



Precision medicine in patients with allergic diseases: Airway diseases and atopic dermatitis—PRACTALL document of the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology

Antonella Muraro, MD,^a Robert F. Lemanske, Jr, MD,^b Peter W. Hellings, MD,^c Cezmi A. Akdis, MD,^d Thomas Bieber, MD,^e Thomas B. Casale, MD,^f Marek Jutel, MD,^g Peck Y. Ong, MD,^h Lars K. Poulsen, PhD,ⁱ Peter Schmid-Grendelmeier, MD,^j Hans-Uwe Simon, MD,^k Sven F. Seys, PhD,^l and Ioana Agache, MD^m
Padua, Italy, Madison, Wis, Leuven, Belgium, Davos and Bern, Switzerland, Bonn, Germany, Tampa, Fla, Wroclaw, Poland, Los Angeles, Calif, Copenhagen, Denmark, and Brasov, Romania

In this consensus document we summarize the current knowledge on major asthma, rhinitis, and atopic dermatitis endotypes under the auspices of the PRACTALL collaboration platform. PRACTALL is an initiative of the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology aiming to harmonize the European and American approaches to best allergy practice and science. Precision medicine is of broad relevance for the management of asthma, rhinitis, and atopic dermatitis in the context of a better selection of treatment responders, risk prediction, and design of disease-modifying strategies. Progress has been made in profiling the type 2 immune response-driven asthma. The endotype driven approach for non-type 2 immune response asthma, rhinitis, and atopic dermatitis is lagging behind. Validation and qualification of biomarkers are needed to facilitate their translation into pathway-specific diagnostic tests. Wide consensus between academia, governmental regulators, and industry for further

development and application of precision medicine in management of allergic diseases is of utmost importance. Improved knowledge of disease pathogenesis together with defining validated and qualified biomarkers are key approaches to precision medicine. (*J Allergy Clin Immunol* 2016;137:1347-58.)

Key words: Precision medicine, personalized care, phenotype, endotype, biomarker, allergic rhinitis, allergic asthma, allergic skin disease

Since the beginning of medicine, patients with similar clinical characteristics, presently termed phenotypes, have been grouped and treated similarly according to the experience of the clinician and, subsequently, evidence-based medicine. However, many patients might not respond to therapy that is considered the standard of care, reinforcing the concept that “one size does not fit

From ^aFood Allergy Referral Centre Veneto Region, Department of Women and Child Health, Padua General University Hospital, Padua; ^bthe Department of Pediatrics, University of Wisconsin School of Medicine and Public Health, Madison; ^cthe Department of Otorhinolaryngology, University Hospitals Leuven; ^dthe Swiss Institute of Allergy and Asthma Research, University of Zurich, Christine Kühne-Center for Allergy Research and Education, Davos; ^ethe Department of Dermatology and Allergy, Christine Kühne-Center for Allergy Research and Education, Friedrich-Wilhelms-University, Bonn; ^fthe Department of Internal Medicine, University of South Florida, Tampa; ^gthe Department of Clinical Immunology, Wroclaw Medical University, and ALL-MED Medical Research Institute, Wroclaw; ^hthe Division of Clinical Immunology and Allergy, Children’s Hospital Los Angeles, and the Department of Pediatrics, Keck School of Medicine, University of Southern California, Los Angeles; ⁱAllergy Clinic Copenhagen University Hospital at Gentofte, Copenhagen; ^jthe Allergy Unit, Department for Dermatology, University of Zurich, Zurich, Switzerland, Christine Kühne-Center for Allergy Research and Education, Davos; ^kthe Institute of Pharmacology, University of Bern; ^lthe Laboratory of Clinical Immunology, University of Leuven; and ^mthe Department of Allergy and Clinical Immunology, Transylvania University, Brasov.

Disclosure of potential conflict of interest: C. A. Akdis serves as a consultant from Actellion, Aventis, Stallergenes, Allergopharma, Circacia; and receives research support from Novartis, The European Commission, Swiss National Science Foundation, and Christine Kühne-Center for Allergy Research. T. Bieber is a member of the board of Astellas, Novartis, L’Oréal, Sanofi, Regeneron, Bioderma, Pfizer, Galderma, and Chugai and serves as a consultant for Astellas, Novartis, L’Oréal, Sanofi,

Regeneron, Bioderma, Pfizer, Galderma, and Chugai; receives payment for lectures from Astellas, Novartis, L’Oréal, Sanofi, Regeneron, and Bioderma. T. B. Casale serves as a consultant for Novartis, Genentech, Teva and AstraZeneca; receives grant funding from Astra Zeneca, Novartis, Genentech, and Sanofi-Regeneron; and is the Executive Vice President of the American Academy of Allergy, Asthma & Immunology. M. Jutel serves as a consultant from Anergis SA and Allergopharma/Merck. L. Poulsen serves as a consultant for Novozymes; receives research support from the EU Commission; and receives travel support from the EAACI. P. Schmid-Grendelmeier serves on the board for Menarini and Novartis. H.-U. Simon receives travel support from the EAACI. S. F. Seys receives travel support from the EAACI and receives research support from Fund for Scientific Research Flanders. The rest of the authors declare that they have no relevant conflicts of interest.

Received for publication January 25, 2016; revised March 14, 2016; accepted for publication March 15, 2016.

Corresponding author: Antonella Muraro, MD, Food Allergy Centre Department of Women and Child Health, Padua General University Hospital, Padua, Italy. E-mail: muraro@centroallergialeimentari.eu.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections

0091-6749

© 2016 The Authors. Published by Elsevier Inc. on behalf of American Academy of Allergy, Asthma & Immunology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

<http://dx.doi.org/10.1016/j.jaci.2016.03.010>

Abbreviations used

AD:	Atopic dermatitis
AR:	Allergic rhinitis
CRTH2:	Chemoattractant receptor-homologous molecule expressed on T _H 2 cells
FENO:	Fraction of exhaled nitric oxide
NO:	Nitric oxide
TSLP:	Thymic stromal lymphopoietin

all” and encouraging the scientific community to unravel the pathophysiologic mechanisms causing the disease.

Currently, it is generally accepted that the clinical differences in treatment responses or disease course over time are related to underlying variations in genetic, pharmacologic, physiologic, biologic, and/or immunologic mechanisms that produce subclasses of phenotypes termed endotypes.¹ This endotype-driven observed heterogeneity in therapeutic response has led to the use of terms, such as precision or personalized medicine (among others), to direct therapy more specifically, when possible. For example, although the phenotype of anemia presents clinically with pallor related to low red blood cell indices, the underlying endotypes responsible for this phenotype are multiple (eg, iron deficiency, G6PD deficiency, and autoimmune disease among others). Thus, for anemia, defining the underlying endotype is critical in more precisely choosing any therapeutic intervention.

To evaluate the latest findings in precisely defining the endotypic profile of the allergic and/or asthmatic patient and the potential for the specialty of allergy/immunology to use this precision medicine approach, the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology have conducted a project focused on this topic. The previously successful PRACTALL approach, in which a panel of experts from these 2 geographic regions reviewed the literature and harmonized the evidence that supported the particular topic being analyzed, was used to conduct these analyses.²

The focus for this PRACTALL was an examination of the potential benefits of applying the concepts of precision medicine to first airway and skin allergic diseases. A second PRACTALL paper soon to be published will cover the precision medicine approach for food allergy and anaphylaxis. Although a number of terms have been used to define this type of approach, the consensus of the writing groups was to use the term precision medicine. As such, according to the National Institutes of Health, precision medicine is an emerging approach for disease treatment and prevention that takes into account individual variability in genes, environment, and lifestyle for each person.¹

PRECISION MEDICINE AT THE LOWER AIRWAYS: ASTHMA

The heterogeneity of asthma in relation to patients’ characteristics (phenotype), underlying pathogenic mechanisms (endotype), and clinically significant outcomes, including response to treatment, has been established beyond any doubt.^{3–6} Better asthma management needs a refined understanding of disease heterogeneity and mechanisms in relation to clinically significant outcomes.

Extended heterogeneous disease-related metabolic, inflammatory, immunologic, and remodeling pathways have been described, and a stable pattern is defined as a disease endotype.

A well-defined endotype should link the key pathogenic mechanism with a clinical phenotype of asthma through biomarkers.^{7,8} There are several benefits of endotyping in a clinical setting, such as stringent consideration of entry criteria for epidemiologic, genetic, or therapeutic trials.

Defining asthma endotypes

Generally, it is considered that a type 2 immune response underlies atopic asthma.⁹ Eosinophilic airway inflammation and an increase in type 2 cytokine levels (eg, induced sputum, bronchoalveolar lavage fluid, and bronchial biopsy specimens) are characteristic of these patients.^{10–15} The type 2 immune response endotype has been related to response to inhaled corticosteroids^{9,16} and disease outcomes, such as exacerbations.^{17,18}

Several subendotypes can exist within the type 2 complex endotype, such as the IL-5–high, IL-13–high, or IgE-high endotypes.¹⁹ Aspirin-exacerbated respiratory disease is also a particular subtype of the type 2 complex endotype, where the hyperactive metabolic pathway shapes the type 2 immune response.²⁰ In this view type 2 immune response endotypes are defined by subgroups of patients who have a beneficial response to treatment targeting the IL-5, IL-13, or IgE pathogenic pathways (Fig 1, A). The type 2 complex endotype can also be identified in patients with allergic rhinitis (AR) as a fundament for the united airways disease concept (Fig 1, B).

Both the innate and acquired immune responses contribute to type 2 immune response endotypes (Fig 1, A). T_H1/T_H17 inflammatory cells^{21–25} and nonallergic mechanisms, such as environmental factors, psychosocial stress, activation of metabolic pathways,^{26–28} resident cells in the remodeled phenotype,^{29,30} or epithelial barrier dysfunction,³¹ further modulate the profile of type 2–driven inflammation. In addition, type 2–driven inflammation is characterized by a high cellular plasticity that enables the cells to adapt to a specific inflammatory milieu. Innate immune response cytokines, such as IL-33 and thymic stromal lymphopoietin (TSLP), modulate the mast cell–driven phenotype, whereas type 2 cytokines promote a particular phenotype involving smooth muscle cells and epithelial and endothelial cells in asthmatic patients. The latter also influence the permissiveness of the epithelium for allergens and of the endothelium for the recruitment of inflammatory cells to inflamed tissues and mucus production.^{32–34}

The mechanisms contributing to the non–type 2 immune response in asthmatic patients are less clear (Fig 2). Two major mechanisms leading to neutrophilic inflammation are postulated: (1) the dysregulated innate immune response, including neutrophil-intrinsic abnormalities, and (2) activation of the IL-17–dependent pathway.^{15,16,35–39} In addition, type 1 immune responses might contribute to asthma severity: high IFN- γ levels in sputum of asthmatic patients have been associated with severe asthma.^{13,40} Several factors, such as metabolic or epigenetic factors, or activation of the epithelial-mesenchymal trophic unit have been identified as modulators. The endotyping of non–type 2 immune response asthma lags behind that of type 2 immune response asthma, and until now, no endotype-driven interventions have been proved effective.

Asthma biomarkers

Currently identified asthma biomarkers are used to predict treatment response in patients with type 2 immune

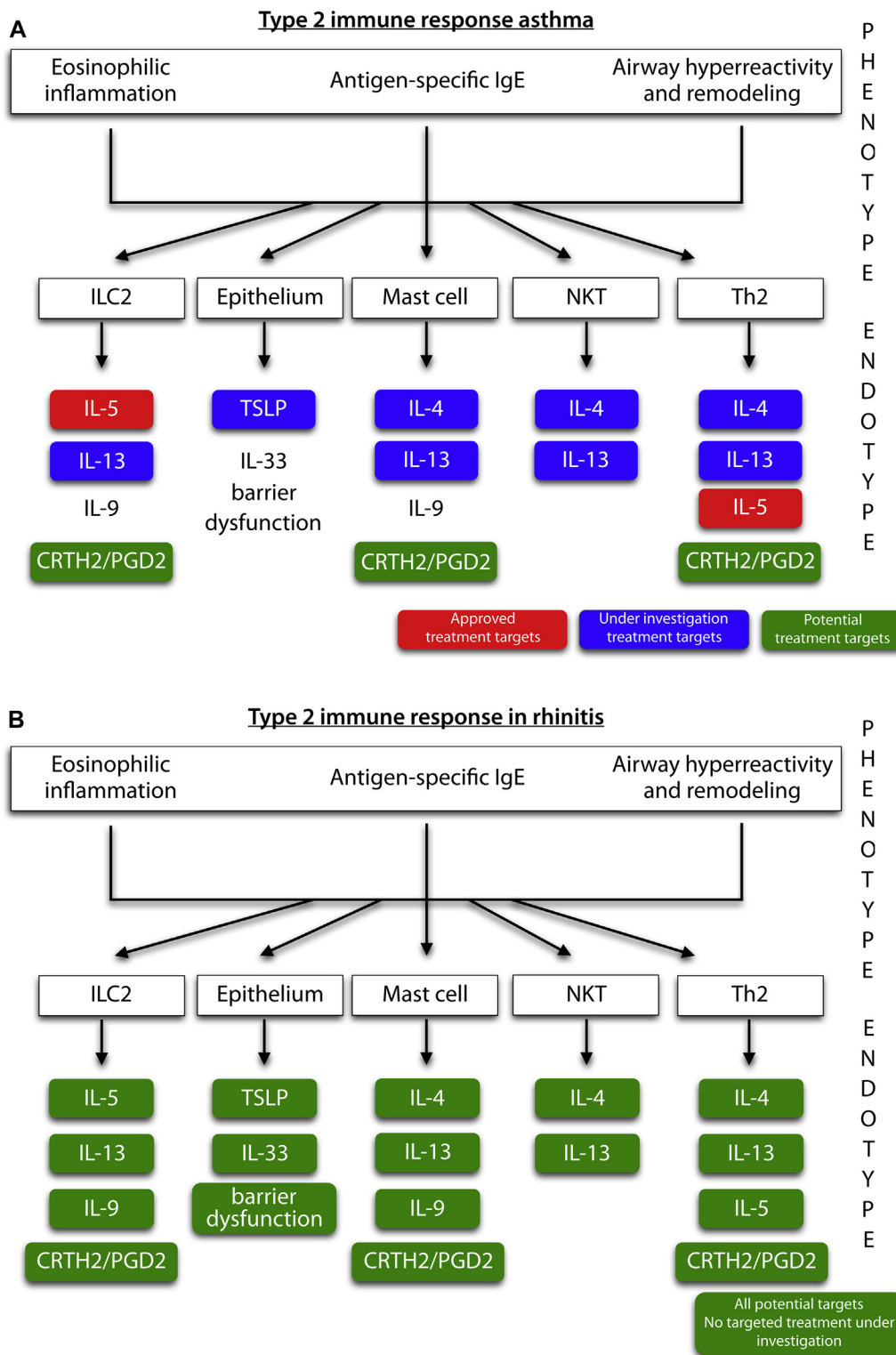


FIG 1. A, Overview of the type 2 immune response in asthmatic patients. Three main phenotypes of type 2 immune response–driven asthma are described: eosinophilic inflammation; allergic sensitization, as depicted by the presence of antigen-specific IgE; and airway hyperreactivity and remodeling. Both the innate and acquired immune responses contribute to type 2 immune response endotypes. Endotype-driven asthma management targets most of the molecular pathways involved in type 2 immune response asthma: *green*, approved treatment targets for asthma; *blue*, under investigation; *red*, potential treatment targets. **B**, Overview of the type 2 immune response in patients with rhinitis. Three main phenotypes of rhinitis are described, which are similar to those of asthma, with the exception of remodeling. Different cellular and molecular players contribute to type 2 immune responses in patients with rhinitis. In contrast to asthma, none of these molecular pathways are under investigation for targeted treatment. *ILC*, Innate lymphoid cell; *NKT*, natural killer T cell; *PGD2*, prostaglandin D₂.

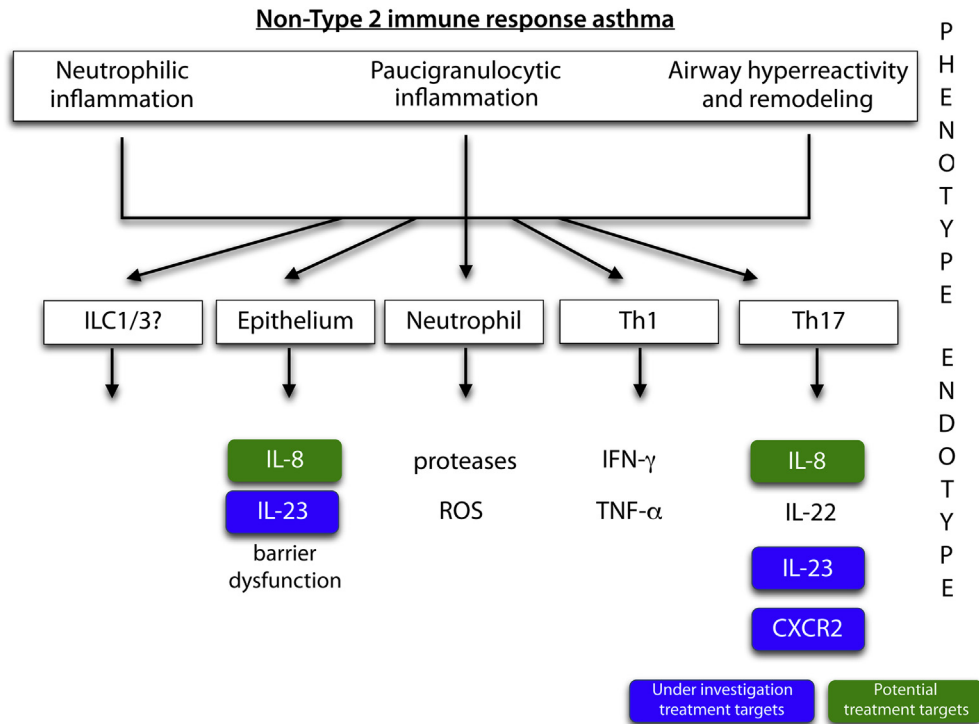


FIG 2. Overview of non-type 2 immune response in asthmatic patients. Three main phenotypes of non-type 2 immune response asthma can be described based on the inflammation pattern and the presence of airway hyperreactivity and remodeling. Some of the described molecular pathways are under investigation (blue) for an endotype-driven approach. Potential treatment targets are indicated in red. *ILC*, Innate lymphoid cell; *ROS*, reactive oxygen species.

response-driven inflammation (Table I). It should be noted that most asthma biomarkers are currently used in research settings and still need to be validated and qualified. A valid biomarker is defined as “a biomarker that is measured in an analytical test system with well-established performance characteristics and for which there is an established scientific framework or body of evidence that elucidates the physiologic, toxicologic, pharmacologic, or clinical significance of the test results.”⁴¹ Validation is the process of assessing the biomarker and its performance characteristics and determining the range of conditions under which the biomarker will produce reproducible and accurate data. Qualification is the evidentiary process of linking a biomarker with biological processes and clinical end points.⁴²⁻⁴⁴

Blood eosinophilia is a well-demonstrated biomarker of type 2 immune response-driven inflammation in asthmatic patients and has been linked to response to corticosteroids and, more recently, anti-IL-4/IL-13-targeted⁴⁵ and anti-IL-5-targeted^{46,47} treatment. Its correlation to sputum or bronchial eosinophilia cannot always be demonstrated; thus blood and airway eosinophils cannot be used interchangeably because they might reflect different type 2 subendotypes. Sputum eosinophil levels have also been useful for predicting response to inhaled steroids¹⁶ and anti-IL-13 and anti-IL-5 therapy.^{17,18,46-48}

The periostin gene has been identified as an IL-13-inducible gene in bronchial brushings from asthmatic patients.^{9,49} Periostin expression in bronchial tissue has been shown to be a biomarker of eosinophilic airway inflammation,⁵⁰ whereas serum periostin levels have been related to the response to anti-IL-13 therapy in patients with mild-to-moderate asthma.⁵¹

Serum dipeptidyl peptidase 4 has also been shown to predict responses to anti-IL-13 therapy.⁵² In another study sputum IL-13 levels were used to identify responders to anti-IL-13 treatment.⁵³

In a *post hoc* analysis a composite biomarker combining blood eosinophils, periostin, and fraction of exhaled nitric oxide (FENO) identified anti-IgE mAb omalizumab responders.⁵⁴ Recent data suggest that blood eosinophils alone might be a useful biomarker to predict responses to omalizumab.⁵⁵

Biomarkers measured in exhaled breath are of particular interest because of their noninvasive character. In steroid-naïve asthmatic patients FENO values correlated well with eosinophilic airway inflammation. Breath analysis by using eNose (volatile organic compounds in exhaled breath) can identify asthmatic patients and can be used to predict their response to steroids with greater accuracy than sputum eosinophil counts or FENO values.^{56,57}

There are several biomarkers predicting poor steroid response in asthmatic patients, such as p38 and MSK1 phosphorylation status of blood monocytes, vanin-1 expression and CpG methylation, the presence of T_H2/T_H17 double-producing cells in bronchoalveolar lavage fluid, and airway expansion of specific gram-negative bacteria. A corticosteroid-responsive endophenotype was recently described.⁵⁸⁻⁶³

Endotype-driven asthma treatment

Early clinical trials with anticytokine therapies in asthmatic patients were not successful because of inclusion of unselected patients. As an example, anti-IL-5 therapy in unselected patients

TABLE I. Asthma biomarkers guiding tailored treatment approaches

Biomarker	Treatment expected to produce a response	Associations	Comments (point of care, variability/fluctuation)
Blood			
Eosinophils	Anti-IL-5	Exacerbations	Easily available
	Anti-IgE Anti-IL-4/IL-13 Corticosteroids CRTH2 antagonists	LF decrease Fixed airway obstruction	Significant fluctuation
Specific IgE	Anti-IgE AIT	Exacerbations AHR (AIT)	
Periostin	Anti-IL-13	LF decline	Research type
DPP-4		Exacerbations	Assay dependent
Induced sputum			
Eosinophils	Anti-IL-5	Exacerbations	Research type
	ICS		Significant fluctuation
IL-13	Anti-IL-13	Unknown	Research type
Exhaled breath			
FENO	Anti-IL-5	Exacerbations, LF decrease	Easily available
	Anti-IgE Anti-IL-13 ICS		Significant fluctuation
Metabolomics (VOC)	ICS	Unknown	Research type

There is significant overlap between biomarkers used to predict response to different endotype-driven strategies. In addition, few biomarkers are easily available, most are subject to significant fluctuation, and none are validated and qualified.

AIT, Allergen immunotherapy; DPP-4, dipeptidyl peptidase 4; ICS, inhaled corticosteroids; LF, lung function; VOC, volatile organic compounds.

TABLE II. Endotype-driven treatment in type 2 immune response-driven asthma

Predictive biomarker	Drug	Target	Effects	Regulatory status
Blood eosinophils Periostin FENO	Omalizumab	IgE	Reduces exacerbations Improves symptoms and quality of life	FDA and EMA approved
Blood/sputum eosinophils FENO	Mepolizumab	IL-5	Reduces eosinophil counts, exacerbations, and OCS Improves FEV ₁	FDA approved EMA under consideration Tested for CRSwNP
Blood eosinophils	Reslizumab	IL-5	Reduces eosinophil counts, exacerbations Improves FEV ₁	FDA under consideration
Blood eosinophils	Benralizumab	IL-5Rα	Reduces eosinophil and basophil counts, exacerbations Improves FEV ₁	Phase III
Blood eosinophils	Dupilumab	IL-4Rα	Reduces exacerbations Improves FEV ₁ Improves symptoms and quality of life	Phase III Tested for CRSwNP, AD, and EoE
Periostin DPP-4	Tralokinumab	IL-13	Reduces eosinophil counts and exacerbations Improves FEV ₁	Phase II
Periostin	Lebrikizumab	IL-13	Reduces exacerbations Improves FEV ₁	Phase III

The IgE, IL-5, and IL-4/IL-13 pathways can be targeted with mAbs. There is a remarkable overlap between the so-called predictive biomarkers and a significant heterogeneity in clinical response.

CRSwNP, Chronic rhinosinusitis with nasal polyps; DPP-4, dipeptidyl peptidase 4; EMA, European Medicines Agency; EoE, eosinophilic esophagitis; FDA, US Food and Drug Administration; IL-4Rα, IL-4 receptor α; IL-5Rα, IL-5 receptor α; OCS, oral corticosteroids.

(lack of evaluation for blood or sputum eosinophilia) did not show significant effects on asthma exacerbations or lung function improvement.^{64,65} A recent tailored approach selecting patients for anti-IL-5–targeted treatment based on their blood or sputum eosinophil counts proved to be more rewarding (Table II).

Several steps need to be taken into account when considering tailored therapy for asthmatic patients (Fig 3). Before assessment of a patient's phenotype and endotype, correct diagnosis of asthma should be ensured. Comorbidities need to be evaluated and treated properly. A crucial step is to unravel which pathophysiologic mechanism or mechanisms are driving the disease, thereby determining the patient's endotype. Translation of

biomarkers into pathway-specific diagnostic tests is essential and should guide the design of future large clinical trials incorporating both longitudinal and mechanism-tailored end points.

Many targeted treatments are in various stages of clinical development for patients with type 2 immune response-driven inflammation: anti-IL-4/IL-13, anti-IL-4, anti-IL-5, and anti-IgE antibodies, as well as CRTH2 antagonists (Fig 1 and Table II).^{66,67} At present, biomarkers are not sufficiently specific to select the subendotype of type 2 immune response asthma specifically responding to a targeted treatment (Table I). For example, blood eosinophils predicted response to anti-IL-4/IL-13, anti-IL-5, and anti-IgE antibodies, as well as CRTH2 antagonists, and the

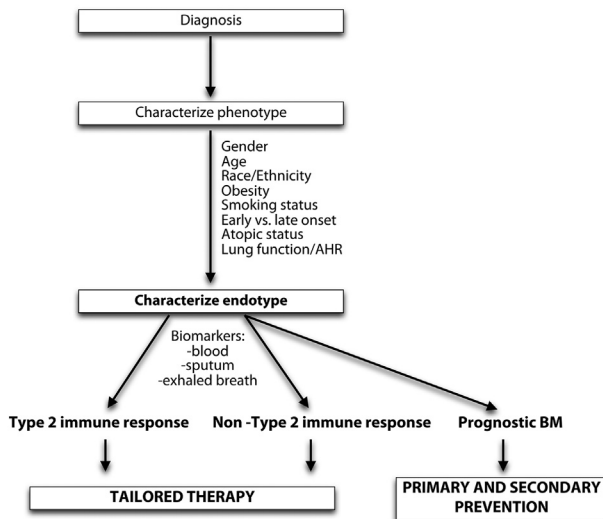
Suggested approach to precision medicine in asthma

FIG 3. Suggested approach to precision medicine in asthmatic patients. First, the correct diagnosis of asthma should be verified, and comorbidities should be treated properly. In a second step phenotype is established based on visible properties. Further characterization of the patient's endotype is crucial to ensure the optimum response to treatment and risk prediction, especially for those with severe and uncontrolled disease. Validation of prognostic biomarkers related to disease severity and risk prediction (including risk to develop asthma) open new pathways for primary and secondary asthma prevention. *AHR*, Airway hyperresponsiveness; *BM*, biomarkers.

clinician will face a conundrum of how best to treat patients with severe asthma with high blood eosinophil counts.⁶⁸

Limited data are available about the long-term efficacy of targeted treatment in asthmatic patients. For example, sputum eosinophil counts and exacerbation frequency increased as soon as 3 months after stopping treatment with the anti-IL-5 mAb mepolizumab.⁶⁹ Current targeted asthma interventions do not seem to influence the natural history of the disease or induce long-term remission.

Another limitation of targeted treatment in asthmatic patients is the interindividual and intraindividual variation in response, which is called the dissociated effect.⁷ This might be due to the (epi)genetic background, the predominant immune-inflammatory pathway, and the contribution from remodeled tissue. Drug efficacy at the target site adds to the observed variability in response.

In summary, recent therapeutic advances have unraveled some of the contributions of phenotype and endotype to the pathogenesis of asthma and the responses to specific therapies. However, more information is needed to better target specific pathways in subjects that will optimize the patient's therapeutic responses while avoiding adverse effects.

PRECISION MEDICINE AT THE UPPER AIRWAYS: RHINITIS

The current definition of rhinitis relies on the combination of history, clinical examination, and allergy diagnostic testing, which allows the distinction of 3 major subgroups: allergic, infectious, and nonallergic noninfectious rhinitis.^{70,71}

Rhinitis phenotypes were described in relation to the severity and duration of symptoms, major presenting symptoms, sensitization pattern, presence of comorbidities, and level of control

after treatment. Rhinitis phenotypes have been the basis of evidence-based treatment algorithms for rhinitis. A phenotype-based strategy for rhinitis implies a trial-and-error approach, with guidance of treatment based on the severity and duration of symptoms. As a consequence, a significant percentage of patients with AR have uncontrolled disease,⁷² highlighting the need for precision medicine in patients with AR. Precision medicine implicates endotype- rather than phenotype-driven treatment added to the prediction of successful therapy, prevention of disease, and participation of the patient.

The first step in the implementation of precision medicine in patients with rhinitis will be to characterize the endotype as a guide to a tailored therapeutic approach. It should be emphasized that patients with rhinitis might have a complex endotype and that the current understanding of cellular and molecular processes giving rise to a certain phenotype require further study. In addition, as described for asthma, there are several modulators of endotype expression, such as the environment, microbiome, lifestyle, and nasal anatomy.

The following endotypes of rhinitis are being proposed (Fig 4).^{73,74}

Type 2 immune response rhinitis

Mast cell-bound specific IgE is cross-linked by absorbed allergen molecules, leading to acute symptoms and influx into the nasal mucosa of eosinophils, basophils, and T and B lymphocytes. This is often accompanied by a systemic immune response dominated by type 2 cytokines produced by CD4⁺ T cells,⁷⁵ type 2 innate lymphoid cells, and basophils, which is associated with blood and nasal eosinophilia. The type 2 immune response endotype usually is attributed to AR⁷⁶; however, occupational/environmental low-molecular-weight substances leading to release of epithelially derived TSLP, IL-33, and IL-25, can initiate or aggravate a type 2 immune response.^{77,78}

Type 1 immune response rhinitis

An innate and adaptive type 1/IL-17 immune response leads to influx of neutrophils and IFN- γ -producing CD4⁺ T cells, usually as the background of infectious rhinitis.⁷⁹

Neurogenic rhinitis

This particular endotype is characterized by a relative overexpression of transient receptor potential channels on trigeminal nerves and high concentrations of substance P and neurokinins and is linked to gustatory rhinitis, rhinitis of the elderly, and idiopathic rhinitis with nasal hyperreactivity.⁸⁰

Epithelial dysfunction

Epithelial dysfunction can be primary or secondary to type 2 or type 1 immune response-induced inflammation. It can be divided roughly into the ciliary dysfunctional pathway (primary vs secondary) and the barrier dysfunctional pathway, with reduced expression of zonula occludens 1 and occludin-1 facilitating subepithelial migration of exogenous immune-stimulating molecules.⁸¹

Several other rhinitis phenotypes, such as drug-related, senile, and hormonal rhinitis, are poorly characterized by the lack of data on biomarkers and the molecular and cellular mechanisms involved.

In clinical practice efforts can be made in endotyping patients with rhinitis by measuring total and allergen-specific IgE levels

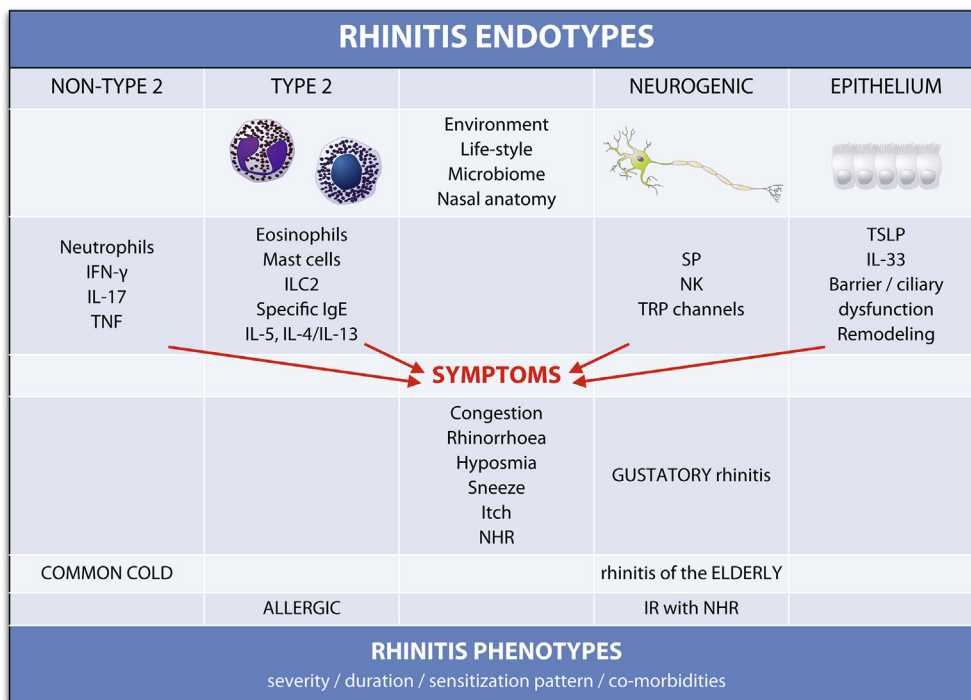


FIG 4. Overview of rhinitis phenotypes and endotypes. Similar to asthma, a type 2 immune response and non-type 2 immune response endotype can be described for rhinitis. Neurogenic and epithelial barrier dysfunction endotypes are particularly relevant for rhinitis. *ILC*, Innate lymphoid cell; *IR*, idiopathic rhinitis; *NHR*, nasal hyperreactivity; *NK*, neurokinin; *SP*, substance P; *TRP*, transient receptor potential.

and blood eosinophil, nasal eosinophil, and neutrophil counts. Several other biomarkers are used in research settings, such as serum IL-5, nasal total and allergen-specific IgE, eosinophil-derived neurotoxin, eosinophil cationic protein, eosinophil peroxidase, IL-5, substance P, neurokinin 1, IL-33, and TSLP levels and staining of mucosal biopsy specimens for TRPV-1, zonula occludens 1, or occludin.

These biomarkers should ideally be supplemented by nasal function measurements, such as nasal flow measurement (to confirm nasal obstruction) and cold dry air provocation (to determine nasal hyperreactivity), nasal nitric oxide measurement (to measure nasal inflammation), nasal allergen provocation (to confirm the clinical relevance of allergens), and evaluation of smell performance (in patients mentioning reduced smell capacity).

The best example of an endotype-driven treatment in rhinitis is the use of allergen-specific immunotherapy in patients in whom an allergen-induced type 2 immune response endotype leads to a clinically relevant exposure-symptom relation.^{82,83} Another example of endotype-driven treatment is the highly successful intervention with capsaicin for the neurogenic endotype.⁸⁰

Precision medicine represents the future of rhinitis care in patients whose symptoms are not fully controlled despite evidence-based treatment. Essential steps toward precision medicine in patients with rhinitis are described in [Table III](#).

PRECISION MEDICINE AT THE SKIN: ATOPIC DERMATITIS

Atopic dermatitis (AD) is a disease with a highly complex pathophysiology ([Fig 5](#)) and heterogeneous phenotypes, which

are illustrated by different features, such as age of disease onset, variable response to allergens, spectrum of severity, potential of IgE autoreactivity, and comorbidities (asthma, rhinitis, food allergy, and infections).⁸⁴

In the field of AD, in contrast to asthma, we are just in the beginning of the development of precision medicine and the attempt to reach a biomarker-based molecular taxonomy. We expect that the complexity of the clinical phenotype is underlined by even more complex profiles of possibly different pathophysiologic pathways⁸⁴⁻⁸⁶ from which we can learn and develop a strategy for discovery, validation, and qualification of biomarkers.⁸⁷

Precision medicine is of broad relevance for the management of AD, which is known to have a diverse natural history ranging from complete remission to relapsing flares to very severe and persistent forms variably associated with comorbidities, such as asthma and AR. Clearly, the discovery and validation of biomarkers with ideally prognostic and predictive value for AD represents a significant unmet need in this field.

The following endotypes of AD are being proposed ([Fig 6](#) and [Table IV](#)): (1) type 2 immune response AD, covering the whole disease spectrum from background inflammation in nonlesional skin to acute disease flares to chronic disease, peaking during acute flares, and (2) non-type 2 immune response AD mixing T_H1 -, T_H17 -, and T_H22 -driven inflammation and epithelial dysfunction.^{84,86,88,89}

In addition to the attempt to identify possible provocative factors, the current approach in AD management is still “one size fits all” based on use of emollients and anti-inflammatory drugs in all patients, although the disease provides a number of

TABLE III. Essential steps for applying precision medicine in patients with rhinitis

- Precise evaluation of the patient's perception of disease severity and effect of the disease on the patient's quality of life, as well as the social and general environment of the patient
- Clear-cut dissection of nasal pathophysiology into mucosal and structural components
- Rigorous assessment of inflammatory components (eg, eosinophilic vs neutrophilic inflammation, IgE, cytokines, and neural mediators) and functional effects (nasal hyperreactivity, smell, and patency)
- Correct evaluation of the risk for disease progression
- Proper information for the patient on the treatment strategy (monotherapy vs combined therapy), involving information on treatment goals, expected benefits and adverse events, and effects of treatment in the long-term together with evaluation of the patient's preference for a particular therapeutic plan

Type 2 and non-type 2 immune responses are common pathogenic pathways and disease endotypes for asthma, rhinitis, and AD. Epithelial dysfunction is of particular relevance for describing disease endotypes in patients with rhinitis and AD, whereas the neurogenic pathway is most prominent for rhinitis.

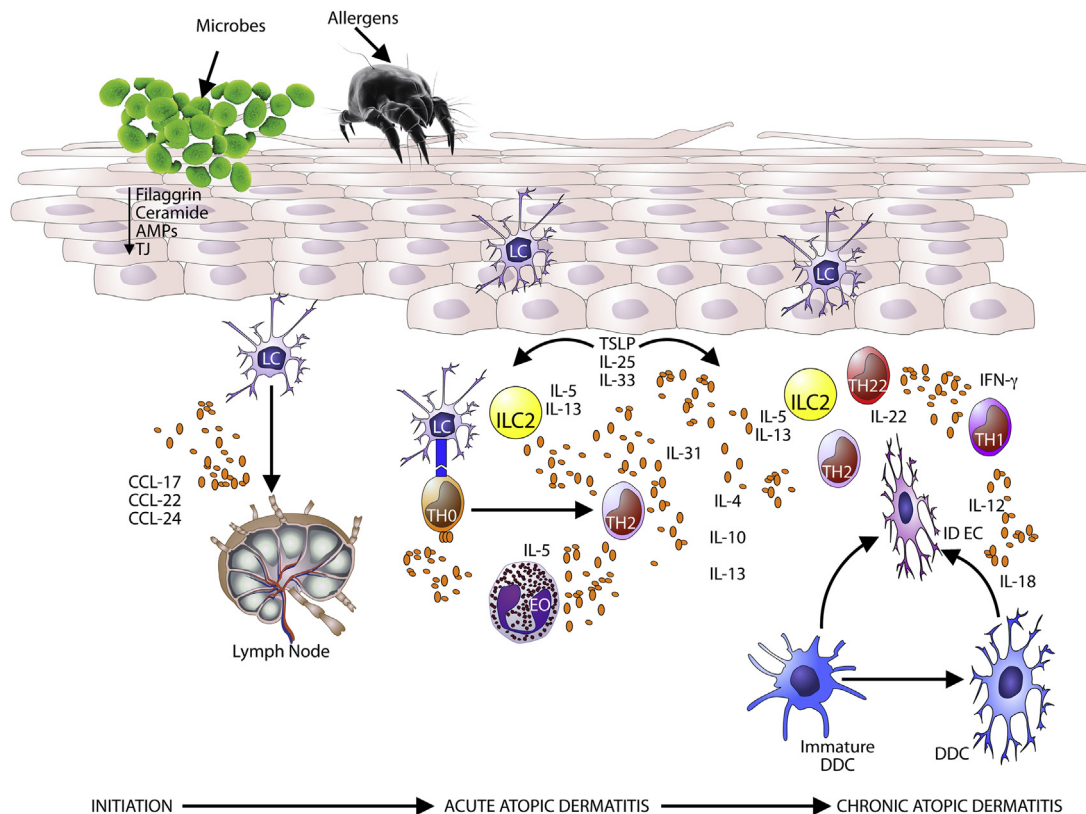


FIG 5. Pathogenesis of AD. The complexity of the clinical phenotype in patients with AD is underlined by complex profiles of different pathophysiologic pathways connecting the innate and the adaptive immune response with epithelial barrier dysfunction and allergic sensitization. *AMPs*, Antimicrobial peptides; *DDC*, dermal dendritic cell; *Eo*, eosinophil; *ID EC*, inflammatory dendritic epidermal cell; *ILC*, innate lymphoid cells; *LC*, Langerhans cell; *TJ*, tight junctions.

opportunities for more personalized management.^{90,91} Thus far, there is no clear evidence for targeted therapy for any kind of approved anti-inflammatory treatment regimen in patients with AD. However, with the emergence of biologics targeting well-defined cytokines and pathways, such as anti-IL-4/IL-13 or anti-IL-31,^{92,93} the need for predictive biomarkers of therapeutic response has to be reconsidered.

Biomarkers could be useful in the management of early-onset disease at different time points throughout the natural history of AD (Fig 7).⁹⁴ Some biomarkers, such as CCL17, have been shown to be a consistent measurement of AD severity in multiple clinical trials. Also, filaggrin deficiency as a potential candidate for prognosis and indoleamine 2,3-dioxygenase as a predictive marker for

viral skin infections leading to eczema herpeticum have been demonstrated.⁹⁵ It is also a common phenomenon to see multiple allergen-specific IgE sensitizations, particularly in patients with moderate-to-severe disease, but their clinical relevance is often questionable for avoidance strategies. It is highly probable that multiple biomarkers will be needed as a signature profile in AD to predict the severity, comorbidities, and treatment response.

Two recent proof-of-concept studies showed that 6 to 8 months of skin barrier therapy prevents the development of AD during this period of time in a significant portion (30% to 50%) of infants born to parents with a history of atopy.^{96,97} This suggests an opportunity for early intervention with a positive effect on the emergence of AD and possibly on the “atopic march,” thus

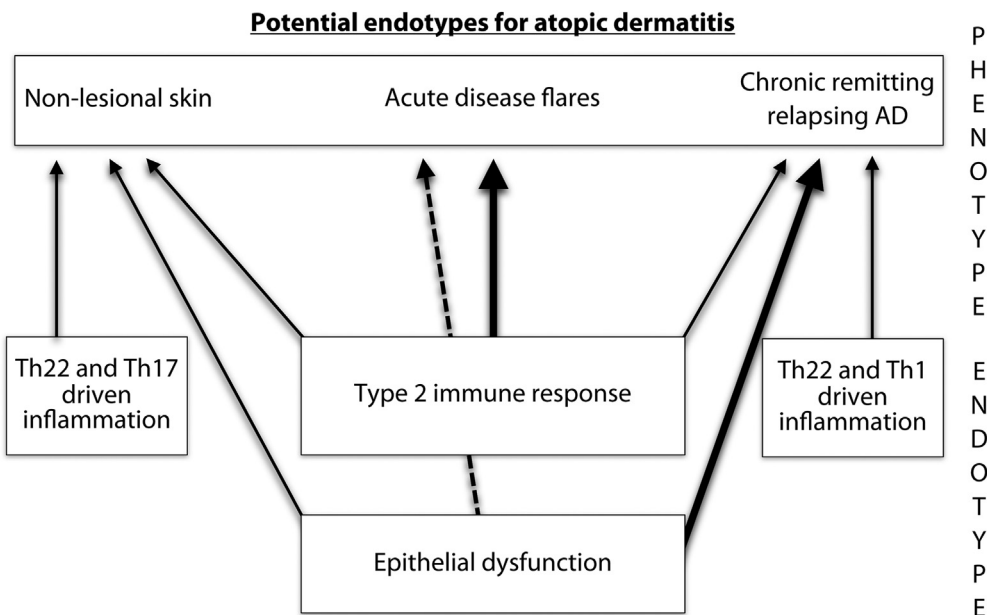


FIG 6. Proposed endotypes for AD. Three main phenotypes of AD are described: nonlesional skin, acute disease flares, and chronic remitting relapsing AD. A type 2 immune response is present in all 3 phenotypes, with a peak in acute disease flares. T_H22 - and T_H17 -driven inflammation adds to the type 2 immune response in the dysregulated immune response present in nonlesional skin, whereas T_H22 - and T_H1 -driven inflammation is prominent in patients with the chronic form of AD. Epithelial dysfunction is a key mechanism partnering with the dysregulated immune response in nonlesional skin and in patients with chronic AD and facilitates acute disease flares.

TABLE IV. Proposed endotypes of asthma, AR, and AD

Asthma	AR	AD
Type 2 immune response	Type 2 immune response	Type 2 immune response
Non-type 2 immune response	Non-type 2 immune response	Non-type 2 immune response
	Epithelial dysfunction Neurogenic	Epithelial dysfunction

representing a disease-modifying strategy.⁹⁰ The selection of these high-risk patients was based solely on family history. The outcome could be substantially improved if we used validated biomarkers to select those infants (the right patient) with high risk assessed not only based on family history but also on biomarker signature.⁹¹ Moreover, it is expected that the early improvement (the right time) of the barrier dysfunction could be substantially enhanced if we have appropriate new emollients (the right drug), including ingredients able to support barrier function, given in the optimal frequency amount (the right dose). These new products and innovative ingredients could be based on the availability of biomarkers unraveling the individual pathophysiologic origin of the barrier dysfunction in a given patient subgroup.

CONCLUSION AND FUTURE PERSPECTIVES

Precision medicine is of broad relevance for the management of asthma, rhinitis, and AD from a better selection of responders to

treatment and design of better clinical trials to risk prediction and disease-modifying strategies. In this PRACTALL we summarized the current knowledge on major asthma, rhinitis, and AD endotypes (Table IV).

For asthma, several steps have been taken in profiling the type 2 immune response-driven asthma, together with endotype-driven strategies. However, more information is needed to better target specific pathways in patients that will optimize patients’ therapeutic responses while avoiding adverse effects. Endotype-driven management of non-type 2 immune response asthma, rhinitis, and AD is clearly an unmet need in the field.

In addition, most biomarkers are currently used in research settings and still need to be validated and qualified. Asthma, rhinitis, and AD biomarkers are complicated by remarkable heterogeneity compared with specific cancer biomarkers. This complexity includes different patterns of onset and clinical presentation and marked variations in the rate of disease remission or progression, together adding to the considerable challenge both in determining the appropriate clinical outcome and in delineating efficacy biomarkers.

A strategy for biomarker validation and qualification needs to be created, including development of reference laboratories and clinical epidemiology and validation centers, as well as networks of cooperative human tissue banks or resources. Open interaction among steering committees of large trials and large cohort studies should be encouraged for the free exchange of ideas and specimens.

Improved knowledge of the pathogenesis of asthma, rhinitis, and AD and information-relating biomarkers with clinically relevant outcomes will permit a better means for assessment of the effects of new interventions. It is evident that there is a shared

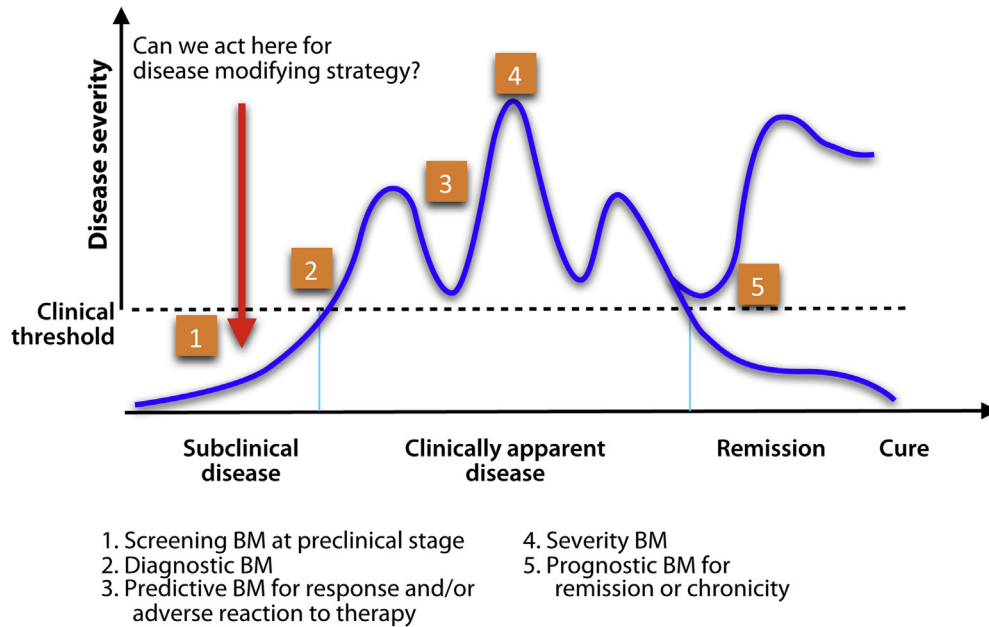


FIG 7. The concept of longitudinal biomarkers (BM) in the management of AD at different time points throughout the natural history of the disease. Early stage BM (stages 1 and 2) allows screening at the preclinical stage and primary prevention of the disease together with early diagnosis. During the clinical course of the disease, biomarkers can predict responses, adverse reactions, or both to treatment and can guide targeted, endotype-driven interventions with an improved safety profile. Prognostic biomarkers relate to disease severity, disease flares, or occurrence of remission.

recognition between academia, government regulators, and industry regarding the need for both the development and application of precision medicine in patients with asthma, rhinitis, and AD.⁶⁶ This is a path other disease areas have taken, and there are experiences, processes, and infrastructure mechanisms in existence on which we can build.

Clinical implications: Improved knowledge of the pathogenesis of asthma, rhinitis, and AD leads to the concept of disease endotypes, thus supporting the potential for the specialty of allergy/immunology to use the precision medicine approach. After a correct diagnosis and proper management of comorbidities, a crucial step is to unravel which pathophysiologic mechanism or mechanisms are driving the disease, thereby determining the endotype of the patient and providing validated pathway-specific diagnostic tests.

REFERENCES

- Collins FS, Varmus H. A new initiative on precision medicine. *N Engl J Med* 2015;372:793-5.
- Akdis CA, Akdis M, Bieber T, Bindslev-Jensen C, Boguniewicz M, Eigenmann P, et al. Diagnosis and treatment of atopic dermatitis in children and adults: European Academy of Allergy and Clinical Immunology/American Academy of Allergy, Asthma and Immunology/PRACTALL Consensus Report. *Allergy* 2006; 61:969-87.
- Agache I, Akdis C, Jutel M, Virchow JC. Untangling asthma phenotypes and endotypes. *Allergy* 2012;67:835-46.
- Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. *Nat Med* 2012;18:716-25.
- Fahy JV. Type 2 inflammation in asthma—present in most, absent in many. *Nat Rev Immunol* 2015;15:57-65.
- Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol* 2015;16: 45-56.
- Agache IO. From phenotypes to endotypes to asthma treatment. *Curr Opin Allergy Clin Immunol* 2013;13:249-56.
- Fajt ML, Wenzel SE. Asthma phenotypes and the use of biologic medications in asthma and allergic disease: the next steps toward personalized care. *J Allergy Clin Immunol* 2015;135:299-311.
- Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, et al. T-helper type 2-driven inflammation defines major subphenotypes of asthma. *Am J Respir Crit Care Med* 2009;180:388-95.
- Brightling CE, Symon FA, Birring SS, Bradding P, Pavord ID, Wardlaw AJ. TH2 cytokine expression in bronchoalveolar lavage fluid T lymphocytes and bronchial submucosa is a feature of asthma and eosinophilic bronchitis. *J Allergy Clin Immunol* 2002;110:899-905.
- Cho S-H, Stanciu LA, Holgate ST, Johnston SL. Increased interleukin-4, interleukin-5, and interferon-gamma in airway CD4+ and CD8+ T cells in atopic asthma. *Am J Respir Crit Care Med* 2005;171:224-30.
- Berry MA, Parker D, Neale N, Woodman L, Morgan A, Monk P, et al. Sputum and bronchial submucosal IL-13 expression in asthma and eosinophilic bronchitis. *J Allergy Clin Immunol* 2004;114:1106-9.
- Truyen E, Coteur L, Dilissen E, Overbergh L, Dupont LJ, Ceuppens JL, et al. Evaluation of airway inflammation by quantitative Th1/Th2 cytokine mRNA measurement in sputum of asthma patients. *Thorax* 2006; 61:202-8.
- Saha SK, Berry MA, Parker D, Siddiqui S, Morgan A, May R, et al. Increased sputum and bronchial biopsy IL-13 expression in severe asthma. *J Allergy Clin Immunol* 2008;121:685-91.
- Peters MC, Mekonnen ZK, Yuan S, Bhakta NR, Woodruff PG, Fahy JV. Measures of gene expression in sputum cells can identify TH2-high and TH2-low subtypes of asthma. *J Allergy Clin Immunol* 2014;133:388-94.
- Green RH, Brightling CE, Woltmann G, Parker D, Wardlaw AJ, Pavord ID. Analysis of induced sputum in adults with asthma: identification of subgroup with isolated sputum neutrophilia and poor response to inhaled corticosteroids. *Thorax* 2002;57:875-9.
- Haldar P, Brightling CE, Hargadon B, Gupta S, Monteiro W, Sousa A, et al. Mepolizumab and exacerbations of refractory eosinophilic asthma. *N Engl J Med* 2009;360:973-84.
- Nair P, Pizzichini MM, Kjarsgaard M, Inman MD, Efthimiadis A, Pizzichini E, et al. Mepolizumab for prednisone-dependent asthma with sputum eosinophilia. *N Engl J Med* 2009;360:985-93.

19. Agache I, Sugita K, Morita H, Akdis M, Akdis C. The complex type 2 endotype in allergy and asthma: from laboratory to bedside. *Curr Allergy Asthma Rep* 2015;15:29.
20. Rajan JP, Wineinger NE, Stevenson DD, White AA. Prevalence of aspirin-exacerbated respiratory disease among asthmatic patients: a meta-analysis of the literature. *J Allergy Clin Immunol* 2015;135:676-81.e1.
21. Steinke JW, Liu L, Huyett P, Negri J, Payne SC, Borish L. Prominent role of IFN- γ in patients with aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol* 2013;134:856-65, e1-3.
22. Seys SF, Grabowski M, Adriaenssens W, Decraene A, Dilissen E, Vanoirbeek JA, et al. Sputum cytokine mapping reveals an "IL-5, IL-17A, IL-25-high" pattern associated with poorly controlled asthma. *Clin Exp Allergy* 2013;43:1009-17.
23. Marijse GS, Seys SF, Schelpe A-S, Dilissen E, Goeminne P, Dupont LJ, et al. Obese individuals with asthma preferentially have a high IL-5/IL-17A/IL-25 sputum inflammatory pattern. *Am J Respir Crit Care Med* 2014;189:1284-5.
24. Lindén A, Dahlén B. Interleukin-17 cytokine signalling in patients with asthma. *Eur Respir J* 2014;44:1319-31.
25. Chambers ES, Nanzer AM, Pfeffer PE, Richards DF, Timms PM, Martineau AR, et al. Distinct endotypes of steroid-resistant asthma characterized by IL-17A(high) and IFN- γ (high) immunophenotypes: potential benefits of calcitriol. *J Allergy Clin Immunol* 2015;136:628-37.e4.
26. Conus S, Bruno A, Simon H-U. Leptin is an eosinophil survival factor. *J Allergy Clin Immunol* 2005;116:1228-34.
27. Holguin F, Comhair SAA, Hazen SL, Powers RW, Khatri SS, Bleecker ER, et al. An association between L-arginine/asymmetric dimethyl arginine balance, obesity, and the age of asthma onset phenotype. *Am J Respir Crit Care Med* 2013;187:153-9.
28. Cowan DC, Taylor DR, Peterson LE, Cowan JO, Palmay R, Williamson A, et al. Biomarker-based asthma phenotypes of corticosteroid response. *J Allergy Clin Immunol* 2015;135:877-83.e1.
29. Xie Y, Jiang H, Nguyen H, Jia S, Berro A, Panettieri RA, et al. Regulator of G protein signaling 2 is a key modulator of airway hyperresponsiveness. *J Allergy Clin Immunol* 2012;130:968-76.e3.
30. Holgate ST. Epithelium dysfunction in asthma. *J Allergy Clin Immunol* 2007;120:1233-46.
31. Xiao C, Puddicombe SM, Field S, Haywood J, Broughton-Head V, Puxeddu I, et al. Defective epithelial barrier function in asthma. *J Allergy Clin Immunol* 2011;128:549-56, e1-12.
32. Dougherty RH, Sidhu SS, Raman K, Solon M, Solberg OD, Caughey GH, et al. Accumulation of intraepithelial mast cells with a unique protease phenotype in TH2-high asthma. *J Allergy Clin Immunol* 2010;125:1046-53.e8.
33. Han N-R, Oh H-A, Nam S-Y, Moon P-D, Kim D-W, Kim H-M, et al. TSLP induces mast cell development and aggravates allergic reactions through the activation of MDM2 and STAT6. *J Invest Dermatol* 2014;134:2521-30.
34. Soyka MB, Wawrzyniak P, Eiwegger T, Holzmann D, Treis A, Wanke K, et al. Defective epithelial barrier in chronic rhinosinusitis: the regulation of tight junctions by IFN- γ and IL-4. *J Allergy Clin Immunol* 2012;130:1087-96.e10.
35. Bullens DM, Truyen E, Coteur L, Dilissen E, Hellings PW, Dupont LJ, et al. IL-17 mRNA in sputum of asthmatic patients: linking T cell driven inflammation and granulocytic influx? *Respir Res* 2006;7:135.
36. Pene J, Chevalier S, Preisser L, Venereau E, Guilleux M-H, Ghannam S, et al. Chronically inflamed human tissues are infiltrated by highly differentiated Th17 lymphocytes. *J Immunol* 2008;180:7423-30.
37. Simpson JL, Gibson PG, Yang IA, Upham J, James A, Reynolds PN, et al. Impaired macrophage phagocytosis in non-eosinophilic asthma. *Clin Exp Allergy* 2013;43:29-35.
38. Baines KJ, Simpson JL, Wood LG, Scott RJ, Fibbens NL, Powell H, et al. Sputum gene expression signature of 6 biomarkers discriminates asthma inflammatory phenotypes. *J Allergy Clin Immunol* 2014;133:997-1007.
39. Raedler D, Ballenberger N, Klucker E, Böck A, Otto R, Prazeres da Costa O, et al. Identification of novel immune phenotypes for allergic and nonallergic childhood asthma. *J Allergy Clin Immunol* 2015;135:81-91.
40. Raundhal M, Morse C, Khare A, Oriss TB, Milosevic J, Trudeau J, et al. High IFN- γ and low SLPI mark severe asthma in mice and humans. *J Clin Invest* 2015;125:3037-50.
41. Guidance for industry—pharmacogenomic data submissions. 2005. <http://www.fda.gov/downloads/RegulatoryInformation/Guidances/UCM126957.pdf>. Accessed March 30, 2016.
42. Wagner JA. Overview of biomarkers and surrogate endpoints in drug development. *Dis Markers* 2002;18:41-6.
43. Wagner JA, Williams SA, Webster CJ. Biomarkers and surrogate end points for fit-for-purpose development and regulatory evaluation of new drugs. *Clin Pharmacol Ther* 2007;81:104-7.
44. Goodsaid FM, Frueh FW, Mattes W. Strategic paths for biomarker qualification. *Toxicology* 2008;245:219-23.
45. Wenzel SE, Wang L, Pirozzi G. Dupilumab in persistent asthma. *N Engl J Med* 2013;369:1276.
46. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl R, Keene ON, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet* 2012;380:651-9.
47. Ortega HG, Liu MC, Pavord ID, Brusselle GG, FitzGerald JM, Chetta A, et al. Mepolizumab treatment in patients with severe eosinophilic asthma. *N Engl J Med* 2014;371:1198-207.
48. Bel EH, Wenzel SE, Thompson PJ, Prazma CM, Keene ON, Yancey SW, et al. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med* 2014;371:1189-97.
49. Woodruff PG, Boushey HA, Dolganov GM, Barker CS, Yang YH, Donnelly S, et al. Genome-wide profiling identifies epithelial cell genes associated with asthma and with treatment response to corticosteroids. *Proc Natl Acad Sci U S A* 2007;104:15858-63.
50. Jia G, Erickson RW, Choy DF, Mosesova S, Wu LC, Solberg OD, et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. *J Allergy Clin Immunol* 2012;130:647-54.e10.
51. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, et al. Lebrikizumab treatment in adults with asthma. *N Engl J Med* 2011;365:1088-98.
52. Brightling CE, Chaney P, Leigh R, O'Byrne PM, Korn S, She D, et al. Efficacy and safety of tralokinumab in patients with severe uncontrolled asthma: a randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet Respir Med* 2015;3:692-701.
53. Piper E, Brightling C, Niven R, Oh C, Faggioni R, Poon K, et al. A phase II placebo-controlled study of tralokinumab in moderate-to-severe asthma. *Eur Respir J* 2013;41:330-8.
54. Hanania NA, Wenzel S, Rosén K, Hsieh H-J, Mosesova S, Choy DF, et al. Exploring the effects of omalizumab in allergic asthma: an analysis of biomarkers in the EXTRA study. *Am J Respir Crit Care Med* 2013;187:804-11.
55. Busse W, Spector S, Rosén K, Wang Y, Alpan O. High eosinophil count: a potential biomarker for assessing successful omalizumab treatment effects. *J Allergy Clin Immunol* 2013;132:485-6.e11.
56. van der Schee MP, Palmay R, Cowan JO, Taylor DR. Predicting steroid responsiveness in patients with asthma using exhaled breath profiling. *Clin Exp Allergy* 2013;43:1217-25.
57. Fens N, van der Sluijs KF, van de Pol MA, Dijkhuis A, Smids BS, van der Zee JS, et al. Electronic nose identifies bronchoalveolar lavage fluid eosinophils in asthma. *Am J Respir Crit Care Med* 2015;191:1086-8.
58. Li L-B, Leung DYM, Goleva E. Activated p38 MAPK in peripheral blood monocytes of steroid resistant asthmatics. *PLoS One* 2015;10:e0141909.
59. Xiao C, Biagini Myers JM, Ji H, Metz K, Martin LJ, Lindsey M, et al. Vanin-1 expression and methylation discriminate pediatric asthma corticosteroid treatment response. *J Allergy Clin Immunol* 2015;136:923-31.e3.
60. Irvin C, Zafar I, Good J, Rollins D, Christianson C, Gorska MM, et al. Increased frequency of dual-positive TH2/TH17 cells in bronchoalveolar lavage fluid characterizes a population of patients with severe asthma. *J Allergy Clin Immunol* 2014;134:1175-86.e7.
61. Lipworth BJ. Biomarkers to predict inhaled corticosteroid response. *J Allergy Clin Immunol* 2015;136:515.
62. Goleva E, Jackson LP, Harris JK, Robertson CE, Sutherland ER, Hall CF, et al. The effects of airway microbiome on corticosteroid responsiveness in asthma. *Am J Respir Crit Care Med* 2013;188:1193-201.
63. Clemmer GL, Wu AC, Rosner B, McGeachie MJ, Litonjua AA, Tantisira KG, et al. Measuring the corticosteroid responsiveness endophenotype in asthmatic patients. *J Allergy Clin Immunol* 2015;136:274-81.e8.
64. Leckie MJ, ten Brinke A, Khan J, Diamant Z, O'Connor BJ, Walls CM, et al. Effects of an interleukin-5 blocking monoclonal antibody on eosinophils, airway hyper-responsiveness, and the late asthmatic response. *Lancet* 2000;356:2144-8.
65. Flood-Page P, Swenson C, Faiferman I, Matthews J, Williams M, Brannick L, et al. A study to evaluate safety and efficacy of mepolizumab in patients with moderate persistent asthma. *Am J Respir Crit Care Med* 2007;176:1062-71.
66. Boyman O, Kaegi C, Akdis M, Bavbek S, Bossios A, Chatzipetrou A, et al. EAACI IG Biologicals task force paper on the use of biologic agents in allergic disorders. *Allergy* 2015;70:727-54.
67. Radonjic-Hoesli S, Valent P, Klion AD, Wechsler ME, Simon H-U. Novel targeted therapies for eosinophil-associated diseases and allergy. *Annu Rev Pharmacol Toxicol* 2015;55:633-56.
68. Pettipher R, Hunter MG, Perkins CM, Collins LP, Lewis T, Baillet M, et al. Heightened response of eosinophilic asthmatic patients to the CRTH2 antagonist OC000459. *Allergy* 2014;69:1223-32.

69. Haldar P, Brightling CE, Singapuri A, Hargadon B, Gupta S, Monteiro W, et al. Outcomes after cessation of mepolizumab therapy in severe eosinophilic asthma: a 12-month follow-up analysis. *J Allergy Clin Immunol* 2014;133:921-3.
70. Bousquet J, Khaltaev N, Cruz AA, Denburg J, Fokkens WJ, Togias A, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) 2008 update (in collaboration with the World Health Organization, GA(2)LEN and AllerGen). *Allergy* 2008; 63(suppl 8):8-160.
71. Brozek JL, Bousquet J, Baena-Cagnani CE, Bonini S, Canonica GW, Casale TB, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines: 2010 revision. *J Allergy Clin Immunol* 2010;126:466-76.
72. Hellings PW, Fokkens WJ, Akdis C, Bachert C, Cingi C, Dietz de Loos D, et al. Uncontrolled allergic rhinitis and chronic rhinosinusitis: where do we stand today? *Allergy* 2013;68:1-7.
73. Papadopoulos NG, Bernstein JA, Demoly P, Dykewicz M, Fokkens W, Hellings PW, et al. Phenotypes and endotypes of rhinitis and their impact on management: a PRACTALL report. *Allergy* 2015;70:474-94.
74. Akdis CA, Bachert C, Cingi C, Dykewicz MS, Hellings PW, Naclerio RM, et al. Endotypes and phenotypes of chronic rhinosinusitis: a PRACTALL document of the European Academy of Allergy and Clinical Immunology and the American Academy of Allergy, Asthma & Immunology. *J Allergy Clin Immunol* 2013;131:1479-90.
75. Bentley AM, Jacobson MR, Cumberworth V, Barkans JR, Moqbel R, Schwartz LB, et al. Immunohistology of the nasal mucosa in seasonal allergic rhinitis: increases in activated eosinophils and epithelial mast cells. *J Allergy Clin Immunol* 1992;89:877-83.
76. Greiner AN, Hellings PW, Rotiroli G, Scadding GK. Allergic rhinitis. *Lancet* 2011;378:2112-22.
77. Castano R, Maghni K, Castellanos L, Trudeau C, Malo J-L, Gautrin D. Proinflammatory mediators in nasal lavage of subjects with occupational rhinitis. *Otolaryngol Neck Surg* 2010;143:301-3.e1.
78. Hox V, Steelant B, Fokkens W, Nemery B, Hellings PW. Occupational upper airway disease: how work affects the nose. *Allergy* 2014;69:282-91.
79. Wang M, Zhang W, Shang J, Yang J, Zhang L, Bachert C. Immunomodulatory effects of IL-23 and IL-17 in a mouse model of allergic rhinitis. *Clin Exp Allergy* 2013;43:956-66.
80. Van Gerven L, Alpizar YA, Wouters MM, Hox V, Hauben E, Jorissen M, et al. Capsaicin treatment reduces nasal hyperreactivity and transient receptor potential cation channel subfamily V, receptor 1 (TRPV1) overexpression in patients with idiopathic rhinitis. *J Allergy Clin Immunol* 2014;133:1332-9, e1-3.
81. Steelant B, Farré R, Wawrzyniak P, Belmans J, Dekimpe E, Vanheel H, et al. Impaired barrier function in patients with house dust mite-induced allergic rhinitis is accompanied by decreased occludin and zonula occludens-1 expression. *J Allergy Clin Immunol* 2016;137:1043-53.e5.
82. Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W, et al. International Consensus on Allergen Immunotherapy II: mechanisms, standardization, and pharmacoeconomics. *J Allergy Clin Immunol* 2016;137:358-68.
83. Jutel M, Agache I, Bonini S, Burks AW, Calderon M, Canonica W, et al. International consensus on allergy immunotherapy. *J Allergy Clin Immunol* 2015;136: 556-68.
84. Noda S, Suárez-Fariñas M, Ungar B, Kim SJ, de Guzman Strong C, Xu H, et al. The Asian atopic dermatitis phenotype combines features of atopic dermatitis and psoriasis with increased TH17 polarization. *J Allergy Clin Immunol* 2015;136: 1254-64.
85. Weidinger S, Novak N. Atopic dermatitis. *Lancet* 2016;387:1109-22.
86. Czarnowicki T, Esaki H, Gonzalez J, Malajian D, Shemer A, Noda S, et al. Early pediatric atopic dermatitis shows only a cutaneous lymphocyte antigen (CLA)(+) TH2/TH1 cell imbalance, whereas adults acquire CLA(+) TH22/TC22 cell subsets. *J Allergy Clin Immunol* 2015;136:941-51.e3.
87. Bieber T. Atopic dermatitis 2.0: from the clinical phenotype to the molecular taxonomy and stratified medicine. *Allergy* 2012;67:1475-82.
88. Czarnowicki T, Gonzalez J, Shemer A, Malajian D, Xu H, Zheng X, et al. Severe atopic dermatitis is characterized by selective expansion of circulating TH2/TC2 and TH22/TC22, but not TH17/TC17, cells within the skin-homing T-cell population. *J Allergy Clin Immunol* 2015;136:104-15.e7.
89. Hammad H, Lambrecht BN. Barrier epithelial cells and the control of type 2 immunity. *Immunity* 2015;43:29-40.
90. Bieber T, Cork M, Reitamo S. Atopic dermatitis: a candidate for disease-modifying strategy. *Allergy* 2012;67:969-75.
91. Bieber T, Nestle F. Personalized treatment options in dermatology. Berlin: Springer; 2015.
92. Beck LA, Thaçi D, Hamilton JD, Graham NM, Bieber T, Rocklin R, et al. Dupilumab treatment in adults with moderate-to-severe atopic dermatitis. *N Engl J Med* 2014;371:130-9.
93. Nemoto O, Furue M, Nakagawa H, Shiramoto M, Hanada R, Matsuki S, et al. The first trial of CIM331, a humanized anti-human IL-31 receptor A antibody, for healthy volunteers and patients with atopic dermatitis to evaluate safety, tolerability and pharmacokinetics of a single dose in a randomised, double-blind, placebo-controlled. *Br J Dermatol* 2016;174:296-304.
94. Bieber T. Stratified medicine: a new challenge for academia, industry, regulators and patients. London: Future Medicine; 2013; 75.
95. Staudacher A, Hinz T, Novak N, von Bubnoff D, Bieber T. Exaggerated IDO1 expression and activity in Langerhans cells from patients with atopic dermatitis upon viral stimulation: a potential predictive biomarker for high risk of eczema herpeticum. *Allergy* 2015;70:1432-9.
96. Simpson EL, Chalmers JR, Hanifin JM, Thomas KS, Cork MJ, McLean WHI, et al. Emollient enhancement of the skin barrier from birth offers effective atopic dermatitis prevention. *J Allergy Clin Immunol* 2014;134: 818-23.
97. Horimukai K, Morita K, Narita M, Kondo M, Kitazawa H, Nozaki M, et al. Application of moisturizer to neonates prevents development of atopic dermatitis. *J Allergy Clin Immunol* 2014;134:824-30.e6.