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UTILIZATION OF NON-CONVENTIONAL SYSTEMS FOR CONVERSION OF
BIOMASS TO FOOD COMPONENTS

RECOVERY OPTIMIZATION AND CHARACTERIZATION
OF
ALGAL PROTEINS AND LIPIDS

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ABSTRACT

Protein isolate obtained from green algae (Scenedesmus obliquus) cultivated under controlled conditions was characterized. Molecular weight determination of fractionated algal proteins using SDS-polyacrylamide gel electrophoresis revealed a wide spectrum of molecular weights ranging from 15,000 to 220,000. Isoelectric points of dissociated proteins were in the range of 3.95 to 6.20. Amino acid composition of protein isolate compared favorably with FAO standards. High content of essential amino acids leucine, valine, phenylalanine and lysine makes algal protein isolate a high quality component of CELSS diets. To optimize the removal of algal lipids and pigments supercritical carbon dioxide extraction (with and without ethanol as a co-solvent) was used. Addition of ethanol to supercritical CO₂ resulted in more efficient removal of algal lipids and produced protein isolate with a good yield and protein recovery. The protein isolate extracted by the above mixture had an improved water solubility.

INTRODUCTION

There is a need for suitable feeding systems for extended manned space missions. The potential utilization of algal biomass in a non-conventional food supply in space, is the subject of our study. Unique multifunctional characteristics of algae such as high photosynthetic activity, lack of need for organic carbon and nitrogen in the growth media and the ability to control algal chemical composition by varying cultivation conditions make algae an attractive system for biological regeneration of the CELSS environment (Casey and Lubitz, 1963; Clement et al., 1967; Lachance, 1968; Lipinsky and Litchfield, 1970; Waslien, 1975; Clement, 1975; Waslien et al., 1978; Oswald, 1980; and Piorreck et al., 1984). Consumption of

untreated and unpurified algal biomass, due to associated physiological problems (e.g. nondigestible components of cell wall and disturbing carbohydrates, excess nucleic acids, unknown toxins and allergens) and poor flavor or color limits the amount of algae in the diet. Conversion of algal biomass into physiologically and organoleptically acceptable components requires suitable methodology. Previously developed methods eliminated or largely reduced three known undesirable components of algae (cell walls, nucleic acids, and pigments) and gave an "algal protein isolate" with a reasonably high yield (Kamarei et al. 1985).

Green algae (Scenedesmus obliquus) grown in Constant Cell Density Apparatus, under controlled conditions were used in this study. They were obtained through the courtesy of Drs. Richard Radmer and Paul Behrens of MARTEK Corp. Columbia, MD. Figure 1 shows the stepwise procedure for preparation of various fractions from algae. Major nutritional components of the algae flour (freeze-dried ruptured cells) were determined. Upon cell rupture and removal of the cell wall, the nucleic acid content of the algae was reduced by 92%, using an enzymatic procedure. This increased the safe consumption level of algal protein concentrate (fraction #6) from 20g to approximately 250g per day and for algal protein isolate (fraction #8A) from 15g to 192g per day. Generally accepted safe level of nucleic acids intake in human is 2g per day (Scrimshaw, 1975). Ethanol extraction of algal pigments (mostly chlorophyll A) from the protein concentrate (fraction #6) resulted in removal of most of the green color and consequently the algal protein isolate (fraction #8A) had a "light olive" color upon freeze-drying. Removal of the pigments enhanced the color and flavor, but caused protein denaturation and insolubility of the final product.

In order to further improve the quality of algal proteins we have experimented with the removal of algal pigments and lipids using supercritical fluids. The results are reported here.

Characterization of isolated algal proteins was a major part of our recent study. This knowledge can be useful in designing further processing and fabrication of algal proteins as a food component in CELSS. The present paper also reports the results of electrophoretic studies and amino acid analyses of algal proteins.

METHODOLOGY

Preparation of algal protein fractions:

Green algae (Scenedesmus obliquus) were grown in Constant Cell Density Apparatus (CCDA). Conditions were: Algal density in medium: 0.55 mg dry weight/ml.; 32°C; pH=7.0; nitrogen source: KNO₃ 2.0g/l; carbon source: 2% CO₂ in air. The algae were grown, harvested, and supplied to us by Martek Corp. (Colombia, MD).

SDS-Polyacrylamide gel electrophoresis (SDS-PAGE):

SDS-PAGE was used to determine the molecular weights (M.W.) of algal proteins. Freeze-dried algal protein concentrate (fraction #6) was used. The experimental details of SDS-PAGE were reported previously (Nakhost and Karel, 1983).

Isoelectric focusing (IEF):

IEF in polyacrylamide gel was used to determine the isoelectric points (pI) of algal proteins. Ampholine pH 3-10 was used to produce pH gradient across the gel. The IEF conditions were as previously reported (Nakhost and Karel, 1983). Freeze-dried algal protein isolate (fraction #6) was used in this study.

Amino acid analysis:

Amino acid composition of algal protein isolate was determined using a recently developed pre-column derivatization method which has been originally demonstrated by Koop et al. (1982) for analysis of free amino acids and later modified by Bidlingmeyer et al. (1984) for application to acid-hydrolysates of proteins.

The method which is called "Pico-Tag" is based on derivatization of amino acids with phenylisothiocyanate (PITC). PITC reacts with free amino acids to yield phenylthiocarbamyl (PTC) amino acid, which can then be separated on a reversed-phase HPLC column. Strong UV absorbance of PTC-amino acids (254 nm) makes it possible to detect quantities as small as 1 picomole (Bidlingmeyer et al. 1984). Because PITC forms the same chromophore with primary and secondary amino acids no extra treatment is necessary for detection of proline. Excellent reproducibility, derivatives stability (6-10 hrs. at R.T.) and rapid analysis (12 min) of amino acids made Pico-Tag the method of choice for our purpose.

Upon HCl hydrolysis destruction of Trp, conversion of Asn and Gln to ASP and Glu., also oxidation of cysteine to cystine and consequently partial destruction of cystine occur (Haschemeyer and Haschemeyer, 1973). Hence, in order to obtain complete amino acid profile, algal protein isolate (fraction #8A) was treated in 3 different ways: 1) HCl hydrolysis and PITC derivatization to detect all PTC-amino acid derivatives except for Cys. and Trp.

2) Performic acid oxidation followed by HCl hydrolysis and derivatization to quantify the amount of cysteine in the form of cysteic acid, and 3) methanesulfonic acid hydrolysis followed by derivatization to obtain the amount of tryptophan. Amino acid standard used was Pierce standard H. Least square treatment of data and calculated regression lines were used for quantification of amino acid content of algal proteins.

Supercritical Fluid Extraction:

Supercritical fluid extraction of algal pigments and lipids was carried out in two steps. The 1st extraction was with supercritical carbon dioxide (SC -CO₂) and the 2nd extraction was with carbon dioxide and anhydrous ethanol (22%) as a co-solvent. Freeze-dried algal protein concentrate (fraction #6), 4.86g was charged alternately between layers of glass wool to an extraction vessel, a 1.8 cm diameter X 30 cm long stainless steel tube (Autoclave Engineers, Inc.), and connected to the system shown in Figure 2; the glass wool served to keep the algae powder from compacting during passage of gas through the extractor. Carbon dioxide (Airco, Inc., Grade 2.8) was supplied at about 87 bar pressure and 313°K to the suction side of a double-end diaphragm compressor (Superpressure, Inc.) and was compressed to the measurement pressure. The pressure was controlled by a back-pressure regulator (Circle Seal, BPR) which diverted the bulk of the compressed gas from the surge tank back to the suction side of the compressor resulting in an almost pulse-free flow of gas to the extractor. The high-pressure gas passing downstream of the compressor was heated in a tube preheater to about 338°K and was passed through the extraction vessel which was maintained at 338°± 2 K by a temperature indicator/controller which measured the temperature via an iron-constantan thermocouple (Superpressure, Inc.) positioned in the bed of algae powder and which regulated power to a heater (Glascol tapes) on the extractor.

The solution (consisting of carbon dioxide and dissolved materials) leaving the extraction vessel was passed through a heated, flow-regulating, pressure let-down valve and was expanded to ambient pressure. The materials which were dissolved by the gas passing through the extraction vessel precipitated during the pressure-reduction step and were separated

from the gas in a U-tube collector (Kimble, 200 mm) whose exit junction was fitted with a glass wool filter to prevent fine-particle solids from passing through the tube; a second U-tube with a more tightly packed glass wool filter was positioned downstream of the first collector and served to trap any fine particles which might have passed through the first filter.

The ambient gas leaving the collection system passed through a rotameter, (Fischer-Porter, Inc., Series 10A 35) for flow rate measurement and through a dry test meter (Singer, Inc., DTM-200) for flow volume integration. The flow rate of the carbon dioxide through the extractor was maintained at 3 SLPM (standard liters per minute). A total of 0.66 g of green semi solid materials, SC-CO₂ extract (fraction #7B) was collected (in the U-tube) during the passage of 720 g of carbon dioxide at 380 bar pressure. After the extraction with carbon dioxide a fraction of the charge of algae (SC-CO₂ residue, fraction #8B) was removed from the vessel to be used for solubility tests, the vessel was resealed and connected to the system. The 2nd extraction was carried out with SC-CO₂ at 380 bar pressure and 338° K, with anhydrous ethanol added as a co-solvent. It was pumped into the system at the entrance to the extractor. The ratio of ethanol to carbon dioxide was 100 to 360 g (22%). The SC-CO₂ + EtOH extract (dark green ethanol solution) was collected in a container (fraction #7C). The SC-CO₂ + EtOH residue (fraction #8C) was used for solubility test (Fig. 3).

Solubility Test:

Algal protein isolate (fraction #8A), SC-CO₂ residue (fraction 8B) and SC-CO₂ + EtOH residue (fraction #8C) were used. To 100 mg of each fraction, 10 ml, 0.05 M phosphate buffer, pH 8.0 (based on pH-solubility profile of plant proteins, Wolf, 1978) was added. The samples were mixed and stored at room temperature for about 1 hr. They were then centrifuged at

15,000 RPM for 10 min. at 2° C. The supernatant was collected and the pellet was washed with another 2 ml of phosphate buffer and recentrifuged. The wash was added to supernatant (kept at 5° C) and the pellet was freeze-dried (Virtis Research Equipment, Gardiner, NY). Protein concentration of the supernatants and the pellets were determined using the microkjeldahl method (A.O.A.C., 1980). Solubility was defined as:

$$\% \text{ Solubility} = \frac{\text{Protein in supernatant (mg)} \times 100}{\text{(Sum of protein in supernatant and pellet) mg}}$$

Absorption Spectra:

Absorption spectra of SC-CO₂ extract (fraction #7B) and SC-CO₂ + EtOH extract (fraction #7C) in the range of 700 to 240 nm were obtained and compared with that of fraction 7A, which is the boiling-ethanol extract. Since SC-CO₂ extract (fraction #7B) is in the form of semi solid materials, an aliquot was dissolved in hexane, filtered (Millex-SLSRO25NS) and used for this study.

Analysis of Algal Lipids:

Isolation of algal lipid components (neutral, glyco and phospholipids) using sequential solvent extraction, gel filtration and thin layer chromatography, as well as determination of fatty acid composition using GC is underway. SC-CO₂ and SC-CO₂ + EtOH extracts (fractions #7B and 7C) and the corresponding residues (fractions #8B and 8C) will be analyzed.

RESULTS AND DISCUSSION

Molecular Weight Determination:

Literature on molecular weights of algal proteins is limited. Lee and Picard (1982) studied the electrophoretic pattern of *Oocystis* algal pro-

teins using three systems: Tris-glycine pH 8.7, SDS-gel and SDS-gel with 2-mercaptoethanol. Using the SDS-gel along with dissociating agents they observed a wide spectrum of molecular weights (M.W.) ranging from 12,500 to 250,000. In our study using SDS-PAGE we also obtained wide range of M.W. for Scenedesmus obliquus proteins. Figure 4 shows the electrophoretic pattern of protein concentrate (fraction #6). The M.W. of proteins ranged from 15,000 (band #20) to 220,000 (band #1). Major protein bands corresponded to bands # 7, 14, 15, 18, 19, and 20 with M.W. of 55,000, 30,000, 28,000, 18,000, 16,500 and 15,000 respectively. Band #21 corresponded to chlorophyll and other algal pigments (M.W. below 1,000) which migrated to the front of the gel. A comparison of SDS-PAGE pattern of protein isolate obtained from ethanol extraction (fraction #8A) with that of protein isolate resulting from supercritical fluid extraction (fractions #8B and 8C) will enable us to detect the extent of changes such as oligomerization and/or partial fragmentation of treated algal proteins.

Isoelectric Point Determination:

There is no report available on isoelectric points of algal proteins. The IEF pattern of algal proteins and pH profile of IEF gel for determination of the PI's of focused proteins are shown in figure 5. The isoelectric points of the algal proteins ranged from 3.95 (band #15) to 6.20 (band #2). Major protein bands corresponded to bands #7, 9, 11 and 13 with PI's of 5.45, 5.20, 4.85 and 4.35 respectively. Band #1 corresponded to chlorophyll and other pigments which did not migrate in the IEF gel.

Amino Acid Composition:

Nutritional value of proteins is primarily determined by their amino acid composition. Hence amino acid profile of algal protein (Scenedesmus obliquus) was studied and aminograms were obtained (figure 6).

The amino acid content of algal proteins compared favorably with FAO standards, Table 1. Analysis of amino acid composition revealed high content of leucine, valine, phenylalanine and lysine and low content of methionine and tryptophan. (The high values for cysteine contrary to reported low values in the literature is because we analysed cysteine in the form of cysteic acid which yielded in about 70% more compared to values we obtained for cystine).

There is no data reported on amino acid composition of Scenedesmus obliquus cultivated under controlled conditions (CCDA). However, there are reports on amino acid composition of Scenedesmus obliquus cultivated under open air conditions according to German technique of Dortmund (Bock and Wunsche, 1967, Soeder et al., 1970, Walz et al., 1975, and Kugler and Pithakpol, 1975); cultivation ponds (Becker, 1981) and unspecified conditions (Waslien, 1975). The comparison of the aminograms from Scenedesmus obliquus cultivated under different conditions revealed a similar pattern of high content of valine, leucine and lysine and low content of methionine and tryptophan. Our results, showed relatively higher values for isoleucine, leucine and phenylalanine.

The nutritional value of green algae depends on digestibility which depends on pretreatment of algae and disruption of the cell wall. Low nutritional value is to be expected if algae is not sufficiently processed (Payer et al., 1980). Nutritional value (biological value, BV, digestibility, D, net protein utilization, NPU, and protein efficiency ratio, PER) of pretreated Scenedesmus obliquus cultivated under open air conditions (Dortmund technique) has been studied in a series of different and independent investigations in animals and also in human volunteers. The results repeatedly and unequivocally confirmed high nutritive value of algal pro-

teins (Pabst, 1978, Kofranyi and Jekat, 1967, Muller-Wesker and Kofranyi, 1973, and Payer et al., 1980). Drum-dried Scenedesmus obliquus was studied as a diet component in rats. High nutritional value due to improved protein digestibility (as a result of cell disruption) was reported, Table 2 (Pabst, 1974, and Richmond, 1983). Fortification of wheat and rice with proteins from Scenedesmus acutus in rats' diets resulted in significantly higher PER value than cereal diet alone (Venkatamaran et al., 1977). They also concluded that Scenedesmus proteins supplemented wheat proteins to a greater extent than rice.

In summary, algal proteins, because of their high content of essential amino acids (valine, leucine, lysine and phenylalanine) upon cell wall rupture and removal of undesirable components, can significantly improve the nutritional value of other plant proteins e.g. cereals (wheat, rice and corn with lysine as their limiting amino acid), legumes, etc. and serve as a high quality component of CELSS diets. Wheat and soybeans are considered as two sources of macronutrients in CELSS diet scenario (Karel and Kamarei, 1984).

Supercritical Fluid Extraction:

In our previous study boiling ethanol was used to remove pigments and lipids. Ethanol treatment of algal proteins (regardless of extraction temperature) improves the color and flavor of the isolate, but causes denaturation and consequently aggregation of algal proteins with loss of solubility. Supercritical fluids are receiving increasing attention as extraction solvents. Because of their pressure-dependent dissolving power properties often display the fractionation ability of multicomponent solutes (Krukonis, 1984). Supercritical carbon dioxide (SC-CO₂) has been considered by many investigators as the ideal fluid for extraction and

separation processes. It is reported to behave very much like a hydrocarbon solvent with very low polarizability (Hyatt, 1984). Therefore, the use of SC-CO₂ for extraction of lipids or lipid soluble materials while giving comparable yields offers several advantages.

Organic solvents are flammable and explosive. They might also contain traces of higher boiling fractions that may be left in the extract and pose a potential health hazard (Christianson et al., 1982). On the other hand carbon dioxide is nontoxic, nonflammable and rather easily separatable from the extracted materials.

There are an increasing number of reports on replacement of lipophilic extractions by SC-CO₂. Extractions of oil from corn germ (Christianson et al., 1982 and Christianson et al., 1984); soybeans (Quirin, 1982, List and Friedrich, 1985 and Eldridge et al., 1986); cottonseed (List et al., 1984); lupine seeds (Stahl et al., 1981) and animal oils and fats (Stahl, 1982) using supercritical CO₂ has been reported.

Comparative study on the storage stability of defatted corn germ flour using hexane and SC-CO₂ showed significant reduction of peroxidase activity (apparently as a result of protein denaturation under the extraction conditions, 344 to 551 bar pressure and 323°K) and high flavor quality upon accelerated storage tests for SC-CO₂ extracted corn germ flour (Christianson et al., 1984). Similar study on oil extraction of soybean flakes using SC-CO₂ (730 to 854 bar pressure, 353-373°K and 5-13% moisture) and hexane also resulted in better flavor quality and shelf life for SC-CO₂ defatted soybean flour (Eldrige et al., 1986). In a comparative study on oxidation stability of "oil" extracted from soybean flakes using either SC-CO₂ at 344 bar and 323°K or hexane, SC-CO₂ extraction yielded a product comparable to a hexane-extracted degummed soybean oil. Phosphatides were

essentially absent in SC-CO₂ and this may be the reason for lower oxidative stability of SC-CO₂ extracted soybean oil (List and Friedrich, 1985).

There is no report on supercritical extraction of algal lipids and/or pigments. The extraction of algal protein concentrate (fraction #6) using (SC-CO₂) resulted in removal of green semi solid materials believed to contain nonpolar lipids and lipid-soluble pigments (such as chlorophyll and carotenoids). Chlorophylls are classified as lipid- and water-soluble and carotenoids as lipid-soluble pigments (Clydesdale and Francis, 1976). The yield of SC-CO₂ extract (fraction #7B) was 7%. The absorption spectra of fraction #7B (dissolved in hexane) indicated the predominance of chlorophyll A (with absorption maximum, λ_{\max} at 410 nm) and some other minor peaks probably due to extracted carotenoids. The yield of SC-CO₂ residue (fraction #8B) which had an "olive-green" color was 44.6%. Further extraction of protein isolate (fraction #8B) using SC-CO₂ and 22% ethanol resulted in a dark green ethanol solution believed to contain polar lipids and chlorophylls (fraction #7C) with the yield of 12%. The absorption spectra of SC-CO₂ + EtOH extract indicated mainly the presence of chlorophyll A and traces of chlorophyll B (λ_{\max} at 470 nm). The absorption spectra of SC-CO₂ + EtOH extract (fraction #7C) were almost identical to those of ethanol extract (fraction #7A). The SC-CO₂ + EtOH residue (fraction #8C) had a "light olive" color similar to ethanol extract residue (fraction #8A). Yield of algal protein isolate obtained from SC-CO₂ + EtOH extraction was 33%. The remaining balance as compared to previously reported value for fraction 8A which was 36.6%, is due to the removal of 19% total lipids (nonpolar and polar) and pigments (fraction #7B + 7C) compared to 15% (fraction #7A). We believe that SC-CO₂ extraction resulted in more efficient removal of non-polar lipids. Isolation and determination of lipid components (neutral, glyco and phospholipids) and fatty acid composition of various extracts

(ethanol, SC-CO₂ and SC-CO₂ + EtOH) and the residues is underway and will be reported in our next paper. Protein concentration of SC-CO₂ + EtOH residue (protein isolate 8C) was 69.1%, similar to the ethanol residue (protein isolate 8A) which was 70.5%. Table 3 shows the yield, protein concentration and recovery of various algal fractions.

Protein Recovery:

Recovery of Oocystis algal proteins was studied by Lee and Picard (1982). Using successive ammonium sulfate fractionations they recovered 44% of algal proteins. The protein recovery increased to 66% when the fractionation was done in the presence of ascorbic acid and insoluble PVP (polyvinylpyrrolidone). The presence of endogenous proteolytic and oxidative enzymes may partially impair the recovery of plant proteins. Phenoloxidase and peroxidase catalyze the reduction of O-diphenols to quinones; subsequently the quinones polymerize and form complexes with proteins thus impairing the solubility (Loomis and Battaile, 1966; Betschart and Kinsella, 1973; Lee and Picard, 1982; Cremer and Van De Walle, 1984). The formation of these complexes is prevented by extracting the plant proteins in the presence of reducing agents (such as ascorbic acid) and by removing the phenols with substances such as PVP which forms hydrogen bonds with phenols. Nonspecific aggregation of plant proteins is also reported in the literature and believed to be the result of sulphhydryl interactions (Stahman, 1963).

Solubility of Algal Proteins:

Solubility of algal proteins is shown in Table 3. Under our experimental conditions, the solubility of algal protein (fraction #6) prior to removal of pigments and lipids was 63%. Upon extraction with ethanol the solubi-

lity decreased to 20% (protein isolate 8A). The solubility of SC-CO₂ residue (protein isolate 8B) was 45% and of SC-CO₂ + EtOH residue was 41%.

Partial protein denaturation as a result of "anhydrous" SC-CO₂ extraction (344 to 551 bar and 323°K) of dry-milled corn germ flour (Christianson et al., 1984) and "humid" SC-CO₂ (730 to 854 bar, at 353 to 373°K and 5-13% moisture) extraction of soybean flakes (Eldridge et al., 1986) has been reported. Christianson et al. (1984) considered denaturation to be the result of high pressure extraction. However, this partial denaturation resulted in reduction of peroxidase activity (heat-resistant oxidative enzyme) and consequently better storage stability of the extracted corn germ flour and soybean flakes. Weder (1980 and 1984) studied the effect of humid supercritical CO₂ and N₂ at 300 bar, 353°K and room temperature on the structure and amino acid composition of ribonuclease and lysozyme. The amino acid composition and TNBS-reactive lysine (trinitrobenzene sulfonic acid) remained unaltered. Partial oligomerization (due to disulfides) and some fragmentation of protein molecules in protein extracted at 353°K was detected. No oligomers were detected using SDS-PAGE when extraction was at room temperature. The occurrence of the changes in the protein molecules was concluded to be the result of heating proteins in the presence of water regardless of the nature or pressure of the gas used. Tryptic digestion of the treated proteins indicated a better digestibility than untreated proteins.

In our system, decrease in the solubility of protein isolate (41%) as compared to protein concentrate (63%) could possibly be due to the extraction temperature (338°K). In our previous study on enzymatic reactions of polyphenol oxidase in supercritical fluids the enzyme was found to be cata-

lytically active at 344 bar, 307°K and 309°K (Hammond et al., 1985) indicating that no protein denaturation has occurred due to the exposure to supercritical fluids.

In summary: SC-CO₂ + EtOH extraction of green algae resulted in; 1) more efficient removal of algal lipids and pigments (19% vs 15% ethanol extract). 2) Fractionation of nonpolar and polar lipids. 3) Protein isolate with a good yield and protein recovery. 4) Increased solubility of the protein.

The above results suggest that supercritical fluid extraction is a useful extraction method to be used in the space habitats.

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Table 1) Amino acid composition of algal protein
isolate (fraction #8A)
(g/100g protein)

Amino acid	FAO Standard	<u>Scenedesmus obliquus</u>
ASP		8.0
Glu		8.3
Ser		3.6
Gly		6.1
His		1.8
Arg		5.8
Thr*	4.0	4.6
Ala		8.3
Pro		5.8
Tyr		3.5
Val*	5.0	7.3
Met*	2.7	2.2
Cys		7.0
Ile*	4.0	5.0
Leu*	7.0	11.1
Phe*	3.4	6.0
Trp*	1.0	
Lys*	5.5	^a 5.7
<u>%Essential</u>	<u>32.6</u>	<u>42.0</u>

*Essential amino acid
a-not reported here

Table 2) Nutritional Value of Algae in Comparison with Some Representative Food Proteins (Liener, 1978)

	PER	BV	NPU	C.S.	limiting a.a.
Egg	3.8	87-97	91-94	100	none
Soybeans	0.7-1.8	58-69	48-61	69	S
Rice	1.9	75	70	57	Lys.
Corn	1.2	60	49-55	55	Lys.
Wheat	1.0	52	52	57	Lys.
Algae(a)	3.21	81	68	57(b)	Met. Trp.

(a) - Drum-dried Scenedesmus obliquus, Richmond, 1983

(b) - Waslien, 1975

PER - Protein Efficiency Ratio

BV - Biological Value

NPU - Net Protein Utilization

C.S.- Chemical Score

S - Sulfur-containing amino acid

Table 3) Yield (%), protein concentration (%) Protein recovery (%) Solubility (%) of various algae fractions without enzymatic treatment.

fraction #	yield (%)	Protein concentration (%)	Protein Recovery (%)	Solubility (%)
2 (algae flour)	100	52.6	100	---
4 (algal crude protein)	65	57.6	71.2	---
6 (algal protein concentrate)	51.6	53.4	52.4	63
8A (algal Protein Isolate)	36.6	70.5	49.1	20
8B (SC-CO ₂ Protein Isolate)	44.6	66.5	56.4	45
8C (SC-CO ₂ + EtOH Protein Isolate)	33	69.1	43.3	41

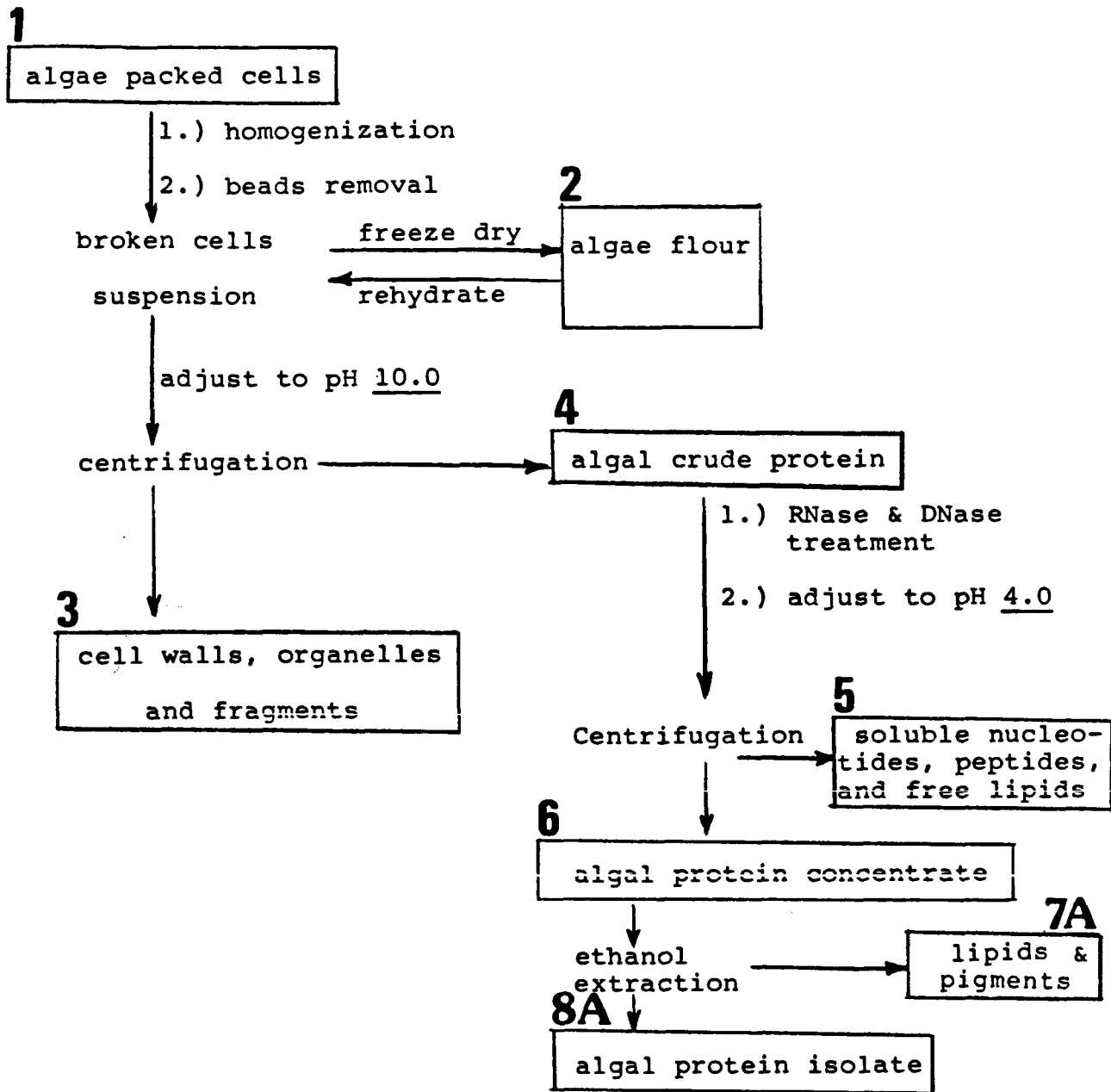


Fig. 1. Procedure for preparation of different fractions from algae.

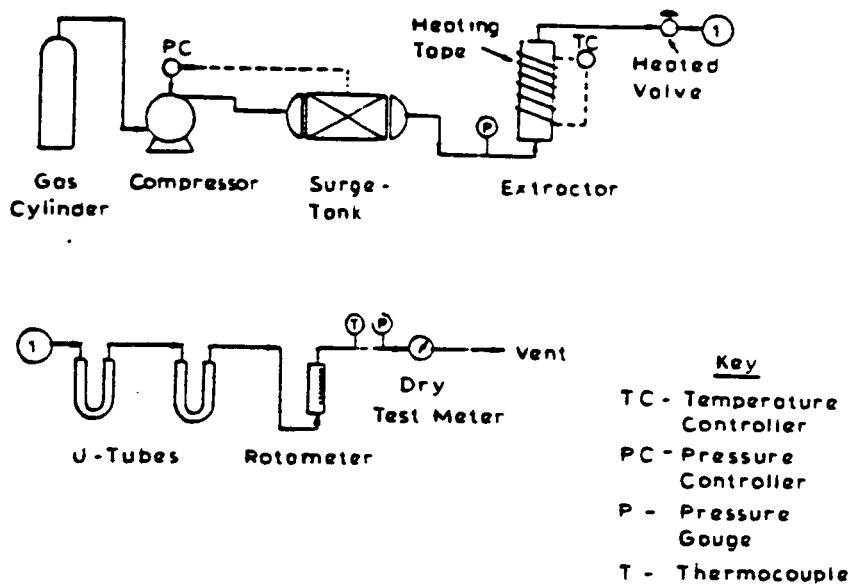


Fig. 2. Schematic diagram of experimental for supercritical fluid extraction (Krukonis and Kurnik, 1985).

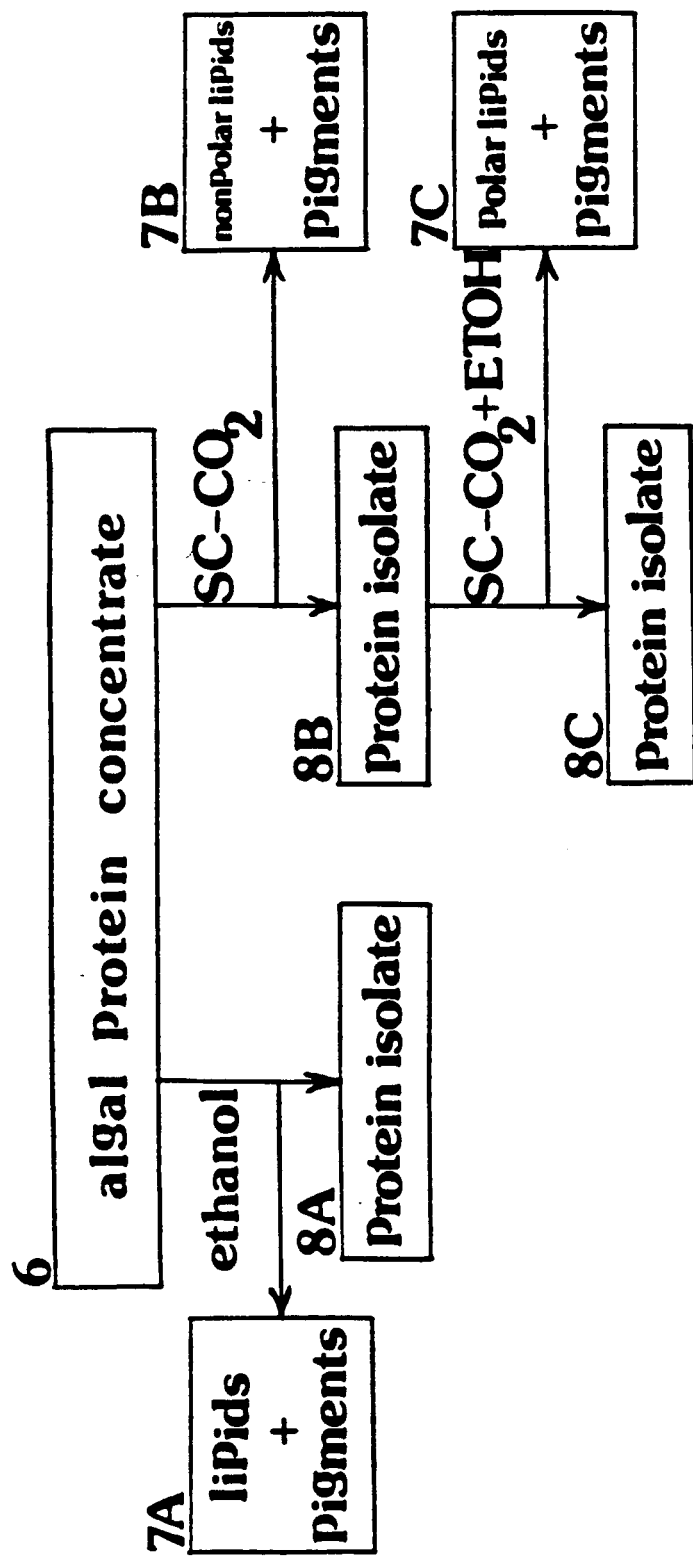


Fig. 3. Procedure for extraction of algal lipids and pigments using supercritical fluids (at about 380 bar pressure and 338°K).

SDS/PAGE

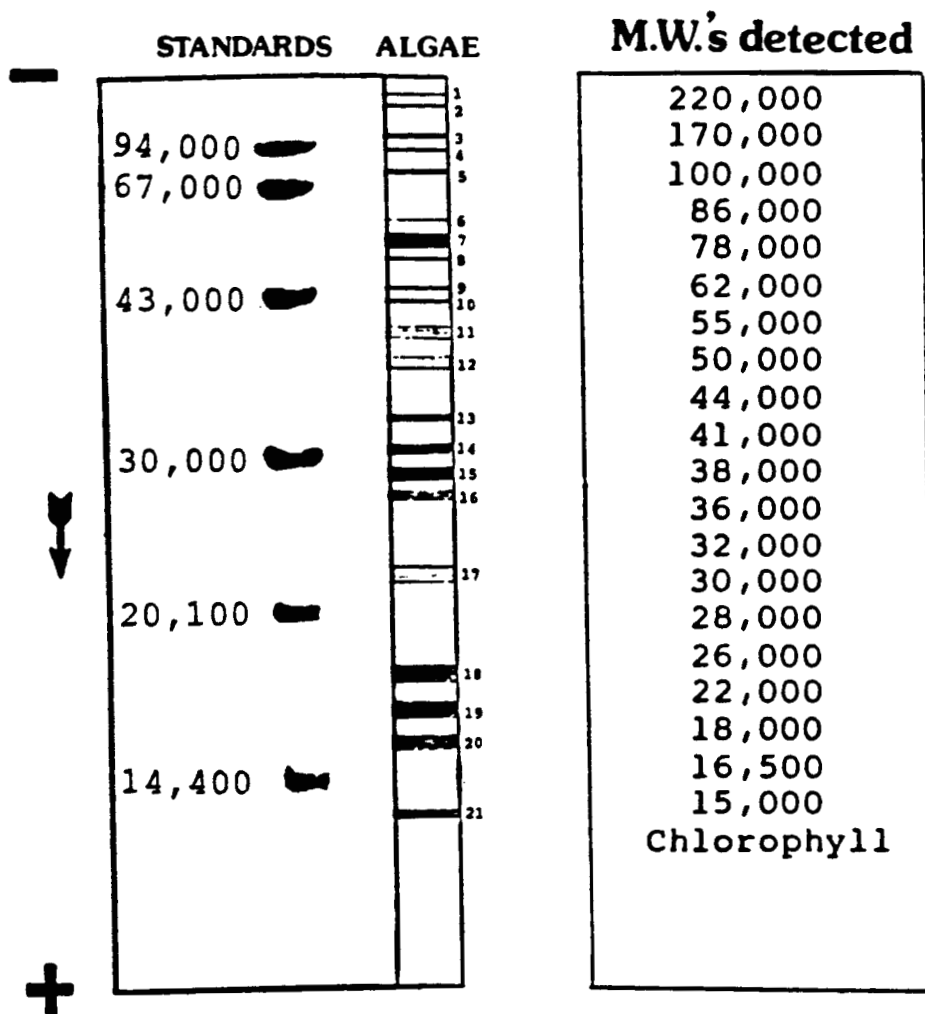


Fig. 4. SDS-PAGE pattern of algal protein concentrate (fraction #6).

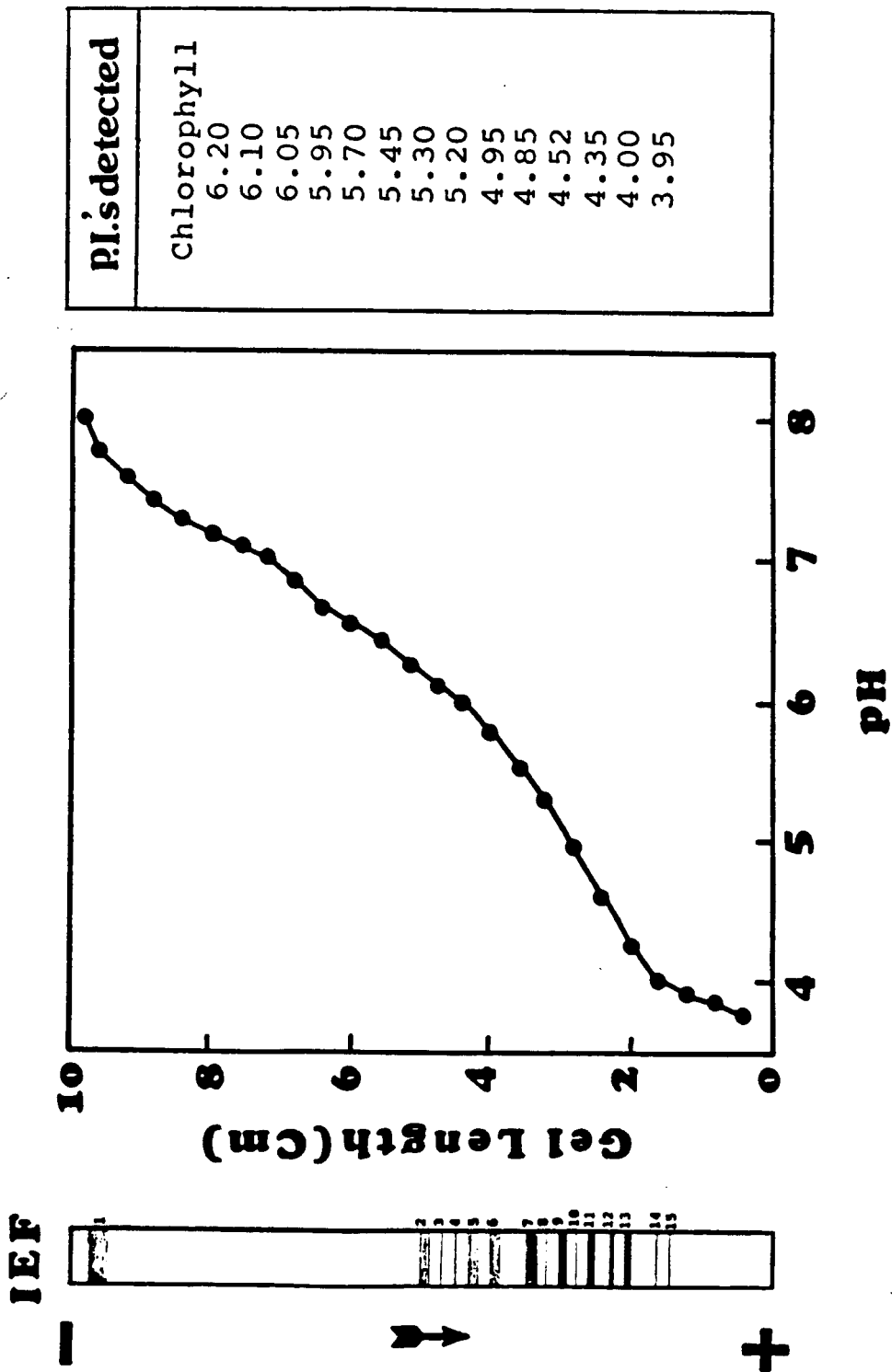
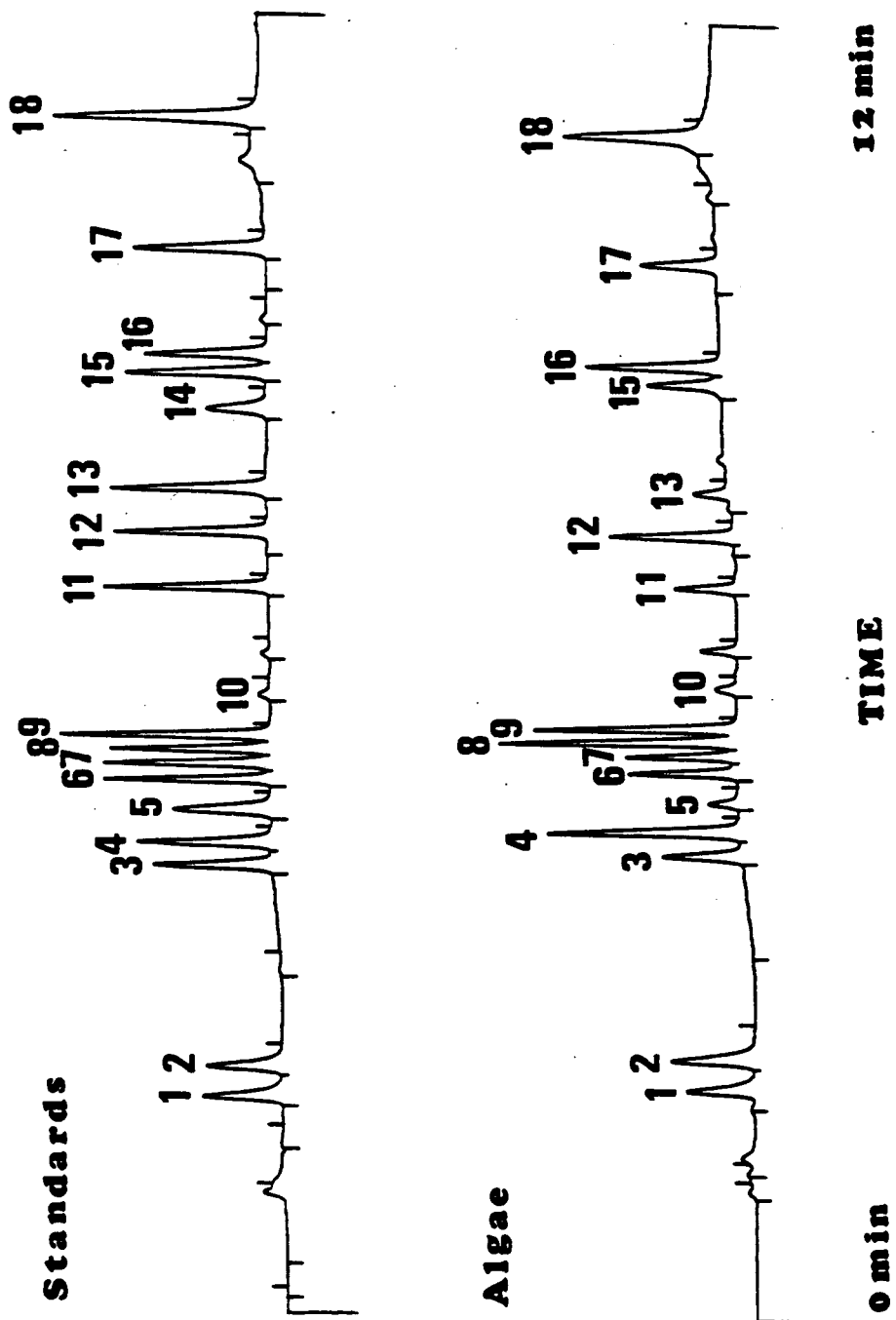


Fig. 5. IEF pattern of algal protein concentrate (fraction #6) and pH profile of IEF gel for determination of pI of focused proteins.



- | | |
|--------|---------------------|
| 1. ASP | 10. NH ₃ |
| 2. GLU | 11. TYR |
| 3. SER | 12. VAL |
| 4. GLY | 13. MET |
| 5. HIS | 14. CYS |
| 6. ARG | 15. ILE |
| 7. THR | 16. LEU |
| 8. ALA | 17. PHE |
| 9. PRO | 18. LYS |

Fig. 6. HPLC chromatograms of amino acid standards (Pierce standard H) and of algal protein isolate (fraction #8A).