



UNIVERSITI PUTRA MALAYSIA

MORPHOLOGICAL AND MOLECULAR CHARACTERISATION OF ETHANOLIC NEEM (AZADIRACHTA INDICA) LEAF EXTRACT IN AN IN VIVO BREAST CANCER MODEL

LAM TSUEY PENG.

IB 2007 4

MORPHOLOGICAL AND MOLECULAR CHARACTERISATION OF ETHANOLIC NEEM (Azadirachta indica) LEAF EXTRACT IN AN IN VIVO BREAST CANCER MODEL

By

LAM TSUEY PENG

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malavsia, in Fulfillment of the Requirements for the Degree of Master of Science

)

October 2007



Specially dedicated to,

My beloved mother, sister, brother, David Chieng, and all my family

members

For their invaluable love, understanding, encouragement and patience

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

MORPHOLOGICAL AND MOLECULAR CHARACTERISATION OF ETHANOLIC NEEM (Azadirachta indica) LEAF EXTRACT IN AN IN VIVO BREAST CANCER MODEL

By

LAM TSUEY PENG

October 2007

Chairman	:	Professor	Fauziah	Othman.	PhD
----------	---	-----------	---------	---------	-----

Faculty : Institute of Bioscience

Breast cancer is the commonest cause of cancer death in women worldwide and Malaysia in all ethnic groups and all age groups. Neem's (*Azadirachta indica*) ability as a medicinal herb is traced as far back as 4500 years ago. Some of the impressive therapeutic qualities have been discovered such as anti-viral, anti-microbial, anti-inflammatory, anti-tumour, anti-bacterial, anti-fungal and anti-hyperglycemic; however the anticancer effect of ethanolic Neem leaves extract against breast cancer has not been documented. Besides this, Neem was found to induce apotosis in MCF-7 breast cancer cell line in local study recently. Thus, this study was done to evaluate the effect of ethanolic Neem leaves extract as apoptosis inducer in *in vivo* 4T1 breast cancer model. Two different concentrations of Neem, 250 mg/kg and 500

jii



mg/kg were tested on 4T1 breast cancer model. The 4T1 breast cancer models were evaluated by light microscopy, transmission electron microscopy for morphological changes, TUNEL assay for apoptotic cell labeling and in situ RT-PCR for c-myc, c-erbB2 and c-fos oncogene expressions. All treatment groups exhibited a higher incidence of apoptosis compared to untreated group from morphological analysis and TUNEL assay. The cancerous mice treated with both different concentration of Neem showed significantly higher value (p<0.05) in mean body weight, mean apoptotic index and mean apoptotic score compared to the control group. At the same time both group were showing a significantly lower value of mean mitotic index in histological evaluation. The mean tumour volume and mass proved that there was evidence of tumour regression in Neem treated mice. However, the overall observation showed that 500 mg/kg of Neem has more significant effect (p <0.05) of inducing apoptosis in the 4T1 breast cancer cells compared to 250 mg/kg of Neem. Furthermore, the 500 mg/kg Neem concentration has significantly lengthened the mean survival time by 44.62% in the 4T1 breast cancer model (p <0.05). Neem 500 mg/kg group also showed a better suppression of c-myc, c-erbB2 and c-fos oncogenes expression in mean distribution and intensity score (p< 0.05) in the 4T1 breast cancer model. By considering all the three down regulated oncogenes (c-myc, c-erbB2 and c-fos) under effect of Neem 500 mg/kg together, it becomes clearer that Neem 500 mg/kg was effective in inducing apoptosis in the 4T1 breast cancer



model. In conclusion, the Neem 500 mg/kg treatment was effective in inducing cell death via apoptosis and regulates cell proliferation in 4T1 breast cancer model. Its effectiveness was proportional to the concentration of Neem treatment given.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KAJIAN MORFOLOGI DAN MOLEKULAR EKSTRAK ETHANOL DAUN NEEM (Azadirachta indica) KE ATAS MODEL KANSER PAYU DARA IN VIVO

Oleh

LAM TSUEY PENG

Oktober 2007

Pengerusi : Profesor Fauziah Othman, PhD

Fakulti : Institut Biosains

Kanser payu dara ialah kanser terkenal yang mengakibatkan kematian bagi wanita sedunia dan Malaysia bagi semua kaum dan kumpulan umur. Keberkesanan Neem (*Azadirachta indica*) sebagai herbal perubatan telah dikaji semenjak 4500 tahun yang lalu. Antara terapeutik kualitinya yang kagum yang telah dijumpai adalah seperti anti-viral, anti-mikrobial, anti-radang, anti-tumor, anti-bakteria, anti-fungus dan anti-hiperglisemic; tetapi kesan anti-kanser dari ekstrak etanol daun Neem terhadap kanser payu dara belum pernah didokumentasi. Di samping itu, Neem telah dikesani bahawa mendorong apoptotis pada MCF-7 kanser sel payu dara oleh kajian tempatan kebelakangan ini. Jadi, kajian ini dijalankan untuk menilai kesan etanol ekstrak daun Neem sebagai pemangkin apoptosis ke atas model kanser payu dara 4T1 kanser payu dara secara *in vivo*. Dua kepekatan Neem



vi

yang berlainan, 250 mg/kg and 500 mg/kg telah diuji ke atas model kanser payu dara 4T1. Model kajian yang diuji telah dinilai melalui mikroskop cahaya, mikroskop transmisi elektron untuk mengkaji perubahan morfologi, ujian TUNEL untuk label sel apoptosis dan in situ RT-PCR untuk mengkaji ekspresi c-myc, c-erbB2 dan c-fos. Semua kumpulan rawatan mempamerkan insiden apoptosis yang lebih tinggi berbanding kepada kumpulan tanpa rawatan di bawah bukti uraian morfologi dan ujian TUNEL. Tikus kanser yang diubati dengan dua jenis penumpuan Neem yang berlainan menunjukkan nilai yang lebih tinggi dan ketara dari segi purata berat badan, purata indeks apoptasis dan purata markah apoptosis berbanding dengan kumpulan kawalan. Dalam kedua-dua kumpulan tersebut masa yang sama, menunjukkan nilai yang lebih rendah dengan ketaranya bagi purata indeks mitotic dalam penilaian histologi. Purata kandungan dan berat tumor telah membuktikan bahawa adanya kemunduran tumor bagi tikus kanser yang menerima perubatan Neem. Tetapi, pemerhatian keseluruhan menunjukkan bahawa 500 mg/kg Neem mempunyai kesan yang lebih ketara (p < 0.05) dalam memangkin apoptosis dalam 4T1 sel kanser payu dara berbanding kepada 250 mg/kg Neem. Tambahan pula, 500 mg/kg Neem telah memanjangkan purata masa hidup sebanyak 44.62 % dalam model kanser payu dara 4T1 dengan ketara (p < 0.05). Kumpulan Neem 500 mg/kg juga menunjukkan penindasan yang lebih bagus bagi ekpresi onkogen c-myc, c-erbB2 dan c-fos bagi purata markah taburan dan kekuatan (p< 0.05) di

vii

dalam model kanser payu dara 4T1. Dengan menimbangkan kesemua tiga onkogen (c-myc, c-erbB2, c-fos) yang ditindas di bawah kesan 500 mg/kg Neem sekali, adalah lebih jelas bahawa 500 mg/kg Neem berupaya untuk menuju ke arah apoptosis di dalam model kanser payu dara 4T1. Kesimpulannya, rawatan 500 mg/kg Neem adalah berkesan dalam mendorong kematian sel melalui apoptosis dan pengawalan pembahagian sel dalam model kanser payu dara 4T1. Tahap keberkesanan tersebut adalah bergantung kepada kepekatan Neem yang diberi dalam rawatan.





ACKNOWLEDGEMENTS

I would like to acknowledge a number of people, without their support the completion of this dissertation would not be possible. First, I would like to thank my research advisor, Prof. Dr Fauziah Othman, for revealing to me the beauty of science and teaching me many valuable scientific techniques; for her great guidance, support, and endless optimism and smile, which have been a constant source of inspiration and encouragement. Special thanks to my graduate committee: Assoc. Prof. Dr. Asmah Rahmat, Dr. Sharida Fakurazi, and Dr. Chong Pei Pei for their critical remarks, guidance and invaluable discussions of my project.

Thanks to my friends and colleagues: Mun Yee, Hanim, Pele, Shahrul, Hernani, Mahani, Lee Yean and Phelim for their unconditional help, support, and friendship throughout all these years. Thanks to everyone in the Microscopy Imaging and Nanoscience Unit, Institute of Biosciences: (Mr. Ho, Azilah, Aini, Rafi, Ida) sharing their priceless experiences and unsparing assistance throughout the entire microscopy work of this research.

My sincere gratitude is also accorded to Prof. Dr Nordin Hj. Lajis, Laboratory of Natural Products, Institute of Biosciences for his kindness in providing the facilities of leaves extraction to start on this study. I would also like to

ix

acknowledge and thank Prof. Datin Paduka Dr Khatijah Mohd. Yusoff from Faculty of Biotechnology, Universiti Putra Malaysia for her generosity in providing the animal house facility to make this study possible.

I would also like to express my love and gratefulness to my mother, Tan Yook Lin, my sister, Lam Tsuey Yun and my brother, Lam Boon Chin, for their patience, love, and support. Not forgetting also my dear colleague at National Blood Centre for their kind understanding and support during my thesis writing. Most importantly, I would like to thank my boy friend, David Chieng for his love, understanding, and encouragement for being a constant source of joy and surprises. I certify that an Examination Committee has met on 22nd October 2007 to conduct the final examination of Lam Tsuey Peng on her Master of Science thesis entitled "Morphological and Molecular Characterisation of Ethanolic Neem (*Azadirachta indica*) Leaf Extract in an *In Vivo* Breast Cancer Model" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the degree of Master of Science.

Members of the Examination Committee were as follows:

Rozita Rosli, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Khatiza Haida Ali, PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Internal Examiner)

Abdah Md. Akim, PhD

Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Internal Examiner)

Yasmin Anum Mohd Yusof, PhD

Associate Professor Faculty of Medicine Universiti Kebangsaan Malaysia (External Examiner)

HASANAH MOHD. GHAZALI, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 22 November 2007





This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Fauziah Othman. PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Asmah Rahmat, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Sharida Fakurazi, PhD

Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Chong Pei Pei, PhD

Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Æ

AINI IDERIS, PhD Professor and Dean School of Graduate Studies Univesiti Putra Malaysia

Date: 13 December 2007





DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

LAM TSUEY PENG

Date: 22 November 2007





TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	Ш
ABSTRAK	vi
ACKNOWLEDGEMENTS	ix
APPROVAL	xiii
DEECLARATION	xiii
LIST OF TABLES	xvi
LIST OF FIGURES	XX
LIST OF ABBREVIATIONS	xxvii

CHAPTER

1	INTROD	UCTION	
	1.1	Breast Cancer	1
	1.2	Neem (Azadirachta indica)	2
	1.3	Objectives of the Study	4
2	LITERA	TURE REVIEW	5
	2.1	Anticancer Agent Research	5
	2.2	Genetic Aspect of Cancer	6
	2.3	Breasts	7
	2.4	Epidemiology of Breast Cancer	9
	2.5	Risk Factors for Breast Cancer	12
		2.5.1 Age	13
		2.5.2 Age at Menarche and Menopause	13
		2.5.3 Age at First Pregnancy	14
		2.5.4 Family History	14
		2.5.5 Previous Benign Breast Disease	16
		2.5.6 Radiation	16
		2.5.7 Lifestyle	17
		2.5.8 Oral Contraceptive	17
		2.5.9 Hormone Replacement Therapy (HRT)	18
	2.6	Pathology of Breast Cancer	19
	2.7	Treatment and Prevention of Breast Cancer	19
		2.7.1Tamoxifen – Commercial Drug for Breast Cancer	21
	2.8	The Cell Cycle	23
	2.9	Proliferation Activity and Tumour Growth	24
		2.9.1 Mitotic Index (MI)	26
	2.10	. ,	27
	2.11	Apoptosis and Breast Cancer	34
	2.12	Oncogenes	34
		2.12.1 C-myc	36

xiv

	2.12.2 C-erbB2	37
	2.12.3 C-fos	39
2.13	Animal Models for Breast Cancer	40
	2.13.1 4T1 breast cancer model	40
2.14	Medicinal Herbs and Cancer Research	41
2.15	Neem (Azadirachta indica)	42
	2.15.1 Botanical Description and Cultivation	42
	2.15.2 Elements in Neem Leaf	43
	2.15.3 Biological Properties of Neem	43
	2.15.4 Potential Anticancer Activity of Neem	45
2.16	TUNEL Assay - In situ Apoptotic Cell Labeling	47
2.17	In situ Reverse Transcription-Polymerase	49
	Chain Reaction (In situ RT-PCR)	
	2.17.1 Internal Control - Housekeeping Gene	50
METHO	DOLOGY	52
3.1	Experimental Study Design	53
3.2	Ethanolic Neem leaf Extract	54
3.3	Cell Culture	55
3.4	Animals	56
	3.4.1 Breast Cancer Induction	57
	3.4.2 Treatments	58
	3.4.3 Mean Survival Time (MST)	59
	3.4.4 Sample Collection	59
3.5	Light Microscopy	60
	3.5.1 Apoptotic / Mitotic Index	62
3.6	Transmission Electron Microscopy (TEM)	62
	3.6.1 Specimen Preparation for TEM	63
	3.6.2 Semithin Sectioning	64
	3.6.3 Ultrathin Sectioning	64
	3.6.4 Contrasting / Staining	65
3.7	Fluorometric TUNEL Assay	66
	3.7.1 Pretreatment of Paraffin-Embedded Tissues	66
	3.7.2 Positive Control	67
	3.7.3 Negative Control	68
	3.7.4 Apoptosis Detection	68
	3.7.5 Analysis of TUNEL Assay	70
3.8	In situ Reverse Transcription- Polymerase	70
5.0	Chain Reaction (RT-PCR)	11
	3.8.1 Tissue Section Preparation	71
	3.8.2 Pretest Preparation and Optimization	71
	3.8.2.1 Primer Designing	71
	, 0 0	74
	3.8.2.2 Isolating Genomic DNA From	14

3

`

xv



		4T1 cells	
		3.8.2.3 Gradient PCR	75
		3.8.2.4 Agarose Gel Electrophoresis	77
		3.8.2.5 PCR Purification by Gel	78
		Extraction	
		3.8.2.6 Automated DNA Sequencing	80
		and DNA Sequence	
		Analysis	
		3.8.2.7 Tissue Processing	80
		3.8.2.8 Proteolytic Digestion	81
		3.8.2.8.1 Proteinase K	81
		Concentration	
		Optimization	
		3.8.2.8.2 Proteolytic Digestion	82
		and DNase Treatment	
		3.8.3 One-step in situ RT-PCR Assay	82
		3.8.4 Immunodetection of PCR Products	84
		3.8.5 Preparing Controls in <i>in situ</i> RT-PCR	85
		3.8.6 Scoring System for in situ RT-PCR	86
4	RESULI	ſS	87
	4.1	Profile of Experimental Animals	87
		4.1.1 Mean Survival Time (MST)	87
		4.1.2 Body Weight Profile	89
		4.1.3 Tumour Volume Profile	81
		4.1.4 Tumour Mass Profile	93
	4.2	Histological Analysis	96
		4.2.1 Quantification of Apoptosis and Mitosis	101
	4.3	Effect of Neem on Ultrastuctural Changes	104
	4.4	Apoptotic Analysis by TUNEL Assay	110
		4.4.1 Controls for TUNEL Assay	110
		4.4.2 Apoptotic Cells by TUNEL Labeling	112
	4.5	4.4.3 Scoring of Apoptotic Cell	116
	4.5	Oncogene Expression	118
		4.5.1 Annealing Temperature Selection by	118
		Gradient PCR	140
		4.5.1.1 ß-actin	118
		4.5.1.2 c-myc 4.5.1.3 c-erbB2	119
		4.5.1.3 C-erbBz 4.5.1.4 c-fos	120 121
		4.5.2 DNA Sequence Analysis of Selected Gene	122
		4.5.3 Optimized Proteinase k Concentration	122
		4.5.3 Optimized Proteinase & Concentration 4.5.4 Scoring for <i>in situ</i> RT-PCR	122
		\neg	144

١

۰,

xvi



	4.5.5 Controls for in situ RT-PCR			125		
	4.5.6	ß-actin mRN/ Control	A Exp	ression as	Internal	126
	4.5.7	Localisation Expression	of	c-myc	mRNA	128
	4.5.8	Localisation Expression	of	c-erbB2	mRNA	130
	4.5.9	Localisation o	f c-fos	s mRNA Ex	pression	132
5	DISCUSSION					133
6	CONCLUSION					147
	6.1 Future	e Research Re	comm	endations		147
R	EFERENCES					149
AF	APPENDICES				160	
BIODATA OF THE AUTHOR			184			
LI	LISTS OF PUBLICATIONS			185		





LIST OF TABLES

Table		Page
2.1	Risk factors for breast cancer (Bennett et al., 2001)	12
2.2	Morphological and biochemical differences between accidental (necrosis) and programmed (apoptosis) cell death (Vermes <i>et al.</i> , 1997)	33
3.1	The reagent involved and the tissue processing time used for the tissue processing by an automated tissue processor (Leica ASP 300)	60
3.2	Reagents involved and its immersing time for the staining with Hematoxylin and Eosin (H&E)	61
3.3	Preparation of rTdT incubation buffer for experimental and positive control reactions	69
3.4	Oligonucleotide primer sequences of c-myc. c-fos. c-erbB2 and ß-actin	73
3.5	The final reaction mix with appropriate volume and concentration for each sample and control during PCR	76
3.6	Thermal cycler condition of Gradient PCR amplification	77
3.7	The final reaction mix with appropriate volume and concentration for each sample RT-PCR	83
3.8	Thermal cycler condition for each set of primers in gradient PCR	84
4.1	MST and percentage of increase in lifespan in experimental groups under effect of Neem	88
C1	Mean body weight of 4T1 breast cancer model treated with Neem	182
C2	Mean tumor volume of 4T1 breast cancer model treated with Neem	183

xviii

- C3 Mean tumor mass of 4T1 breast cancer model treated 183 with Neem
- C4 Mean percentage of MI and AI in Neem treated 4T1 184 mouse model according to sampling time
- C5 Mean apoptotic score with different sampling time 185 among the studied groups
- C6 mRNA Expression of the positive reaction and intensity 186 for positive cells for different, c-myc, c-erbB2, c-fos and ß-actin mRNA by breast tumor cells in 4T1 mouse model as detected by *in situ* RT-PCR

;



LIST OF FIGURES

Figure		Page
1.1	A photo of Neem leaves	2
2.1	Sagital view of the breast. The breast consist of lobules, ducts (connect the lobules to the nipple), and stroma (fatty tissue and connective tissue surrounding the ducts and lobules. (American cancer society, 2004)	8
2.2	Age-standardized incidence and mortality rates for breast cancer. Data shown per 100,000 (Parkin <i>et al.</i> , 2005)	11
2.3	Illustration of cell number in normal condition of cell cycle. The proliferative index is balanced with apoptotic rate	25
2.4	Illustration of cell number of tumour tissue in cell cycle. The expansion of tumour cells is achieved by an increased proliferative index and by a decreased apoptotic rate	25
2.5	Schematic representation of necrotic cell death. Upon being triggered to undergo necrosis (generally a pathological or severely injurious stimulus), a normal cell (A) begins to swell as a consequence of an increase in cell membrane permeability (B), which is followed by high amplitude swelling of the nucleus and organelles and flocculation of the nuclear chromatin (C). Finally, lysis of the cell occurs, thereby spilling its contents into the extracellular space potentially damaging neighbouring cells and provoking inflammation (D) (Seamus and Douglas, 1995)	31
2.6	Schematic representation of apoptotic cell death. Upon being triggered to undergo apoptosis (generally a physiological or mild pathological stimulus), a normal	32

cell (A) condenses its cytoplasm and DNA (B), and proceeds to fragment into many intact vesicles (apoptotic bodies), many containing fragments of



хх

condensed chromatin and morphologically normal organelles (C), that are then recognized and engulfed by neighbouring phagocytes (D). Apoptotic cells generally do not provoke an inflammatory response (Seamus and Douglas, 1995)

- 3.1 The overview of experimental study design. Seven 53 experimental groups were created to study the effect of Neem against 4T1 breast cancer model.
- 3.2 Number of female balb/c mice used in various 56 treatment groups.
- 4.1 The effect of Neem on mean body weight changes in 90 4T1 breast cancer model
- 4.2 The effect of Neem on mean tumour volume changes 92 in 4T1 breast cancer model
- 4.3 The effect of Neem on mean tumour mass changes in 93 4T1 breast cancer model
- 4.4 The effect of Neem on mean body weight changes in 94
 4T1 breast cancer model without tumour in last sampling
- 4.5 Appearance of 4T1 cells induced breast cancer tumour 95 in Balb/C mice with its respective harvested tumour of the cancerous groups. Note the regression of tumour size in CN 500 group (showed in ruler scale cm) compared to CC and CN 250 groups.
- 4.6 Light microscopy of breast tissue from normal group. A 98 typical mammary duct (arrow) lined with low cuboidal epithelium surrounded fat tissue. Note the artificial tearing effect of the section (Magnification x200)
- 4.7 Light microscopy of breast tissue from normal with 98 treatment of 250 mg/kg of Neem group. Note the mammary ductule was normal without any inflammation found (Magnification x200).

2

4.8 Light microscopy of breast tissue from normal with 98 treatment of 500 mg/kg of Neem group. A typical mammary duct surrounded by the myoepithelial cells

xxi

(arrow). Note the cells were normal without any inflammation (Magnification x200)

- 4.9 Light microscopy of breast tumour from cancer control 99 group. Noted most of the cancer cells were mitotic with prominent swollen nucleus associated with chromatin clot (blue arrowheads). The staining was dense compared to the cancer treated groups (Magnification x 200). Enlargement: (i) Mitotic cell in metaphase (ii) Mitotic cell in early anaphase
- 4.10 Light microscopy of breast tumour from cancer with 99 treatment of 250 mg/kg of Neem group. Noted the mitotic figure (blue arrowheads) were dominant than apoptotic figure (green arrowheads). An apoptotic figure of halo shape was observed (Magnification x200). Enlargement: (i) Apoptotic cell with halo shape resulted by segregation from neighbor cells (ii) Mitotic cell in late anaphase
- 4.11 Light microscopy of breast tumour from cancer with 100 treatment of 500 mg/kg Neem group. Note the frequency of apoptotic figure (green arrowheads) were increase than apoptotic figure in CN 250. However, the mitotic figure also visible (blue arrowheads) (Magnification x200). Enlargement: (i) & (ii) Apoptotic cell halo shape
- 4.12 Light microscopy of breast tumour from cancer with 100 treatment of 0.5 µg/mL tamoxifen citrate group. Note the apoptotic figures (green arrowheads) were dominant in the plane of view (Magnification x200).
 Enlargement: (i) & (ii) Apoptotic cell with nucleus condensation
- 4.13 Mean apoptotic index changes of 4T1 breast cancer 102 mouse model under effect of Neem
- 4.14 Mean mitotic index changes of 4T1 breast cancer 103 mouse model under effect of Neem
- 4.15 Transmission electron microscopy of cells around the 104 mammary ductule of the mice in normal control group with a mirovillus (arrow) on the outer layer of the low

XXII



cuboidal epithelium

- 4.16 Transmission electron microscopy of a mitotic 4T1 cell 105 in cancer control group at the stage of early anaphase.
 Note the hairy extensions of the chromosomes clots in the nucleus which was started to divide into two portions (red arrow)
- 4.17 Transmission electron micrscopy of an apoptotic 4T1 107 cell from 250 mg/kg of Neem treated group. Note the condensed and vacuolized nucleus of the cell.
- 4.18 Transmission electron microscopy of late apoptotic 108 4T1 cells treated with 500 mg/kg of Neem. Note a piece of cytoplasm (apoptotic body) was tearing apart from the originate cell (arrow). Nucleus of the neighboring cell (*) was vacuolated.
- 4.19 Transmission electron microscopy of enlarged nucleus 108 vacuolization found in Figure 4.15 at higher magnification (*)
- 4.20 Transmission electron microscopy of apoptotic 4T1 cell 109 treated with tamoxifen citrate. Note the cell was isolated from the neighbouring cells. The membrane of the cell was blebbed with highly condensed and vacuolized nucleus
- 4.21 TUNEL labeling of positive control of 4T1 breast 111 cancer tissue pretreated with DNase 1 (Magnification x 400)
- 4.22 TUNEL labeling of negative control of 4T1 breast 111 cancer tissue without rTdT enzyme (Magnification x 400)
- 4.23 TUNEL labeling of (a) normal group; (b) Normal with 114 250 mg/kg of Neem treatment; (c) Normal with 500 mg/kg of Neem treatment. No green fluorescent stain was noted on the section (Magnification x 600)
- 4.24 TUNEL labeling of cancer control group. Note the 115 propidium iodide staining was dominant on the section. Mitotic figure in anaphase (arrow) was observed

xxiii



(Magnification x 600)

- 4.25 TUNEL labeling of cancer with treatment of 250 mg/kg 115 of Neem group. Note small dots of green fluorescent stained among the chromosomes clot in the nucleus overlapping the PI red fluorescent staining for the DNA nucleus (Magnification x 600)
- 4.26 TUNEL labeling of cancer with treatment of 500 mg/kg 115 of Neem group. Note the some of the cells was stained with dense green fluorescent (arrow) (Magnification x 600)
- 4.27 TUNEL labeling of cancer treatment with tamoxifen 115 citrate group. Note the nuclei were stained with dense red and green fluorescent (Magnification x 600)
- 4.28 Mean apoptotic score changes of 4T1 breast cancer 114 mouse model under effect of Neem
- 4.29 Gel electrophoresis of gradient PCR amplification of 118 ß-actin gene 4T1 DNA template. The optimized annealing temperature for ß-actin was 52.8°C (circle) with the target of 540 base pairs
- 4.30 Gel electrophoresis of gradient PCR amplification of 119 c-myc gene 4T1 DNA template. The optimized annealing temperature for c-myc was 52.8°C (circle) with the target of 281 base pairs
- 4.31 Gel electrophoresis of gradient PCR amplification of 120 c-erbB2 gene 4T1 DNA template. The optimized annealing temperature for c-erbB2 was 60.0°C (circle) with the target of 570 base pairs
- 4.32 Gel electrophoresis of gradient PCR amplification of 121 c-fos gene 4T1 DNA template. The optimized annealing temperature for c-fos was 60.0°C (circle) with the target of 241 base pairs
- 4.33 Mean distribution score of breast cancer oncogenes 123 signals in 4T1 breast cancer mouse model under effect of Neem

xxiv

