Rag proteins regulate amino-acid-induced mTORC1 signalling

Yasemin Sancak*† and David M. Sabatini*†‡¹

*Whitehead Institute for Biomedical Research and Massachusetts Institute of Technology Department of Biology, Nine Cambridge Center, Cambridge, MA 02142, U.S.A., †MIT Center for Cancer Research, 77 Massachusetts Avenue, Cambridge, MA 02139, U.S.A., and ‡Broad Institute, Seven Cambridge Center, Cambridge, MA 02142, U.S.A.

Abstract

The serum- and nutrient-sensitive protein kinase mTOR (mammalian target of rapamycin) is a master regulator of cell growth and survival. The mechanisms through which nutrients regulate mTOR have been one of the major unanswered questions in the mTOR field. Identification of the Rag (Ras-related GTPase) family of GTPases as mediators of amino acid signalling to mTOR is an important step towards our understanding of this mechanism.

The mTOR (mammalian target of rapamycin) kinase is an evolutionarily conserved protein that regulates cell growth, survival and metabolism. mTOR participates in two biochemically and functionally distinct protein complexes: mTORC (mTOR complex) 1 and 2 [1]. mTOR and mLST8 (mammalian lethal with sec-13) are common members of both complexes; raptor (regulatory associated protein of mTOR) and PRAS40 (proline-rich Akt substrate of 40 kDa) participate in mTORC1 specifically; and rictor (rapamycininsensitive companion of mTOR), sin1 and protor (protein observed with rictor) are unique members of mTORC2 [1-13]. mTORC1 is downstream of both growth factor and nutrient signalling (glucose and amino acids), and integrates the two so as to ensure cell growth only when conditions are ideal. Given the central role of mTORC1 in cell growth, it is not surprising that many upstream regulators of mTORC1 are involved in disease, and understanding how it is regulated is of great interest. The identification of the small GTPase Rheb as a potent mTORC1 activator, and the finding that TSC (tuberous sclerosis complex) 1/2 is the GAP (GTPase-activating protein) for Rheb advanced our understanding of mTORC1

regulation significantly [14–20]. Both growth factors and cellular energy levels regulate TSC1/2 activity, which in turn modulates the level of Rheb-GTP, which binds to and activates mTORC1 directly [7,21–27]. In addition, two mTORC1 members, PRAS40 and raptor, are phosphorylated in response to mitogenic stimuli and cellular stress, leading to mTORC1 activation and inhibition respectively [6,7,28,29].

Although Rheb is necessary for amino-acid-induced mTORC1 activation and Rheb overexpression can overcome amino-acid-starvation-induced mTORC1 inhibition, TSC2-null MEFs (mouse embryonic fibroblasts) are sensitive

Abbreviations used: MEF, mouse embryonic fibroblast; mTOR, mammalian target of rapamycin; mTORC1, mTOR complex 1; PRAS40, proline-rich Akt substrate of 40 kDa; Rag, Ras-related GTPase; raptor, regulatory associated protein of mTOR; rictor, rapamycin-insensitive companion of mTOR; TORC1, target of rapamycin complex 1; TSC, tuberous sclerosis complex.

¹To whom correspondence should be addressed (email sabatini@wi.mit.edu).

to amino acid starvation [30]. This observation suggested that there are additional important players in the amino acid regulation of mTORC1. Recently, the finding that Rag (Ras-related GTPase) proteins mediate amino acid signalling to mTORC1 provided new insights into this problem [31,32].

The Rag proteins are a unique family of GTPases with a canonical N-terminal Ras-like GTPase domain and a unique C-terminal RagA conserved region. In mammals, there are four Rag genes (*Rraga*, *Rragb*, *Rragc* and *Rragd*), whereas yeast and fruitflies have one RagA-like gene and one RagC-like gene.

We identified RagC by MS mass spectrometric analysis as a raptor-interacting protein [32]. Kim et al. [31] found that the Rag proteins are important for amino-acid-induced TORC1 (target of rapamycin complex 1) activation in *Drosophila* cells using an RNAi (RNA interference) screen. Both groups showed that knocking down the Rag proteins impairs amino acid signalling to TORC1, and that overexpression of constitutively GTP-bound RagA-like mutant makes TORC1 insensitive to amino acid deprivation. Moreover, Kim et al. [31], monitoring cell and organ size, as well as autophagy, observed that Rag overexpression or deletion in *Drosophila* directly parallels the effects of dTOR (*Drosophila* target of rapamycin) activation or inhibition respectively.

Although we do not completely understand how Rag proteins activate mTORC1 in detail, the critical observation that amino acid stimulation induces a change in mTORC1 localization prompted us to hypothesize that Rag proteins may regulate mTORC1 localization. Supporting our hypothesis, when Rag proteins are knocked down, amino-acidinduced mTORC1 localization change is ablated. Similarly, when a constitutively GTP-bound RagB mutant is expressed, mTORC1 localization resembles the amino-acid-induced state, even in the absence of amino acids. We also showed that, after amino acid stimulation, mTORC1 moves to Rab7-containing vesicles, where its activator Rheb is thought to reside. On the basis of these observations, we proposed a model: upon amino acid stimulation, Rag proteins initiate a localization change of mTORC1, taking it to Rheb-containing

Key words: amino acid, localization, mammalian target of rapamycin (mTOR), Ras-related GTPase (Rag), Rheb.

vesicles, leading to its activation. This model explains (i) how overexpressed and thus mislocalized Rheb can overcome amino acid starvation, and (ii) why TSC2-null MEFs are sensitive to amino acid starvation: even though Rheb is always in a GTP-bound state, mTORC1 is not in the same subcellular compartment as its activator when amino acids are not present.

The identification of Rag proteins as members of the mTORC1 pathway is an important first step towards deciphering the molecular events that signal nutrient availability to mTORC1 and evokes many interesting questions. How amino acid availability is sensed and communicated to the Rag proteins, how mTOR localization contributes to its activity, and whether Rag-related signalling can be targeted in disease are some of the exciting questions for which mTOR biologists will be seeking answers.

Acknowledgements

We thank former and current members of the laboratory.

Funding

This work was supported by the National Institutes of Health [grant numbers R01 CA103866 and AI47389], Department of Defense [grant number W81XWH-07-0448] and W.M. Keck Foundation to D.H.S.

References

- 1 Frias, M.A., Thoreen, C.C., Jaffe, J.D., Schroder, W., Sculley, T., Carr, S.A. and Sabatini, D.M. (2006) mSin1 is necessary for Akt/PKB phosphorylation, and its isoforms define three distinct mTORC2s. Curr. Biol. **16**, 1865–1870
- 2 Hara, K., Maruki, Y., Long, X., Yoshino, K., Oshiro, N., Hidayat, S., Tokunaga, C., Avruch, J. and Yonezawa, K. (2002) Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. Cell. **110**, 177–189
- 3 Kim, D.H., Sarbassov, D.D., Ali, S.M., Latek, R.R., Guntur, K.V., Erdjument-Bromage, H., Tempst, P. and Sabatini, D.M. (2003) $G\beta$ L, a positive regulator of the rapamycin-sensitive pathway required for the nutrient-sensitive interaction between raptor and mTOR. Mol. Cell **11**, 895–904
- 4 Kim, D.H., Sarbassov, D.D., Ali, S.M., King, J.E., Latek, R.R., Erdjument-Bromage, H., Tempst, P. and Sabatini, D.M. (2002) mTOR interacts with raptor to form a nutrient-sensitive complex that signals to the cell growth machinery. Cell **110**, 163–175
- 5 Sarbassov, D.D., Ali, S.M., Kim, D.H., Guertin, D.A., Latek, R.R., Erdjument-Bromage, H., Tempst, P. and Sabatini, D.M. (2004) Rictor, a novel binding partner of mTOR, defines a rapamycin-insensitive and raptor-independent pathway that regulates the cytoskeleton. Curr. Biol. 14, 1296–1302
- 6 Fonseca, B.D., Smith, E.M., Lee, V.H., MacKintosh, C. and Proud, C.G. (2007) PRAS40 is a target for mammalian target of rapamycin complex 1 and is required for signaling downstream of this complex. J. Biol. Chem. **282**, 24514–24524
- 7 Sancak, Y., Thoreen, C.C., Peterson, T.R., Lindquist, R.A., Kang, S.A., Spooner, E., Carr, S.A. and Sabatini, D.M. (2007) PRAS40 is an insulinregulated inhibitor of the mTORC1 protein kinase. Mol. Cell 25, 903–915
- 8 Wang, L., Harris, T. and Lawrence, Jr, J.C. (2008) Regulation of proline-rich Akt substrate of 40 kDa (PRAS40) function by mammalian target of rapamycin complex 1 (mTORC1)-mediated phosphorylation. J. Biol. Chem. 283, 15619–15627
- 9 Wang, L., Harris, T.E., Roth, R.A. and Lawrence, Jr, J.C. (2007) PRAS40 regulates mTORC1 kinase activity by functioning as a direct inhibitor of substrate binding. J. Biol. Chem. 282, 20036–20044

- 10 Jacinto, E., Facchinetti, V., Liu, D., Soto, N., Wei, S., Jung, S., Huang, Q., Qin, J. and Su, B. (2006) SIN1/MIP1 maintains rictor-mTOR complex integrity and regulates Akt phosphorylation and substrate specificity. Cell **127**, 125–137
- 11 Woo, S.Y., Kim, D.H., Jun, C.B., Kim, Y.M., Haar, E.V., Lee, S.I., Hegg, J.W., Bandhakavi, S., Griffin, T.J. and Kim, D.H. (2007) PRR5, a novel component of mTOR complex 2, regulates platelet-derived growth factor receptor β expression and signaling. J. Biol. Chem. **282**, 25604–25612
- 12 Yang, Q., Inoki, K., Ikenoue, T. and Guan, K.L. (2006) Identification of Sin1 as an essential TORC2 component required for complex formation and kinase activity. Genes Dev. **20**, 2820–2832
- Pearce, L.R., Huang, X., Boudeau, J., Pawlowski, R., Wullschleger, S., Deak, M., Ibrahim, A.F., Gourlay, R., Magnuson, M.A. and Alessi, D.R. (2007) Identification of Protor as a novel Rictor-binding component of mTOR complex-2. Biochem. J. 405, 513–522
- 14 Saucedo, L.J., Gao, X., Chiarelli, D.A., Li, L., Pan, D. and Edgar, B.A. (2003) Rheb promotes cell growth as a component of the insulin/TOR signalling network. Nat. Cell Biol. 5, 566–571
- 15 Stocker, H., Radimerski, T., Schindelholz, B., Wittwer, F., Belawat, P., Daram, P., Breuer, S., Thomas, G. and Hafen, E. (2003) Rheb is an essential regulator of S6K in controlling cell growth in *Drosophila*. Nat. Cell Biol. 5, 559–565
- 16 Inoki, K., Li, Y., Xu, T. and Guan, K.L. (2003) Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. Genes Dev. 17, 1829–1834
- 17 Tee, A.R., Manning, B.D., Roux, P.P., Cantley, L.C. and Blenis, J. (2003) Tuberous sclerosis complex gene products, Tuberin and Hamartin, control mTOR signaling by acting as a GTPase-activating protein complex toward Rheb. Curr. Biol. **13**, 1259–1268
- 18 Zhang, Y., Gao, X., Saucedo, L., Ru, B., Edgar, B. and Pan, D. (2003) Rheb is a direct target of the tuberous sclerosis tumour suppressor proteins. Nat. Cell Biol. 5, 578–581
- 19 Gao, X. and Pan, D. (2001) TSC1 and TSC2 tumor suppressors antagonize insulin signaling in cell growth. Genes Dev. **15**, 1383–1392
- 20 Potter, C.J., Huang, H. and Xu, T. (2001) *Drosophila* Tsc1 functions with Tsc2 to antagonize insulin signaling in regulating cell growth, cell proliferation, and organ size. Cell **105**, 357–368
- 21 Inoki, K., Li, Y., Zhu, T., Wu, J. and Guan, K.L. (2002) TSC2 is phosphorylated and inhibited by Akt and suppresses mTOR signalling. Nat. Cell Biol. **4**, 648–657
- 22 Potter, C.J., Pedraza, L.G. and Xu, T. (2002) Akt regulates growth by directly phosphorylating Tsc2. Nat. Cell Biol. **4**, 658–665
- 23 Manning, B.D., Tee, A.R., Logsdon, M.N., Blenis, J. and Cantley, L.C. (2002) Identification of the tuberous sclerosis complex-2 tumor suppressor gene product tuberin as a target of the phosphoinositide 3-kinase/Akt pathway. Mol. Cell **10**, 151–162
- 24 Long, X., Lin, Y., Ortiz-Vega, S., Yonezawa, K. and Avruch, J. (2005) Rheb binds and regulates the mTOR kinase. Curr. Biol. **15**, 702–713
- 25 Inoki, K., Zhu, T. and Guan, K.L. (2003) TSC2 mediates cellular energy response to control cell growth and survival. Cell **115**, 577–590
- 26 Lee, D.F., Kuo, H.P., Chen, C.T., Hsu, J.M., Chou, C.K., Wei, Y., Sun, H.L., Li, L.Y., Ping, B., Huang, W.C. et al. (2007) IKK β suppression of TSC1 links inflammation and turnour angiogenesis via the mTOR pathway. Cell **130**, 440–455
- 27 Ma, L., Chen, Z., Erdjument-Bromage, H., Tempst, P. and Pandolfi, P.P. (2005) Phosphorylation and functional inactivation of TSC2 by Erk: implications for tuberous sclerosis and cancer pathogenesis. Cell **121**, 179–193
- 28 Oshiro, N., Takahashi, R., Yoshin, K., Tanimura, K., Nakashima, A., Eguchi, S., Miyamoto, T., Hara, K., Takehana, K., Avruch, J. et al. (2007) The proline-rich Akt substrate of 40 kDa (PRAS40) is a physiological substrate of mammalian target of rapamycin complex 1. J. Biol. Chem. 282, 20329–20339
- 29 Gwinn, D.M., Shackelford, D.B., Egan, D.F., Mihaylova, M.M., Mery, A., Vasquez, D.S., Turk, B.E. and Shaw, R.J. (2008) AMPK phosphorylation of raptor mediates a metabolic checkpoint. Mol. Cell **30**, 214–226
- 30 Smith, E.M., Finn, S.G., Tee, A.R., Browne, G.J. and Proud, C.G. (2005) The tuberous sclerosis protein TSC2 is not required for the regulation of the mammalian target of rapamycin by amino acids and certain cellular stresses. J. Biol. Chem. 280, 18717–18727
- 31 Kim, E., Goraksha-Hicks, P., Li, L., Neufeld, T.P. and Guan, K.L. (2008) Regulation of TORC1 by Rag GTPases in nutrient response. Nat. Cell Biol. 10, 395–345
- 32 Sancak, Y., Peterson, T.R., Shaul, Y.D., Lindquist, R.A., Thoreen, C.C., Bar-Peled, L. and Sabatini, D.M. (2008) The Rag GTPases bind raptor and mediate amino acid signaling to mTORC1. Science **320**, 1496–1501

Received 18 September 2008 doi:10.1042/BST0370289