77 [0.68–0.87]	4.49 × 10 <sup>-5</sup>
Hypertension	0.92 [0.89-0.96]	$5.11 \times 10^{-10}$	0.99 [0.97–1.02]	0.175

**Table 2** Comparison of effect estimates between observational associations and Mendelian randomization (MR) estimates from inverse variance weighted (IVW) method.

Observational associations adjust for age, age<sup>2</sup>, and 25-level smoking. MR estimates use selected variants previously identified to be associated with mLOY susceptibility (Thompson et al. [46])

diseases, most effect estimates were higher in magnitude in people with a prior cancer diagnosis (Additional file 1: Table S1); this is despite much larger variation in these strata due to smaller sample sizes. Sensitivity analyses with additional adjustment of genetic ancestry resulted in the same statistically significant findings in autosomal mCAs, except for C93 monocytic leukemia (Additional file 1: Table S2). In this case, the model of monocytic leukemia failed to converge suggesting uneven ancestral distribution of monocytic leukemia among UK Biobank participants. PheWAS on each autosome found associations with a very similar set of diseases and demonstrated that blood organ malignancies were often associated with mCAs on multiple autosomes (Additional file 1: Tables S3, S4).

For mLOX events in women, we found an increased risk of C91 lymphoid leukemia (OR 2.45,  $p = 1.40 \times 10^{-6}$ ) as well as an unrelated outcome, J03 acute tonsillitis (OR 1.78,  $p = 4.85 \times 10^{-6}$ ) (Table 1, Fig. 1b). mLOY had an entirely different spectrum of disease associations from mLOX and autosomal mCAs. Not only was there an absence of an association with blood organ and tissue-related diseases, we observed protective associations between mLOY and a series of metabolic conditions: E11 type II diabetes (OR 0.80,  $p = 3.67 \times 10^{-22}$ ), M10 gout (OR 0.87,  $p = 3.74 \times 10^{-5}$ ), E66 obesity (OR 0.87,  $p = 1.46 \times 10^{-9}$ ), and I10 primary hypertension (OR 0.91,  $p = 2.91 \times 10^{-11}$ ) (Table 1, Fig. 1c).

## Examining the potential protective effects of mLOY with incident metabolic diseases

To investigate connections with a range of metabolic diseases, we performed additional analyses to better examine the impact of established common underlying confounders (e.g., prior cancer history or BMI). A sensitivity analysis in a subset excluding individuals with prior cancer history, did not find evidence that prior history of cancer is a confounder in our study (Additional file 1: Table S5). However, adjustment for BMI completely attenuated most of the negative associations between mLOY and metabolic diseases. The retinal disorders (OR 0.65,  $p=6.36 \times 10^{-8}$ ) and primary hypertension (OR 0.94,  $p=1.35 \times 10^{-6}$ ) remained negatively associated after BMI adjustment, albeit with attenuated effect sizes (Additional file 1: Table S6). We examined potential underlying causes of retinal diseases by investigating other comorbidities in the group of patients with incident retinal disease (n=3752). Among individuals with retinal disorders, 3408 (90.8%) were diagnosed with non-insulin dependent diabetes, 3103 (82.7%) had primary hypertension, and 3073 (81.9%) had unspecified diabetes. In total, 98.8% of all patients suffering retinal disorders had at least one of the three aforementioned diseases suggesting the association of mLOY with retinal disease may be a result of these underlying conditions that predispose to retinal disease.

To further examine the observed protective association between Y loss and the two traits that remained significant after BMI adjustment (hypertension and retinal diseases), Mendelian randomization (MR) was performed on previously-reported germline variants associated with mLOY [46]. We used Steiger filter to remove SNPs which could be associated with mLOY through effects on hypertension or abnormal blood sugar (see Methods). Table 2 shows the effect estimates from the observational study and MR (IVW method). The Mendelian Randomization (MR) analyses for retinal diseases remained statistically significant, and the MR effect estimate was in the same direction suggesting that mLOY could be an independent protective factor for retinal disease, although further investigation is needed.

### Exploratory analyses of mosaicism and prevalent disease

We also tested for associations between the three types of detectable mosaicism (autosomal, LOX, and LOY) and prevalent disease. We observed 39 phenome-wide significant (Bonferroni corrected p <  $3.88 \times 10^{-5}$ ) disease associations with autosomal mosaic events, 62 (p <  $4.43 \times 10^{-5}$ ) disease associations with mLOX, and 149 phenome-wide significant (p <  $4.39 \times 10^{-5}$ ) disease associations with mLOY (Additional file 1: Table S7, Fig S2). Caution

should be exercised in interpretation of these prevalent disease associations due to unknown timing of both the primary diagnoses and detectible mosaicism as well as the fact that important confounders were measured at study enrollment and might not reflect the status of participants when diseases and mosaicism first occurred.

We observed several prevalent diseases that were associated with mosaic events across autosomes and both sex chromosomes. The most frequently observed disease associations were in diseases related to prevalent cardiovascular or metabolic disease. A number of ageing-related conditions, like coxarthrosis, other arthrosis (M19 other arthrosis includes M19.0 primary arthrosis, M19.1 posttraumatic arthrosis, M19.2 other secondary arthrosis, and M19.8 other specified arthrosis, and M19.9 unspecified arthrosis), osteoporosis without fracture, and spondylosis, were also associated with all forms of mosaicism. Other observed prevalent disease associations included contraceptive management, vasomotor and allergic rhinitis.

Prevalent conditions associated with both autosomal mosaicism and mLOY in men included a history of malignant neoplasm, hyperplasia or neoplasm of prostate, atherosclerosis, and inguinal hernia. In women, autosomal mCAs and mLOX were both negatively associated with prevalent excessive, frequent and irregular menstruation.

A number of conditions shared between mLOY and mLOX were not associated with autosomal mCAs. Diseases of the respiratory and digestive systems were the most frequently found to be positively associated with sex chromosome loss in both sexes. We also observed associations for sex chromosome loss with diseases in the urinary system, including chronic renal failure and unspecified hematuria.

## Observed associations between mosaicism and medication ATC codes

UK Biobank participant medications were converted to WHO Anatomic Therapeutic Chemical (ATC) codes and screened for associations for mCAs at different ATC code levels. Figure 2 and Additional file 1: Table S8 demonstrate that while we did not identify statistically significant associations between ATC level 3 medications and autosomal mCAs ( $p < 3.27 \times 10^{-4}$ ) or mLOX ( $p < 3.27 \times 10^{-4}$ ), mLOY ( $p < 3.27 \times 10^{-4}$ ) were shown to be negatively associated with blood glucose lowering drugs (OR 0.73,  $p = 4.24 \times 10^{-25}$ ), insulins and analogues (OR 0.78,  $p = 9.18 \times 10^{-6}$ ), as well as calcium channel blockers (OR 0.89,  $p = 2.42 \times 10^{-8}$ ), beta blockers (OR 0.91,  $p = 4.29 \times 10^{-6}$ ), and antigout preparations (OR 0.68,  $p = 1.25 \times 10^{-18}$ ). These results are consistent with the protective associations with incident diseases.

### Discussion

In this study, we examined plausible associations between detectible mCAs and first occurrence of a range of diseases in a large, well-characterized population of approximately 500,000 individuals. We utilized existing high-confidence mosaic alteration calls [28, 29, 46] and created a comprehensive aggregation of first occurrence of diseases by integrating primary care, inpatient data, death registry, cancer registry as well as self-reported disease history obtained through interview at study enrollment. Our analyses identified mCA-disease associations, many of which were associated with specific types of mCA, suggesting distinct mechanisms or perturbations of key pathways in their pathogenesis. Examination of potential confounders using sensitivity analysis and adjustment demonstrated the critical importance of careful covariate adjustment in CH-disease association studies to avoid reporting spurious associations.

We observed robust evidence for known associations between autosomal mCAs and hematologic cancer risk. We observed the strongest associations with lymphoid malignancies, and also noted substantial evidence for myeloid malignancies. This suggests that autosomal mCAs predispose to cancer risk in both the lymphoid and myeloid compartments. Consistent with prior reports, we observed evidence for potential associations between autosomal mCAs and various blood organrelated diseases as well as infections possibly linked to cancer treatment [54]. These findings indicate that clonal expansion of aberrant mCA clones can have an overall effect on select blood components that manifest in distinct detectable and diagnosable diseases. Such clonal expansion of autosomal mCAs could promote blood tissue disease risk by reducing stem cell diversity, impacting blood cell differentiation and altering leukocyte function due to copy-number or loss of heterozygosity (LOH) induced changes in gene regulation and expression. Similar to the autosomes, mLOX was associated with increased risk of lymphoid leukemia suggesting mLOX events may also predispose to hematologic cancer risk. Likewise, we observed an association between mLOX and acute tonsillitis indicating mLOX could play a role in infection or immune response as well.

The spectrum of incident diseases associated with mLOY differed substantially from autosomal mCAs and mLOX. As mLOY is the most commonly observed mCA in men, the UK Biobank male population had increased power for detecting incident disease associations with mLOY. Despite this improved power for detecting associations, we observed a paucity of evidence for incident disease associations with mLOY. It is notable that previously published mLOY associations were not confirmed

Fig. 2 Manhattan plots of ATC medication codes (level 3) and mCAs in the UK Biobank. All associations are adjusted for age, age<sup>2</sup>, detailed 25-level smoking, and sex (only for autosomal mCA analyses). Colors are used to separate medications in different ATC level 1 categories. The red line represents the Bonferroni-corrected, PheWAS significance level (0.05/number of medication codes tested). The blue dashed line represents the minimum, suggestive threshold (1/ number of medication codes tested). A maximum of the ten most significant medication codes above the suggestive threshold are labeled. PheWAS associations are reported for: a Autosomal mCAs, 169 medication codes tested (PheWAS significance:  $p < 2.96 \times 10^{-4}$ suggestive threshold:  $5.92 \times 10^{-3}$ ); L01X Other antineoplastic agents. b mLOX, 164 medication codes tested (PheWAS significance level  $p < 3.05 \times 10^{-4}$ , suggestive threshold: 6.10  $\times 10^{-3}$ ); D03B Enzymes for treatment of wounds and ulcers and M09A Other drugs for disorders of the musculo-skeletal system. c mLOY, 153 medication codes tested (PheWAS significance level  $p < 3.27 \times 10^{-4}$ , suggestive threshold:  $6.54 \times 10^{-3}$ ); A10A Insulins and analogues, A10B Blood glucose lowering drugs, excluding insulin, A11C Vitamin A and D, including combinations of the two, C02C Antiandrenergic agents, peripherally acting, C07A Beta blocking agents, C08C Selective calcium channel blockers with mainly vascular effects, C10A Lipid modifying agents, plain, M04A Antigout preparations, R01A Decongestants and other nasal preparations for topical use, and S01X Other opththalmologicals

after Bonferroni correction ( $p < 4.39 \times 10^{-5}$ ), such as solid tumor risk [13, 26], Alzheimer's disease [7], major cardiovascular events [18], age-related macular degeneration [17], autoimmune thyroiditis [39], primary biliary cirrhosis [24], testicular germ cell tumor [32], and abdominal aortic aneurysm [45]. While these findings could indicate limited evidence for associations with mLOY and incidence of these diseases in the UK Biobank male population, we noted the multiple testing correction threshold for significance was stringent and that the generally healthy participant pool in the UK Biobank male participants [15] may not have been ideal to investigate associations with these disease outcomes.

To further compare our mLOY findings with those previously reported, we restricted our analyses to male participants over the age of 65 years old but did not observe evidence for associations of mLOY with Alzheimer's disease, major cardiovascular events or age-related macular degeneration (Additional file 1: Table S9). Our detection method also included phase-based data to increase sensitivity to detect lower frequency mLOY clones, which may have reduced the effect size of associations with disease risk. To further explore the impact of clonal frequency on disease risk, we performed sensitivity analyses using different cell fraction thresholds (0.03, 0.1, and 0.2), but did not observe evidence for mLOY associations with incidence of closely matching disease codes of the previously reported diseases above passing phenome-wide significance level (Additional file 1: Table S10). Finally, our modeling approach investigated incident disease risk in



which some previously reported mLOY associations were for prevalent disease and adjusted for an expanded list of confounders in an attempt to as comprehensively as possible rule out potential biases. For example, we derived and adjusted for more granular smoking covariates (25 levels, Additional file 1: Table S11) as well as modeled age with both age and age<sup>2</sup> to account for potential non-linear relationships with age (Additional file 1: Tables S12).

Interestingly, we observed negative associations between mLOY and several metabolic-related diseases. We performed a series of exploratory analyses to investigate these protective associations further and found that adjustment for BMI substantially attenuated many of these associations; however, risk for primary hypertension and retinal diseases associated with metabolic syndrome remained statistically significant. MR analyses using previously reported mLOY variants discovered by GWAS [46] added additional supporting evidence to confirm the associations with primary hypertension and retinal diseases. While it is reassuring that the association between retinal diseases and mLOY were confirmed in an MR analysis, we note that the mLOY discovery GWAS and this PheWAS contain sample overlap which could result in bias of the MR results toward the observed PheWAS result. Biologic mechanisms linking mLOY with hypertension and retinal diseases are unclear and future studies in independent populations are needed to confirm these observed relationships.

In our PheWAS, we compared three models with respect to smoking adjustment: (1) no smoking adjustment, (2) adjusting for 3 smoking categories (never, former, and current), and (3) with a 25-level detailed smoking adjustment. The results in Additional file 1: Table S11 demonstrates that some observed associations between mLOY and diseases using models that did not carefully adjust for smoking behavior are due to residual confounding, for example some behavioral outcomes were completely confounded by smoking. Similarly, we also ran models further examining the impact of age adjustment. We ran the following models: (1) unadjusted for age, (2) only adjusting for linear age, and (3) adjusting for both a linear and squared age term. Results with no adjustment for age demonstrated a host of spurious associations due to confounding by age (Additional file 1: Table S12). While linear age adjustment removed several of these spurious findings, a few associations such as mental and behavioral disorders due to opioids, arthrosis, and polyuria remained nominally significant indicating evidence of residual confounding, perhaps due to age. Our results suggest that including both a linear and squared term for age adjustment resulted in robust statistical adjustment for potential confounding due to age.

We conducted exploratory analyses that included prevalent disease. While we identified several potential associations between prevalent disease and mCAs, the incident disease associations did not always support these findings. In addition, as the timing of prevalent disease diagnosis is unknown relative to the onset of mosaicism, it is not possible to time the relationship between disease and detectible mCA in the UK Biobank (i.e., the order could not be conclusively defined for diseases). We included preliminary results from a PheWAS of prevalent conditions as exploratory analyses for hypothesis generation and initial evidence for designing future investigations. We highlighted similarities across autosomal and sex chromosome mosaicism, but recommend caution be taken in the interpretation of these findings and stress the critical need for further replication in independent datasets.

Likewise, we performed PheWAS on medication use, but it is difficult to separate the relationship between medications and diagnoses in these results. For example, someone with a diabetes diagnosis is likely to be taking a diabetes medication such as metformin. While the mechanism of metformin is still being studied, it is possible that metformin-induced inhibition of gluconeogenesis and the mTOR pathway [35, 38] could have impacts on mCA clonal expansion [10, 47].

There are limitations of our study that should be taken into account when interpreting the associations reported herein between mCAs and diseases. First, although the date of diagnosis provides a certain level of temporality between mosaic events and diseases, we are unaware of the time period when initial symptoms and onset of diseases occurred. The onset of disease might predate the onset of mosaicism even though the diagnoses occurred much later. Second, we chose the first occurrence of each disease as a defining marker of onset. For diseases or treatments that are ongoing or chronically reoccur, factoring in multiple episodes of that particular disease might further increase the power to detect associations. Third, there may be some inaccuracies or underreporting in the first occurrence data in the UK Biobank due to (1) the primary care data is still an interim release with  $\sim$  45% of the participants available, (2) the completeness of inpatient data varies by geographical location with England dating back to 1997, Wales to 1998, and Scotland to 1981, and (3) self-report information from participants are susceptible to inaccuracies and biases. Finally, conditions which do not result in primary care or inpatient admissions are inevitably missing and not captured in the analyses.

### Conclusion

In the current investigation, we report evidence for a broad spectrum of associations between mCAs and first occurrence of diseases that varies by type of mCA and highlight the critical importance of careful covariate adjustment to minimize confounding. Our findings suggest mCAs could be linked to a spectrum of health outcomes and as such future more focused studies are needed to identify important etiologic relationships with potential clinical utility for assessment of disease risk.

## **Supplementary Information**

The online version contains supplementary material available at https://doi. org/10.1186/s13578-021-00651-z.

Additional file 1. Additional figures and tables.

#### Acknowledgements

We thank Dr. Gretchen Gierach for her critical input on the manuscript.

### Authors' contributions

MJM. conceived and designed the study. MJM, SHL, DWB, and SMK contributed to the acquisition and management of data. BR and SHL analyzed the data. SHL, MJM, and BR drafted the first version of the manuscript. SHL, DWB, BR, FD, OWL, SMK, JH, SJC, JRBP, MJM contributed to the interpretation of analyses, critically revised the manuscript and approved the final text. All authors read and approved the final manuscript.

#### Funding

Open Access funding provided by the National Institutes of Health (NIH). This work was supported by the Intramural Research Program of the National Cancer Institute (NCI). This research was conducted using the UK Biobank resource (application 21552). The UK Biobank was established by the Wellcome Trust, the Medical Research Council, the United Kingdom Department of Health, and the Scottish Government. The UK Biobank has also received funding from the Welsh Assembly Government, the British Heart Foundation, and Diabetes UK.

### Availability of data and materials

The data reported in this paper are available by application directly to the UK Biobank. Statistically significant associations between mosaicism and health outcomes as well as medication are provided in the manuscript. Software code in R for the analyses is available upon request.

### Declarations

### Ethics approval and consent to participate

The UK Biobank received ethical approval from the research ethics committee (REC reference for UK Biobank 21552) and all participants provided signed informed consent at enrollment and all research was performed in accordance with relevant guidelines/regulations. All data used in this analysis is available through application to the UK Biobank.

### **Consent for publication**

Not applicable.

### **Competing interests**

The authors declare no competing interests.

### Author details

<sup>1</sup> Division of Cancer Epidemiology and Genetics, National Cancer Institute, 9609 Medical Center Drive, Rockville, MD 20892, USA. <sup>2</sup>University of Miami Miller School of Medicine, Miami, FL 33136, USA. <sup>3</sup>MRC Epidemiology Unit, University of Cambridge School of Clinical Medicine, Cambridge, UK.

Received: 23 April 2021 Accepted: 6 July 2021 Published online: 23 July 2021

#### References

- Bonnefond A, Skrobek B, Lobbens S, Eury E, Thuillier D, Cauchi S, Lantieri O, Balkau B, Riboli E, Marre M, Charpentier G, Yengo L, Froguel P. Association between large detectable clonal mosaicism and type 2 diabetes with vascular complications. Nat Genet. 2013;45:1040–3.
- Carroll RJ, Bastarache L, Denny JC. R PheWAS: data analysis and plotting tools for phenome-wide association studies in the R environment. Bioinformatics. 2014;30:2375–6.
- Categorising UK Biobank Self-Reported Medication Data using Text Matching (2019) Available at: https://www.researchsquare.com/article/ rs-9729/v1. Accessed Dec 30 2020.
- Chen C-Y, Pollack S, Hunter DJ, Hirschhorn JN, Kraft P, Price AL. Improved ancestry inference using weights from external reference panels. Bioinformatics. 2013;29:1399–406.
- Dai JY, Peters U, Wang X, Kocarnik J, Chang-Claude J, Slattery ML, Chan A, Lemire M, Berndt SI, Casey G, Song M, Jenkins MA, Brenner H, Thrift AP, White E, Hsu L. Diagnostics for pleiotropy in Mendelian Randomization Studies: global and individual tests for direct effects. Am J Epidemiol. 2018;187:2672–80.
- Denny JC, Ritchie MD, Basford MA, Pulley JM, Bastarache L, Brown-Gentry K, Wang D, Masys DR, Roden DM, Crawford DC. PheWAS: demonstrating the feasibility of a phenome-wide scan to discover gene-disease associations. Bioinformatics. 2010;26:1205–10.
- Dumanski JP, Lambert J-C, Rasi C, Giedraitis V, Davies H, Grenier-Boley B, Lindgren CM, Campion D, Dufouil C, European Alzheimer's Disease Initiative Investigators, Pasquier F, Amouyel P, Lannfelt L, Ingelsson M, Kilander L, Lind L, Forsberg LA. Mosaic loss of chromosome Y in blood is associated with Alzheimer disease. Am J Hum Genet. 2016;98:1208–19.
- 8. Elliott P, Peakman TC, on behalf of UK Biobank, . The UK Biobank sample handling and storage protocol for the collection, processing and archiving of human blood and urine. Int J Epidemiol. 2008;37:234–44.
- 9. Evan GI, Vousden KH. Proliferation, cell cycle and apoptosis in cancer. Nature. 2001;411:342–8.
- Fernandes H, Moura J, Carvalho E. mTOR signaling as a regulator of hematopoietic stem cell fate. Stem Cell Rev Rep . 2021. https://doi.org/10. 1007/s12015-021-10131-z.
- Forsberg LA. Loss of chromosome Y (LOY) in blood cells is associated with increased risk for disease and mortality in aging men. Hum Genet. 2017;136:657–63.
- Forsberg LA, Gisselsson D, Dumanski JP. Mosaicism in health and disease clones picking up speed. Nat Rev Genet. 2017;18:128–42.
- Forsberg LA, Rasi C, Malmqvist N, Davies H, Pasupulati S, Pakalapati G, Sandgren J, Diaz de Ståhl T, Zaghlool A, Giedraitis V, Lannfelt L, Score J, Cross NCP, Absher D, Janson ET, Lindgren CM, Morris AP, Ingelsson E, Lind L, Dumanski JP. Mosaic loss of chromosome Y in peripheral blood is associated with shorter survival and higher risk of cancer. Nat Genet. 2014;46:624–8.
- Forsberg LA, Rasi C, Razzaghian HR, Pakalapati G, Waite L, Thilbeault KS, Ronowicz A, Wineinger NE, Tiwari HK, Boomsma D, Westerman MP, Harris JR, Lyle R, Essand M, Eriksson F, Assimes TL, Iribarren C, Strachan E, O'Hanlon TP, Rider LG, et al. Age-related somatic structural changes in the nuclear genome of human blood cells. Am J Hum Genet. 2012;90:217–28.
- Fry A, Littlejohns TJ, Sudlow C, Doherty N, Adamska L, Sprosen T, Collins R, Allen NE. Comparison of sociodemographic and health-related characteristics of UK Biobank participants with those of the general population. Am J Epidemiol. 2017;186:1026–34.
- Genovese G, Kähler AK, Handsaker RE, Lindberg J, Rose SA, Bakhoum SF, Chambert K, Mick E, Neale BM, Fromer M, Purcell SM, Svantesson O, Landén M, Höglund M, Lehmann S, Gabriel SB, Moran JL, Lander ES, Sullivan PF, Sklar P, et al. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. N Engl J Med. 2014;371:2477–87.
- Grassmann F, Kiel C, den Hollander AI, Weeks DE, Lotery A, Cipriani V, Weber BHF, International Age-related Macular Degeneration Genomics Consortium (IAMDGC). Y chromosome mosaicism is associated with agerelated macular degeneration. Eur J Hum Genet. 2019;27:36–41.
- Haitjema S, Kofink D, van Setten J, van der Laan SW, Schoneveld AH, Eales J, Tomaszewski M, de Jager SCA, Pasterkamp G, Asselbergs FW, den Ruijter HM. Loss of Y chromosome in blood is associated with major cardiovascular events during follow-up in men after carotid endarterectomy. Circ Cardiovas Genet. 2017;10:e001544.

- 19. Hoeijmakers JH. Genome maintenance mechanisms for preventing cancer. Nature. 2001;411:366–74.
- Jacobs KB, Yeager M, Zhou W, Wacholder S, Wang Z, Rodriguez-Santiago B, Hutchinson A, Deng X, Liu C, Horner M-J, Cullen M, Epstein CG, Burdett L, Dean MC, Chatterjee N, Sampson J, Chung CC, Kovaks J, Gapstur SM, Stevens VL, et al. Detectable clonal mosaicism and its relationship to aging and cancer. Nat Genet. 2012;44:651–8.
- 21. Jaiswal S, Ebert BL. Clonal hematopoiesis in human aging and disease. Science. 2019;366:eaan4673.
- Jaiswal S, Fontanillas P, Flannick J, Manning A, Grauman PV, Mar BG, Lindsley RC, Mermel CH, Burtt N, Chavez A, Higgins JM, Moltchanov V, Kuo FC, Kluk MJ, Henderson B, Kinnunen L, Koistinen HA, Ladenvall C, Getz G, Correa A, et al. Age-related clonal hematopoiesis associated with adverse outcomes. N Engl J Med. 2014;371:2488–98.
- Laurie CC, Laurie CA, Rice K, Doheny KF, Zelnick LR, McHugh CP, Ling H, Hetrick KN, Pugh EW, Amos C, Wei Q, Wang L, Lee JE, Barnes KC, Hansel NN, Mathias R, Daley D, Beaty TH, Scott AF, Ruczinski I, et al. Detectable clonal mosaicism from birth to old age and its relationship to cancer. Nat Genet. 2012;44:642–50.
- Lleo A, Oertelt-Prigione S, Bianchi I, Caliari L, Finelli P, Miozzo M, Lazzari R, Floreani A, Donato F, Colombo M, Gershwin ME, Podda M, Invernizzi P. Y chromosome loss in male patients with primary biliary cirrhosis. J Autoimmun. 2013;41:87–91.
- Loftfield E, Zhou W, Graubard BJ, Yeager M, Chanock SJ, Freedman ND, Machiela MJ. Predictors of mosaic chromosome Y loss and associations with mortality in the UK Biobank. Sci Rep. 2018;8:12316.
- Loftfield E, Zhou W, Yeager M, Chanock SJ, Freedman ND, Machiela MJ. Mosaic Y loss is moderately associated with solid tumor risk. Can Res. 2019;79:461–6.
- Loh P-R, Danecek P, Palamara PF, Fuchsberger C, Reshef A, Y., K Finucane, H., Schoenherr, S., Forer, L., McCarthy, S., Abecasis, G.R., Durbin, R. & L Price, A. . Reference-based phasing using the Haplotype Reference Consortium panel. Nat Genet. 2016;48:1443–8.
- Loh P-R, Genovese G, Handsaker RE, Finucane HK, Reshef YA, Palamara PF, Birmann BM, Talkowski ME, Bakhoum SF, McCarroll SA, Price AL. Insights into clonal haematopoiesis from 8,342 mosaic chromosomal alterations. Nature. 2018;559:350–5.
- 29. Loh P-R, Genovese G, McCarroll SA. Monogenic and polygenic inheritance become instruments for clonal selection. Nature. 2020;584:1–6.
- Loh P-R, Palamara PF, Price AL. Fast and accurate long-range phasing in a UK Biobank cohort. Nat Genet. 2016;48:811–6.
- Machiela MJ, Chanock SJ. Detectable clonal mosaicism in the human genome. Semin Hematol. 2013;50:348–59.
- Machiela MJ, Dagnall CL, Pathak A, Loud JT, Chanock SJ, Greene MH, McGlynn KA, Stewart DR. Mosaic chromosome Y loss and testicular germ cell tumor risk. J Hum Genet. 2017;62:637–40.
- 33. Machiela MJ, Zhou W, Karlins E, Sampson JN, Freedman ND, Yang Q, Hicks B, Dagnall C, Hautman C, Jacobs KB, Abnet CC, Aldrich MC, Amos C, Amundadottir LT, Arslan AA, Beane-Freeman LE, Berndt SI, Black A, Blot WJ, Bock CH, et al. Female chromosome X mosaicism is age-related and preferentially affects the inactivated X chromosome. Nat Commun. 2016;7:11843.
- Machiela MJ, Zhou W, Sampson JN, Dean MC, Jacobs KB, Black A, Brinton LA, Chang I-S, Chen C, Chen C, Chen K, Cook LS, Crous Bou M, De Vivo I, Doherty J, Friedenreich CM, Gaudet MM, Haiman CA, Hankinson SE, Hartge P, et al. Characterization of large structural genetic mosaicism in human autosomes. Am J Hum Genet. 2015;96:487–97.
- Mao, Z. & Zhang, W. (2018) Role of mTOR in Glucose and Lipid Metabolism. *International Journal of Molecular Sciences*, **19**, https://www.ncbi.nlm. nih.gov/pmc/articles/PMC6073766/. Accessed Apr 1 2021.
- Martincorena I, Campbell PJ. Somatic mutation in cancer and normal cells. Science. 2015;349:1483–9.
- Notini AJ, Craig JM, White SJ. Copy number variation and mosaicism. Cytogenet Genome Res. 2008;123:270–7.
- Pernicova I, Korbonits M. Metformin—mode of action and clinical implications for diabetes and cancer. Nat Rev Endocrinol. 2014;10:143–56.
- Persani L, Bonomi M, Lleo A, Pasini S, Civardi F, Bianchi I, Campi I, Finelli P, Miozzo M, Castronovo C, Sirchia S, Gershwin ME, Invernizzi P. Increased loss of the Y chromosome in peripheral blood cells in male patients with autoimmune thyroiditis. J Autoimmun. 2012;38:J193-196.

- 40. Poduri A, Evrony GD, Cai X, Walsh CA. Somatic mutation, genomic variation, and neurological disease. Science. 2013;341:1237758.
- 41. R Core Team (2014)R: A language and environment for statistical computing. Vienna, Austria. http://www.R-project.org/.
- Rodríguez-Santiago B, Malats N, Rothman N, Armengol L, Garcia-Closas M, Kogevinas M, Villa O, Hutchinson A, Earl J, Marenne G, Jacobs K, Rico D, Tardón A, Carrato A, Thomas G, Valencia A, Silverman D, Real FX, Chanock SJ, Pérez-Jurado LA. Mosaic uniparental disomies and aneuploidies as large structural variants of the human genome. Am J Hum Genet. 2010;87:129–38.
- 43. Strachan T, Read A. Human molecular genetics. 4th ed. New York: Garland Science; 2010.
- 44. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, Downey P, Elliott P, Green J, Landray M, Liu B, Matthews P, Ong G, Pell J, Silman A, Young A, Sprosen T, Peakman T, Collins R. UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. PLoS medicine. 2015;12:e1001779.
- Tang D, Han Y, Lun Y, Jiang H, Xin S, Duan Z, Zhang J. Y chromosome loss is associated with age-related male patients with abdominal aortic aneurysms. Clin Interv Aging. 2019;14:1227–41.
- 46. Thompson DJ, Genovese G, Halvardson J, Ulirsch JC, Wright DJ, Terao C, Davidsson OB, Day FR, Sulem P, Jiang Y, Danielsson M, Davies H, Dennis J, Dunlop MG, Easton DF, Fisher VA, Zink F, Houlston RS, Ingelsson M, Kar S, et al. Genetic predisposition to mosaic Y chromosome loss in blood. Nature. 2019;575:652–7.
- Wang X, Chu Y, Wang W, Yuan W. mTORC signaling in hematopoiesis. Int J Hematol. 2016;103:510–8.
- Werner B, Case J, Williams MJ, Chkhaidze K, Temko D, Fernández-Mateos J, Cresswell GD, Nichol D, Cross W, Spiteri I, Huang W, Tomlinson IPM, Barnes CP, Graham TA, Sottoriva A. Measuring single cell divisions in human tissues from multi-region sequencing data. Nat Commun. 2020;11:1035.
- Wickham H. ggplot2: elegant graphics for data analysis. Cham: Springer; 2016.
- Wright DJ, Day FR, Kerrison ND, Zink F, Cardona A, Sulem P, Thompson DJ, Sigurjonsdottir S, Gudbjartsson DF, Helgason A, Chapman JR, Jackson SP, Langenberg C, Wareham NJ, Scott RA, Thorsteindottir U, Ong KK, Stefansson K, Perry JRB. Genetic variants associated with mosaic Y chromosome loss highlight cell cycle genes and overlap with cancer susceptibility. Nat Genet. 2017;49:674–9.
- Wu Y, Byrne EM, Zheng Z, Kemper KE, Yengo L, Mallett AJ, Yang J, Visscher PM, Wray NR. Genome-wide association study of medication-use and associated disease in the UK Biobank. Nat Commun. 2019;10:1891.
- Yavorska OO, Burgess S. MendelianRandomization: an R package for performing Mendelian randomization analyses using summarized data. Int J Epidemiol. 2017;46:1734–9.
- 53. Youssoufian H, Pyeritz RE. Mechanisms and consequences of somatic mosaicism in humans. Nat Rev Genet. 2002;3:748–58.
- Zekavat SM, Lin S-H, Bick AG, Liu A, Paruchuri K, Uddin MM, Ye Y, Yu Z, Liu X, Kamatani Y, Pirruccello JP, Pampana A, Loh P-R, Kohli P, McCarroll SA, Neale B, Engels EA, Brown DW, Smoller JW, Green R, et al. Hematopoietic mosaic chromosomal alterations and risk for infection among 767,891 individuals without blood cancer. medRxiv. 2020. https://doi.org/10.1101/ 2020.11.12.20230821.
- Zhou W, Lin S-H, Khan SM, Yeager M, Chanock SJ, Machiela MJ. Detectable chromosome X mosaicism in males is rarely tolerated in peripheral leukocytes. Sci Rep. 2021;11:1193.
- 56. Zhou W, Machiela MJ, Freedman ND, Rothman N, Malats N, Dagnall C, Caporaso N, Teras LT, Gaudet MM, Gapstur SM, Stevens VL, Jacobs KB, Sampson J, Albanes D, Weinstein S, Virtamo J, Berndt S, Hoover RN, Black A, Silverman D, et al. Mosaic loss of chromosome Y is associated with common variation near TCL1A. Nat Genet. 2016;48:563–8.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.