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4	The Pathogenesis of Bisphosphonate Related Osteonecrosis of the Jaw: So Many		
5	Hypotheses so Few Data		
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29 Abstract

30 Bisphosphonate-related osteonecrosis of the jaw (BRONJ) has generated great interest in the 31 medical and research communities yet remains an enigma given its unknown pathogenesis. The 32 goal of this review is to summarize the various proposed hypotheses underlying BRONJ. While a 33 role of the oral mucosa has been proposed, the bone is likely the primary tissue of interest for 34 BRONJ. The most popular BRONJ hypothesis, manifestation of necrotic bone resulting from 35 bisphosphonate-induced remodeling suppression, is supported mostly by indirect evidence 36 although recent data has shown bisphosphonates significantly reduce remodeling in the jaw. 37 Remodeling suppression would be expected, and has been shown, to allow accumulation of non-38 viable osteocytes while a more direct cytotoxic effect of bisphosphonates on osteocytes has also 39 been proposed. Bisphosphonates have anti-angiogenic effects, leading to speculation that this 40 could contribute to the BRONJ pathogenesis. Compromised angiogenesis would most likely be 41 involved in post-intervention healing although other aspects of the vasculature (e.g. blood flow) 42 could contribute to BRONJ. Despite infection being present in many BRONJ patients, there is no 43 clear evidence as to whether infection is a primary or secondary event in the pathophysiology. In 44 addition to these main factors proposed in the pathogenesis, numerous co-factors associated 45 with BRONJ (e.g. diabetes, smoking, dental extraction, concurrent medications), could interact 46 with bisphosphonates and affect remodeling, angiogenesis/blood flow, and/or infection. As our 47 lack of knowledge concerning BRONJ pathogenesis is due to a lack of data, it is only through the 48 initiation of hypothesis-driven studies that significant progress will be made to understand this 49 serious and debilitating condition.

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51 Key words: Osteonecrosis of the jaw, bisphosphonates, bone remodeling, osteocytes,

52 angiogenesis

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54 Introduction

55 Bisphosphonate-related osteonecrosis of the jaw (BRONJ) has become one of the most 56 prominent enigmas not only in the dental community, but in the field of skeletal biology as a 57 whole. Confusion surrounding BRONJ exists for several reasons, including a lack of 58 understanding about how and why this condition manifests ¹. Since brought to light in 2003/2004 59 ²⁻⁵, well over 400 manuscripts have been published concerning BRONJ. Despite this large 60 volume of work there remain few data, yet many hypotheses, concerning the underlying 61 pathophysiology. The goal of this review is to summarize some of the various proposed 62 hypotheses for the pathophysiology of BRONJ.

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64 The starting point for BRONJ: Bone or soft tissue?

65 As its name implies, BRONJ is often assumed to be primarily a bone condition. The hallmark of 66 BRONJ is the existence of exposed bone with the majority of cases manifesting following dental 67 intervention¹. As epithelialization is an essential step in post-intervention wound healing^{6,7}, it 68 has been hypothesized that the soft tissue of the oral mucosa could play a significant role in 69 BRONJ. Specifically, it has been proposed that bisphosphonates, which accumulate in the bone, 70 have direct toxic effects on the oral epithelium and inhibit normal healing of soft tissue lesions caused either by dental intervention or some other trauma^{8,9}. The failure of soft tissue to heal 71 72 would result in exposure of the bone, which then becomes necrotic. Although dental extraction is a significant risk factor for BRONJ¹⁰, the condition does occur in the absence of dental 73 74 intervention¹.

The effect of localized high concentration on the oral mucosa is most clearly illustrated by the case report of stomatitis in a patient who held their bisphosphonate medication in their mouth ¹¹. However, use of bisphosphonates in gel form for treatment of periodontal lesions, although not widely studied and only used short-term, likely present the oral mucosa with high local concentrations and have not shown any adverse effects ¹². Beyond this, however, little is known about bisphosphonates and the oral mucosa. 81 One key unknown for the hypothesis of soft-tissue toxicity is whether the oral mucosa, 82 comprised of epithelial and vascular tissue, is exposed to sufficient bisphosphonate levels in vivo 83 to disrupt its normal physiology. Since bisphosphonates only accumulate in the bone ¹³, a 84 scenario would most likely have to exist in which large amounts of drug are liberated either all at 85 once, or at sufficiently high concentrations over a prolonged duration. It has been suggested that 86 this would occur during dental intervention due to physical disruption of the bone^{8,9}, although this 87 has not been assessed and would not explain the occurrence of BRONJ in the absence of dental 88 procedures. Alternatively, sufficient concentrations of bisphosphonates in the saliva, or gingival 89 crevicular fluid could expose the oral mucosa to high levels of drug. Whether or not the several 90 BRONJ risk factors, such as diabetes, smoking, and concurrent medications, play a role in 91 compromising the oral mucosa is also unclear. Thus, while the hypothesis remains intriguing and 92 is worth further study, the skeleton seems most likely to serve as the central factor for initiation of 93 BRONJ.

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95 Bone cells and BRONJ: Where to focus attention

96 The physiological effects of bisphosphonates on bone cells - osteoblasts, osteoclasts, and osteocytes - have recently been expertly reviewed ^{14, 15}. Osteoclasts (Figure 1a) are the main 97 98 cellular target of bisphosphonates ¹⁶. Specifically, through disruption of intracellular pathways, 99 bisphosphonates suppress osteoclast-mediated bone remodeling. As remodeling plays a vital 100 role in tissue renewal and bone healing, bisphosphonate-induced remodeling suppression imparts significant effects on various tissue-level properties ¹⁷⁻²². The effects of bisphosphonates 101 102 on osteocytes (Figure 1b), the most abundant of the bone cells, are less clear and more 103 controversial. Evidence exists for both direct and indirect effects, most of which are centered around the viability and integrity of these cells and their environment ^{20, 23-26}. Osteoblasts 104 105 (Figure 1c) appear to be the least affected of the bone cells. . While systemic bone formation is 106 reduced in the presence of bisphosphonates, this is primarily an indirect consequence of 107 remodeling suppression and the coupling between resorption and formation. At the level of the individual basic multicellular unit osteoblast activity appears unaffected ^{27, 28}. Reports from small 108

animal models suggest that bisphosphonates may suppress osteoblastic bone formation directly
 on those surface undergoing bone formation without prior resorption (i.e. formation modeling)²⁹,
 although large animal models do not show a similar suppressive effect on periosteal surfaces³⁰⁻
 ³². Collectively, this evidence points towards the osteoclasts and/or osteocytes as the main cells
 of interest for BRONJ pathogenesis.

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115 Remodeling suppression and BRONJ: The basic premise of the most popular hypothesis 116 Nearly every report and review of BRONJ points to bisphosphonate-induced remodeling 117 suppression as a likely mechanism. The basic premise of this hypothesis is that the jaw has a 118 high remodeling rate and bisphosphonates suppress remodeling. There is no debate about the 119 latter as this is the principal mechanism of action of the bisphosphonates ^{14, 15}. It is also clear that 120 remodeling, specifically within the intracortical envelope, is considerably higher in the jaw 121 compared to other skeletal sites. As opposed to remodeling that occurs on bone surfaces, 122 intracortical (osteonal) remodeling occurs within cortical bone of humans and many large animals 123 ³³. In humans, intracortical remodeling rates of the jaw are 10-20 times higher than within the 124 cortex of the iliac crest ^{34, 35}. Animal studies support the limited human data, consistently showing 125 that remodeling rates in the jaw are significantly higher than in the long bones ^{36, 37}. The BRONJ 126 hypothesis thus follows the idea that since remodeling is high in the jaw, and bisphosphonates 127 suppress remodeling, this likely plays a role in the pathophysiology of BRONJ.

128 Bisphosphonate effects on intracortical bone remodeling of the mandible have only 129 recently been documented ²⁰. Following three years of daily treatment with doses of oral 130 alendronate that were either equivalent to the clinical dose for osteoporosis, or 5x higher, the 131 level of intracortical remodeling was histologically assessed in skeletally mature beagle dogs. 132 Consistent with previous work by others, untreated animals had mandible remodeling rates, 133 specifically within the alveolar region, that were >10-fold higher than within long bones. Daily oral 134 alendronate significantly suppressed intracortical bone formation rate of the mandible compared to vehicle, mainly due to suppression in the alveolar bone region (Figure 2)²⁰. These data 135 136 represent clear evidence of bisphosphonate-induced turnover suppression in the jaw and

137 although consistent with both components of the remodeling-suppression hypothesis for BRONJ138 they do not establish a clear cause-effect relationship.

139 The incidence of BRONJ is significantly higher in cancer patients compared to those 140 treated for osteoporosis ¹ suggesting differences should exist in the degree of remodeling 141 suppression between these two populations. Compared to treatment regimens used for 142 osteoporosis, treatment regimens in cancer patients use bisphosphonates with higher potency 143 and binding affinity (zoledronate and pamidronate) and involve both higher doses and more 144 frequent dosing schedules. Differences in potencies and binding affinities among the 145 bisphosphonates are known to affect the degree of remodeling suppression ¹⁴, and both 146 treatment dose and duration are associated with BRONJ^{10, 38-40}. Thus, it is reasonable to 147 hypothesize that cancer patients may experience a greater remodeling suppression within the 148 jaw, although such measurements have never been made even in pre-clinical models.

149 If remodeling suppression is part of the pathophysiology of BRONJ, one might expect it to 150 occur during the course of treatment with other anti-catabolic therapies. Although no BRONJ 151 cases have been reported in patients treated with other anti-remodeling agents (hormone 152 replacement therapy, selective estrogen receptor modulators, and calcitonin), these agents 153 typically do not suppress turnover by more than 50%. Denosumab, a monoclonal anti-RANKL 154 antibody currently in clinical trials for both osteoporosis and cancer populations, has been shown 155 to suppress remodeling an equal or greater extent than bisphosphonates ^{41, 42}. Although no 156 cases of BRONJ have been reported in patients treated with denosumab, increases in the 157 number of treated patients, as well as the duration of treatment, will help to answer questions 158 surrounding the role of remodeling suppression in BRONJ.

Individuals with genetic mutations affecting osteoclast activity provide a means of studying the effects of significant levels of remodeling suppression ⁴³. Several of these genetic conditions have been reported to produce BRONJ-like symptoms ^{44, 45}, supporting the idea of remodeling-suppression in the pathophysiology. For example, patients with inactivating mutations in the chloride channel 7 gene have autosomal dominant osteopetrosis (ADO), a condition in which osteoclast resorption is significantly compromised ^{45, 46}. Jaw osteomyelitis was noted in 13% of patients with ADO, compared to a complete absence of osteomyelitis in the
control population ⁴⁶. Interestingly, 5 of the ADO patients (8%) had draining fistulas and/or
obvious bony destruction resulting in visible defects in the jaw or palate, a similar clinical
presentation to that of BRONJ ⁴⁷. Patients with a different genetic condition, pyknodysostosis, an
autosomal recessive mutation in the cathepsin-K gene which inhibits osteoclast activity, have also
been shown to develop exposed bone in the oral cavity ^{48, 49}.

171 Perhaps the most intriguing reports supporting the remodeling suppression hypothesis of 172 BRONJ concern resolution of the condition subsequent to treatment with agents that stimulate 173 remodeling. In three separate case reports, patients with confirmed BRONJ were treated with 174 teriparatide (recombinant human parathyroid hormone (1-34)), an FDA approved agent for 175 treating post-menopausal osteoporosis which acts through stimulation of bone remodeling ⁵⁰⁻⁵². 176 While each of these cases involved numerous other interventions (including cessation of 177 bisphosphonate treatment, debridement, and anti-bacterial washes), the resolution of BRONJ 178 came only after introduction of teriparatide treatment.

The strongest challenge to the remodeling suppression hypothesis comes from children with osteogenesis imperfecta. These patients are routinely treated with high doses of bisphosphonates and to date there have been no reports of BRONJ ^{53, 54}. It is unclear if or why bisphosphonates differentially affect remodeling in the jaw of young and old subjects.

183 An important aspect of the remodeling suppression hypothesis is that much of the focus 184 has been on the pre-existing bone which may not be the true site of interest. Similar to fracture 185 healing, after dental extraction the socket fills with woven bone which over time is remodeled into 186 lamellar bone ^{6, 7, 55}. The fracture healing literature clearly shows woven bone formation is not 187 compromised in the presence of bisphosphonates, yet remodeling of this callus is significantly 188 delayed ^{22, 56-58}. Thus, it may be that bone formed early during oral wound healing is not 189 remodeled in a timely fashion and this in turn develops into BRONJ. Although some investigation has occurred looking at extractions in the presence of bisphosphonates ^{59, 60}, these studies have 190 191 focused on the preservation of the alveolar bone structure. There exist no data to describe how 192 bisphosphonates affect bone that is formed post-extraction or how it is remodeled over time.

The hypothesis of remodeling suppression as a factor in the pathophysiology of BRONJ makes sense and is supported by some data. Yet a key unanswered question is how the suppression of remodeling, even at a site with high turnover, results in necrotic bone and why this seems specific to the high doses of intravenous bisphosphonates.

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198 Remodeling suppression and BRONJ: Focus on the osteocyte

199 There exists limited histological assessment of BRONJ tissue, yet that which exists almost 200 universally notes the presence of empty lacunae - void of their resident osteocytes. Osteocytes, 201 the most abundant bone cells, form an intricate communication network throughout the 202 mineralized matrix (Figure 3) and play a key role in skeletal physiology ^{61, 62}. While generally 203 considered a long-lived cell, the lifespan of the osteocyte is finite and therefore over time these cells undergo natural death 63-65. Under normal physiological conditions, loss of osteocytes and 204 205 the associated changes to tissue can likely be held in check by bone remodeling ⁶⁴. Yet as 206 bisphosphonates suppress remodeling, regions of non-viable osteocytes would be expected to 207 accumulate.

208 Focal loss of viable bone matrix has been documented in a pre-clinical animal model. 209 Following three years of treatment with oral bisphosphonates, mandibles of beagle dogs contained significant regions of non-viable bone matrix²⁰. Using en bloc basic fuchsin staining 210 211 which fills all voids within the matrix (microdamage, Haversian canals, osteocyte lacunae and 212 canaliculi 66, 67), regions of non-viable bone matrix were identified by the absence of stain 213 suggesting the osteocyte network had filled with mineral (Figure 4). Non-viable matrix was noted 214 in a fraction of bisphosphonate-treated animals (~30%), most often in the alveolar portion of the bone, yet was not observed in any control animals ²⁰. Using this same basic fuchsin technique, 215 216 regions of matrix necrosis can be observed in pathological samples from BRONJ patients (Figure 217 5). Previous studies on samples from patients with BRONJ, using more standard methods of 218 histological evaluation with hematoxylin and eosin staining, have also observed areas of bone tissue with empty lacuna interspersed among areas of vital bone ⁶⁸. Although it remains unclear if 219

or how these areas of focal matrix necrosis play into BRONJ ⁶⁹ these findings suggest the
 osteocyte could have a central role in the pathophysiology.

222 The accumulation of non-viable osteocytes in association with bisphosphonate treatment 223 could manifest through indirect or direct mechanisms. As outlined above, osteocyte death is a 224 natural process and through suppression of remodeling it would be expected that regions with 225 non-viable cells would be more prevalent. This accumulation would have little to do with 226 bisphosphonates, per se, but rather would be a result of suppressed remodeling. If this 227 hypothesis is correct, it would be expected that other anti-remodeling agents, or other conditions 228 which result in remodeling suppression, would also result in an accumulation of non-viable bone, 229 and that it would be dose- or potency-dependent. Additionally, it would be expected that regions 230 of non-viable bone would not be confined to the mandible but would be present throughout the 231 skeleton.

232 An alternate hypothesis for the accumulation of non-viable osteocytes with 233 bisphosphonates is through a direct effect of these drugs on the osteocytes ^{9, 69}. It is well 234 accepted that bisphosphonates become embedded in the skeleton and therefore accumulate 235 over time ^{70, 71}. It has recently been demonstrated that systemically administered 236 bisphosphonates have access to, and become embedded in, the osteocyte lacunae⁷². As such, 237 it is possible that osteocytes could be exposed to high concentrations of bisphosphonates over 238 time which in turn could affect cell viability. 239 The effects of bisphosphonates on osteoblast/osteocyte viability have been

240 predominantly investigated in vitro. Through connexin (Cx)-43 hemichannel transduction of 241 extracellular signal regulated kinases (ERKs), low concentrations of bisphosphonates have been 242 shown to suppress osteocyte apoptosis through maintenance of cellular connections ^{26, 73-75}. 243 These results have translated well to in vivo models where bisphosphonates have also been shown to suppress prednisone-induced ²⁶ and mechanically-induced ²⁵ osteocyte apoptosis. 244 245 However, the anti-apoptotic effects in vitro appear to be dose-dependent such that higher concentrations increase osteocyte apoptosis²⁴. This establishes a plausible scenario where 246 247 osteocytes are initially exposed to low levels of bisphosphonates which prolong osteocyte

248 longevity yet with continued treatment, particularly at high doses, concentrations of drug249 accumulate near the osteocyte which results in cell death.

250 Whether these direct or indirect pathways connecting bisphosphonates to loss of 251 osteocyte viability play a role in BRONJ is unclear. Of the two, the direct pathway is more 252 consistent with numerous clinical aspects of the condition. BRONJ is more prominent in patients 253 treated with high-doses of intravenous pamidronate or zoledronate, as compared to those treated 254 at lower doses for osteoporosis¹. Intravenous administration results in a higher skeletal 255 accumulation as compared to oral administration ¹³, while pamidronate and zoledronate have the highest mineral binding affinities among all of the bisphosphonates ^{76,77}. The increased risk of 256 257 BRONJ associated with treatment duration is also consistent with the accumulation of drug over 258 time.

Given the central role of osteocytes in the regulation of the skeleton, understanding how pharmacological agents affect their physiology is essential. The effects of bisphosphonates on osteocytes are only now beginning to be understood. Despite indirect evidence that the loss of osteocyte viability could play a role in the pathophysiology of BRONJ, the paucity of data results in this remaining a hypothesis.

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265 BRONJ and vasculature: The anti-angiogenic effects of bisphosphonates.

Prior to the emergence of BRONJ, much of what was known concerning osteonecrosis centered
on two conditions which manifest due to disruptions in vasculature. Avascular necrosis of the hip
occurs secondary to disruption of the vasculature ⁷⁸. Similarly, osteoradionecrosis, most
prominently of the jaw, occurs following radiation-induced disruption of the vasculature ^{68, 79, 80}.
The existence of these conditions, and the clear role of disrupted vasculature in their
pathophysiology, has led to the hypothesis that the vasculature plays a key role in
pathophysiology of BRONJ.

A role of the vasculature in BRONJ has been mostly fueled by studies showing antiangiogenic properties of bisphosphonates. Indeed, bisphosphonates are emerging as a potential means of suppressing angiogenesis associated with tumor growth ^{81, 82}. Numerous studies have 276 documented anti-angiogenic effects of bisphosphonates in vitro while a smaller number have 277 shown similar effects in vivo. The latter include suppression of angiogenesis in subcutaneously implanted tissue chambers⁸³, reduced testosterone-induced prostate tissue re-vascularization 278 279 following castration⁸⁴, and significant reductions in marrow vessel number of iliac crest biopsies after six months of clodronate treatment for Paget's disease⁸⁴. Conversely, early in vivo studies 280 281 with high doses of bisphosphonates did not document altered vascular invasion near the growth 282 plates ⁸⁵. There have been no systematic studies assessing the vascular pattern in BRONJ. 283 Qualitatively, the vasculature has been reported to be intact in a series of BRONJ cases ⁸⁶ while 284 a separate series reported 'vessel obliteration' in some BRONJ specimens ^{68, 87}. In the dog 285 model of matrix necrosis the vasculature appears to be patent and intact even in regions devoid 286 of viable osteocytes ²⁰.

Recently, two cases of exposed bone in the mandible, similar in nature to BRONJ, have
been reported in cancer patients treated with Bevacizumab, a recombinant human monoclonal
antibody that binds to vascular endothelial growth factor (VEGF) and inhibits angiogenesis ⁸⁸.
These patients were not treated with bisphosphonates and did not undergo any dental
intervention. This provides the strongest evidence to date suggesting a role of the vasculature in
BRONJ.

Reduced angiogenesis within bone would actually be expected to occur with bisphosphonates, due to a suppression of remodeling. Each remodeling unit receives its nutrients by a vessel ⁸⁹; therefore bone remodeling and angiogenesis are intimately linked. Bisphosphonate-induced reductions in remodeling should be associated with reduced angiogenesis yet the reduction would be a secondary effect. It remains unclear if this has relevance to BRONJ.

Perhaps the most intriguing role of altered angiogenesis with bisphosphonates may be related to wound healing ⁹⁰. Following tooth extraction, a major precipitating event in BRONJ ¹⁰, the extraction site undergoes a well-defined series of healing steps which include an initial clot formation, conversion of clot to granulation tissue, formation of connective tissue and preosseous tissue, and finally filling of the extraction socket with bone ^{7, 91-93}. Disruption of this 304 normal process at any stage, particularly the formation of the provisional matrix that occurs early 305 during treatment, could compromise the entire process ⁹⁰. Furthermore, disruption of the 306 remodeling of this extraction site by osteoclasts, which normally occurs via an accelerated rate of 307 modeling and remodeling ^{55, 93}, may potentially play into the lack of healing that is a prominent 308 feature of BRONJ. The sole evidence on this topic comes from an in vivo study in mice which 309 showed that bisphosphonates did not affect angiogenesis associated with endochondral 310 ossification, a process that is similar to that which occurs with skeletal wound healing ⁹⁴.

311 Another aspect related to vasculature, but not as routinely discussed in the BRONJ 312 literature as angiogenesis, is potential effects of bisphosphonates on blood flow ⁹⁰. Tissue blood 313 flow is directly proportional to its metabolic activity with the bone receiving ~4-7% of total cardiac 314 output at rest ⁹⁵ compared to ~17% for skeletal muscle ⁹⁶. Blood flow distribution throughout the 315 skeleton is heterogeneous and varies by a factor of ten among bones ⁹⁷. Given its high 316 remodeling rate, the mandible would be expected to have high blood flow rates. The lone data 317 concerning mandible blood flow show values for the mandible that are similar to long bones ^{97, 98}. 318 Importantly, however, these flow rates in the mandible are probably underestimated as the teeth 319 likely account for a significant portion of the mass, yet do not directly receive blood. Regardless 320 of basal blood flow, it would be expected that blood flow to the mandible would be reduced with 321 bisphosphonates due to the suppression of remodeling (which would lower the metabolic 322 demand). This reduced blood flow would lead to vascular remodeling ⁹⁹ with the skeletal vessels 323 becoming smaller and thus less able to accommodate the demands for skeletal perfusion that are known to exist post-extraction or with infection ^{100, 101}. The inability to raise blood flow in these 324 325 circumstances could compromise tissue viability and play a role in BRONJ.

326

327 BRONJ and Infection: It's there but does it contribute to the pathophysiology

Numerous bacteria have been reported in patients with BRONJ yet there is nearly a universal presence of Actinomyces ^{68, 86, 87 102}. Actinomyces species, most commonly *Actinomyces israelii*, are the most prominent of the over 500 microflora in the oral cavity ¹⁰³. Through their formation of a biofilm on the bone/tooth/mucosal surface, Actinomyces perpetuate the adherence of other microflora which results in a heterogeneous population of bacteria primed for development of
 infection ¹⁰³. Despite the presence of these bacterial conglomerates in many patients with
 BRONJ, there is no clear evidence to address the question of whether infection is a primary or
 secondary event in BRONJ pathophysiology.

336 One plausible mechanism through which infection could contribute to BRONJ is by 337 enhancing osteoclast-independent bone resorption. BRONJ tissue consistently shows a prevalence of scalloped bone surfaces ^{68, 86, 87} (Figure 5), a seemingly paradoxical property given 338 339 the effect of bisphosphonates on bone resorption. Bacteria and associated fibroblast-like cells 340 have the capacity to directly resorb bone, independent of osteoclasts, by liberating various acids 341 and proteases ^{102, 104, 105}. As osteoclasts signal osteoblasts during normal bone remodeling ^{106,} 342 ¹⁰⁷, resorption that occurs independent of osteoclasts would likely lack osteoblast-mediated bone 343 formation. Whether such resorption could factor into BRONJ is unclear but seems worth 344 exploring.

345

Other hypotheses of BRONJ

347 In addition to the hypotheses outlined above, numerous others exist mostly related to the role of 348 various co-factors in the pathophysiology of BRONJ. Co-morbidities (e.g. diabetes ¹⁰⁸), lifestyle factors (e.g. smoking ¹⁰⁹ and obesity ¹⁰⁹), interventions (e.g. dental extraction ¹⁰), and concurrent 349 medications (e.g. corticosteroids (¹¹⁰)) have all been associated with BRONJ. With all of these 350 351 factors, the proposed mechanism for contribution to BRONJ relates back to the main 352 mechanisms outlined above - remodeling, angiogenesis/blood flow, and infection. As dental 353 manifestations similar to BRONJ have not been observed with any of these co-factors in the 354 absence of bisphosphonates, it suggests either these co-factors don't play a significant role or 355 that it is the interaction between the co-factors and bisphosphonates that is the key to the 356 pathophysiology.

357

358 Future Directions: Data anyone?

359 Above all else, the field of BRONJ needs data. The amount of data, excluding those concerning 360 incidence/prevalence/risk factors, is appalling given the five years that have passed since the 361 initial descriptions of this condition. Without undertaking hypothesis-driven studies to tease apart 362 the potential pathophysiology we simply won't get any closer to understanding this condition. 363 Recently, the American Society for Bone and Mineral Research organized a multi-disciplinary 364 task force concerning BRONJ which put forward several questions/areas of study, ranging from 365 clinical to molecular, that the field needs to advance¹. While this provides an excellent starting 366 point, the topics outlined are not all encompassing; other important areas related to BRONJ 367 surely exist. The key is that we need to start generating data, without which interest in BRONJ 368 within the research community will wane as the field will simply not move forward. We are 369 dangerously close to this happening and for the sake of the patients with BRONJ we must do 370 everything we can to understand all that we can. 371 372 373 **Figure Legends** 374 375 Figure 1. The pathophysiology of BRONJ likely involves one or more of the bone cell 376 populations. Osteoclasts (A), seen here stained with tartrate-resistant acid phosphatase, 377 function to resorb bone; suppression of their activity is the mechanism underlying the 378 effectiveness of bisphosphonate treatment. Osteocytes (B), entombed within the mineralized 379 matrix, are connected to each other and to the bone surface by an intricate cell process network 380 (seen here stained with basic fuchsin); the effect of bisphosphonates on these cells remains 381 controversial. Osteoblasts (C), seen here as tall cuboidal cells actively forming osteoid (the thin 382 pale blue seam adjacent to the bone surface), are less active in the presence of bisphosphonates 383 although this is predominately an indirect effect of reduced bone remodeling. Scale bars = 50 µm. 384

Figure 2. Bisphosphonates reduce mandible bone remodeling. Following three years of
 daily treatment with oral alendronate (ALN), at the dose used for osteoporosis treatment (ALN)

387 0.2) or a dose 5x higher (ALN1.0), intracortical bone formation rate was assessed in the mandible 388 of female beagle dogs. There was a significant reduction in the overall bone formation rate of the 389 mandible with both doses of alendronate compared to age-matched animals treated with vehicle. 390 The greatest suppression of turnover was noted in the alveolar portion of the mandible with no 391 significant effect of ALN treatment on turnover suppression in the non-alveolar portion. *p < 0.05392 versus VEH. Adapted from J Oral Maxillofac Surg, 66(5), MR Allen and DB Burr, Mandible Matrix 393 Necrosis in Beagle Dogs After 3 Years of Daily Oral Bisphosphonate Treatment, 2008, with 394 permission from Elsevier.

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Figure 3. The osteocyte lacunar-canalicular network. Using acid etching of plastic embedded
specimens, the intricate nature of the lacunar-canalicular system can be revealed. Disruption of
this network could play a significant role in the pathophysiology of BRONJ. Scale bar = 50 µm.
Image complements of Daniel Kubek, Indiana University School of Medicine.

400

401 Figure 4. Mandible matrix necrosis following bisphosphonate treatment in a pre-clinical 402 model. Following three years of daily treatment with alendronate (ALN), regions of focal matrix 403 necrosis existed in the mandibles of beagle dogs. Using en bloc basic fuchsin staining, which 404 passively diffuses and fills all void spaces (blood vessels, lacunae, canaliculi), viable bone matrix 405 tissue can easily be identified by the presence of stain; the absence of stain indicates the lack of 406 permeability to a given region. In this representative photomicrograph of a mandible from an 407 ALN-treated animal, the central region is noticeably void of stain and therefore considered to be 408 non-viable tissue. Peripheral to the central region of non-viable bone matrix, tissue that is 409 sufficiently stained (and therefore considered viable) can be observed. The upper right of the 410 photomicrograph shows the tooth, below which is the periodontal ligament (which is heavily 411 stained with fuchsin). Scale bar = $500 \mu m$.

412

413 Figure 5. Non-viable bone matrix in BRONJ specimens. Using en bloc basic fuchsin staining,

414 regions of non-viable bone matrix can be observed in a pathological specimen from a patient with

415 BRONJ (courtesy of Dr. Salvatore Ruggiero). Similar to that observed in bisphosphonate-treated

416 beagle dogs, a region void of fuchsin stain, and therefore considered non-viable, is surrounded by

417 stained (viable) tissue. Scale bar = $100 \mu m$.

418

419 **Figure 6. Extensive scalloped bone surfaces in BRONJ tissue.** Using high-resolution micro-

420 computed tomography (Skyscan 1172, 5 µm resolution), the extent of eroded surfaces (examples

421 shown by arrows) in a sequestrum from a patient with BRONJ (courtesy of Dr. Salvatore

422 Ruggiero) can be visualized. Such extensive erosion would be unexpected in patients treated

- 423 with bisphosphonates, suggesting osteoclast-independent mechanisms of bone resorption may
- 424 be active in BRONJ. Scale bar = 1 mm.
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427 References

- Khosla, S., D. Burr, J. Cauley, et al. Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. J Bone Miner Res. 22(10): 1479-91. 2007.
- 431
 431
 432
 Marx, R.E. Pamidronate (Aredia) and zoledronate (Zometa) induced avascular necrosis of the jaws: a growing epidemic. J Oral Maxillofac Surg. 61(9): 1115-7. 2003.
- 433
 434
 434
 435
 Mehrotra, B., J. Fantasia, S. Nissel-Horowitz, et al. Osteonecrosis of the maxilla: an unusual complication of prolonged bisphosphonate therapy. a case report. . Proc Am Soc Clin Oncol. 22: (abstr 3194). 2003.
- 436
 4. Ruggiero, S.L., B. Mehrotra, T.J. Rosenberg, et al. Osteonecrosis of the jaws associated
 with the use of bisphosphonates: a review of 63 cases. J Oral Maxillofac Surg. 62(5):
 527-34. 2004.
- 439 5. Wang, J., N.M. Goodger, and M.A. Pogrel. Osteonecrosis of the jaws associated with cancer chemotherapy. J Oral Maxillofac Surg. 61(9): 1104-7. 2003.
- Amler, M.H., P.L. Johnson, and I. Salman. Histological and histochemical investigation of human alveolar socket healing in undisturbed extraction wounds. J Am Dent Assoc. 61: 32-44. 1960.
- 4447.Cardaropoli, G., M. Araujo, and J. Lindhe. Dynamics of bone tissue formation in tooth445extraction sites. An experimental study in dogs. J Clin Periodontol. 30(9): 809-18. 2003.
- 4468.Reid, I.R., M.J. Bolland, and A.B. Grey. Is bisphosphonate-associated osteonecrosis of
the jaw caused by soft tissue toxicity? Bone. 41(3): 318-20. 2007.
- 448 9. Reid, I.R. and T. Cundy. Osteonecrosis of the jaw. Skeletal Radiol. 2008.
- 449 10. Mavrokokki, T., A. Cheng, B. Stein, et al. Nature and frequency of bisphosphonate450 associated osteonecrosis of the jaws in Australia. J Oral Maxillofac Surg. 65(3): 415-23.
 451 2007.
- 45211.Rubegni, P. and M. Fimiani. Images in clinical medicine. Bisphosphonate-associated
contact stomatitis. N Engl J Med. 355(22): e25. 2006.

454 12. Reddy, G.T., T.M. Kumar, and Veena. Formulation and evaluation of Alendronate 455 Sodium gel for the treatment of bone resorptive lesions in Periodontitis. Drug Deliv. 12(4): 456 217-22. 2005. 457 13. Lin, J.H. Bisphosphonates: a review of their pharmacokinetic properties. Bone. 18(2): 75-458 85. 1996. 459 14. Russell, R.G., N.B. Watts, F.H. Ebetino, et al. Mechanisms of action of bisphosphonates: 460 similarities and differences and their potential influence on clinical efficacy. Osteoporos 461 Int. 2008. 462 Russell, R.G., Z. Xia, J.E. Dunford, et al. Bisphosphonates: an update on mechanisms of 15. 463 action and how these relate to clinical efficacy. Ann N Y Acad Sci. 1117: 209-57. 2007. 464 16. Rodan, G.A. and H.A. Fleisch. Bisphosphonates: mechanisms of action. J Clin Invest. 465 97(12): 2692-6. 1996. 466 Allen, M.R., K. Iwata, R. Phipps, et al. Alterations in canine vertebral bone turnover, 17. 467 microdamage accumulation, and biomechanical properties following 1-year treatment 468 with clinical treatment doses of risedronate or alendronate. Bone. 39(4): 872-9. 2006. 469 18. Mashiba, T., C.H. Turner, T. Hirano, et al. Effects of suppressed bone turnover by 470 bisphosphonates on microdamage accumulation and biomechanical properties in 471 clinically relevant skeletal sites in beagles. Bone. 28(5): 524-31. 2001. 472 19. Allen, M.R. and D.B. Burr. Three years of alendronate treatment results in similar levels 473 of vertebral microdamage as after one year of treatment. J Bone Miner Res. 22(11): 474 1759-65. 2007. 475 20. Allen, M.R. and D.B. Burr. Mandible matrix necrosis in beagle dogs after 3 years of daily 476 oral bisphosphonate treatment. J Oral Maxillofac Surg. 66(5): 987-94. 2008. 477 21. Boivin, G.Y., P.M. Chavassieux, A.C. Santora, et al. Alendronate increases bone strength 478 by increasing the mean degree of mineralization of bone tissue in osteoporotic women. 479 Bone. 27(5): 687-94. 2000. 480 22. McDonald, M.M., S. Dulai, C. Godfrey, et al. Bolus or weekly zoledronic acid 481 administration does not delay endochondral fracture repair but weekly dosing enhances 482 delays in hard callus remodeling. Bone. 2008. 483 23. Plotkin, L.I., J.I. Aguirre, S. Kousteni, et al. Bisphosphonates and estrogens inhibit 484 osteocyte apoptosis via distinct molecular mechanisms downstream of extracellular 485 signal-regulated kinase activation. J Biol Chem. 280(8): 7317-25. 2005. 486 24. Idris, A.I., J. Rojas, I.R. Greig, et al. Aminobisphosphonates cause osteoblast apoptosis 487 and inhibit bone nodule formation in vitro. Calcif Tissue Int. 82(3): 191-201. 2008. 488 25. Follet, H., J. Li, R.J. Phipps, et al. Risedronate and alendronate suppress osteocyte 489 apoptosis following cyclic fatigue loading. Bone. 40(4): 1172-7. 2007. 490 26. Plotkin, L.I., R.S. Weinstein, A.M. Parfitt, et al. Prevention of osteocyte and osteoblast 491 apoptosis by bisphosphonates and calcitonin. J Clin Invest. 104(10): 1363-74. 1999. 492 27. Boyce, R.W., C.L. Paddock, J.R. Gleason, et al. The effects of risedronate on canine 493 cancellous bone remodeling: three-dimensional kinetic reconstruction of the remodeling 494 site. J Bone Miner Res. 10(2): 211-21. 1995. 495 28. Eriksen, E.F., F. Melsen, E. Sod, et al. Effects of long-term risedronate on bone quality 496 and bone turnover in women with postmenopausal osteoporosis. Bone. 31(5): 620-5. 497 2002. 498 29. Iwata, K., J. Li, H. Follet, et al. Bisphosphonates suppress periosteal osteoblast activity 499 independent of resorption in rat femur and tibia. Bone. 39(5): 1053-8. 2006. 500 30. Allen, M.R., H. Follet, M. Khurana, et al. Anti-remodeling agents influence osteoblast 501 activity differently in modeling- and remodeling-associated bone formation Calcified 502 Tissue International. 79(4): 255-61. 2006. 503 31. Allen, M.R., S. Reinwald, and D.B. Burr. Alendronate reduces bone toughness of ribs 504 without significantly increasing microdamage accumulation in dogs following 3 years of 505 daily treatment. Calcif Tissue Int. 82(5): 354-60. 2008. 506 32. Mashiba, T., T. Hirano, C.H. Turner, et al. Suppressed bone turnover by 507 bisphosphonates increases microdamage accumulation and reduces some 508 biomechanical properties in dog rib. J Bone Miner Res. 15(4): 613-20. 2000.

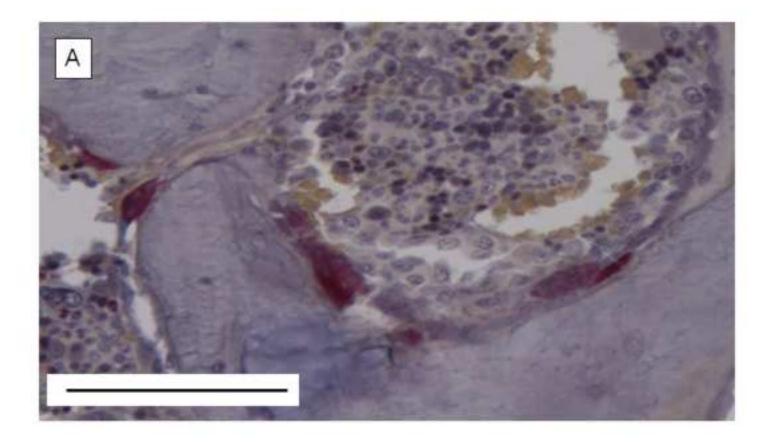
509 33. Reinwald, S. and D. Burr. Review of nonprimate, large animal models for osteoporosis 510 research. J Bone Miner Res. 23(9): 1353-68. 2008. 511 Garetto, L.P., J. Chen, J.A. Parr, et al. Remodeling dynamics of bone supporting rigidly 34. fixed titanium implants: a histomorphometric comparison in four species including 512 513 humans. Implant Dent. 4(4): 235-43. 1995. 514 Han, Z.H., S. Palnitkar, D.S. Rao, et al. Effects of ethnicity and age or menopause on the 35. 515 remodeling and turnover of iliac bone: implications for mechanisms of bone loss. J Bone 516 Miner Res. 12(4): 498-508. 1997. 517 Huja, S.S., S.A. Fernandez, K.J. Hill, et al. Remodeling dynamics in the alveolar process 36. in skeletally mature dogs. Anat Rec A Discov Mol Cell Evol Biol. 288(12): 1243-9. 2006. 518 519 37. Garetto, L.P. and N.D. Tricker. 1998 Remodeling of bone surrounding the implant 520 interface., in Bridging the Gap Between Dental & Orthopaedic Implants, 3rd Annual 521 Indiana Conference, L.P. Garetto, et al., Editors: Indianapolis, IN. 522 Bamias, A., E. Kastritis, C. Bamia, et al. Osteonecrosis of the jaw in cancer after 38. 523 treatment with bisphosphonates: incidence and risk factors. J Clin Oncol. 23(34): 8580-7. 524 2005. 525 39. Dimopoulos, M.A., E. Kastritis, A. Anagnostopoulos, et al. Osteonecrosis of the jaw in 526 patients with multiple myeloma treated with bisphosphonates: evidence of increased risk 527 after treatment with zoledronic acid. Haematologica. 91(7): 968-71. 2006. 528 40. Corso, A., M. Varettoni, P. Zappasodi, et al. A different schedule of zoledronic acid can 529 reduce the risk of the osteonecrosis of the jaw in patients with multiple myeloma. 530 Leukemia. 2007. 531 41. Lewiecki, E.M., P.D. Miller, M.R. McClung, et al. Two-year treatment with denosumab 532 (AMG 162) in a randomized phase 2 study of postmenopausal women with low BMD. J 533 Bone Miner Res. 22(12): 1832-41. 2007. 534 42. Lipton, A., G.G. Steger, J. Figueroa, et al. Randomized active-controlled phase II study of 535 denosumab efficacy and safety in patients with breast cancer-related bone metastases. J 536 Clin Oncol. 25(28): 4431-7. 2007. 537 43. Helfrich, M.H. Osteoclast diseases and dental abnormalities. Arch Oral Biol. 50(2): 115-538 22. 2005. 539 44. Barry, C.P., C.D. Ryan, and L.F. Stassen. Osteomyelitis of the maxilla secondary to 540 osteopetrosis: a report of 2 cases in sisters. J Oral Maxillofac Surg. 65(1): 144-7. 2007. 541 45. Johnston, C.C., Jr., N. Lavy, T. Lord, et al. Osteopetrosis. A clinical, genetic, metabolic, 542 and morphologic study of the dominantly inherited, benign form. Medicine (Baltimore). 543 47(2): 149-67. 1968. 544 46. Waguespack, S.G., D.L. Koller, K.E. White, et al. Chloride channel 7 (CICN7) gene 545 mutations and autosomal dominant osteopetrosis, type II. J Bone Miner Res. 18(8): 1513-546 8. 2003. 547 47. Ruggiero, S.L. and S.J. Drew. Osteonecrosis of the jaws and bisphosphonate therapy. J 548 Dent Res. 86(11): 1013-21. 2007. 549 Bathi, R.J. and V.N. Masur. Pyknodysostosis -- a report of two cases with a brief review of 48. 550 the literature. Int J Oral Maxillofac Surg. 29(6): 439-42. 2000. 551 49. Dimitrakopoulos, I., C. Magopoulos, and T. Katopodi. Mandibular osteomyelitis in a 552 patient with pyknodysostosis: a case report of a 50-year misdiagnosis. J Oral Maxillofac 553 Surg. 65(3): 580-5. 2007. 554 Harper, R.P. and E. Fung. Resolution of bisphosphonate-associated osteonecrosis of the 50. 555 mandible: possible application for intermittent low-dose parathyroid hormone [rhPTH(1-556 34)]. J Oral Maxillofac Surg. 65(3): 573-80. 2007. 557 Lau, A.N., S.H. Ali, and J.D. Adachi. Resolution of osteonecrosis of the jaw after 51. 558 teriparatide [recombinant human PTH (1-34)] therapy. Osteoporos Int. 19 (Suppl 1): S80. 559 2008. 560 52. Wang, H.L., D. Weber, and L.K. McCauley. Effect of long-term oral bisphosphonates on 561 implant wound healing: literature review and a case report. J Periodontol. 78(3): 584-94. 562 2007.

563 564 565	53.	Malmgren, B., E. Astrom, and S. Soderhall. No osteonecrosis in jaws of young patients with osteogenesis imperfecta treated with bisphosphonates. J Oral Pathol Med. 37(4): 196-200. 2008.
566	54.	Schwartz, S., C. Joseph, D. lera, et al. Bisphosphonates, osteonecrosis, osteogenesis
567 568 569	55.	imperfecta and dental extractions: a case series. J Can Dent Assoc. 74(6): 537-42. 2008. Kingsmill, V.J. Post-extraction remodeling of the adult mandible. Crit Rev Oral Biol Med. 10(3): 384-404. 1999.
570 571	56.	Li, C., S. Mori, J. Li, et al. Long-term effect of incadronate disodium (YM-175) on fracture healing of femoral shaft in growing rats. J Bone Miner Res. 16(3): 429-36. 2001.
572 573	57.	Li, J., S. Mori, Y. Kaji, et al. Concentration of bisphosphonate (incadronate) in callus area and its effects on fracture healing in rats. J Bone Miner Res. 15(10): 2042-51. 2000.
574 575	58.	Peter, C.P., W.O. Cook, D.M. Nunamaker, et al. Effect of alendronate on fracture healing and bone remodeling in dogs. J Orthop Res. 14(1): 74-9. 1996.
576 577	59.	Altundal, H. and O. Guvener. The effect of alendronate on resorption of the alveolar bone following tooth extraction. Int J Oral Maxillofac Surg. 33(3): 286-93. 2004.
578 579	60.	Olson, H.M. and A. Hagen. Inhibition of post-extraction alveolar ridge resorption in rats by dichloromethane diphosphonate. J Periodontal Res. 17(6): 669-74. 1982.
580 581	61.	Bonewald, L.F. Osteocytes as dynamic multifunctional cells. Ann N Y Acad Sci. 1116: 281-90. 2007.
582 583	62.	Turner, C.H., A.G. Robling, R.L. Duncan, et al. Do bone cells behave like a neuronal network? Calcif Tissue Int. 70(6): 435-42. 2002.
584 585	63.	Weinstein, R.S., R.L. Jilka, A.M. Parfitt, et al. Inhibition of osteoblastogenesis and promotion of apoptosis of osteoblasts and osteocytes by glucocorticoids. Potential
586		mechanisms of their deleterious effects on bone. J Clin Invest. 102(2): 274-82. 1998.
587	64.	Enlow, D.H. Functions of the Haversian system. Am J Anat. 110: 269-305. 1962.
588	65.	Frost, H.M. In vivo osteocyte death. J Bone Joint Surg Am. 42-A: 138-43. 1960.
589 590	66.	Burr, D.B. and M. Hooser. Alterations to the en bloc basic fuchsin staining protocol for the demonstration of microdamage produced in vivo. Bone. 17(4): 431-3. 1995.
591	67.	Frost, H.M. Micropetrosis. J Bone Joint Surg Am. 42-A: 144-50. 1960.
592	68.	Hansen, T., M. Kunkel, A. Weber, et al. Osteonecrosis of the jaws in patients treated with
593 594		bisphosphonates - histomorphologic analysis in comparison with infected
594 595	69.	osteoradionecrosis. J Oral Pathol Med. 35(3): 155-60. 2006.
595 596	09.	Allen, M.R. Bisphosphonates and Osteonecrosis of the Jaw: Moving From the 'Bedside' to the 'Bench'. Cells, Tissues, Organs. In Press. 2008.
590 597	70.	Masarachia, P., M. Weinreb, R. Balena, et al. Comparison of the distribution of 3H-
598	70.	alendronate and 3H-etidronate in rat and mouse bones. Bone. 19(3): 281-90. 1996.
599	71.	Lin, J.H., D.E. Duggan, I.W. Chen, et al. Physiological disposition of alendronate, a
600		potent anti-osteolytic bisphosphonate, in laboratory animals. Drug Metab Dispos. 19(5):
601		926-32. 1991.
602	72.	Roelofs, A.J., F.P. Coxon, F.H. Ebetino, et al. Use of a fluorescent analogue of
603		risedronate to study localization and cellular uptake of bisphosphonates in vivo. Bone. 42:
604		S85. 2008.
605	73.	Plotkin, L.I., S.C. Manolagas, and T. Bellido. Transduction of cell survival signals by
606		connexin-43 hemichannels. J Biol Chem. 277(10): 8648-57. 2002.
607	74.	Plotkin, L.I., S.C. Manolagas, and T. Bellido. Dissociation of the pro-apoptotic effects of
608		bisphosphonates on osteoclasts from their anti-apoptotic effects on
609		osteoblasts/osteocytes with novel analogs. Bone. 39(3): 443-52. 2006.
610	75.	Plotkin, L.I., V. Lezcano, J. Thostenson, et al. Connexin 43 Is Required for the Anti-
611 612		Apoptotic Effect of Bisphosphonates on Osteocytes and Osteoblasts In Vivo. J Bone
613	76.	Miner Res. 2008. Nancollas, G.H., R. Tang, R.J. Phipps, et al. Novel insights into actions of
614	10.	bisphosphonates on bone: Differences in interactions with hydroxyapatite. Bone. 2005.
615	77.	Leu, C.T., E. Luegmayr, L.P. Freedman, et al. Relative binding affinities of
616		bisphosphonates for human bone and relationship to antiresorptive efficacy. Bone. 38(5):
617		628-36. 2006.

618 619	78.	Kim, H.K. Introduction to osteonecrosis of the femoral head (OFH) and osteonecrosis of the jaw (ONJ). J Musculoskelet Neuronal Interact. 7(4): 350-3. 2007.
620	79.	Store, G. and M. Boysen. Mandibular osteoradionecrosis: clincal behavious and
621		diagnostic aspects. Clin. Otolaryngal. 25: 378-384. 2000.
622 623	80.	Store, G. and G. Grandstrom. Osteoradionecrosis of the mandible: a microradiographic study of cortical bone. Scan J Plast Reconstr Hand Surg. 33: 307-314. 1999.
623	01	
625	81.	Guise, T.A. Antitumor effects of bisphosphonates: promising preclinical evidence. Cancer Treat Rev. 34 Suppl 1: S19-24. 2008.
626	82.	Lipton, A. Emerging role of bisphosphonates in the clinicantitumor activity and
620 627	02.	
628	83.	prevention of metastasis to bone. Cancer Treat Rev. 34 Suppl 1: S25-30. 2008.
629	03.	Wood, J., K. Bonjean, S. Ruetz, et al. Novel antiangiogenic effects of the bisphosphonate compound zoledronic acid. J Pharmacol Exp Ther. 302(3): 1055-61. 2002.
630	84.	Fournier, P., S. Boissier, S. Filleur, et al. Bisphosphonates inhibit angiogenesis in vitro
631	04.	and testosterone-stimulated vascular regrowth in the ventral prostate in castrated rats.
632		Cancer Res. 62(22): 6538-44. 2002.
633	85.	Schenk, R., W.A. Merz, R. Muhlbauer, et al. Effect of ethane-1-hydroxy-1,1-
634	05.	diphosphonate (EHDP) and dichloromethylene diphosphonate (Cl 2 MDP) on the
635		calcification and resorption of cartilage and bone in the tibial epiphysis and metaphysis of
636		rats. Calcif Tissue Res. 11(3): 196-214. 1973.
637	86.	Hellstein, J.W. and C.L. Marek. Bisphosphonate osteochemonecrosis (bis-phossy jaw): is
638	00.	this phossy jaw of the 21st century? J Oral Maxillofac Surg. 63(5): 682-9. 2005.
639	87.	Hansen, T., M. Kunkel, E. Springer, et al. Actinomycosis of the jawshistopathological
640	07.	study of 45 patients shows significant involvement in bisphosphonate-associated
641		osteonecrosis and infected osteoradionecrosis. Virchows Arch. 451(6): 1009-17. 2007.
642	88.	Estilo, C.L., M. Fornier, A. Farooki, et al. Osteonecrosis of the jaw related to
643	00.	bevacizumab. J Clin Oncol. 26(24): 4037-8. 2008.
644	89.	Parfitt, A.M. The mechanism of coupling: a role for the vasculature. Bone. 26(4): 319-23.
645	00.	2000.
646	90.	Somerman, M.J. and L.K. McCauley. Bisphosphonates: Sacrificing the jaw to save the
647		skeleton? BoneKEy-Osteovision. 3(9): 12-18. 2006.
648	91.	Adeyemo, W.L., A.L. Ladeinde, and M.O. Ogunlewe. Clinical evaluation of post-extraction
649		site wound healing. J Contemp Dent Pract. 7(3): 40-9. 2006.
650	92.	Amler, M.H. Disturbed healing of extraction wounds. J Oral Implantol. 25(3): 179-84.
651		1999.
652	93.	Trombelli, L., R. Farina, A. Marzola, et al. Modeling and remodeling of human extraction
653		sockets. J Clin Periodontol. 35(7): 630-9. 2008.
654	94.	Deckers, M.M., E.R. Van Beek, G. Van Der Pluijm, et al. Dissociation of angiogenesis
655		and osteoclastogenesis during endochondral bone formation in neonatal mice. J Bone
656		Miner Res. 17(6): 998-1007. 2002.
657	95.	Tothill, P. and J.N. MacPherson. The distribution of blood flow to the whole skeleton in
658		dogs, rabbits and rats measured with microspheres. Clin Phys Physiol Meas. 7(2): 117-
659		23. 1986.
660	96.	Rowell, L.B. 1993 Human Cardiovascular Control. New York: Oxford University Press.
661	97.	Colleran, P.N., M.K. Wilkerson, S.A. Bloomfield, et al. Alterations in skeletal perfusion
662		with simulated microgravity: a possible mechanism for bone remodeling. J Appl Physiol.
663		89(3): 1046-54. 2000.
664	98.	Bloomfield, S.A., H.A. Hogan, and M.D. Delp. Decreases in bone blood flow and bone
665	00	material properties in aging Fischer-344 rats. Clin Orthop Relat Res. (396): 248-57. 2002.
666	99.	Delp, M.D., P.N. Colleran, M.K. Wilkerson, et al. Structural and functional remodeling of
667		skeletal muscle microvasculature is induced by simulated microgravity. Am J Physiol
668	400	Heart Circ Physiol. 278(6): H1866-73. 2000.
669 670	100.	Lurie, A.G. and S.R. Matteson. 99MTc-diphosphonate bone imaging and uptake in
670	101	healing rat extraction sockets. J Nucl Med. 17(8): 688-92. 1976.
671	101.	McDougall, I.R. Skeletal scintigraphy. West J Med. 130(6): 503-14. 1979.

671 101. McDougall, I.R. Skeletal scintigraphy. West J Med. 130(6): 503-14. 1979.

- 672 102. Sedghizadeh, P.P., S.K. Kumar, A. Gorur, et al. Identification of microbial biofilms in osteonecrosis of the jaws secondary to bisphosphonate therapy. J Oral Maxillofac Surg. 66(4): 767-75. 2008.
- 103. Yeung, M.K. Molecular and genetic analyses of Actinomyces spp. Crit Rev Oral Biol Med.
 10(2): 120-38. 1999.
- 677104.Nair, S.P., S. Meghji, M. Wilson, et al. Bacterially induced bone destruction: mechanisms678and misconceptions. Infect Immun. 64(7): 2371-80. 1996.
- 679 105. Pap, T., A. Claus, S. Ohtsu, et al. Osteoclast-independent bone resorption by fibroblast680 like cells. Arthritis Res Ther. 5(3): R163-73. 2003.
- 681 106. Mundy, G.R. and F. Elefteriou. Boning up on ephrin signaling. Cell. 126(3): 441-3. 2006.
- 682107.Zhao, C., N. Irie, Y. Takada, et al. Bidirectional ephrinB2-EphB4 signaling controls bone683homeostasis. Cell Metab. 4(2): 111-21. 2006.
- 684108.Khamaisi, M., E. Regev, N. Yarom, et al. Possible association between diabetes and685bisphosphonate-related jaw osteonecrosis. J Clin Endocrinol Metab. 92(3): 1172-5. 2007.
- Wessel, J.H., T.B. Dodson, and A.I. Zavras. Zoledronate, smoking, and obesity are
 strong risk factors for osteonecrosis of the jaw: a case-control study. J Oral Maxillofac
 Surg. 66(4): 625-31. 2008.
- 689 110. American Association of Oral and Maxillofacial Surgeons position paper on
 690 bisphosphonate-related osteonecrosis of the jaws. J Oral Maxillofac Surg. 65(3): 369-76.
 691 2007.
- 692 693



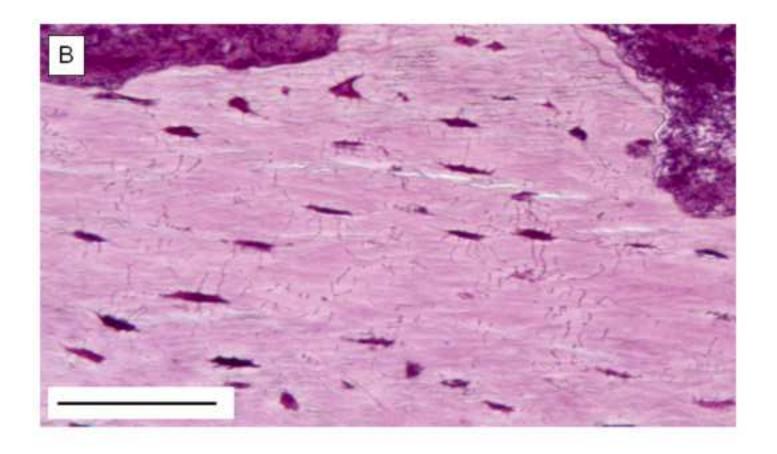


Figure 1b

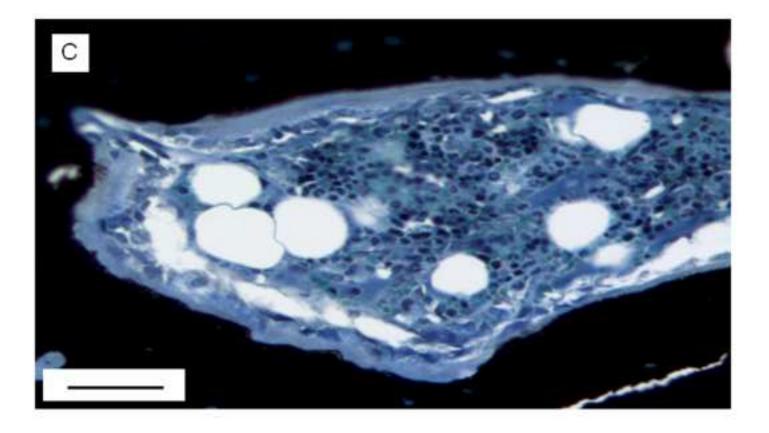


Figure 1c

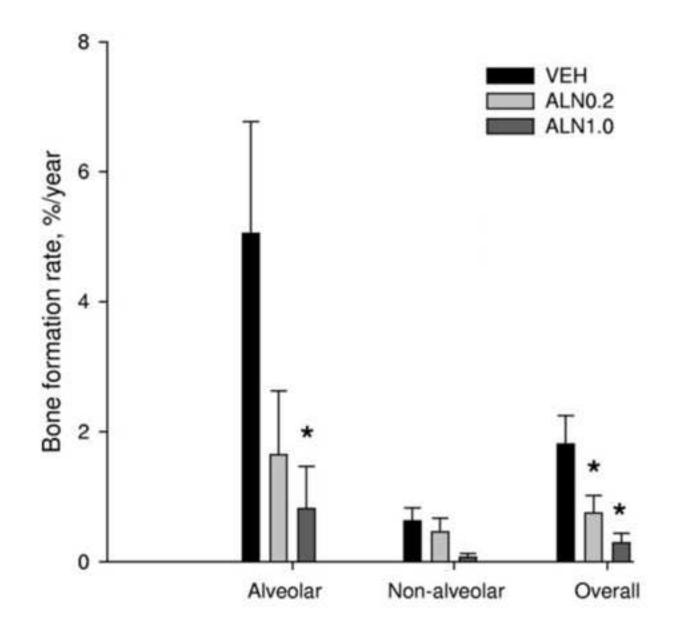
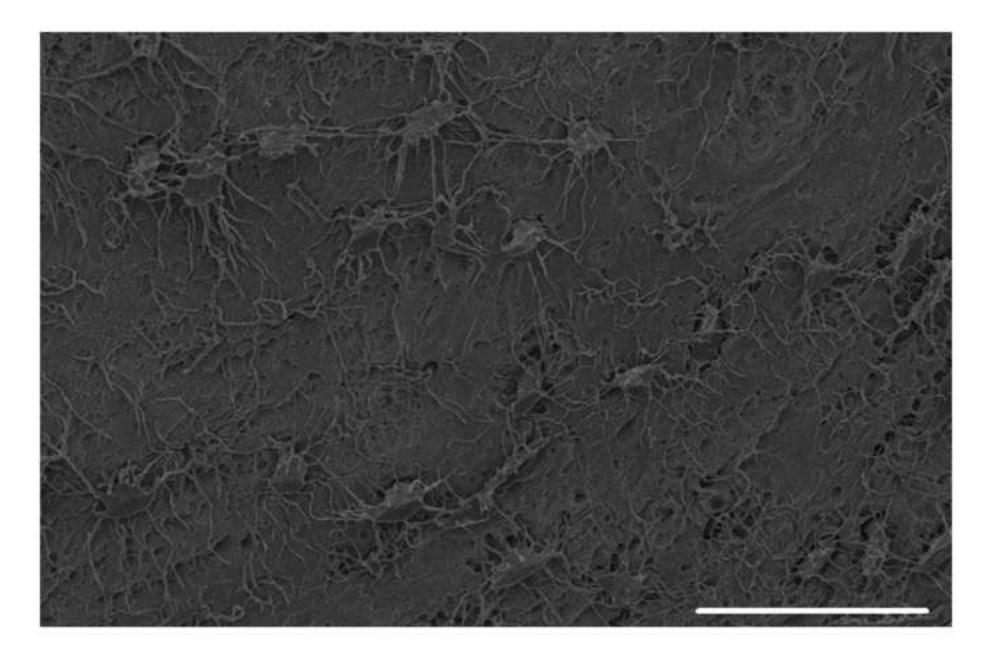
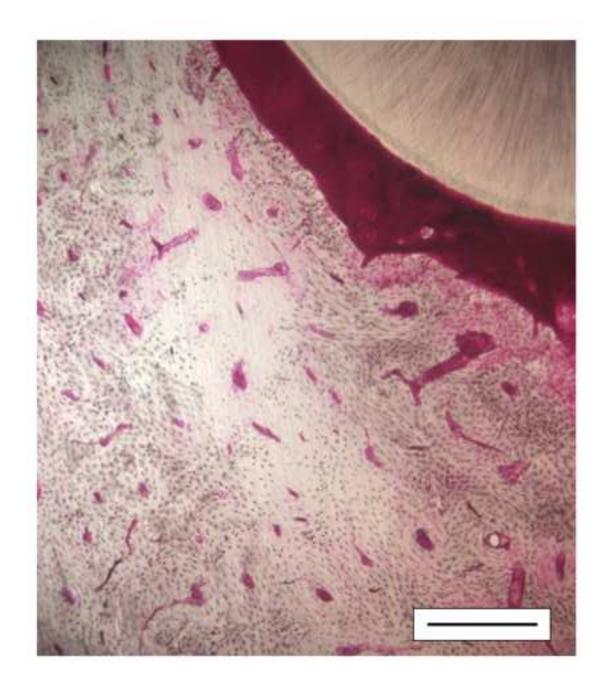
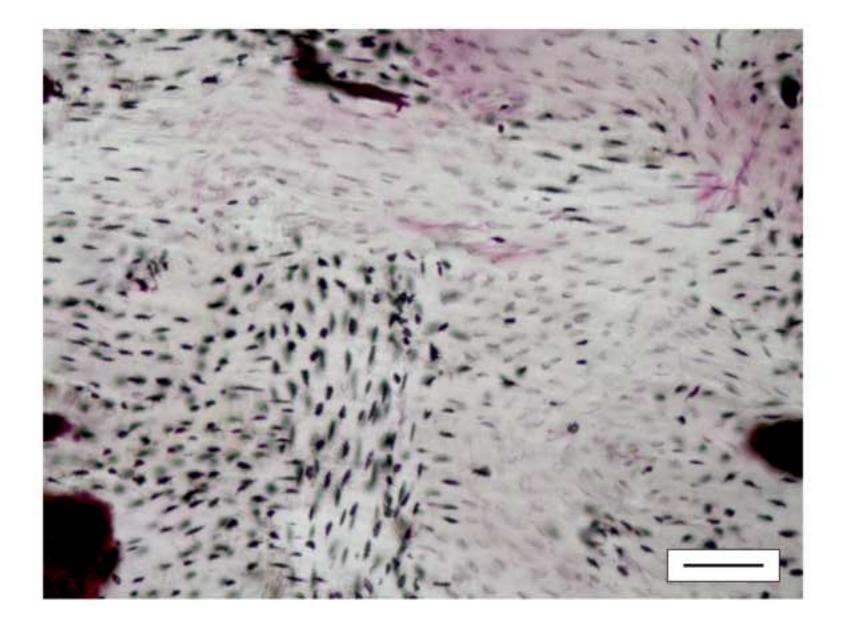


Figure 2







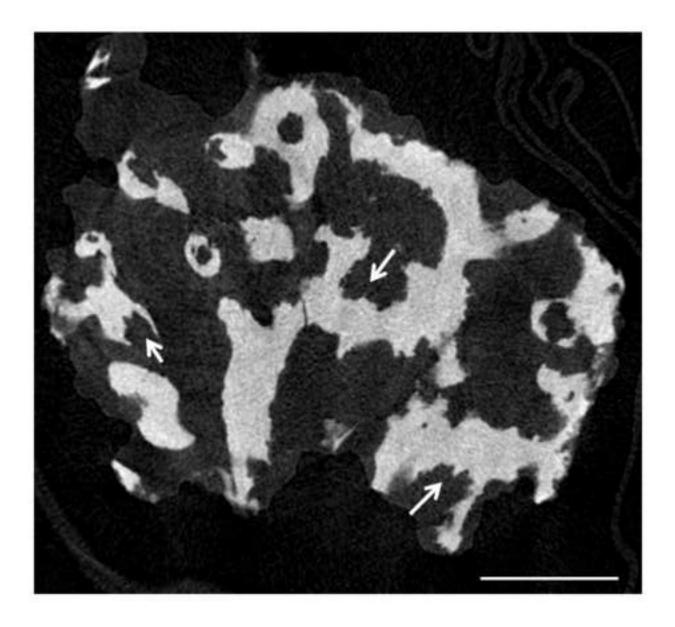


Figure 6