View metadata, citation and similar papers at core.ac.uk

NASA TECHNICAL MEMORANDUM

NASA TM X-62,411

(NASA-TM-X-62411) AMINO ACIDS IN A FISCHER N75-21034 TROPSCH TYPE SYNTHESIS (NASA) 10 p HC \$3.25 CSCL 06C Unclas G3/55 14684

AMINO ACIDS IN A FISCHER TROPSCH TYPE SYNTHESIS

Donald L. Brown Department of Chemistry Santa Clara University Santa Clara, Calif. 95053

and

NASA TM X-62,411

James G. Lawless Ames Research Center Moffett Field, Calif. 94035



August 1974

1 Report No	2 Government Acces	ing No	3 Recipient's Catalon	No			
NASA TM X-62,411							
4. Title and Subtitle	5. Report Date						
AMINO ACIDS IN A FISCHER	6. Performing Organiz	ation Code					
7. Author(s)	· · · · · · · · · · · · · · · · · · ·	8. Performing Organiz	ation Report No.				
Donald L. Brown and James	· .	A-5909					
9. Performing Organization Name and Address	10. Work Unit No. $102 \text{ FE} - 61 - 07$						
Department of Chemistry	192-55-01-0	No.					
Santa Clara University	х						
Santa Clara, Calli. 95053		13. Type of Report and Period Covered					
12. Sponsoring Agency Name and Address		Technical Memorandum					
National Aeronautics and Washington, D.C. 20546	14. Sponsoring Agency	Code					
15. Supplementary Notes				· .			
• • • • • • • • • • • • • • • • • • •		,					
		·					
16. Abstract							
that they were formed during the condensation of the solar nebula. A viable laboratory simulation of these conditions can be modeled after the industrial Fischer Tropsch reaction, which is known to produce organic compounds called hydrocarbons. In this simulation, a mixture of carbon monoxide, hydrogen and ammonia is heated in the presence of iron meteorite. We examined the reaction products for amino acids, a class of organic compounds important to life. A large number of these compounds is found in meteorites and other chemical evolution experiments. Only small quantities of a few amino acids were found in the present simulation work. These results are at odds with the existing literature in which many amino acids were reported. Because of the relevance of this work, future laboratory simulations will be designed to maximize the yields of these compounds.							
17. Key Words (Suggested by Author(s))		18. Distribution Statement					
Amino acids		Unclassified - Unlimited					
Chemical evolution	Chemical evolution						
Meteorites		STA	R Category -	55			
10 Descriter Classif (of this suggest)	20 Security Classif 4	of this page)	21. No. of Pages	22, Price*			
19. Security Gasuri, (or this report)	Line 2 and 2 for			¢7.05			
Unclassified		ea	/	<u> </u>			

*For sale by the National Technical Information Service, Springfield, Virginia 22151

AMINO ACIDS IN A FISCHER TROPSCH TYPE SYNTHESIS

Donald L. Brown and James G. Lawless. Ames Research Center

The Oparin-Haldane (1,2) hypothesis of chemical evolution postulates that simple precursors formed complex organic molecules before life appeared on earth, and much experimental data has been obtained in the laboratory to support this hypothesis (3). This hypothesis could be further substantiated by a natural evidence for chemical evolution, and recently, a number of publications have appeared which indicate the indigenous nature of organic compounds in extraterrestrial samples, particularly meteorites (4-7). For the first time it appears possible to begin to define the organic compounds characteristic to an extraterrestrial abiotic synthesis. Because of the importance of amino acids to terrestrial life, much of the meteorite analyses has focused on these compounds (4,5,7). The apparent racemic nature of amino acids and the random distribution of structural isomers suggest that they are indigenous, and are not the result of contamination. These findings have induced others to look in more detail at the constituent amino acids produced by electric discharge through an atmosphere of CH_4 , NH_3 , H_2O , NH_4C1 and N_2 . (8,9) Many of the amino acids present in the meteorite were found in the electric discharge reaction, and it was suggested that the amino acids found in carbonaceous chondrites could have been produced on the meteorite parent body by a similar mechanism.

An alternative postulation is that these organic compounds were formed in the solar nebula during accretion (10). Anders and co-workers believed that a viable laboratory simulation of solar nebular conditions might be modeled after the industrial Fischer Tropsch reaction, which is known to produce hydrocarbons. They have therefore studied Fischer-Tropsch Type (FTT) reactions in which they heated a mixture of CO, H_2 , and NH_3 in the presence of iron meteorite. A number of amino acids have been identified as products of FTT reactions (11), however, in light of the more complex suites of amino acids found in meteorites and discharge experiments, we have examined several FTT reactions to determine if meteoritic amino acids could be accounted for by this process.

Experimental

The FTT reactions were carried out in a quartz tube equipped with a 500 ml bulb, 4 ml vacuum stopcock, and a side tube for introducing catalyst and removing reaction products. During heating, the quartz tube was submerged to a depth of 9 cm in a tubular furnace while the remainder of the reaction flask was outside the furnace where reaction products could condense.

The catalysts, 300 mg of alumina and 200 mg of Canyon Diablo (CD) iron meteorite, were placed in the quartz tube and oxidized-reduced to remove organic contaminants. First oxygen gas at 1 atm was placed in the reaction flask and the catalysts were baked at 700°C for 7 hr. After reaction the oxygen was pumped off and hydrogen gas was introduced at 1 atm and the catalysts were reduced at 400°C for 4 hr. The Canyon Diablo meteorite was supplied by

1

Dr. R. Hayatsu, University of Chicago; the alumina was "Baker Analyzed" reagent grade chromatography support.

After oxidation-reduction of the catalysts, the reaction gases, CO:H₂:NH₃, were mixed in the reaction flask in a 1:1:1 molar ratio with a total pressure of 1 atm. For each FTT run, the tubular furnace was preheated to 700°C and the reaction flask was submerged in the furnace for 15 min. The flask was then quenched in air to about 200°C while at the same time the furnace was cooled to 70°C. Each flask was then reinserted into the furnace and allowed to bake at 70°C for 2-6 days. Table 1 lists the 70°C baking times for the seven FTT runs.

After heating, the reaction products were treated in three ways. The reaction flask was either extracted with water, extracted with acid, or extracted sequentially with both water and acid (see Table 1). In acid extraction, 50 ml of 6N HCl was refluxed in the reaction flask overnight under nitrogen atmosphere. Water extraction was carried out by introducing 50 ml of water into the flask and refluxing overnight. The supernatant was then separated from the solids and, in all but one run, the water extract was then hydrolyzed with acid. Hydrolysis of the water extract involved evaporating the water soluble fraction under reduced pressure and refluxing with 6N HCl overnight under nitrogen.

After acid extraction it was necessary to remove inorganic salts that formed from attack of the acid on the catalysts. Desalting was accomplished with cation exchange on Bio-Rad Dowex AG50W-X8, 50-100 mesh resin. The FTT samples were applied to a 65 ml column at a pH of 3.5; the column was washed with water until a test for chloride was negative, and then amino acids were eluted with 2N NH₄OH. Ammonia was removed by evaporating the sample under reduced pressure.

After removal of inorganic salts the amino acid samples were derivatized and analyzed by gas chromatography. Derivatization involved two steps (12). First the butyl ester was formed by reaction of the amino acids with anhydrous 6N HCl-n-butanol. The acid-butanol was refluxed with the amino acids for 1 hr. Excess acid-butanol was removed by evaporation under reduced pressure. In the second step the amino group was acylated using triflouroacetic anhydride (TFAA) and CH_2Cl_2 as solvent with a 10 min reflux.

In the derivatized form tentative identification of amino acids was made by gas chromatography (GC), on a 5% OV-17 glass packed column mounted in a Hewlett-Packard 5750 research chromatograph. Positive identification was obtained using a Perkin Elmer 990 gas chromatograph, equipped with a UCON 75H90000 capillary column, combined with a Dupont 21-491 mass spectrometer (GC-MS) by means of a membrane separator. A Decision, Inc., data system was used to acquire, store, and process the data.

Contamination was checked by simulating several FTT reactions without heat. This involved oxidation-reduction of the catalysts, introduction of the reaction gases as before, but without a heating mode, extraction of the reaction flask, and analysis of the extract for amino acids. A chromatogram of the water extract of a FTT blank run is shown in figure 1A.

The sensitivity of the analytical technique used in the FTT project was challenged by derivatizing 10 nanomoles of alanine, adjusting the total volume of solvent (CH_2Cl_2) to 50 microliters to yield an alanine concentration of 0.2 nanomoles/microliter, and injecting 1.0 microliter onto the gas chromatography column. One small peak was observed at the retention time corresponding to alanine.

Results & Discussion

In run 1 the reaction flask was extracted with water only. It was assumed that the reaction products had condensed on the surface of the flask and catalyst, and that a simple water extract would remove them. The water extract was not hydrolized with acid. The extract was simply evaporated, derivatized, and analyzed by gas chromatography. No amino acids were observed.

In runs 2, 3 and 4 the reaction flasks were extracted with water. Each water extract was then hydrolyzed with acid to convert suspected precursors into corresponding amino acids and improve the yields of these compounds. Gas chromatograms showed small peaks, corresponding to amino acid retention values, but yields were so small that positive identification with mass spectrometry could not be made.

Run 5 was aborted due to contamination.

In run 6 the reaction flask was extracted sequentially with both water and acid. The water extract was then acid hydrolyzed, derivatized, and analyzed with gas chromatography. Figure 1B shows the chromatogram of the water soluble fraction of run 6. Amino acid peaks appear to be present but yields are too small for positive identification on mass spectrometry. The subsequent acid extraction of run 6 was performed in order to more thoroughly leach the reaction flask and catalyst (7).

In run 7 only an acid extraction was performed, which was then desalted, derivatized, and analyzed by gas chromatography. Yields were an order of magnitude greater so that positive identification using gas chromatography combined with mass spectrometry could be made. Figure 2 shows the total ion plot as a function of scan number for the acid extracts of runs 6 and 7. Positive identifications were made by matching mass spectra of unknowns and standards. Figure 3B shows the mass spectrum associated with scan number 285 of an unknown G.C. peak in run 7. This has been identified as N-trifluoroacetyl glycine-n-butyl ester by comparison with the mass spectrum of a known (figure 3A). In a similar manner several other peaks were identified. The total ion plots for the acid extracts of runs 6 and 7 are quite similar, however corresponding peaks don't occur at identical scan numbers because chromatographic conditions differed. In figure 2, peak 1 is trifluoracetamide in both plots; peak 2 is alanine; peak 3 is leucine; and peak 4 is glycine in both plots. The large unidentified peak may be urea, but positive identification was not made using a standard. The presence of leucine, and none of the 4 or 5 carbon amino acids, suggests that run 7 might have some amino acid contamination.

A FTT reaction prepared by Hayatsu was also investigated by E. Peterson at the Chemical Evolution Branch, Ames Research Center. The deuterated reaction gases CO, D₂, and ND₃ were used to distinguish synthesized (deuterated) amino acids from contaminants. Only small amounts of deuterated alanine and glycine could be identified by GC-MS.

The presence of only small amounts of a few protein amino acids and no non-protein amino acids in our FTT experiments and Hayatsu's FTT reaction is at odds with the literature (11). The reasons for this discrepancy are at present not understood, but because this type of experiment is important for our understanding of the nature of organic compounds present in meteorites, future laboratory simulations should be designed to maximize the yields of amino acids.

Acknowledgments. We thank Ms. J. Mazzurco and Mr. F. Church for their assistance.

References

- 1. A. I. Oparin, "Proischogdenie Zhiani." (Moscow Robotchii, Moscow, 1924).
- 2. J.B.S. Haldane, "Rationalist Ann." 148, (1928).
- 3. R.H. Lemmon, "Chemical Reviews" 70, 95 (1970).
- K.A. Kvenvolden, J.G. Lawless, K. Pering, E. Peterson, J. Flores, C. Ponnamperuma, C. Moore, "Science" 228, 923 (1970).
- 5. J. Lawless, K. Kvenvolden, E. Peterson, C. Ponnamperuma, C. Moore, "Science" 173, 626 (1971).
- 6. C.E. Folsome, J. Lawless, M. Romiez, C. Ponnamperuma, "Nature" 232, 108 (1971).
- 7. J. Oro, J. Gilbert, H. Lichtenstein, S. Wilkstrom, D.A. Flory, "Nature" 230, 105 (1971).
- D. Ring, Y. Wolman, N. Friedmann and S.L. Miller, "Proc. Nat. Acad. Sci." USA, 69, 765 (1972).
- 9. Y. Wolman, W.J. Haverland, and S.L. Miller, "Proc. Nat. Acad. Sci." USA, 69, 809 (1972).
- 10. E. Anders, R. Hayatsu, M. H. Studier, "Science" 182, 781-790 (1973).
- 11. D. Yoshino, R. Hayatsu, E. Anders, "Geochimica et Cosmochimica Acta" <u>35</u>, 927 (1971).
- 12. R.W. Zumwalt, D. Roach, and C.W. Gehrke, "Chromatog." 53, 171 (1970).

4

			Baking time	Extraction Conditions		Water Extract Hydrolysis	-
	Cat CD(mg)	alyst Al(mg)	(in hr.) at 70°C	water t(hr)	6NHC1 t(hr)	(6 N HC1) Conditions	
R1	185.05	266.76	42	17			
R2	225,53	315,18	117	27		16	
R3	229.56	253.22	144	18		16	
R4	201.53	340.46	65	16		16	
R5	·			 ,			
R6	200.85	305.26	64	48	16	264	
R7	250.0	339.7	48	~-	16	·	

TABLE 1.- EXPERIMENTAL CONDITIONS

-.--

·

.

.

· ·

. .

.

.





(a) Derivatized HC1 extract of FTT run 6.

(b) Derivatized HC1 extract of FTT run 7.





(a) Glycine standard as the n-butyl, N-trifluoro-acetyl derivative.(b) Derivatized unknown from FTT run 7 (scan number 285).

Figure 3.- Mass spectrum.

.