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Relative telomere length: a novel non-invasive biomarker for the risk of non-cirrhotic hepatocellular carcinoma in patients with chronic hepatitis B infection.

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**Relative telomere length: a novel non-invasive biomarker for the risk of
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ABSTRACT

Abnormal telomere maintenance has been significantly implicated in tumor development and progression. Telomere length has recently emerged as a promising predictor for the risk of various cancers including hepatocellular carcinoma (HCC). Nonetheless, the vast majority of these studies measured telomere length in hepatocytes and one in peripheral blood lymphocytes with conflicting results. Moreover, no studies have been reported on using circulating DNA telomere length as a non-invasive biomarker of HCC risk. In this nested case-control study, we used real-time quantitative PCR to determine the relative telomere length (RTL) in serum DNA samples from 140 hepatitis B virus (HBV)-related HCC (HBV-HCC) cases and 280 frequency-matched cancer-free HBV controls. We found that HBV-HCC cases had a significantly longer RTL (median, 0.31; range, 0.02-2.31) versus HBV controls (median, 0.20; range, 0.01-1.60) ($P=0.003$). Using the median RTL value in controls as the cutoff point, individuals with longer RTLs exhibited a significantly increased risk of HCC compared to those with short RTLs in an univariate logistic regression analysis (odds ratio [OR]=1.55, 95% confidence interval [CI] 1.02-2.33, $P=0.038$). However, the strength of this association attenuated after multivariate adjustment (OR=1.40, 95% CI 0.90-2.19, $P=0.132$). In a quartile analysis, a significant dose-response relationship was noted in the univariate analysis ($P_{\text{trend}}=0.017$) which was attenuated in the multivariate analysis ($P_{\text{trend}}=0.079$). Further analyses revealed that the significant association between serum RTL and HCC risk was only evident in non-cirrhotic (OR=3.54, 95% CI 1.58-7.93 $P=0.002$), but not in cirrhotic (OR=0.95, 95% CI 0.55-1.64, $P=0.860$) HBV patients. Moreover, the significantly increased HCC risk conferred by cirrhosis seemed to be modulated by RTL with a significant interaction effect (P for interaction=0.013). Collectively, our data suggested that RTL in circulating cell-free serum DNA could potentially be used as a novel non-invasive biomarker for non-cirrhotic HCC. Prospective

cohort studies are warranted to further validate this finding and assess its clinical significance in HCC prevention.

INTRODUCTION

In the United States, the incidence of new acute HBV infection decreased in the past two decades, but the number of patients living with chronic HBV infection has been growing significantly. This development is partly due to the trend of increasing immigration from areas with high HBV endemicity such as Asian and Sub-Arab countries [1]. Among the more than 40 million Americans born outside of the United States, about 1.5 million have chronic HBV infection. Moreover, over 60% of current HBV patients in the United States are relatively young and as these patients age, significant increases in HBV-induced hepatocellular carcinoma (HCC) and corresponding medical costs are expected to occur [1]. Thus, continuous monitoring of these patients and identification of high-risk individuals for more targeted and intensive intervention are important to the reduction of HBV-induced burden.

Telomeres consist of small tandem nucleotide repeats (TTAGGG in humans) that form the physical ends of eukaryotic chromosomes [2]. The main function of telomeres is to stabilize the linear chromosome ends and protect them from degradation and other insults resulting from DNA damage response as well as end-to-end fusion [3, 4]. Due to the end replication inefficiency of DNA polymerase, telomeres shorten by 50 to 200 base pairs within each cell division cycle in normal somatic cells [5]. When the shortening of telomeres reaches a critical point, loss of telomere protection will lead to replicative senescence and/or apoptosis in normal cells, which in turn prevents tumor initiation caused by genomic instability [6, 7]. In addition to the end replication issue, other factors such as oxidative stress, chronic inflammation, and loss of telomere binding proteins may also contribute to telomere shortening and lead to the tumorigenesis of many solid cancers including HCC [8-10]. Furthermore, several studies have demonstrated a significant correlation between decreased telomere length of tissue DNAs and promotion of hepatocarcinogenesis [11-14].

Recently, Liu et al. conducted a case-control study in a Chinese population and reported that longer telomeres in peripheral blood leukocytes (PBLs) conferred an increased risk of HCC [15]. This finding was consistent with several epidemiologic reports indicating a positive correlation between longer telomere length in blood cells and a greater susceptibility to breast cancer [16], melanoma [17], and non-Hodgkin lymphoma [18], whereas contradictory to the findings of other studies demonstrating that shorter telomeres were associated with increased risk in various cancers [19]. In HCC, mounting evidence has substantiated the correlation between short telomere length in liver tissues and increased cancer risk [20]. These paradox findings regarding telomere length in cancer predisposition suggest that the previously observed associations might be tissue and/or tumor specific, which warrants further evaluation.

Biomarkers based on circulating cell-free serum or plasma samples have unique advantages over other specimens due to their non-invasive nature, especially in prospective studies that have been followed-up for extended time periods but only collected serum or plasma samples at the time of study initiation. Circulating serum DNA as non-invasive predictive, diagnostic, and prognostic biomarkers has been extensively investigated in many cancers [21-23]. In the current study, we sought to evaluate the association between serum DNA telomere length and the risk of HBV-related HCC (HBV-HCC) using a nested case-control approach in a clinic-based cohort of Korean HBV patient population.

MATERIALS AND METHODS

Study population

The subjects in this study were selected from an existing and ongoing clinic-based cohort. The patients were consecutively enrolled from those who visited the Liver Disease Prevention Center at Thomas Jefferson University Hospital for treatment of liver

diseases, such as chronic HBV or HCV infection, fibrosis, cirrhosis, or HCC. There were no restrictions on age, sex, ethnicity, and disease etiology in patient enrollment. Enrollment started from 1988 and is still ongoing. As of October 2010, the cohort included more than 2600 patients, of which 90% were of Korean ancestry. More than 90% of the patients in this cohort had HBV infection. To minimize confounding effects from disease etiology and ethnicity, we restricted the current study to Korean HBV patients. This study has been approved by the Institutional Review Board of Thomas Jefferson University.

Epidemiologic and clinical data collection

Demographic and clinical data were obtained for each patient through medical chart review and consulting with the treating physicians. Demographic variables collected in this research included age, sex, ethnicity, smoking status, drinking status, cirrhosis, and family history of cancer. An individual who smoked more than 100 cigarettes in lifetime was defined as an ever smoker, otherwise as a never smoker. Never drinkers were defined as those who never consumed alcohol or consumed less than or equal to one drink per month. Ever drinkers were those who consumed more than one drink per month. Liver cirrhosis and HCC were determined by the combined use of clinical diagnosis and imaging studies (ultrasound, computed tomography, or magnetic resonance imaging). Blood sample was drawn for each participant for clinical laboratory test, and the remaining serum sample was stored at -80 °C for research purpose.

Measurement of relative telomere length

Circulating cell-free serum DNA was extracted from 200 μ L serum sample using QIAamp DNA Blood Mini kit (Qiagen, CA) according to the manufacturer's protocol. The relative telomere length (RTL) of each DNA sample was determined by quantitative real-time polymerase chain reaction (qRT-PCR) using a protocol described by Cawthon [24],

which measured the ratio of the copy number of telomere repeats to the copy number of a human single copy gene, *36B4*, with minor modifications. Briefly, the PCR reaction (10 μ L) for the telomere or *36B4* amplification consisted of 1 X SYBR Green Master Mix (Applied Biosystems), 200 nmol/L each telomere or *36B4* specific primers, and 2 μ L purified serum DNA sample. The thermal cycling conditions for both telomere and *36B4* were 95 °C for 10 min followed by 40 cycles of 95 °C for 15 seconds and 58 °C for 1 min with signal collection. The primer sequences were as follows: forward telomere primer (Tel-1), 5'-CGGTTGTTTGGGTTTGGGTTTGGGTTTGGGTTTGGGTT-3'; reverse telomere primer (Tel-2), 5'-GGCTTGCCTTACCCTTACCCTTACCCTTACCCTTACCCT-3'; forward human *36B4* primer, 5'-CAGCAAGTGGGAAGGTGTAATCC-3'; reverse human *36B4* primer, 5'-CCCATTCTATCATCAACGGGTACAA-3'. All samples were assayed in duplicate on a 0.1 mL fast plate using a StepOnePlus real-time PCR system (Applied Biosystems, CA). The same negative controls and calibrator DNAs were included on each plate for quality control and calibration of PCR efficiency. A reference DNA sample was used to construct a standard curve for RTL measurement on each plate. The reference DNA was extracted from 5 ml pooled serum from 25 randomly selected patients. For each standard curve, the reference DNA sample was diluted by using a 3-fold increment per dilution to produce a seven-point standard curve, between 40 and 0.05 ng DNA in each reaction. The R^2 for each standard curve was ≥ 0.98 , with acceptable standard deviation set at 0.25 (for the C_t values). If the RTL data for a tested sample was found to be out of the acceptable range of the standard curve, the sample was repeated.

Statistical analysis

All statistical analyses were performed using the SAS software version 9.2 (SAS Institute, Cary, NC). The differences in the distribution of host characteristics between

cases and controls were compared using the chi-square test for categorical variables, and the Student's *t* test for continuous variables. Wilcoxon rank-sum test was used to evaluate the differences of RTL between cases and controls. RTL was analyzed as categorical variable based on a cut-off point at the median, tertile, or quartile value of the control group. The association between RTL and HCC risk was estimated using unconditional univariate and multivariate logistic regressions to determine the unadjusted and adjusted odds ratios (ORs), respectively, and 95% confidence intervals (95% CIs). The multivariate analysis controlled for age, sex, smoking status, drinking status, cirrhosis, and family history of cancer, where appropriate. The test for interaction between demographic factors and RTL on HCC risk was conducted by including a cross-product term into the logistic regression model. All statistical tests were two sided, and $P < 0.05$ was considered as the threshold of statistical significance.

RESULTS

Demographic characteristics of the study subjects

The distributions of patient characteristics were summarized in Table 1. This study included 140 HBV-HCC cases and 280 cancer-free HBV controls that were 1:2 frequency-matched to cases on age and gender. All participants were restricted to Korean HBV patients to control the confounding effect of ethnicity and HCC etiology. As shown in Table 1, there was no significant differences between cases and controls on age (mean \pm standard deviation [SD], 55.3 ± 8.8 years vs. 53.7 ± 9.6 years, respectively, $P = 0.099$) and gender ($P = 0.327$). No significant differences were identified between cases and controls with regard to smoking status ($P = 0.096$) and drinking status ($P = 0.334$). As expectedly, a significant higher percentage of cirrhotic patients were identified in HCC cases (75.0%) versus controls (41.4%) ($P < 0.001$). Additionally, there

was a significantly higher percentage of patients with a family history of cancer among cases (47.9%) than controls (32.9%) ($P=0.003$).

The distribution of RTL in cases and controls by host characteristics

As shown in Table 2, the overall serum DNA RTL was significantly longer in HCC cases than in cancer-free HBV controls [median, 0.31 (range, 0.02-2.31) vs. median, 0.20 (range, 0.01-1.60), $P=0.003$]. We further conducted a stratified analysis to evaluate the RTL distribution differences between cases and controls by host characteristics. The differences of RTL between cases and controls remained at least borderline significant in patients regardless of age, smoking status, and drinking status. In comparison, the difference was only evident in males ($P=0.001$) but not females ($P=0.757$), in non-cirrhotic patients ($P=0.007$) but not cirrhotic patients ($P=0.423$), and in patients without a family history of cancer ($P=0.002$) but not in those with a family history of cancer ($P=0.264$). Moreover, we also compared the RTL differences between the strata of each variable in cases and controls separately, and identified a significant difference between cirrhotic and non-cirrhotic HBV controls. The median (range) of RTL was 0.28 (0.01-1.60) and 0.18 (0.01-1.11) in cirrhotic and non-cirrhotic HBV controls, respectively ($P=0.004$) (Table 2).

RTL and HCC risk

We estimated the association between serum DNA RTL and the risk on HCC by univariate and multivariate logistic regression analyses, through treating RTL as a categorical variable based on a cut off value of median, tertile or quartile distribution in cancer-free HBV controls. As shown in Table 3, individuals with longer RTL by median cut-off had a significantly increased risk of HCC in univariate analysis (unadjusted OR=1.55, 95% CI, 1.02-2.33, $P=0.038$), but not in multivariate analysis after adjusting all host variables including age, gender, smoking status, drinking status, cirrhosis, and family history of cancer (adjusted OR=1.40, 95% CI 0.90-2.19, $P=0.132$). When patients

were further categorized according to the tertile or quartile distribution of RTL values in controls, we found that, using the first (shortest RTL) group as reference, there was a significant dose-response relationship between longer RTL and increased HCC risk in the univariate analysis ($P_{\text{trend}}=0.013$ by tertile; $P_{\text{trend}}=0.017$ by quartile), which was attenuated in the multivariate analysis ($P_{\text{trend}}=0.067$ by tertile; $P_{\text{trend}}=0.079$ by quartile). The OR for the second or third group in the tertile analysis was 1.15 (95% CI 0.67-1.96, $P=0.615$) and 1.85 (95% CI 1.12-3.06, $P=0.017$), respectively in the univariate analysis, and 1.18 (95% CI 0.67-2.08, $P=0.577$) and 1.64 (95% CI 0.96-2.82, $P=0.072$), respectively in the multivariate analysis. Similar results were obtained in the quartile analysis (Table 3). Because we suspected that the attenuated association between RTL and HCC risk was due to a strong confounding from cirrhosis, we conducted another multivariate analysis adjusting all other host variables but not cirrhosis. The result of this analysis was similar to that of the univariate analysis: individuals with longer TRL by median cut-off had a significantly increased HCC risk (OR=1.59; 95% CI 1.04-2.42, $P=0.031$) and a significant dose-response effect was observed between longer RTL and increased HCC risk in the tertile and quartile analysis ($P_{\text{trend}}=0.007$ and 0.012, respectively). The results of similar multivariate analysis without adjusting other host characteristics including age, gender, smoking status, drinking status, and family history of cancer yielded similar results to that of the multivariate analysis adjusting all the variables (data not shown).

The association of RTL with HCC risk stratified by host characteristics

To further address the apparent attenuation of the significance of the observed association in the multivariate analysis, we conducted a stratified analysis to evaluate the potential modulating effect on the association between RTL and HCC risk by host variables. Consistent with the results of Tables 2 and 3, we found a significant association between long RTL and increased HCC risk that was observed only in non-

cirrhotic HBV patients (adjusted OR=3.54, 95%CI 1.58-7.93, $P=0.002$), but not in cirrhotic HBV patients (adjusted OR=0.95, 95% CI 0.55-1.64, $P=0.860$) (Table 4). A significant association was also observed in never drinkers (adjusted OR=1.93, 95%CI 1.04-3.60, $P=0.038$). In addition, borderline significant associations were observed in males ($P=0.076$), and patients without a family history of cancer ($P=0.078$) (Table 4).

Association between cirrhosis and HCC risk modulated by serum DNA RTL

Because longer RTL conferred a significantly increased risk of HCC in non-cirrhotic but not in cirrhotic patients, we sought to evaluate whether the effect of cirrhosis on HCC risk was also modulated by RTL. As shown in Table 5, as expected, cirrhosis was significantly associated with an increased HCC risk (adjusted OR=4.11, 95% CI 2.60-6.51, $P=1.48 \times 10^{-9}$). This effect was much more prominent in patients with short RTL (adjusted OR=8.04, 95% CI 3.82-16.90, $P=3.89 \times 10^{-8}$), compared to those with long RTL (adjusted OR=2.35, 95% CI 1.27-4.33, $P=0.006$). Moreover, we observed a statistically significant interaction effect between cirrhosis and RTL on the prediction of the risk of HBV-HCC (P for interaction=0.013).

DISCUSSION

In the current study, we evaluated the association between RTL in circulating cell-free serum DNA and the risk of HBV-related HCC in a clinic-based patient population. We found that longer RTL was associated with an increased risk on HBV-HCC, an effect that was much more evident in patients without cirrhosis than those with cirrhosis. Further analysis revealed a significant interaction effect between RTL and cirrhosis in the modulation of HCC risk in HBV patients.

The use of telomere length as a potential predictor for cancer risk and prognosis has been extensively investigated during the past decade. The vast majority of these studies have been focused on evaluating telomere length in tumor tissues, whereas

increasing studies were conducted recently assessing this relationship through measuring telomere length in PBLs using qRT-PCR [24, 25]. In HCC, many studies that measured telomere length in tumor and surrounding normal tissue with mixed results. The majority of these studies reported that telomere shortening was involved in hepatocarcinogenesis, whereas there were also some studies showing that longer telomeres contribute to increased HCC risks [11-13, 20, 26-28]. In addition to these discrepant findings, these studies were focused mainly on the liver tissues, which reflect more on the primary tumor characteristics than on the genetic background of the patients. Recently, Liu et al. measured RTL in PBLs in a Chinese population, and found that longer RTL was significantly associated with an increased risk of HBV-HCC, comparing to both healthy controls and cancer-free controls with chronic liver diseases such as cirrhosis or fibrosis [15]. As a surrogate specimen for individual's genetic background, PBL has been widely used in epidemiological studies. However, the use of PBL as non-invasive biomarker has been limited, although they carry considerable clinical value, especially in those prospective cohort studies with several decades' clinical follow-up that only collected serum or plasma samples at the time of study initiation. To the best of our knowledge, there has not been a population-based study determining the association of serum DNA telomere length with cancer risk or clinical outcome. In the current study, we measured the RTL in circulating cell-free serum DNA in a Korean American HBV-infected population, and observed a significant correlation between long RTL and increased risk of HBV-HCC. This finding is consistent with the results of the study of Liu et al [15]. The overall median value of RTL in our study was lower than that of Liu et al., which is possibly accounted for by the difference between circulating cell-free DNA and PBL DNA. Circulating cell-free DNA is a heterogeneous mixture including DNA released from various sources with broad variations in contents and concentrations. It consists of DNA from a wide spectrum of cells such as necrotic

and apoptotic cells, active blood cells, and circulating tumor cells in cancer patients [22], In comparison, the DNA from PBLs is highly homogeneous and to a larger extent reflects the constitutional genetic background of the subjects.

Cirrhosis is a major risk factor for HCC and between 60-90% of patients with primary HCC have underlying cirrhosis [29-31]. In our study, the increased HCC risk conferred by long serum DNA RTL was only evident in non-cirrhotic patients. This could be due to the possibility that the strong effect of cirrhosis on HCC might have overshadowed the association conferred by long telomere length in our study population. Alternatively, there may be different molecular mechanisms underlying the development of HCC with and without cirrhosis. For instance, HBV viral genome can directly integrate into the host human genome and act as an oncogenic factor, a process that is independent of the chronic inflammation that commonly characterizes cirrhosis [32]. In line with this notion, recent *in vitro* and *in vivo* studies suggested that the HBx protein, encoded by the HBV viral genome, increases both the expression of telomere reverse transcriptase and telomerase activities, the enzyme responsible for the maintenance of telomere length, thus prolonging the lifespan of hepatocytes and contributing to malignant transformation [33, 34]. Partly due to these properties of HBV genome, liver cirrhosis in patients with chronic HBV infection is only present in about 60-70% of HBV-HCC cases and not a prerequisite step for tumorigenesis [35]. These lines of evidence are consistent with our data linking telomere length and the risk of non-cirrhotic HBV-HCC. However, the molecular mechanism underlying these observations still needs further investigations.

Mixed findings have been identified from the numerous studies that have extensively investigated the relationship between altered telomere length and cancer risk. Wentzensen et al. conducted a meta-analysis and found that long telomere length was associated with the increased risk of breast cancer, lung cancer, colorectal cancer,

and Non-Hodgkin's lymphoma, but reduced risk of bladder cancer, esophageal cancer, gastric cancer, head and neck cancer, ovarian cancer, and renal cell carcinoma [19]. In HCC, our finding was consistent with the study of Liu et al., who conducted a case-control analysis in a Chinese population and reported that HCC patients had significantly longer RTL measured using DNA samples of peripheral blood leukocytes, compared to both liver disease-free normal controls and controls with chronic liver diseases such as chronic HBV infection and liver cirrhosis [15]. However, the findings are contradictory to several previous studies reporting that cancerous liver tissues had a shorter telomere length compared to paired normal tissues or non-cancerous liver tissues [14, 27, 36, 37]. Several potential reasons may account for these discrepancies. Ours and Liu's studies were conducted in Asians whereas almost all other studies used Caucasian subjects. It has been demonstrated there were significant racial differences in the distribution of telomere length [38-40]. Whether these differences have an impact on the role of telomere length in the progression of HBV-HCC remains a task of evaluation. Moreover, our study used circulating cell-free DNA from serum samples and Liu' study used PBLs, whereas the majority of other studies used hepatocytes [14, 27, 36, 37]. The RTL measurement in hepatocytes from liver tissue was more likely a reflection of the characteristics of telomere homeostasis, rather than a simple constitutive non-invasive biomarker of risk prediction measured from leukocytes or serum [15]. Consistent with this notion and the results of our study, Wiemann et al. reported that in cirrhotic patients, telomere shortening was mainly restricted to hepatocytes of cirrhotic liver tissues whereas lymphocytes and satellite cells in areas of cirrhosis had significantly longer telomere length [14]. Furthermore, telomere dysfunction has been reported to have dual roles in liver carcinogenesis. Short telomere shortening appears to have a tumor-initiating effect through inducing chromosomal instability [27, 41]. In comparison, the elongation of telomeres by the activation of telomerase has also been reported to

contribute to tumor growth and progression [20]. Various independent studies have demonstrated a significantly elevated activity of telomerase at the stage of severe liver diseases such as cirrhosis and HCC [42-44], which is consistent with the findings of our and Liu's studies that telomere length increases along with liver disease progression. Collectively, these lines of evidence indicate the requirement of a balanced state of telomere length in the normal physiological liver functions, since either extremely short or long telomeres leads to liver pathogenesis. Additional biomarker and basic studies are needed to further explain these paradoxical findings and reveal the underlying mechanisms.

The modulating effect of telomere length by environmental factors such as cigarette smoking and alcohol use have been reported in HBV-HCC [15], and some other solid tumors [45, 46]. In our study, we only observed a borderline significant association between an increased HCC risk with long serum RTL among never drinkers but not ever drinkers (Table 4). In addition, no significant interaction effect was identified between RTL and smoking or drinking status (data not shown). These results may likely be explained by the adequate matching between cases and controls on smoking and drinking status. Moreover, due to the limitation of the retrospective clinic-based study, we did not have detailed data of the intensities of smoking and drinking which could also contribute to the paradox findings. Further larger prospectively designed cohort studies are warranted to test the cumulative and interactions effects between RTL and these environmental exposures on the risk of HCC.

A major strength of this study is the unique and highly homogenous HBV patient population. Our study is restricted to Korean American HBV patients only, eliminating the confounding effects of ethnicity and disease etiology. The vast majority of patients were infected with HBV at birth or childhood, making this population an ideal resource to study the long-term outcome of HBV infection at the population level. Strict matching criteria

were implemented between cases and controls to minimize the potential confounding effects from other major risk factors. To the best of our knowledge, this is the first study to demonstrate that RTL in circulating cell-free serum DNA could potentially be used as a simple inexpensive and non-invasive biomarker of HCC risk and the findings are highly consistent with that obtained using PBLs [15]. This proof-of-concept finding makes serum a valuable resource for RTL-based biomarker research, especially in prospective longitudinal studies that have been followed-up for extended time periods but only collected serum samples at the time of study initiation.

There are also limitations in this study. First, because of the heterogeneous nature of circulating serum DNA, the mechanism underlying the significant associations observed in this study remains a task for further evaluation. Second, due to the nature of retrospective case-control design, our study also has the reverse-causation limitation that is inherent in most case-control studies, and may not differentiate the causal relationship between RTL alteration and HCC development. Prospective and longitudinal studies are needed to address this issue.

In conclusion, our study reported for the first time that longer RTL in circulating cell-free serum DNA was significantly associated with an increased risk of non-cirrhotic HBV-HCC, a finding that warrants further retrospective and prospective validations, in-depth molecular characterizations, and assessments to determine the clinical value in the risk prediction and diagnosis of HCC.

References

1. Kim, W.R., *Epidemiology of hepatitis B in the United States*. Hepatology, 2009. **49**(5 Suppl): p. S28-34.
2. Blackburn, E.H., *Structure and function of telomeres*. Nature, 1991. **350**(6319): p. 569-73.
3. Blackburn, E.H., *Telomeres*. Trends Biochem Sci, 1991. **16**(10): p. 378-81.
4. Lundblad, V., *DNA ends: maintenance of chromosome termini versus repair of double strand breaks*. Mutat Res, 2000. **451**(1-2): p. 227-40.
5. Klapper, W., R. Parwaresch, and G. Krupp, *Telomere biology in human aging and aging syndromes*. Mech Ageing Dev, 2001. **122**(7): p. 695-712.
6. Blasco, M.A., *Telomeres and human disease: ageing, cancer and beyond*. Nat Rev Genet, 2005. **6**(8): p. 611-22.
7. Hackett, J.A. and C.W. Greider, *Balancing instability: dual roles for telomerase and telomere dysfunction in tumorigenesis*. Oncogene, 2002. **21**(4): p. 619-26.
8. Houben, J.M., et al., *Telomere length assessment: biomarker of chronic oxidative stress?* Free Radic Biol Med, 2008. **44**(3): p. 235-46.
9. Gilley, D., H. Tanaka, and B.S. Herbert, *Telomere dysfunction in aging and cancer*. Int J Biochem Cell Biol, 2005. **37**(5): p. 1000-13.
10. Lechel, A., M.P. Manns, and K.L. Rudolph, *Telomeres and telomerase: new targets for the treatment of liver cirrhosis and hepatocellular carcinoma*. J Hepatol, 2004. **41**(3): p. 491-7.
11. Kitada, T., et al., *Telomere shortening in chronic liver diseases*. Biochem Biophys Res Commun, 1995. **211**(1): p. 33-9.
12. Urabe, Y., et al., *Telomere length in human liver diseases*. Liver, 1996. **16**(5): p. 293-7.
13. Miura, N., et al., *Progressive telomere shortening and telomerase reactivation during hepatocellular carcinogenesis*. Cancer Genet Cytogenet, 1997. **93**(1): p. 56-62.
14. Wiemann, S.U., et al., *Hepatocyte telomere shortening and senescence are general markers of human liver cirrhosis*. FASEB J, 2002. **16**(9): p. 935-42.
15. Liu, J., et al., *Longer leukocyte telomere length predicts increased risk of hepatitis b virus-related hepatocellular carcinoma: A case-control analysis*. Cancer, 2011.
16. Gramatges, M.M., et al., *Longer relative telomere length in blood from women with sporadic and familial breast cancer compared with healthy controls*. Cancer Epidemiol Biomarkers Prev, 2010. **19**(2): p. 605-13.
17. Han, J., et al., *A prospective study of telomere length and the risk of skin cancer*. J Invest Dermatol, 2009. **129**(2): p. 415-21.
18. Lan, Q., et al., *A prospective study of telomere length measured by monochrome multiplex quantitative PCR and risk of non-Hodgkin lymphoma*. Clin Cancer Res, 2009. **15**(23): p. 7429-33.
19. Wentzensen, I.M., et al., *The Association of Telomere Length and Cancer: a Meta-analysis*. Cancer Epidemiol Biomarkers Prev, 2011. **20**(6): p. 1238-50.
20. Satyanarayana, A., M.P. Manns, and K.L. Rudolph, *Telomeres and telomerase: a dual role in hepatocarcinogenesis*. Hepatology, 2004. **40**(2): p. 276-83.

21. Gormally, E., et al., *Circulating free DNA in plasma or serum as biomarker of carcinogenesis: practical aspects and biological significance*. *Mutat Res*, 2007. **635**(2-3): p. 105-17.
22. Fleischhacker, M. and B. Schmidt, *Circulating nucleic acids (CNAs) and cancer-- a survey*. *Biochim Biophys Acta*, 2007. **1775**(1): p. 181-232.
23. Lo, Y.M. and R.W. Chiu, *Next-generation sequencing of plasma/serum DNA: an emerging research and molecular diagnostic tool*. *Clin Chem*, 2009. **55**(4): p. 607-8.
24. Cawthon, R.M., *Telomere measurement by quantitative PCR*. *Nucleic Acids Res*, 2002. **30**(10): p. e47.
25. Svenson, U. and G. Roos, *Telomere length as a biological marker in malignancy*. *Biochim Biophys Acta*, 2009. **1792**(4): p. 317-23.
26. Oh, B.-K., et al., *High telomerase activity and long telomeres in advanced hepatocellular carcinomas with poor prognosis*. *Lab Invest*, 2007. **88**(2): p. 144-152.
27. Plentz, R.R., et al., *Hepatocellular telomere shortening correlates with chromosomal instability and the development of human hepatoma*. *Hepatology*, 2004. **40**(1): p. 80-6.
28. Lee, Y.H., et al., *Chromosomal instability, telomere shortening, and inactivation of p21(WAF1/CIP1) in dysplastic nodules of hepatitis B virus-associated multistep hepatocarcinogenesis*. *Mod Pathol*, 2009. **22**(8): p. 1121-31.
29. Llovet, J.M., A. Burroughs, and J. Bruix, *Hepatocellular carcinoma*. *Lancet*, 2003. **362**(9399): p. 1907-17.
30. Fattovich, G., et al., *Hepatocellular carcinoma in cirrhosis: Incidence and risk factors*. *Gastroenterology*, 2004. **127**(5, Supplement 1): p. S35-S50.
31. Simonetti, R.G., et al., *Hepatocellular carcinoma. A worldwide problem and the major risk factors*. *Dig Dis Sci*, 1991. **36**(7): p. 962-72.
32. Buendia, M.A., *Hepatitis B viruses and hepatocellular carcinoma*. *Adv Cancer Res*, 1992. **59**: p. 167-226.
33. Kew, M.C., *Hepatitis B virus x protein in the pathogenesis of hepatitis B virus-induced hepatocellular carcinoma*. *J Gastroenterol Hepatol*, 2011. **26 Suppl 1**: p. 144-52.
34. Zhang, X., et al., *Effects of hepatitis B virus X protein on human telomerase reverse transcriptase expression and activity in hepatoma cells*. *J Lab Clin Med*, 2005. **145**(2): p. 98-104.
35. Di Bisceglie, A.M., *Hepatitis B and hepatocellular carcinoma*. *Hepatology*, 2009. **49**(5 Suppl): p. S56-60.
36. Plentz, R.R., et al., *Telomere shortening and inactivation of cell cycle checkpoints characterize human hepatocarcinogenesis*. *Hepatology*, 2007. **45**(4): p. 968-76.
37. Hartmann, D., et al., *Telomerase gene mutations are associated with cirrhosis formation*. *Hepatology*. **53**(5): p. 1608-17.
38. Zhu, H., et al., *Leukocyte telomere length in healthy Caucasian and African-American adolescents: relationships with race, sex, adiposity, adipokines, and physical activity*. *J Pediatr*. **158**(2): p. 215-20.

39. Eisenberg, D.T., et al., *Substantial variation in qPCR measured mean blood telomere lengths in young men from eleven European countries*. Am J Hum Biol. **23**(2): p. 228-31.
40. Roux, A.V., et al., *Race/ethnicity and telomere length in the Multi-Ethnic Study of Atherosclerosis*. Aging Cell, 2009. **8**(3): p. 251-7.
41. Satyanarayana, A., et al., *Telomere shortening impairs organ regeneration by inhibiting cell cycle re-entry of a subpopulation of cells*. EMBO J, 2003. **22**(15): p. 4003-13.
42. Lee, J.E., et al., *Telomeric 3' overhangs in chronic HBV-related hepatitis and hepatocellular carcinoma*. Int J Cancer, 2008. **123**(2): p. 264-72.
43. Zhang, C., et al., *CpG island methylator phenotype association with upregulated telomerase activity in hepatocellular carcinoma*. Int J Cancer, 2008. **123**(5): p. 998-1004.
44. Saini, N., et al., *Telomerase activity, telomere length and human telomerase reverse transcriptase expression in hepatocellular carcinoma is independent of hepatitis virus status*. Liver Int, 2009. **29**(8): p. 1162-70.
45. Hou, L., et al., *Telomere length in peripheral leukocyte DNA and gastric cancer risk*. Cancer Epidemiol Biomarkers Prev, 2009. **18**(11): p. 3103-9.
46. McGrath, M., et al., *Telomere length, cigarette smoking, and bladder cancer risk in men and women*. Cancer Epidemiol Biomarkers Prev, 2007. **16**(4): p. 815-9.

Table 1. Distribution of host characteristics in all cases and controls

Variable	HCC Cases (n=140)	HBV controls (n=280)	P value
Age: Mean \pm SD	55.3 \pm 8.8	53.7 \pm 9.6	0.099
Gender			
Female	22(15.7)	55(19.6)	
Male	118(84.3)	225(80.4)	0.327
Smoking status			
Never	69(49.3)	162(57.9)	
Ever	71(50.7)	118(42.1)	0.096
Drinking status			
Never	67(47.9)	148(52.9)	
Ever	73(52.1)	132(47.1)	0.334
Cirrhosis			
No	35(25.0)	164(58.6)	
Yes	105(75.0)	116(41.4)	< 0.001
Family cancer			
No	73(52.1)	188(67.1)	
Yes	67(47.9)	92(32.9)	0.003

Table 2. RTL distribution by host characteristics in cases and controls

Variable	RTL: Median (Range)		P value
	HCC cases (n=140)	HBV controls (n=280)	
Overall	0.31(0.02-2.31)	0.20(0.01-1.60)	0.003
Age, y			
≤ 54.1	0.32(0.02-2.22)	0.22(0.02-1.15)	0.071
> 54.1	0.30(0.02-2.31)	0.18(0.01-1.60)	0.016
P value	0.998	0.414	
Gender			
Female	0.28(0.02-2.22)	0.26(0.01-1.60)	0.757
Male	0.32(0.02-2.31)	0.19(0.01-1.15)	0.001
P value	0.866	0.102	
Smoking status			
Never	0.29(0.02-2.30)	0.20(0.01-1.60)	0.045
Ever	0.32(0.02-2.31)	0.21(0.02-1.15)	0.021
P value	0.997	0.674	
Drinking status			
Never	0.28(0.02-2.21)	0.20(0.01-1.60)	0.077
Ever	0.34(0.02-2.31)	0.22(0.02-1.15)	0.011
P value	0.611	0.912	
Cirrhosis			
No	0.29(0.02-2.21)	0.18(0.01-1.11)	0.007
Yes	0.33(0.02-2.31)	0.28(0.01-1.60)	0.423
P value	0.904	0.004	
Family Cancer			
No	0.34(0.02-2.30)	0.20(0.01-1.60)	0.002
Yes	0.27(0.02-2.31)	0.20(0.01-1.35)	0.264
P value	0.120	0.947	

Table 3. HCC risk as estimated by telomere length on HBV controls								
RTL	HCC cases	HBV controls	Unadjusted		Multivariate-adjusted ¹		Multivariate-adjusted ²	
			OR (95%CI)	P Value	OR (95%CI)	P Value	OR (95%CI)	P Value
By median								
Short	55	140	1.00		1.00		1.00	
Long	85	140	1.55(1.02-2.33)	0.038	1.40(0.90-2.19)	0.132	1.59(1.04-2.42)	0.031
By tertile								
First tertile	35	93	1.00		1.00		1.00	
Second tertile	41	95	1.15(0.67-1.96)	0.615	1.18(0.67-2.08)	0.577	1.16(0.68-2.01)	0.583
Third tertile	64	92	1.85(1.12-3.06)	0.017	1.64(0.96-2.82)	0.072	1.99(1.18-3.32)	0.009
P trend				0.013		0.067		0.007
By quartile								
First quartile	26	70	1.00		1.00		1.00	
Second quartile	29	70	1.12(0.60-2.08)	0.732	1.26(0.65-2.46)	0.492	1.15(0.61-2.17)	0.665
Third quartile	35	70	1.35(0.73-2.47)	0.336	1.43(0.75-2.72)	0.283	1.37(0.74-2.54)	0.316
Fourth quartile	50	70	1.92(1.08-3.43)	0.027	1.72(0.93-3.19)	0.085	2.06(1.14-3.72)	0.017
P trend				0.017		0.079		0.012

Notes: ¹Adjusted for age, gender, smoking status, drinking status, cirrhosis and family history of cancer. ²Adjusted for age, gender, smoking status, drinking status, and family history of cancer.

Table 4. HCC risk estimated by RTL on HBV controls stratified by host characteristics

Variable	RTL (by median in controls)	HCC cases	HBV controls	OR (95%CI) ¹	P value
Age, y					
≤ 53.1	Short	27	77	1.00	0.574
	Long	35	72	1.21(0.63- 2.32)	
> 53.1	Short	31	75	1.00	0.120
	Long	47	56	1.64(0.88- 3.04)	
Gender					
Female	Short	10	27	1.00	0.939
	Long	12	28	0.96(0.30- 3.07)	
Male	Short	48	125	1.00	0.076
	Long	70	100	1.55(0.96- 2.52)	
Smoking status					
Never	Short	29	88	1.00	0.200
	Long	40	74	1.49(0.81- 2.76)	
Ever	Short	29	64	1.00	0.312
	Long	42	54	1.40(0.73- 2.70)	
Drinking status					
Never	Short	28	84	1.00	0.038
	Long	39	64	1.93(1.04- 3.60)	
Ever	Short	30	68	1.00	0.857
	Long	43	64	1.06(0.55- 2.06)	
Cirrhosis					
No	Short	12	101	1.00	0.002
	Long	23	63	3.54(1.58- 7.93)	
Yes	Short	46	51	1.00	0.860
	Long	59	65	0.95(0.55- 1.64)	
Family cancer					
No	Short	28	101	1.00	0.078
	Long	45	87	1.71(0.94- 3.09)	
Yes	Short	30	51	1.00	0.587
	Long	37	41	1.21(0.61- 2.38)	

Note: ¹Adjusted for age, gender, smoking status, drinking status, cirrhosis and family history of cancer, where appropriate.

Table 5. Association of cirrhosis with the risk of HBV-HCC modulated by RTL

cirrhosis and RTL	HCC cases	HBV controls	OR (95%CI) ¹	P value
In all patients				
Non-cirrhosis	35	164	1.00	
cirrhosis	105	116	4.11(2.60-6.51)	1.48 x 10⁻⁹
In patients with short RTL				
Non-cirrhosis	12	101	1.00	
cirrhosis	46	51	8.04 (3.82-16.90)	3.89 x 10⁻⁸
In patients with long RTL				
Non-cirrhosis	23	63	1.00	
cirrhosis	59	65	2.35(1.27-4.33)	0.006
<i>P</i> _{interaction}				0.013

Note: ¹Adjusted for age, gender, smoking status, drinking status, and family history of cancer.