

# Ecology and Evolution

Open Access

## Oxidative stress and life histories: unresolved issues and current needs

John R. Speakman<sup>1,2</sup>, Jonathan D. Blount<sup>3</sup>, Anne M. Bronikowski<sup>4</sup>, Rochelle Buffenstein<sup>5</sup>, Caroline Isaksson<sup>6</sup>, Tom B. L. Kirkwood<sup>7</sup>, Pat Monaghan<sup>8</sup>, Susan E. Ozanne<sup>9</sup>, Michaël Beaulieu<sup>10</sup>, Michael Briga<sup>11</sup>, Sarah K. Carr<sup>9</sup>, Louise L. Christensen<sup>1</sup>, Helena M. Cochemé<sup>12</sup>, Dominic L. Cram<sup>13</sup>, Ben Dantzer<sup>14</sup>, Jim M. Harper<sup>15</sup>, Diana Jurk<sup>7</sup>, Annette King<sup>7</sup>, Jose C. Noguera<sup>8</sup>, Karine Salin<sup>8</sup>, Elin Sild<sup>6</sup>, Mirre J. P. Simons<sup>16</sup>, Shona Smith<sup>8</sup>, Antoine Stier<sup>17</sup>, Michael Tobler<sup>6</sup>, Emma Vitikainen<sup>3</sup>, Malcolm Peaker<sup>18</sup> & Colin Selman<sup>8</sup>

<sup>1</sup>Institute of Biological and Environmental Sciences, University of Aberdeen, Tillydrone Avenue, Aberdeen AB24 2TZ, UK

<sup>2</sup>State Key Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, China

<sup>3</sup>Centre for Ecology and Conservation, University of Exeter, Penryn Campus, Cornwall TR10 9FE, UK

<sup>4</sup>Department of Ecology, Evolution and Organismal Biology, Iowa State University, 251 Bessey Hall, Ames, Iowa 50011

<sup>5</sup>Physiology, Barshop Institute for Aging and Longevity Research, UTHSCSA, 15355 Lambda Drive, San Antonio, Texas 78245

<sup>6</sup>Department of Biology, Lund University, Solvegatan 37, Lund 223 62, Sweden

<sup>7</sup>The Newcastle University Institute for Ageing, Institute for Cell & Molecular Biosciences, Campus for Ageing and Vitality, Newcastle upon Tyne NE4 5PL, UK

<sup>8</sup>Institute of Biodiversity, Animal Health and Comparative Medicine, University of Glasgow, Graham Kerr Building, Glasgow G12 8QQ, UK

<sup>9</sup>University of Cambridge Metabolic Research Laboratories and MRC Metabolic Diseases Unit, Level 4, Wellcome Trust-MRC Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge CB2 0QQ, UK

<sup>10</sup>Zoological Institute and Museum, University of Greifswald, Johann-Sebastian Bach Str. 11/12, Greifswald 17489, Germany

<sup>11</sup>Behavioral Biology, University of Groningen, Nijenborgh 7, Groningen 9747 AG, The Netherlands

<sup>12</sup>MRC Clinical Sciences Centre, Imperial College London, Hammersmith Hospital Campus, Du Cane Road, London W12 0NN, UK

<sup>13</sup>Department of Zoology, University of Cambridge, Cambridge CB2 3EJ, UK

<sup>14</sup>Department of Psychology, University of Michigan, Ann Arbor, Michigan 48109

<sup>15</sup>Department of Biological Sciences, Sam Houston State University, 1900 Avenue I, LDB 100B, Huntsville, Texas 77341

<sup>16</sup>Department of Animal and Plant Sciences, University of Sheffield, Alfred Denny Building, Western Bank, Sheffield S10 2TN, UK

<sup>17</sup>Department Ecology, Physiology et Ethology, University of Strasbourg - IPHC (UMR7178), 23, rue Becquerel, Strasbourg 67087, France

<sup>18</sup>Rushmere, 13 Upper Crofts, Alloway KA7 4QX, UK

### Keywords

Aging, disposable soma theory, free radicals, life-history theory, oxidative stress.

### Correspondence

John R. Speakman, Institute of Biological and Environmental Sciences, University of Aberdeen, Tillydrone Avenue, Aberdeen AB24 2TZ, UK.

Tel: +1224 272879; Fax: +1224 272396;

E-mail [j.speakman@abdn.ac.uk](mailto:j.speakman@abdn.ac.uk)

and Colin Selman, Institute of Biodiversity, Animal Health and Comparative Medicine, College of Medical, Veterinary and Life Sciences, Graham Kerr Building, University of Glasgow, Glasgow G12 8QQ, UK.

Tel: +44 141 330 6077;

Fax: +44 141 330 5971;

E-mail: [Colin.Selman@glasgow.ac.uk](mailto:Colin.Selman@glasgow.ac.uk)

### Funding Information

This study was the result of a week-long workshop sponsored by the Rank Prize Funds attended by all the authors

### Abstract

Life-history theory concerns the trade-offs that mold the patterns of investment by animals between reproduction, growth, and survival. It is widely recognized that physiology plays a role in the mediation of life-history trade-offs, but the details remain obscure. As life-history theory concerns aspects of investment in the soma that influence survival, understanding the physiological basis of life histories is related, but not identical, to understanding the process of aging. One idea from the field of aging that has gained considerable traction in the area of life histories is that life-history trade-offs may be mediated by free radical production and oxidative stress. We outline here developments in this field and summarize a number of important unresolved issues that may guide future research efforts. The issues are as follows. First, different tissues and macromolecular targets of oxidative stress respond differently during reproduction. The functional significance of these changes, however, remains uncertain. Consequently there is a need for studies that link oxidative stress measurements to functional outcomes, such as survival. Second, measurements of oxidative stress are often highly invasive or terminal. Terminal studies of oxidative stress in wild animals, where detailed life-history information is available, cannot generally be performed without compromising the aims of the studies that generated the life-history data. There is a need therefore for novel non-invasive measure-

Received: 9 September 2015; Accepted: 20 September 2015

*Ecology and Evolution* 2015; 5(24): 5745–5757

doi: 10.1002/ece3.1790

ments of multi-tissue oxidative stress. Third, laboratory studies provide unrivaled opportunities for experimental manipulation but may fail to expose the physiology underpinning life-history effects, because of the benign laboratory environment. Fourth, the idea that oxidative stress might underlie life-history trade-offs does not make specific enough predictions that are amenable to testing. Moreover, there is a paucity of good alternative theoretical models on which contrasting predictions might be based. Fifth, there is an enormous diversity of life-history variation to test the idea that oxidative stress may be a key mediator. So far we have only scratched the surface. Broadening the scope may reveal new strategies linked to the processes of oxidative damage and repair. Finally, understanding the trade-offs in life histories and understanding the process of aging are related but not identical questions. Scientists inhabiting these two spheres of activity seldom collide, yet they have much to learn from each other.

## Background

First formulated in the 1950s (Harman 1956), the free radical damage theory of aging enjoyed a golden period as the predominant mechanistic theory of aging (Beckman and Ames 1998). In the new millennium, however, its luster has been somewhat tarnished, with a series of studies providing data contrary to its main theoretical expectations [e.g., (Yang *et al.* 2007; Doonan *et al.* 2008; Jang *et al.* 2009; Perez *et al.* 2009c; Zhang *et al.* 2009)]. These findings have led to a number of review articles pronouncing the theory dead, moribund, or at the very least suffering a mid-life crisis (Buffenstein *et al.* 2008; Gems and Doonan 2009; Perez *et al.* 2009a; Speakman and Selman 2011; Stuart *et al.* 2014 but see Barja 2013a). Many gerontologists now question the idea that oxidative stress is the principal, generalized mechanism underlying aging, although most agree it probably plays some role. On the other hand, eco-physiologists have embraced this concept as providing a physiological mechanism that might play a key role in determining the outcome of life-history trade-offs between activities, such as growth or reproduction, and body maintenance and hence future survival or reproduction (Costantini 2008; Dowling and Simmons 2009; Metcalfe and Alonso-Alvarez 2010; Isaksson *et al.* 2011a,b). Over the last decade, there has been a proliferation of studies investigating whether oxidative stress plays a role in life-history evolution. Such investigations have been largely correlative, but sometimes experimental, and undertaken across a diversity of organisms, studied under natural, semi-natural, and laboratory conditions (summarized and subjected to meta-analysis in Blount *et al.* (2015)). The results, however, have often been contradictory. The current absence of consensus and direction has led to the need to crystallize the main issues in the field, to avoid the area drifting into a shambles of conflicting data, and to resolve contradictory claims about

what the data actually mean. This manuscript emerged from a workshop attended by the authors and held in April 2014, funded by the Rank Prize Organization. It reflects a consensus on the main unresolved issues in this field, as perceived by the participants of the workshop, and the theoretical and empirical approaches required to resolve them. Our hope is that this commentary will help to move the field forward positively by stimulating researchers to refine and expand the scope of existing studies so that we do not simply continue to accumulate the same types of data, resulting in repeated additions to the confusion, as opposed to genuine illumination. In a wider context, we hope that by focussing the attention of the wide variety of researchers in this area it may additionally rejuvenate the interests that gerontologists have in oxidative stress – who were perhaps too hasty to declare the idea deceased (Kirkwood and Kowald 2012). We identify 6 Themes that are not mutually exclusive and are not listed in any perceived order of importance. These are (1) to understand the significance of different responses to reproduction observed in different tissues, (2) to refine the methodology for measuring oxidative stress to allow studies in the field that do not require precious animals in longitudinal studies of life history to be sacrificed, (3) to make laboratory studies more representative of conditions in the wild, (4) to develop good theoretical models enabling refinement of predictions of what we expect experiments to show, and the development of good alternative ideas, (5) to expand the range of species and processes that are studied to gain new perspectives and insights, and (6) to foster greater communication between evolutionary/physiological ecologists and biogerontologists. We stress that it is entirely possible for the management of oxidative stress to play a role in life-history evolution and that the need to manage, minimize, or repair oxidative damage might vary among different kinds of tissues and organisms. That such management occurs

might well mean that oxidative stress, at least in simple terms of accumulated damage to macromolecules, also has a role in the aging process.

### **Theme 1: variable responses of different tissues and different macromolecular targets during reproduction**

As data have accumulated, it has become clear that different studies have utilized an extensive range of assays to directly (and indirectly) quantify oxidative stress in different tissues. Much has been written about the need to include direct measures of oxidative damage and/or reactive oxygen species (ROS), rather than only measure antioxidants as a proxy measure, as it is the accumulated damage that is presumed to be harmful (Monaghan *et al.* 2009; Isaksson *et al.* 2011a,b; Selman *et al.* 2012; Metcalfe and Monaghan 2013). In response, a growing number of studies have measured markers of oxidative damage in relation to reproduction. Much of this research has focussed on mammals, mostly rodents, and almost exclusively on females. While some mammalian studies have supported the idea that investment in offspring production (during pregnancy and lactation) causes oxidative damage (Sainz *et al.* 2000; Bergeron *et al.* 2011; Stier *et al.* 2012; Fletcher *et al.* 2013), others have found no effect or that oxidative damage is actually reduced in lactating compared to nonreproducing females (Garratt *et al.* 2011, 2013; Oldakowski *et al.* 2012; Schmidt *et al.* 2014). The biological relevance of this diversity in response is unclear and has often been explained by individual, species, tissue or assay differences, and the difference between studies in the laboratory and field (see Themes 2 and 3 below). However, it has recently become apparent, by applying multiple assays across multiple tissues, that it is possible, in exactly the same individuals, to find some tissues and macromolecules where reproduction elevates damage, others where there is no impact, and yet others where damage is reduced (Garratt *et al.* 2011, 2013; Oldakowski *et al.* 2012; Yang *et al.* 2013). These complex patterns may result from individuals selecting which tissues to protect and which to leave vulnerable (Garratt *et al.* 2013; Yang *et al.* 2013). An intriguing example of this complexity is the lack of an effective antioxidant system in mammalian pancreatic beta-cells, which renders them susceptible to oxidative stress. Rashidi *et al.* (Rashidi *et al.* 2009) have shown how this phenomenon can be explained in terms of life-history trade-offs whereby reactive oxygen species (ROS) in beta-cells, by their negative effect on insulin synthesis/secretion, play a fitness-enhancing role for the whole organism.

One key problem is that we do not know what the long-term consequences (if any) of different forms of

oxidative damage are, for both organ function and ultimately survival, and whether changes in oxidative damage during reproduction are reversible/repairable. Furthermore, we do not know whether the consequences of oxidative damage differ across the life course. An automobile analogy may help conceptualize this problem. In the course of being driven, cars get dirty, and it is easy to imagine that oxidative damage is akin to cellular 'dirt'. Where exactly the dirt lands on a car may have very different consequences. For example, dirt on the body work may be unsightly, and easy to quantify, but it has little impact on the functionality of the car. Dirt on the windscreen/windshield may be more serious, and impair the performance of someone driving the car. But dirt in the fuel line or carburetor could completely prevent the car from operating. At present, we do not know which parts of an animal, that become oxidatively damaged during reproduction, or throughout life, are analogous to the body work (which can be restored to pristine form if washed and waxed), and which are the fuel lines and carburetor. There is a strong need therefore for studies that associate different measurements of damage with functional outcomes such as organ function and fitness outcomes such as survival and future reproductive performance.

We do not wish to completely discourage people from making studies where single tissues are sampled, and single markers of damage and protection are employed; in studies of natural populations of animals, this is sometimes the only possible sampling strategy (Selman *et al.* 2012) (see also Theme two). However, it is important to recognize that the correlation of responses across different tissues in the same individual can be poor, at least based on the relatively small number of studies that have explored such associations during reproduction (Yang *et al.* 2013; Schmidt *et al.* 2014; Xu *et al.* 2014) or in response to other stressors (Selman *et al.* 2002a,b, 2008; Kaushik and Kaur 2003; Veskoukis *et al.* 2009; Meitern *et al.* 2013). Hence, interpreting such studies in the absence of a clear link to a functional outcome is fraught with difficulty. In addition, we sorely need more studies that expand the range of markers that are used and the range of tissues that are measured, in response to experimental manipulations of both reproductive status (allocated to breed or not breed) and reproductive effort (number of raised offspring or investment per offspring). It is important when making such manipulations to quantify the impact of the manipulation on the investment by the female, as simply adding or subtracting offspring does not necessarily cause an impact on female investment (Speakman and Garratt 2014). This is because individuals may continue to work at the same rate they would have without the manipulation, generally with the

outcome of the manipulation being felt more acutely on the offspring than the adult. At present, we have no idea whether conservation exists across different organisms when examining multi-tissue and multi-target patterns of damage and protection. Presumably, organisms allocate protection and repair to those things that are most critical, which may itself vary with life stage. Hence, patterns of protection and repair may provide useful clues as to the targets most likely to be functionally significant for survival. At present, we do not know whether the same targets will be universally important across all organisms, or whether different taxa will have unique points of vulnerability. Moreover, it is unclear whether the effects will be the same across both sexes, in particular as males and females may show pronounced differences in markers of oxidative stress during reproduction (Alonso-Alvarez *et al.* 2004; Wiersma *et al.* 2004; Isaksson *et al.* 2011a,b; Stier *et al.* 2012; Isaksson 2013), and in their rates of aging. Resolving these issues will be an important future aim as our knowledge expands.

**Theme two: the dilemma of field studies. Longitudinal data on life histories make animals too valuable to sample destructively for oxidative stress assays**

Linking together information on oxidative stress and life-history strategies across the lifespan requires detailed knowledge of individuals at different ages/stages of their life histories. Longitudinal studies and experimental manipulations of either status or effort provide the best way to understand the nature of the underlying associations because cross-sectional surveys and correlational studies may be prone to artifacts through differential survival and reproduction (Nussey *et al.* 2009; Bouwhuis *et al.* 2012). This might, for example, lead to the illusion that oxidative damage declines in relation to organismal age, because those individuals with the highest levels of damage die sooner. Additionally, this could suggest oxidative damage is reduced by reproduction, because only those individuals with pre-existing low damage initiate attempts to reproduce [e.g., Beaulieu *et al.* 2014]. Field studies that meticulously follow individuals throughout their life course, quantifying reproduction and growth, are growing in number. Repeatedly measuring oxidative stress markers within individuals over time seems an obvious way to test the role of oxidative stress on life histories (Nussey *et al.* 2009; van de Crommenacker *et al.* 2011). There is, however, a major dilemma because in general such studies were not initiated as tests of the oxidative stress theory, and they have alternative goals and aims that may be conflicting. For example, the currently available assays, apart from measurements in

blood and minor biopsies, require destructive testing. Euthanizing animals that are part of long-term monitored populations is, however, generally incompatible with the primary goals of such longitudinal field studies. This might not be a problem if it turns out that the markers measured in blood are indeed the markers of greatest functional significance (see Theme 1 above). This would be indeed fortuitous. However, if these markers only reflect “dirt on the chassis” then the opportunity to exploit the thousands of person hours devoted to longitudinal quantification of the life-history parameters of known individuals will be lost. There is a major demand therefore for the development of assays in accessible samples that will inform us about the multi-tissue, multi-target nature of protection and repair processes without the need to kill (or severely impair) animals in the process. For example, standard tissue biopsies may be sufficiently traumatic to animals that even though the animal survives the tissue collection, it may have a significantly elevated risk of mortality when placed back into the field, such that its contribution to the population demographic data may be compromised. Alternatively, its behavior might be altered leading to reduced likelihood of it being resampled at a later age. Moreover, tracking, trapping, capturing, and restraining wild animals may affect measures of oxidative status. Nevertheless, some tissues (e.g., feathers, hair, and small epidermal biopsies) may be readily sampled repeatedly with minimal invasiveness, and considerable information can also be gleaned from urine and fecal samples. We envisage that technological advances in the future may come in several different forms. First, it is already clear that compounds in the urine/feces may provide an indication of oxidative damage (e.g., levels of f2 isoprostanes, 8-OHdG). As metabolite profiling of urine/feces becomes more sophisticated with the capacity to monitor simultaneously thousands of compounds, the prospect that multiple urinary markers that may provide a “damage fingerprint” seems feasible. Second, animals may be injected with compounds that are metabolized in relation to aspects of “damage” or “protection/repair” capacity and the products of such reactions in blood or urine monitored. It may be possible to use radioactive markers to localize tissue damage or protection in given tissues using PET scanning or other imaging technology – although admittedly applying such technology in the field would prove challenging. Finally, as the costs of transcriptomic profiling by RNAseq decline, it is now possible to generate profiles of gene expression from nucleated blood cells, mouth swabs and/or very small biopsy samples, allowing simultaneous evaluation of the status of repair, turnover, and protection systems, in multiple tissues. Development of such methods would provide significant benefits for both field- and laboratory-based studies. A

major point, however, that we wish to emphasize is that it is not a trivial matter to bring assays from the laboratory to the field, and needs to be done with great care. Assays may be highly sensitive to environmental conditions and individual factors that while easy to control in the laboratory are much more problematical to control in the field – including for example time as last meal (or the composition of the meal itself), whether or not the individual encountered a predator or a dominant individual in the recent past, and the almost unavoidable influence of pollutants in all but the most pristine environments. Just because a particular assay works well under controlled laboratory settings, we should not assume that it will work equally well in the field.

### Theme three: considerations for laboratory studies

Laboratory studies provide unrivaled opportunities to perform controlled experiments, with multiple detailed measurements and terminal assays that are seldom possible in the field. Yet these studies come with their own drawbacks, chief among which is the question whether the situation in the laboratory adequately mimics the situation in the field with respect to the key elements that generate the trade-off between reproduction and survival. An important question is whether animals in the laboratory with *ad libitum* food supplies are in a situation where it is necessary to trade-off somatic protection against reproduction (e.g., Selman *et al.* 2012; Metcalfe and Monaghan 2013). This could be a serious issue if animals in the field are routinely food limited. However, if animals in the field during peak reproduction are operating at an intrinsically determined physiological capacity, then this issue is less important because both laboratory and field animals will be likely to be working at this same limiting physiological capacity – that is the energy budget within which different costly functions must be accommodated will be the same irrespective of food supply (Speakman and Garratt 2014). There is considerable evidence to suggest that some strains of laboratory mice and other rodents are operating at a physiologically imposed maximum when they are at peak lactation (reviewed in Speakman and Krol 2011). However, the nature of this ceiling is disputed. Similarly in the field, there is evidence that some species of bird are operating at a physiological ceiling (Daan *et al.* 1996; Tinbergen and Verhulst 2000; Welcker *et al.* 2010; Elliott *et al.* 2014), although the causes and definition of this ceiling are also unclear. How commonly physiological factors limit expenditure, as opposed to the level of food supply is uncertain, but studies of the responses of animals to food supplementation indicate that the response is seldom to elevate energy

expenditure (Boutin 1990), instead opting to reduce the time required to ingest a given level of resource, perhaps indicating that physiological limitations may be the norm.

Nevertheless, there are other aspects of the laboratory situation that could compromise experiments to establish the oxidative costs of reproduction, and or the implications of oxidative damage for survival. Among such issues are the levels of antioxidants in laboratory rodent chow, when compared with wild foods (assuming dietary antioxidants do play a role in damage mitigation – which is currently unclear). This could be a problem in different ways. Manufacturers of rodent chow generally add large amounts of antioxidants, not as a nutritional additive but to improve the shelf life of the product. Such levels may be unnaturally high relative to foods available in the wild. As food intake is greatly elevated during lactation, it is possible that reproducing laboratory animals may inadvertently ingest enormous levels of exogenous antioxidants in their diets, and this may explain why a common observation in laboratory studies is that oxidative damage at peak lactation appears to be reduced. Should animals studied in a laboratory context therefore be fed diets with extremely low levels of antioxidants? The problem is that we do not presently know the extent to which lactating mammals, or chick-rearing birds in the wild, modulate their dietary intake to enhance their antioxidant intake or intake fatty acids and other nutritional components that are less susceptible to attack by free radicals (Beaulieu and Schaefer 2013). There is a need for information on the dietary intakes of antioxidants of the diet in wild animals at critical times of peak reproductive activity so that diets could be formulated to allow laboratory studies to more closely mimic the situation in the wild [see also Isaksson *et al.* (2011a,b)].

Another issue relates to the ambient temperatures and photoperiods at which laboratory animals are maintained. In the wild, animals are exposed to fluctuating temperatures and photoperiods which place them under variable thermoregulatory demands (endotherms) or constraints (ectotherms). This might compromise the ability to invest in reproduction [e.g., Krol and Speakman (2003)]. Yet in the laboratory, studies are commonly performed at room temperature (c. 21–23°C) and a fixed photoperiod with minimal fluctuations being part of the legal housing requirements in many countries. The stability of this temperature and its level and invariant photoperiod may be completely inappropriate to mimic the experience of animals in the wild. Levels of many hormones are driven by photoperiod changes, and these often prepare the organism for the change in reproductive status altering not only the hypothalamic–pituitary gonad axis but also metabolic rate, membrane composition appetite, and stress responses. Moreover, melatonin, which directly depends on photoperiod, and has powerful antioxidant



properties, can affect the overall oxidative status of the organism (Tan *et al.* 2010). For some animals which routinely occupy hypoxic environments in the wild, for example, fossorial animals or those from high altitude, well-ventilated facilities may actually expose them to unnaturally high levels of oxygen and hence risk of oxidative stress. A classic example in this regard are the high levels of oxidative damage in naked mole rats (*Heterocephalus glaber*) (Andziak and Buffenstein 2006; Andziak *et al.* 2006), which is an exceptionally long-lived rodent. Naked mole rats naturally live in large family groups in sealed underground burrows where they experience profound hypoxia and hypercapnia (Peterson *et al.* 2012). In the laboratory, high levels of oxidative stress may be a result of the relatively hyperoxic captive conditions. The fact naked mole-rats in captivity tolerate this chronic high level of oxidative damage and their extraordinary longevity is based upon captive data (Lewis *et al.* 2013) may suggest oxidative stress is not a key element of their enhanced longevity. An under-recognized issue is also the possibility that laboratory animals live continually in a state of chronic stress, even in the best-managed facilities. Even such apparently subtle changes such as rotation of staff caring for the animals [e.g., Sorge *et al.* 2014] may influence the stress experienced by captive animals.

Finally, if the intention is to establish the linkage between aspects of damage and survival, it is important to recognize that oxidative stress may only be a survival issue under certain environmental conditions. For example, if animals sustain damage to their skeletal muscle, this might only compromise individuals if they have to perform physical exercise to obtain their food or avoid predation – as is generally the case in the wild, but seldom so in captivity. Another example is if oxidative stress acts to reduce the functionality of the immune system. However, for animals raised in specific pathogen-free (SPF) facilities such an effect would be unlikely to translate into a survival impact.

#### **Theme four: the lack of specificity in the theoretical predictions or of good alternative hypotheses**

The idea that the link between reproduction and future survival may be mediated by oxidative stress predicts that oxidative damage should increase as a consequence of reproduction and that this should limit future survival or future reproductive performance. These predictions are not the same as the predictions surrounding the role of oxidative stress as a mediator of aging. Hence, tests of the free radical theory of aging are not necessarily equivalent to tests of the mediating role of oxidative damage in life histories. Moreover, a problem with this prediction is that

it is completely nonspecific regarding the locations and targets of such damage. The organism is viewed as a single integrated and homogeneous unit that is predicted to be damaged or not damaged. In reality, organisms comprise a complex set of organs that perform very different functions, have different metabolic rates, different levels of oxidants and antioxidants, repair mechanisms and ROS, and these organs are potentially affected very differently by the process of reproduction. For example, for mammals and birds, a major consequence of reproduction is elevated food intake. This food needs to be digested by the alimentary tract and the digested substrates then processed by the liver. We might imagine major impacts of reproduction on the metabolic activities of these tissues. Indeed in mice, at peak lactation the liver doubles in size and the small intestine grows longer by around 50% (Johnson *et al.* 2001). In contrast, the brain needs to function continuously whether the organism reproduces or not. It seems unlikely that major changes occur in brain metabolic rate during reproduction, and its size is unaffected. These changes in size and potentially also the metabolic rate of the component body tissues during reproduction may be coupled with alterations in where the animals decide to selectively allocate their protection and repair processes. However, we do not have any clear predictions regarding what the consequences of these differences might be. Hence, it is difficult to evaluate the data that are being generated. Does the theory predict that damage should be uniformly elevated across all tissues? If so we probably have enough data to reject it already. However, perhaps the model does not predict this. For example, the proliferation of newly constructed, and presumably undamaged, liver tissue during reproduction may mean that the average concentration of measured damage is reduced – and this would be consistent with the empirical observations in several previous studies (Garratt *et al.* 2011, 2013; Oldakowski *et al.* 2012; Yang *et al.* 2013; Xu *et al.* 2014). In fact, many organs in lactation show hyperplasia and this may explain reduced damage reported more widely in other tissues such as the kidney (Oldakowski *et al.* 2012; da Silva *et al.* 2013). Hence, these reduced levels of damage may not be inconsistent with the life-history predictions, despite on the face of it being in the completely opposite direction to the simplistic prediction that “damage will be increased”. Furthermore, even within the cell, there may be nonuniformity in the distribution of sites at which oxidative damage occurs, is detected by conventional assays, and actually matters. There is a strong need therefore for a more refined conceptualization of the idea, and derivation of testable mathematical models which make predictions at the tissue/organ level, at a particular stage of development and with respect to different macromolecu-

lar targets of damage. We envisage these models may generate different predictions depending on the actual mechanics of reproduction. For example, pregnancy and lactation might be expected to involve very different physiological processes and consequences, than oviparity and chick feeding in birds. Viviparous and oviparous reptiles may similarly differ in the patterns of oxidative stress that are predicted by the theory.

A second element of the current theoretical shortcomings is that not only are the predictions of the oxidative stress model for life-history effects poorly defined, but there are few alternative hypotheses to be evaluated. Even the changes that are anticipated under the null hypothesis remain uncertain, as for example the changes in tissue proliferation during reproduction highlighted above might be expected to alter apparent measured damage, even if there was no actual change in the underlying processes. Hence, the required modeling needs not only to refine the predictions of the oxidative stress model but also the expectations under the null and alternative hypotheses. One promising potential alternative is the “oxidative shielding theory” (Blount *et al.* 2015). This idea suggests that there may be major consequences of oxidative damage early in life. Hence, a priority of the parents may be to ensure that damage to their soma(s) is not transferred to their offspring. Even though oxidative protection and repair may be costly, it may be more beneficial for parents to upregulate these during reproduction to reduce their own oxidative damage, thereby minimizing damage to their offspring, because the fitness benefits in offspring quality, offset the sacrifice in terms of offspring numbers. Clearly, transfer of somatic damage from parents to offspring may be more likely during some aspects of reproduction (e.g., in pregnancy where the fetus and mother are in close physiological contact and the mother passes chemicals from her body to the fetus) compared with others like chick rearing in birds where there is no direct physiological contact of parents and offspring. This may suggest shift in focus toward egg production and laying (Vezina and Williams 2002; Williams *et al.* 2009) might be warranted (see also Velando *et al.* 2008).

### **Theme five: a wider diversity of animals and experimental conditions on which to test the ideas may provide useful insights**

Tests of the oxidative stress theory for the trade-off in life-history evolution have been performed on a wide variety of vertebrate species including small mammals (Bergeron *et al.* 2011; Garratt *et al.* 2011; Fletcher *et al.* 2013; Cram *et al.* 2015), including bats (Wilhelm *et al.* 2007; Schneeberger *et al.* 2014), large mammals (Castillo

*et al.* 2005; Nussey *et al.* 2009; Rizzo *et al.* 2013), reptiles (Robert *et al.* 2007; Isaksson *et al.* 2011a,b), and birds (Wiersma *et al.* 2004; Costantini *et al.* 2006, 2010; Bize *et al.* 2008; Marko *et al.* 2011; Isaksson 2013). Yet, despite this large range, there are a wide variety of unstudied reproductive strategies within the endotherms, including semelparity, and brood parasites where contrasting behaviors of the participants may make useful tests of the model. Moreover, within ectotherms, the growing number of long-term mark/recapture studies (e.g., Robert and Bronikowski 2010; Schwanz *et al.* 2011) should allow a more complete understanding of oxidative stress and damage with respect to physiological mode and temperature sensitivity of the animals. Moreover, future analyses may enable partition of the contrasting results between mammals and birds into causes due to phylogeny and physiology (reviewed in Schwanz *et al.* 2011).

Expanding the diversity of models used may provide unexpected insights into the processes involved that are not immediately obvious from the limited set of animals studied thus far. Insights from the long-lived social breeding naked mole rat, for example, have been particularly important regarding the complexity of the process of damage and protection, as the extant levels of oxidative damage are surprisingly high for such a long-lived organism (Andziak *et al.* 2006): if the oxidative stress theory of aging is correct. Moreover, contrasting the role of oxidative stress in life histories, levels of damage are similar among breeding females and nonbreeding subordinates, despite a 3–5 fold change in metabolic rate during pregnancy and lactation (Urison and Buffenstein 1995). High levels of oxidative damage even in young animals have also been observed in long-lived bats and birds (Hamilton *et al.* 2001; Hermes-Lima and Zenteno-Savin 2002; Buffenstein *et al.* 2008) and suggests an understudied strategy for coping with oxidative stress may just be to tolerate the damage by mitigating its functional impact (Lewis *et al.* 2013) (see Theme 1). Tolerance of damage may be attributed to subcellular localization of the oxidatively damaged macromolecules, a phenomenon seldom measured in studies of oxidative damage and life history. Naked mole-rats, in particular, have an enhanced network of mechanisms to maintain protein stability when challenged with oxidative stressors, as indicated by findings that their liver proteins are significantly more resistant to urea-induced protein damage (Perez *et al.* 2009b). Moreover, it appears that certain proteins bear the brunt of the oxidative damage (e.g., triosephosphate isomerase and peroxiredoxin 1), yet their functionality does not appear to be compromised (De Waal *et al.* 2013). These proteins may serve as oxidative sinks, which lead to better protection of the cytosolic environment from the formation of harmful oligomers or larger aggregates (Rodriguez *et al.* 2014).

Although different species have different life-history strategies and these life-history differences may engender different responses to oxidative stress, comparisons across species are accompanied by a suite of still-debated difficulties, such as the problems of how to deal with body size effects and phylogenetic independence (Speakman 2005; Chamberlain *et al.* 2012; Barja 2013b). Studying the co-variation of life-history strategies and oxidative damage within single species avoids the allometry and scaling issues that arise when making comparisons across species. For example, contrasts between “fast” and “slow” living garter snakes have been particularly informative with respect to the evolution of physiological divergence including patterns of oxidative and other stressors (Schwartz and Bronikowski 2011, 2013a,b). Their contrasting reproductive strategies and stress responses could expand our understanding of the universality of oxidative stress as mediator of life histories, particularly interpreted within a coherent theoretical framework (Theme 4) that generates contrasting predictions of what we might expect to happen. Overall, we see distinct advantages to broadening the scope of existing studies to include a wider diversity of organisms exploiting a more expansive diversity of life-history strategies.

### **Theme six: evolutionary/physiological ecologists and gerontologists need to speak to each other more deeply and more often**

The idea that oxidative stress might underpin the phenomenon of aging was the dominant mechanistic theory of aging for almost 50 years. Although the idea that oxidative stress may play a role in life-history trade-offs is not the same, it is clear that the two are related, as reproductive lifespan, reproductive senescence, and mortality risks, hence lifespan, are all components of life histories. However, these theories do make different predictions, and testing them requires different approaches. The popularity of the “free radical oxidative damage” theory of aging among gerontologists waned, almost in parallel with the increase in popularity of the role of oxidative stress in influencing the outcome of life-history trade-offs among eco-physiologists. It is apparent from the literature cited in many ecophysiology texts, which frequently include the original paper on free radicals by Harman (1956) and even the rate of living theory by Pearl (Pearl 1928), that many of the more recent contributions to the literature by biogerontologists, which have fueled the reduction in the popularity of the idea in that field, are either not being read, or are being selectively ignored. Examples include the impressive body of work by Arlan Richardson and colleagues showing that knocking out or over expressing many of the major protective enzymes against oxidative

stress, such as superoxide dismutase, catalase and glutathione peroxidase, leads to the expected changes in oxidative damage, but with no impact on lifespan (Perez *et al.* 2008, 2009c; Jang *et al.* 2009; Zhang *et al.* 2009) (but recall discussion above under Theme 3 regarding the problems of laboratory studies). However, when these genetically manipulated models are subjected to high-fat diets or other unfavorable environmental conditions, increased oxidative stress accelerates pathology and shortens lifespan, and reduced oxidative stress and overexpression of antioxidants reduces age-associated pathology and increases lifespan (Salmon *et al.* 2010). These examples clearly suggest that the role that oxidative stress plays in aging depends on environmental conditions. Under optimized laboratory, husbandry conditions animals can tolerate oxidative damage. However, under environmental conditions including suboptimal nutrition, oxidative damage, in keeping with the oxidative stress theory of aging, may overwhelm the cytoprotective defences with concomitant effects on age-related diseases and lifespan (Salmon *et al.* 2010).

Moreover, many studies by both evolutionary/physiological ecologists and biogerontologists still reiterate the idea that increases in energy expenditure lead to increases in free radical production, despite the evidence in favor of that being at best equivocal. Papers predicting, on the basis of our improved knowledge of mitochondrial functioning, that the opposite is likely to pertain in many circumstances were published 15 years ago (Brand 2000) and empirical studies showing that high metabolism is linked to greater longevity or is unrelated to lifespan, in both correlational and experimental settings, have been published also within the last decade (Speakman 2004; Selman *et al.* 2008; Keipert *et al.* 2011). It seems that the precise mitochondrial pathway responsible for the increased energy expenditure (*i.e.*, mitochondrial proton leak *vs.* ATP synthesis) is likely to determine the sign of the relationship between energy expenditure and oxidative stress levels (Stier *et al.* 2014). This selective blindness to contrary evidence occurs despite the fact that several reviews of the field (*e.g.*, Monaghan *et al.* 2009; Selman *et al.* 2012; Speakman and Garratt 2014) have pointed out the problem with the assumption of a direct positive link in considerable detail.

Recent findings in gerontology that have emphasized the roles of particular signaling pathways in the process of aging and senescence (such as the insulin/IGF-1 signaling, sirtuin, and mTOR pathways), which play highly conserved roles in growth metabolism and reproduction (Kenyon 2011; Gems and Partridge 2013), and the potential role of multiplex resistance to stressors based on responses of cultured primary fibroblasts (Salmon *et al.* 2005; Harper *et al.* 2007, 2011) have much to offer ecophysiologicalists as they strive to understand the physiologi-



cal basis of life-history trade-offs. In addition, many biogerontology laboratories have access to equipment for quantifying various aspects of stress that are beyond the reach of a dedicated ecophysiology laboratory, and hence, collaborations between the two fields may be extremely useful. But the flow of information and technology need not be only in one direction. Furthermore, it is now becoming clearer that ROS themselves participate in signaling pathways. In addition to the long-recognized role of ROS in host defences against pathogens, ROS have also been shown to be an important feature in the regulation of the entry of mammalian cells to the state of replicative senescence. The senescent state of the cell is not merely the result of intrinsic failure of the mitotic machinery, but appears to be a highly regulated outcome that results from activation of a pathway that integrates input from mitochondrial dysfunction, telomere erosion, and DNA damage (Passos *et al.* 2010).

Biogerontologists may have been too hasty to dismiss the role of oxidative stress as an important functional component of aging, based on studies of model organisms in protected laboratory environments (Theme 3). Studies of wild animals living in the field with all its wonderful complexity and challenges may help redress this balance and re-ignite interest in this idea among the community from whence it emerged. After all more than 99.999% of all organisms on earth live and age in the field, and not in a protected laboratory, and the processes that cause them to do so may indeed include oxidative stress in one form or another. So studies taking advantage of the huge diversity in aging rate and lifespan in free-living animals may be particularly relevant and important for gerontologists to be aware of. Finally, free radicals not only play a role in damage, and potentially aging, but are also essential positive components of living systems involved in signaling pathways and in immune function (bactericidal killing). Understanding these functions more clearly may be important in interpreting the confusion of current data. Therefore, meeting people working in these areas to discuss research aims and findings as well may be equally important. However, this cross-fertilization of disciplines will not happen unless there is a council by which such interactions might be facilitated. There is an urgent need for a forum where ecophysiologicals, biogerontologists, and other scientists working on oxidative stress can meet and exchange ideas and data with respect to aging, physiology, ecology, and the evolution of life histories.

## Summary

The role of oxidative stress as a factor influencing aging and longevity, and in driving the evolution of optimal investment patterns between reproduction and survival

(life-history theory) are distinct areas of research with differing underlying theoretical bases. Yet they are interconnected at many different levels. We have formulated a series of questions and propose the collection of some different types of data that will enhance our ability to understand the role of oxidative stress in life histories, that will also impact our understanding of oxidative stress and aging. The six themes highlighted here provide a framework for moving forwards along this alternative path. Significant synergies would be enabled, new insights gained, and much confusion eliminated if there was more dialogue between evolutionary/physiological ecologists and biogerontologists.

## Acknowledgments

This study was the result of a week-long workshop sponsored by the Rank prize funds attended by all the authors. We are grateful to the Rank prize funds for supporting this meeting.

## Conflict of Interest

None of the authors has any conflict of interests to declare.

## References

- Alonso-Alvarez, C., S. Bernas, G. Devevey, J. Prost, B. Faivre, and G. Sorci. 2004. Increased susceptibility to oxidative stress as proximate cost of reproduction. *Ecol. Lett.* 7:363–368.
- Andziak, B., and R. Buffenstein. 2006. Disparate patterns of age-related changes in lipid peroxidation in long-lived naked mole-rats and shorter-lived mice. *Aging Cell* 5:525–532.
- Andziak, B., T. P. O'Connor, W. Qi, E. M. DeWaal, A. Pierce, A. R. Chaudhuri, *et al.* 2006. High oxidative damage levels in the longest-living rodent, the naked mole-rat. *Aging Cell* 5:463–471.
- Barga, G. 2013a. Updating the mitochondrial free-radical theory of aging: an integrated view, key aspects and confounding concepts. *Antioxid. Redox Signal.* 19:1420–1445.
- Barja, G. 2013b. Correlations with longevity and body size: to correct or not correct? *J. Gerontol. A Biol. Med. Sci.* 69:1096–1098.
- Beaulieu, M., and H. M. Schaefer. 2013. Rethinking the role of dietary antioxidants through the lens of self-medication. *Anim. Behav.* 86:17–24.
- Beaulieu, M., S. Mboumba, E. Willaume, P. M. Kappeler, and M. J. E. Charpentier. 2014. The oxidative cost of unstable social dominance. *J. Exp. Biol.* 217:2629–2632.
- Beckman, K. B., and B. N. Ames. 1998. The free radical theory of aging matures. *Physiol. Rev.* 78:547–581.

- Bergeron, P., V. Careau, M. M. Humphries, D. Reale, J. R. Speakman, and D. Garant. 2011. The energetic and oxidative costs of reproduction in a free-ranging rodent. *Funct. Ecol.* 25:1063–1071.
- Bize, P., G. Devevey, P. Monaghan, B. Doligez, and P. Christe. 2008. Fecundity and survival in relation to resistance to oxidative stress in a free-living bird. *Ecology* 89:2584–2593.
- Blount, J. D., E. I. Vitikainen, I. Stott, and M. A. Cant. 2015. Oxidative shielding and the cost of reproduction. *Biol. Rev. Camb. Philos. Soc.* 28; 9:e103286.
- Boutin, S. 1990. Food supplementation experiments with terrestrial vertebrates - patterns, problems, and the future. *Can. J. Zool.* 68:203–220.
- Bouwhuis, S., R. Choquet, B. C. Sheldon, and S. Verhulst. 2012. The forms and fitness cost of senescence: age-specific recapture, survival, reproduction, and reproductive value in a wild bird population. *Am. Nat.* 179:E15–E27.
- Brand, M. D. 2000. Uncoupling to survive? The role of mitochondrial inefficiency in ageing. *Exp. Gerontol.* 35:811–820.
- Buffenstein, R., Y. H. Edrey, T. Yang, and J. Mele. 2008. The oxidative stress theory of aging: embattled or invincible? Insights from non-traditional model organisms. *Age (Dordr)* 30:99–109.
- Castillo, C., J. Hernandez, A. Bravo, M. Lopez-Alonso, V. Pereira, and J. L. Benedito. 2005. Oxidative status during late pregnancy and early lactation in dairy cows. *Vet. J.* 169:286–292.
- Chamberlain, S. A., S. M. Hovick, C. J. Dibble, N. L. Rasmussen, B. G. Van Allen, B. S. Maitner, et al. 2012. Does phylogeny matter? Assessing the impact of phylogenetic information in ecological meta-analysis. *Ecol. Lett.* 15:627–636.
- Costantini, D. 2008. Oxidative stress in ecology and evolution: lessons from avian studies. *Ecol. Lett.* 11:1238–1251.
- Costantini, D., S. Casagrande, S. De Filippis, G. Brambilla, A. Fanfani, J. Tagliavini, et al. 2006. Correlates of oxidative stress in wild kestrel nestlings (*Falco tinnunculus*). *J. Comp. Physiol. B.* 176:329–337.
- Costantini, D., L. Carello, and A. Fanfani. 2010. Relationships among oxidative status, breeding conditions and life-history traits in free-living Great Tits *Parus major* and Common Starlings *Sturnus vulgaris*. *The Ibis* 152:793–802.
- Cram, D. L., J. D. Blount, and A. J. Young. 2015. Oxidative status and social dominance in a wild cooperative breeder. *Funct. Ecol.* 29:229–238.
- van de Crommenacker, J., J. Komdeur, and D. S. Richardson. 2011. Assessing the cost of helping: the roles of body condition and oxidative balance in the Seychelles warbler (*Acrocephalus sechellensis*). *PLoS ONE* 6:e26423.
- Daan, S., C. Deerenberg, and C. Dijkstra. 1996. Increased daily work precipitates natural death in the kestrel. *J. Anim. Ecol.* 65:539–544.
- De Waal, E. M., H. Liang, A. Pierce, R. T. Hamilton, R. Buffenstein, and A. R. Chaudhuri. 2013. Elevated protein carbonylation and oxidative stress do not affect protein structure and function in the long-living naked-mole rat: a proteomic approach. *Biochem. Biophys. Res. Commun.* 434:815–819.
- Doonan, R., J. J. McElwee, F. Matthijssens, G. A. Walker, K. Houthoofd, P. Back, et al. 2008. Against the oxidative damage theory of aging: superoxide dismutases protect against oxidative stress but have little or no effect on life span in *Caenorhabditis elegans*. *Genes Dev.* 22:3236–3241.
- Dowling, D. K., and L. W. Simmons. 2009. Reactive oxygen species as universal constraints in life-history evolution. *Proc. Biol. Sci.* 276:1737–1745.
- Elliott, K. H., M. Le Vaillant, A. Kato, A. J. Gaston, Y. Ropert-Coudert, J. F. Hare, et al. 2014. Age-related variation in energy expenditure in a long-lived bird within the envelope of an energy ceiling. *J. Anim. Ecol.* 83:136–146.
- Fletcher, Q. E., C. Selman, S. Boutin, A. G. McAdam, S. B. Woods, A. Y. Seo, et al. 2013. Oxidative damage increases with reproductive energy expenditure and is reduced by food-supplementation. *Evolution* 67:1527–1536.
- Garratt, M., A. Vasilaki, P. Stockley, F. McArdle, M. Jackson, and J. L. Hurst. 2011. Is oxidative stress a physiological cost of reproduction? An experimental test in house mice. *Proc. Biol. Sci.* 278:1098–1106.
- Garratt, M., N. Pichaud, E. D. King, and R. C. Brooks. 2013. Physiological adaptations to reproduction. I. Experimentally increasing litter size enhances aspects of antioxidant defence but does not cause oxidative damage in mice. *J. Exp. Biol.* 216:2879–2888.
- Gems, D., and R. Doonan. 2009. Antioxidant defense and aging in *C. elegans*: is the oxidative damage theory of aging wrong? *Cell Cycle* 8:1681–1687.
- Gems, D., and L. Partridge. 2013. Genetics of longevity in model organisms: debates and paradigm shifts. *Annu. Rev. Physiol.* 75:621–644.
- Hamilton, M. L., H. Van Remmen, J. A. Drake, H. Yang, Z. M. Guo, K. Kewitt, et al. 2001. Does oxidative damage to DNA increase with age? *Proc. Natl. Acad. Sci. USA* 98:10469–10474.
- Harman, D. 1956. Aging: a theory based on free radical and radiation chemistry. *J. Gerontol.* 11:298–300.
- Harper, J. M., A. B. Salmon, S. F. Leiser, A. T. Galecki, and R. A. Miller. 2007. Skin-derived fibroblasts from long-lived species are resistant to some, but not all, lethal stresses and to the mitochondrial inhibitor rotenone. *Aging Cell* 6:1–13.
- Harper, J. M., M. Wang, A. T. Galecki, J. Ro, J. B. Williams, and R. A. Miller. 2011. Fibroblasts from long-lived bird species are resistant to multiple forms of stress. *J. Exp. Biol.* 214:1902–1910.
- Hermes-Lima, M., and T. Zenteno-Savin. 2002. Animal response to drastic changes in oxygen availability and

- physiological oxidative stress. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 133:537–556.
- Isaksson, C. 2013. Opposing effects on glutathione and reactive oxygen metabolites of sex, habitat, and spring date, but no effect of increased breeding density in great tits (*Parus major*). *Ecol. Evol.* 3:2730–2738.
- Isaksson, C., B. Sheldon, and T. Uller. 2011a. The challenges of integrating oxidative stress into life-history biology. *Bioscience* 61:194–202.
- Isaksson, C., G. M. While, M. Olsson, J. Komdeur, and E. Wapstra. 2011b. Oxidative stress physiology in relation to life history traits of a free-living vertebrate: the spotted snow skink, *Niveoscincus ocellatus*. *Integr. Zool.* 6:140–149.
- Jang, Y. C., V. I. Perez, W. Song, M. S. Lustgarten, A. B. Salmon, J. Mele, et al. 2009. Overexpression of Mn superoxide dismutase does not increase life span in mice. *J. Gerontol. A Biol. Sci. Med. Sci.* 64:1114–1125.
- Johnson, M. S., S. C. Thomson, and J. R. Speakman. 2001. Limits to sustained energy intake II. Inter-relationships between resting metabolic rate, life-history traits and morphology in *Mus musculus*. *J. Exp. Biol.* 204:1937–1946.
- Kaushik, S., and J. Kaur. 2003. Chronic cold exposure affects the antioxidant defense system in various rat tissues. *Clin. Chim. Acta* 333:69–77.
- Keipert, S., A. Voigt, and S. Klaus. 2011. Dietary effects on body composition, glucose metabolism, and longevity are modulated by skeletal muscle mitochondrial uncoupling in mice. *Aging Cell* 10:122–136.
- Kenyon, C. 2011. The first long-lived mutants: discovery of the insulin/IGF-1 pathway for ageing. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 366:9–16.
- Kirkwood, T. B., and A. Kowald. 2012. The free-radical theory of ageing—older, wiser and still alive: modelling positional effects of the primary targets of ROS reveals new support. *BioEssays* 34:692–700.
- Krol, E., and J. R. Speakman. 2003. Limits to sustained energy intake. VII. Milk energy output in laboratory mice at thermoneutrality. *J. Exp. Biol.* 206:4267–4281.
- Lewis, K. N., B. Andziak, T. Yang, and R. Buffenstein. 2013. The naked mole-rat response to oxidative stress: just deal with it. *Antioxid. Redox Signal.* 19:1388–1399.
- Marko, G., D. Costantini, G. Michl, and J. Torok. 2011. Oxidative damage and plasma antioxidant capacity in relation to body size, age, male sexual traits and female reproductive performance in the collared Xycatcher (*Ficedula albicollis*). *J. Comp. Physiol. B* 181:73–81.
- Meitern, R., E. Sild, K. Kilk, R. Porosk, and P. Horak. 2013. On the methodological limitations of detecting oxidative stress: effects of paraquat on measures of oxidative status in greenfinches. *J. Exp. Biol.* 216:2713–2721.
- Metcalfe, N. B., and C. Alonso-Alvarez. 2010. Oxidative stress as a life-history constraint: the role of reactive oxygen species in shaping phenotypes from conception to death. *Funct. Ecol.* 24:984–996.
- Metcalfe, N. B., and P. Monaghan. 2013. Does reproduction cause oxidative stress? An open question. *Trends Ecol. Evol.* 28:347–350.
- Monaghan, P., N. B. Metcalfe, and R. Torres. 2009. Oxidative stress as a mediator of life history trade-offs: mechanisms, measurements and interpretation. *Ecol. Lett.* 12:75–92.
- Nussey, D. H., J. M. Pemberton, J. G. Pilkington, and J. D. Blount. 2009. Life history correlates of oxidative damage in a free-living mammal population. *Funct. Ecol.* 23:809–817.
- Oldakowski, L., Z. Piotrowska, K. M. Chrzaszcik, E. T. Sadowska, P. Koteja, and J. R. Taylor. 2012. Is reproduction costly? No increase of oxidative damage in breeding bank voles. *J. Exp. Biol.* 215:1799–1805.
- Passos, J. F., G. Nelson, C. Wang, T. Richter, C. Simillion, C. J. Proctor, et al. 2010. Feedback between p21 and reactive oxygen production is necessary for cell senescence. *Mol. Syst. Biol.* 6:347.
- Pearl, R. L. 1928. *The rate of living*. Alfred Knopf, New York, USA.
- Perez, V. I., C. M. Lew, L. A. Cortez, C. R. Webb, M. Rodriguez, Y. Liu, et al. 2008. Thioredoxin 2 haploinsufficiency in mice results in impaired mitochondrial function and increased oxidative stress. *Free Radic. Biol. Med.* 44:882–892.
- Perez, V. I., A. Bokov, H. Van Remmen, J. Mele, Q. Ran, Y. Ikeno, et al. 2009a. Is the oxidative stress theory of aging dead? *Biochim. Biophys. Acta* 1790:1005–1014.
- Perez, V. I., R. Buffenstein, V. Masamsetti, S. Leonard, A. B. Salmon, J. Mele, et al. 2009b. Protein stability and resistance to oxidative stress are determinants of longevity in the longest-living rodent, the naked mole-rat. *Proc. Natl. Acad. Sci. USA* 106:3059–3064.
- Perez, V. I., H. Van Remmen, A. Bokov, C. J. Epstein, J. Vijg, and A. Richardson. 2009c. The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell* 8:73–75.
- Peterson, B. L., J. Larson, R. Buffenstein, T. J. Park, and C. P. Fall. 2012. Blunted neuronal calcium response to hypoxia in naked mole-rat hippocampus. *PLoS ONE* 7:e31568.
- Rashidi, A., T. B. Kirkwood, and D. P. Shanley. 2009. Metabolic evolution suggests an explanation for the weakness of antioxidant defences in beta-cells. *Mech. Ageing Dev.* 130:216–221.
- Rizzo, A., E. Ceci, M. Pantaleo, M. Mutinati, M. Spedicato, G. Minoia, et al. 2013. Evaluation of blood and milk oxidative status during early postpartum of dairy cows. *Animal* 7:118–123.
- Robert, K. A., and A. M. Bronikowski. 2010. Evolution of senescence in nature: physiological evolution in populations of garter snake with divergent life histories. *Am. Nat.* 175:147–159.
- Robert, K. A., A. Brunet-Rossinni, and A. M. Bronikowski. 2007. Testing the ‘free radical theory of aging’ hypothesis:

- physiological differences in long-lived and short-lived colubrid snakes. *Aging Cell* 6:395–404.
- Rodriguez, K. A., P. A. Osmulski, A. Pierce, S. T. Weintraub, M. Gaczynska, and R. Buffenstein. 2014. A cytosolic protein factor from the naked mole-rat activates proteasomes of other species and protects these from inhibition. *Biochim. Biophys. Acta* 1842:2060–2072.
- Sainz, R. M., R. J. Reiter, J. C. Mayo, J. Cabrera, D. X. Tan, W. Qi, et al. 2000. Changes in lipid peroxidation during pregnancy and after delivery in rats: effect of pinealectomy. *J. Reprod. Fertil.* 119:143–149.
- Salmon, A. B., S. Murakami, A. Bartke, J. Kopchick, K. Yasumura, and R. A. Miller. 2005. Fibroblast cell lines from young adult mice of long-lived mutant strains are resistant to multiple forms of stress. *Am. J. Physiol. Endocrinol. Metab.* 289:E23–E29.
- Salmon, A. B., A. Richardson, and V. I. Perez. 2010. Update on the oxidative stress theory of aging: does oxidative stress play a role in aging or healthy aging? *Free Radic. Biol. Med.* 48:642–655.
- Schmidt, C. M., J. D. Blount, and N. C. Bennett. 2014. Reproduction is associated with a tissue-dependent reduction of oxidative stress in eusocial female damara land mole-rats (*Fukomys damarensis*). *PLoS ONE* 28; 9: e103286.
- Schneeberger, K., G. A. Czirjak, and C. C. Voigt. 2014. Frugivory is associated with low measures of plasma oxidative stress and high antioxidant concentration in free-ranging bats. *Naturwissenschaften* 101:285–290.
- Schwanz, L., D. A. Warner, S. McGaugh, R. Di Terlizzi, and A. Bronikowski. 2011. State-dependent physiological maintenance in a long-lived ectotherm, the painted turtle (*Chrysemys picta*). *J. Exp. Biol.* 214:88–97.
- Schwartz, T. S., and A. M. Bronikowski. 2011. Molecular stress pathways and the evolution of life histories in reptiles. Pp. 193–209 *in* T. Flatt and A. Heyland, eds. *Mechanisms of Life History Evolution*, Oxford University Press, Oxford, UK.
- Schwartz, T. S., and A. M. Bronikowski. 2013a. Dissecting molecular stress networks: identifying nodes of divergence between life-history phenotypes. *Mol. Ecol.* 22:739–756.
- Schwartz, T. S., and A. M. Bronikowski. 2013b. Plasticity and evolution of stress response networks in divergent life-history phenotypes. *Integr. Comp. Biol.* 53:E192–E192.
- Selman, C., T. Grune, A. Stolzing, M. Jakstadt, J. S. McLaren, and J. R. Speakman. 2002a. The consequences of acute cold exposure on protein oxidation and proteasome activity in short-tailed field voles, *Microtus agrestis*. *Free Radic. Biol. Med.* 33:259–265.
- Selman, C., J. S. McLaren, A. R. Collins, G. G. Duthie, and J. R. Speakman. 2002b. Antioxidant enzyme activities, lipid peroxidation, and DNA oxidative damage: the effects of short-term voluntary wheel running. *Arch. Biochem. Biophys.* 401:255–261.
- Selman, C., J. S. McLaren, A. R. Collins, G. G. Duthie, and J. R. Speakman. 2008. The impact of experimentally elevated energy expenditure on oxidative stress and lifespan in the short-tailed field vole *Microtus agrestis*. *Proc. Biol. Sci.* 275:1907–1916.
- Selman, C., J. D. Blount, D. H. Nussey, and J. R. Speakman. 2012. Oxidative damage, ageing, and life-history evolution: where now? *Trends Ecol. Evol.* 27:570–577.
- da Silva, A. C. A., T. B. Salomon, C. S. Behling, J. Putti, F. S. Hackenhaar, P. V. G. Alabarse, et al. 2013. Oxidative stress in the kidney of reproductive female rats during aging. *Biogerontology* 14:411–422.
- Sorge, R. E., L. J. Martin, K. A. Isbester, S. G. Sotocinal, S. Rosen, A. H. Tuttle, et al. 2014. Olfactory exposure to males, including men, causes stress and related analgesia in rodents. *Nat. Methods* 11:629–632.
- Speakman, J. R. 2004. Oxidative phosphorylation, mitochondrial proton cycling, free-radical production and aging. Pp 35–68 *in* M.P. Mattson, eds. *Energy metabolism and lifespan determination*. Elsevier series *Advances in Cell Aging and Gerontology*. Elsevier, Amsterdam.
- Speakman, J. R. 2005. Body size, energy metabolism and lifespan. *J. Exp. Biol.* 208:1717–1730.
- Speakman, J. R., and M. Garratt. 2014. Oxidative stress as a cost of reproduction: beyond the simplistic trade-off model. *BioEssays* 36:93–106.
- Speakman, J. R., and E. Krol. 2011. Limits to sustained energy intake. XIII. Recent progress and future perspectives. *J. Exp. Biol.* 214:230–241.
- Speakman, J. R., and C. Selman. 2011. The free-radical damage theory: accumulating evidence against a simple link of oxidative stress to ageing and lifespan. *BioEssays* 33:255–259.
- Stier, A., S. Reichert, S. Massemin, P. Bize, and F. Criscuolo. 2012. Constraint and cost of oxidative stress on reproduction: correlative evidence in laboratory mice and review of the literature. *Front. Zool.* 9.
- Stier, A., P. Bize, C. Habold, F. Bouillaud, S. Massemin, and F. Criscuolo. 2014. Mitochondrial uncoupling prevents cold-induced oxidative stress: a case study using UCPI knockout mice. *J. Exp. Biol.* 217:624–630.
- Stuart, J. A., L. A. Maddalena, M. Merilovich, and E. L. Robb. 2014. A midlife crisis for the mitochondrial free radical theory of aging. *Longev. Healthspan* 3:4.
- Tan, D. X., R. Hardeland, L. C. Manchester, S. D. Paredes, A. Korkmaz, R. M. Sainz, et al. 2010. The changing biological roles of melatonin during evolution: from an antioxidant to signals of darkness, sexual selection and fitness. *Biol. Rev. Camb. Philos. Soc.* 85:607–623.
- Tinbergen, J. M., and S. Verhulst. 2000. A fixed energetic ceiling to parental effort in the great tit? *J. Anim. Ecol.* 69:323–334.
- Urison, N. T., and R. B. Buffenstein. 1995. Metabolic and body-temperature changes during pregnancy and lactation



- in the naked mole-rat (*Heterocephalus glaber*). *Physiol. Zool.* 68:402–420.
- Velando, A., R. Torres, and C. Alonso-Alvarez. 2008. Avoiding bad genes: oxidatively damaged DNA in the germ line and mate choice. *BioEssays* 30:1212–1219.
- Veskoukis, A. S., M. G. Nikolaidis, A. Kyparos, and D. Kouretas. 2009. Blood reflects tissue oxidative stress depending on biomarker and tissue studied. *Free Radic. Biol. Med.* 47:1371–1374.
- Vezina, F., and T. D. Williams. 2002. What drives the metabolic costs of egg production in birds? *Integr. Comp. Biol.* 42:1327–1328.
- Welcker, J., B. Moe, C. Bech, M. Fyhn, J. Schultner, J. R. Speakman, et al. 2010. Evidence for an intrinsic energetic ceiling in free-ranging kittiwakes *Rissa tridactyla*. *J. Anim. Ecol.* 79:205–213.
- Wiersma, P., C. Selman, J. R. Speakman, and S. Verhulst. 2004. Birds sacrifice oxidative protection for reproduction. *Proc. Biol. Sci.* 271(Suppl 5):S360–S363.
- Wilhelm, D., S. L. Althoff, A. L. Dafre, and A. Boveris. 2007. Antioxidant defenses, longevity and ecophysiology of South American bats. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 146:214–220.
- Williams, T. D., F. Vezina, and J. R. Speakman. 2009. Individually variable energy management during egg production is repeatable across breeding attempts. *J. Exp. Biol.* 212:1101–1105.
- Xu, Y. C., D. B. Yang, J. R. Speakman, and D. H. Wang. 2014. Oxidative stress in response to natural and experimentally elevated reproductive effort is tissue dependent. *Funct. Ecol.* 28:402–410.
- Yang, W., J. Li, and S. Hekimi. 2007. A measurable increase in oxidative damage due to reduction in superoxide detoxification fails to shorten the life span of long-lived mitochondrial mutants of *Caenorhabditis elegans*. *Genetics* 177:2063–2074.
- Yang, D. B., Y. C. Xu, D. H. Wang, and J. R. Speakman. 2013. Effects of reproduction on immuno-suppression and oxidative damage, and hence support or otherwise for their roles as mechanisms underpinning life history trade-offs, are tissue and assay dependent. *J. Exp. Biol.* 216:4242–4250.
- Zhang, Y. Q., Y. Ikeno, W. B. Qi, A. Chaudhuri, Y. Li, A. Bokov, et al. 2009. Mice deficient in both Mn superoxide dismutase and glutathione peroxidase-1 have increased oxidative damage and a greater incidence of pathology but no reduction in longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* 64:1212–1220.