# Efficiency of N Use by Wheat as a Function of Influx and Efflux of NO<sub>3</sub>

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## EFFICIENCY OF N USE BY WHEAT AS A FUNCTION OF INFLUX AND EFFLUX OF NO3

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#### ABSTRACT

Since N assimilation is one of the most costly functions of a plant, its efflux before assimilation results in a serious energy cost and loss in efficiency which could decrease yields. Efficient crop production is critical to CELSS. Our objective is to determine the extent of efflux of the N species  $NO_3$ ,  $NH_4$ ,  $NO_2$  and urea after uptake, and possible means of regulation. We found that  $NO_3$  efflux became serious as its substrate level increased. Efflux/Influx (E/I) of  $NO_3$  was greater in darkness (35%) than in light (14%) and the ratio greatly increased with increased substrate  $NO_3$ , (up to 45% at 10 mM). It seems advantageous to use the lowest possible nutrient concentration of  $NO_3$ . The feasibility of using  $CIO_3$  as a trapping agent (competitive inhibitor of  $NO_3$  uptake) for effluxed  $NO_3$  was assessed and its toxicity determined.

#### INTRODUCTION

Crop production during extended space flights requires the development of procedures leading to the optimum use of the available energy. Optimizing the utilization of N by crop plants represents an area where significant progress can be made in the CELSS program.

It is well recognized that ion uptake requires ATP and electron flow. The cost of  $NO_3^-$  assimilation is particularly high not only due to the energy requirement of uptake but also because of the need for 10 electrons to reduce it to the level of glutamate. In addition are the costs of maintaining pH balance, since each reduction of  $NO_3^-$  to the level of  $NH_4^+$  produces an  $OH_3^-$ . Thus it is not surprising that estimates of respiratory costs of ion absorption range up to 50% of total root respiration and 20% of total plant respiration (1).

Recent reports in the literature indicate that  $NO_3^-$  efflux can begin very rapidly after its influx (2,3,4,5,6). In addition, prolonged leakage of  $NO_3^-$  occurs from root storage pools dependent upon the N-status of the roots

(7,8,9,10,11). Lee and Clarkson (6) estimated that efflux, at external concentrations above 1 mM, could account for up to 40% of the influxed  $NO_3^-$ . Thus  $NO_3^-$  efflux could represent a significant additional cost to an already energy costly assimilatory pathway. Little is known concerning the regulation of  $NO_3^-$  efflux. Recent work of Briskin (12) showed evidence that efflux of  $NO_3^-$  utilizes ATP, which could greatly add to the energy cost of  $NO_3^-$  assimilation.

Measurement of  $NO_3^-$  efflux has been hampered by the lack of easy, rapid, and less costly analytical techniques for detecting the very low concentrations involved in a process with a half-life  $(t_{1/2})$  of minutes. The use of  $^{15}N$  as a tracer is laborious and has problems of sensitivity.  $^{13}N$  has only a 10 min half-life, must be used at the site of generation (cyclotron), and is very costly to use. We developed an HPLC method which has the sensitivity, ease and low cost required. In addition, we evaluated  $ClO_3^-$  as a trapping agent for  $NO_3^-$ .

This report presents results estimating  $NO_3^-$  influx, efflux, and net uptake across several mechanisms of  $NO_3^-$  uptake.

#### MATERIALS AND METHODS

Plant growth. Wheat seedlings were grown hydroponically in a 1/4 strength Hoagland's solution for 8 days in an environmentally controlled growth chamber at 400  $\mu$ E/m<sup>2</sup>sec, at 18°C, and 80% relative humidity (13,14). On the 7th day they were transferred into 1/4 strength Hoagland's (loading solution) containing NO<sub>3</sub><sup>-</sup> at the concentration to be used in the efflux study (specified in each experiment).

Measurement of  $\mathrm{NO_3}^-$  efflux. After removal from the loading solution, 10 seedlings were placed in 300 ml of the efflux solution for various periods of time (specified below) containing 0.06 mM Pi at pH 5.8, 0.2 mM  $\mathrm{CaSO_4}$ , and with or without  $\mathrm{ClO_3}^-$  at the specified concentrations. The seedlings were rinsed

for 2 sec in 300 ml of efflux solution, then placed in 60 ml of efflux solutions for the following times: 10 sec, 30 sec, 1, 2, 5, 10, 15, and 20 min and the amount of  $NO_3^-$  released at each time from the roots was determined.

Measurement of  $NO_3^-$  and  $C1O_3^-$ . These compounds were measured by HPLC as described previously for  $NO_3^-$  (13,14).  $C1O_3^-$  was also measured at 210 nm with a UV monitor.

Uptake rates of  $NO_3^-$  and  $C1O_3^-$ . Uptake rates were determined as previously described by determining rates of depletion of  $NO_3^-$  from substrate solutions, then fitting the rate curves to best fit curves by polynomial analysis using a computer (13).

#### RESULTS

Mechanisms of NO<sub>3</sub> uptake. The results in Figures 1 and 2 show several mechanisms for NO<sub>3</sub> uptake. Uptake as a function of NO<sub>3</sub> concentration can be determined by either step up or step down, on continuous depletion experiments; Figures 1 and 2 are the results of continuous depletion experiments. In Figure 1, one mechanism is readily seen between about 0.2 and 0.7 mM. This is commonly referred to as Mechanism I in the literature (14). The rates above 0.7 mM are largely undefined but are referred to in the literature as Mechanism II. Another mechanism is indicated in Figure 1 at concentrations below 0.1 mM and it is readily seen when the data are plotted between 0 and 0.1 mM (Fig. 2).

Comparison of uptake of  $NO_3^-$  and  $ClO_3^-$ . The comparative uptake of  $NO_3^-$  and  $ClO_3^-$  is shown in Figures 3 and 4 at initial concentrations of 0.5 and 1 mM. Wheat plants deplete the  $NO_3^-$  concentration very efficiently to near zero (Fig. 1), whereas depletion of  $ClO_3^-$  is not straight forward. The ability to take up  $ClO_3^-$  is continuously lost with time (Fig. 4).

Toxic effects of  ${\rm ClO_3}^-$  on  ${\rm NO_3}^-$  uptake. The increasingly toxic effects of  ${\rm ClO_3}^-$  on  ${\rm NO_3}^-$  uptake with time is seen in Table 1.

Comparative effects of pretreatments of  ${\rm C10_3}^-$  and  ${\rm NO_3}^-$  on their uptakes. The comparative effects of pretreatments of  ${\rm C10_3}^-$  and  ${\rm NO_3}^-$  on their uptakes are shown in Table 2. Pretreatments varying in time had little effect on subsequent uptake of  ${\rm NO_3}^-$ , whereas increasing time of pretreatments greatly decreased  ${\rm C10_3}^-$  uptake.

 ${\rm Cl0_3}^-$  as a competitive inhibitor of  ${\rm NO_3}^-$  uptake. Double reciprocal plots of rates vs concentrations show evidence that  ${\rm Cl0_3}^-$  is a competitive inhibitor of  ${\rm NO_3}^-$  uptake (Fig. 5).

 $NO_3^-$  efflux. Figure 6 shows a typical example of a determination of  $NO_3^-$  efflux. Our results matched quite closely those reported in the literature showing two different early losses of  $NO_3^-$ , one with a  $t_{1/2}$  of less than 10 sec and another with a  $t_{1/2}$  of minutes. After one min, the second set of rates approximate an apparent first order reaction. Extrapolation to  $t_0$  gives an estimate of the rate of  $NO_3^-$  efflux. At a concentration of 1 mM  $NO_3^-$ , efflux varied consistently between 2.0 and 2.5  $\mu$ mol/gxh.

Effect of increasing concentrations of  $NO_3^-$  and  $C1O_3^-$  on  $NO_3^-$  efflux. Efflux in the presence of increasing concentrations of  $NO_3^-$  and  $C1O_3^-$  is shown in Figure 7. As expected, efflux increased with increasing concentrations of  $NO_3^-$  and  $C1O_3^-$ .

Influx, efflux, and net uptake of  $NO_3^-$ . Comparative rates of the three kinetic components of  $NO_3^-$  absorption are shown in Table 3. Efflux and influx greatly increased between 0.2 and 10 mM external  $NO_3^-$ , whereas net uptake remained about the same. Efflux/influx increased from 15 to 45% with increasing concentration of  $NO_3^-$ .

Effect of light and dark on influx, efflux, and net uptake of  $NO_3^-$ .  $NO_3^-$  efflux was similar in plants in darkness and in light; however, influx and net uptake were much greater in light (Table 4). Thus, in darkness 35% of the influxed  $NO_3^-$  was effluxed, while in light the proportion effluxed was reduced to 14%.

#### DISCUSSION

It is important to continue to develop information concerning the mechanisms of uptake of nutrient ions both to understand the reactions of the plants to changing concentrations and also for planning optimum concentrations of nutrient solutions for maximum efficiency.

Mechanisms of  $NO_3^-$  uptake. At least three mechanisms of  $NO_3^-$  uptake are present, one between 0 and about 0.05 to 0.08 mM with a  $K_m$  of ca 0.012 to 0.018 mM, and another between 0.1 and about 0.7 mM with a  $k_m$  of ca 0.025 to 0.04 mM (Figs. 1 and 2). The latter is the typical mechanism reported in the literature (14). At concentrations above 1 mM, the mechanisms are largely undefined and are difficult to determine because efflux becomes such an important component (discussed below).

Comparison of uptake of  $NO_3^-$  and  $ClO_3^-$ . In a determination of  $NO_3^-$  efflux, very low concentrations of  $NO_3^-$  are present in the external solution. At these low levels of  $NO_3^-$  (see Fig. 3), the wheat plant can very efficiently absorb the  $NO_3^-$  as it is effluxed into the external solution. Hence,  $ClO_3^-$  has been used as a trapping agent for the effluxed  $NO_3^-$ .

Although much work has been reported on the effects of  ${\rm C10_3}^-$  on  ${\rm NO_3}^-$  uptake, the analytical procedures reported to separate  ${\rm C10_3}^-$  from  ${\rm NO_3}^-$  in the solutions were in some cases non specific, i.e., ion electrodes (3). Radio-active  ${\rm C10_3}^-$  was also used (3) which presents problems of low specific activity and sticking to glassware. We developed an HPLC method which effectively separates  ${\rm NO_3}^-$  from  ${\rm C10_3}^-$  and both can be measured simultaneously. In addition, much of the reported literature did not discriminate between kinetic effects and toxic effects of  ${\rm C10_3}^-$  on  ${\rm NO_3}^-$  uptake (15).

Toxic effects of  ${\rm C10_3}^-$ . The results showed that toxicity symptoms, as shown by decreased rates of uptake of  ${\rm N0_3}^-$  and  ${\rm C10_3}^-$ , were apparent after 1 h (Fig. 4, Tables 1 and 2). Toxic effects of  ${\rm C10_3}^-$  were greater toward its own

uptake than towards  $NO_3$  uptake. We found that little toxicity occurred during the 20 min period of the efflux experiments.

 $C10_3^-$  as a competitive inhibitor of  $NO_3^-$  uptake. The results verified that  $C10_3^-$  was a competitive inhibitor of  $NO_3^-$  uptake as earlier reported (3) (Fig. 5). Apparently the  $NO_3^-$  transporter discriminated effectively between  $NO_3^-$  and  $C10_3^-$  since proportionately much larger concentrations of  $C10_3^-$  were required to inhibit  $NO_3^-$  uptake. In summary,  $C10_3^-$  could be used as a trapping agent for effluxed  $NO_3^-$  for short time experiments.

 $NO_3^-$  efflux. The method for determining efflux was based on some excellent work done by Lee and Clarkson (6) and Shone and Flood (16) who used cereal roots to determine the kinetic parameters required to measure efflux of  $NO_3^-$ . They found that the combined  $t_{1/2}$  of release of  $NO_3^-$  in the surface film attached to the roots from the external solution and from the root free space was ca 7 sec. In addition, their results showed that the  $t_{1/2}$  for cytoplasmic release of  $NO_3^-$  was ca 4 min. A semilog plot of efflux rate vs time resulted in a linear regression line after 1 min, since at that time the first two kinetic parameters had passed through ca 9 half-lives. In our results, the relative contributions of these parameters would be very small in relation to the amount of  $NO_3^-$  efflux. The rates of efflux we measured are similar to those reported for cereals by workers using  $^{13}N-NO_3^-$  at  $NO_3^-$  concentrations of 1.5 to 5 mM (3,4,6).

Effect of increasing concentrations  $NO_3^-$  and  $ClO_3^-$  on  $NO_3^-$  efflux. As expected, efflux increased with increasing concentrations of  $NO_3^-$  (Fig. 7). In the region of Mechanism II of uptake (0.2 mM), efflux was a significant deterrent to N use efficiency. Here 15% of the  $NO_3^-$  influxed was effluxed. Net  $NO_3^-$  uptake remained quite constant as external  $NO_3^-$  increased beyond 0.2 mM. This occurred because, although influx increased, efflux correspondingly increased. The ratio of efflux/influx, expressed as a percentage, increased up to 45% at 10 mM  $NO_3^-$ . These results help explain results from Raper's

laboratory which showed that net uptake changed little as  $NO_3$  concentrations increased to high levels (11). These workers also presented evidence for significant levels of efflux at increasing concentrations of  $NO_3$ .

Effect of light and dark on influx, efflux, and net uptake of  $NO_3^-$ . Since efflux is a function of the concentration of  $NO_3^-$  in the cytoplasm, light and dark periods had a profound effect on the ratio of efflux/influx (Table 4). Although efflux was the same in light or dark, influx was greatly increased in light along with the rate of  $NO_3^-$  reduction (17,18). Therefore, the relative percentage of  $NO_3^-$  effluxed is much less in light.

Effect of increasing concentrations  ${\rm C10_3}^-$  on  ${\rm NO_3}^-$  efflux. Higher rates of  ${\rm NO_3}^-$  efflux were detected as  ${\rm C10_3}^-$  increased in the efflux solution (Fig. 7). This could be because of  ${\rm C10_3}^-$  serving as a trapping agent or because of an unknown effect. The increase in efflux rate with 5 and 10 mM  ${\rm C10_3}^-$  was sufficiently large to be somewhat uncertain. We are currently evaluating this result.

Effect of  $\mathrm{NH_4}^+$  on  $\mathrm{NO_3}^-$  efflux. The literature is confusing on this issue with results of little if any effect of  $\mathrm{NH_4}^+$  on  $\mathrm{NO_3}^-$  efflux (5,6), and of large effect (4). We have not yet examined the interactions of the different N species on  $\mathrm{NO_3}^-$  efflux.

Ramifications. As the concentration of  $NO_3^-$  increases in the external solution, efflux becomes an increasingly important energy cost to the plant. Not only is energy needed for  $NO_3^-$  influx, it now appears that  $NO_3^-$  efflux may utilize ATP (12). This results in an almost doubling of the cost of absorption at higher concentrations of  $NO_3^-$ . On this basis, it would seem that a nutrient solution for crop growth in CELSS should be optimized at the lowest concentrations possible. In addition, it is increasingly critical to determine the regulation of efflux of N compounds since it is not known how  $NO_3^-$ ,  $NH_4^+$ ,  $NO_2^-$ , and urea influence each others efflux. Will total efflux

decrease if the total concentration of N in the nutrient solution is made up of the four species instead of only  $NO_3^-$  and  $NH_4^{+}$ ?

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Table 1. Effect of pretreatment of  ${\rm ClO_3}$  on  ${\rm NO_3}$  uptake. The initial concentration of substrate  ${\rm NO_3}$  was 0.6mM. See Materials and Methods for procedures.

	[C:	10 <sub>3</sub> ] 5mM
HOURS PRETREATMENT	<u>NO<sub>3</sub> 1</u>	<u>UPTAKE</u> l/gxh)
0	7.3	7.3
1	6.2	5.3
2	5.3	3.0
3	A Q	2 2

Table 2. Effect of pretreatment of ClO<sub>3</sub> or NO<sub>3</sub> on their uptakes. See Materials and Methods for procedures.

	PRETREATMENT (1mM)		
	ио3	clo <sub>3</sub>	
HOURS PRETREATMENT	<u>UPTAKE (at 1mM)</u> (μmol/gxh)		
0	6.3	3.5	
3		1.1	
5	6.8	0.5	

Table 3. Efflux, net uptake and influx of NO<sub>3</sub>. See Materials & Methods for procedures.

### influx, efflux, and net uptake of $no_3$

	EFFLUX	NET UPTAKE	INFLUX	EFFLUX INFLUX
[NO <sub>3</sub> ] (mM)	#** j	μmol/gxh	1	*
0.2	1.1	6.5	7.6	15
1.0	2.5	8.5	11.0	23
10.0	7.7	9.5	17.2	45

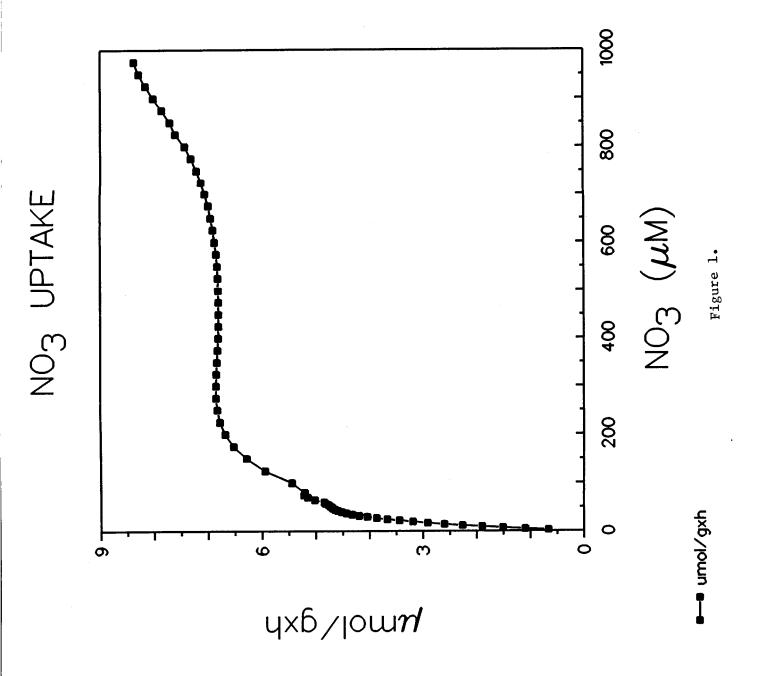
Table 4. Effect of light and dark treatments (24h) on  $NO_3$  efflux, net uptake and influx. See Material and Methods for procedures.

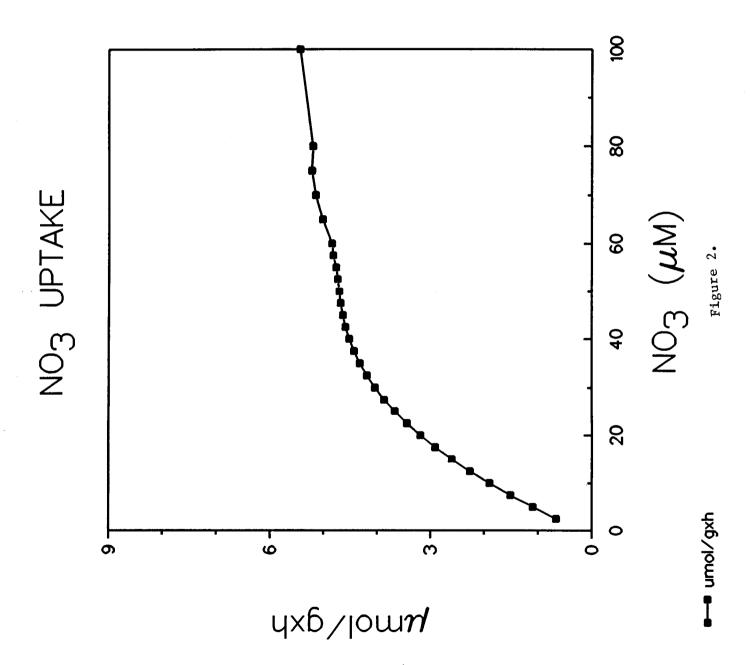
TREATMENT	<u>EFFLUX</u>	NET <u>UPTAKE</u>	INFLUX	EFFLUX INFLUX
		(µmol/gxl	h)	8
LIGHT*	1.3	7.9	9.2	14
DARK	1.0	1.8	2.8	35

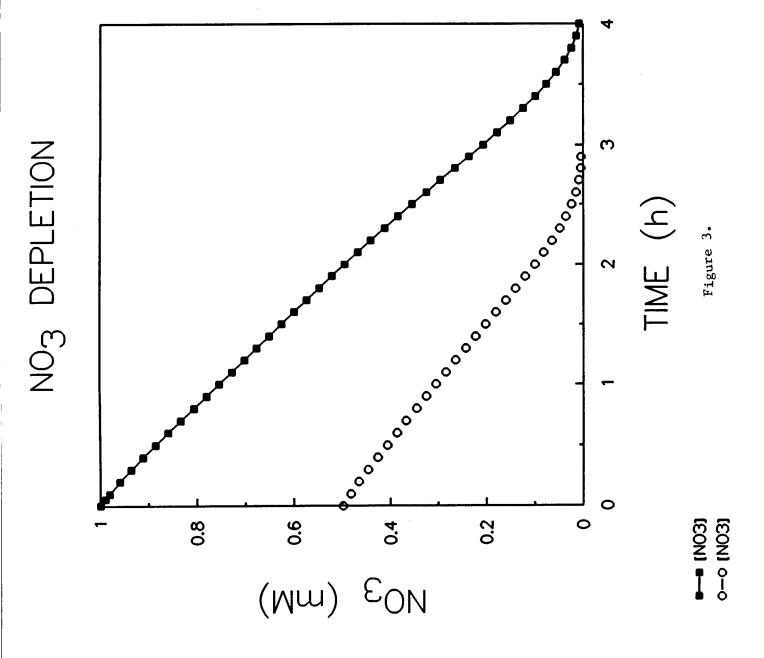
<sup>\*</sup>plants induced in 0.2mM NO<sub>3</sub> for 24h light or dark

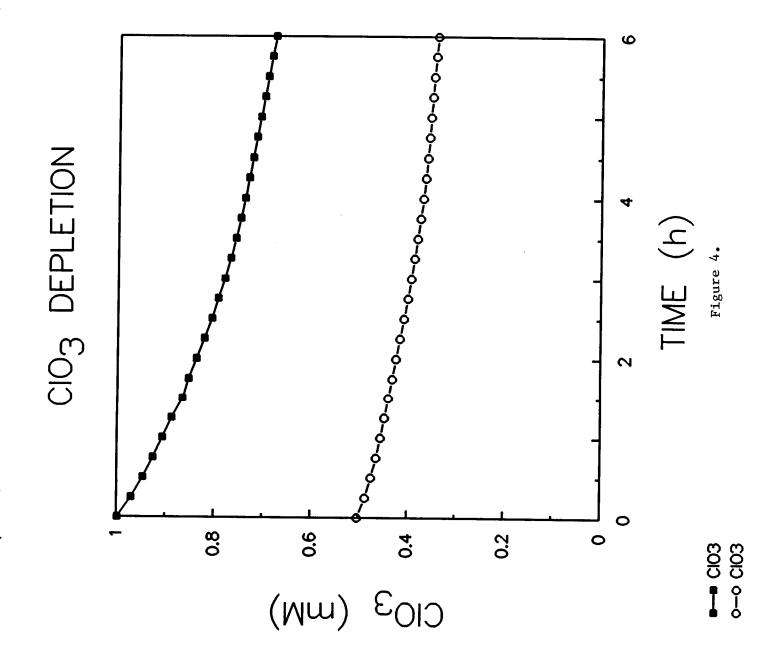
#### FIGURE LEGENDS

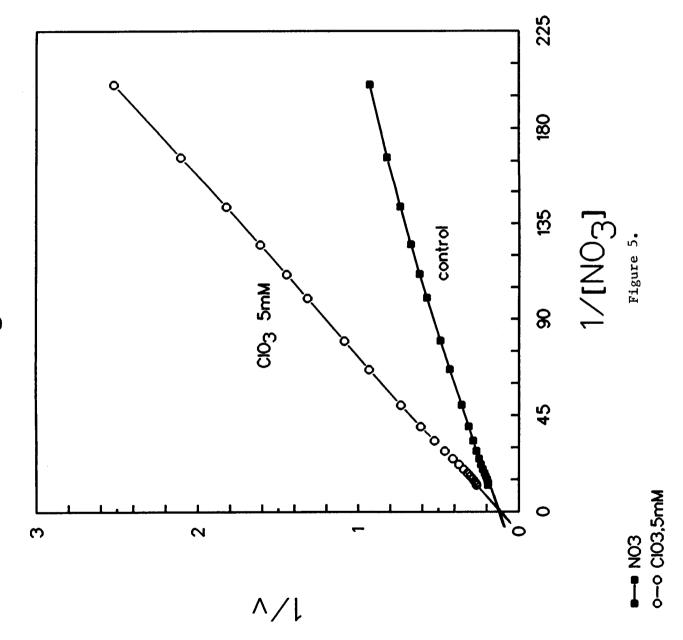
- Fig. 1.  ${\rm NO_3}$  uptake across Mechanisms I and II. See Materials and Methods for procedures.
- Fig. 2.  ${\rm NO_3}$  uptake across Mechanism I. See Materials and Methods for procedures.
- Fig. 3. Depletion of  $NO_3$  from 0.5 and 1.0mM substrate solutions. See Materials and Methods for procedures.
- Fig. 4. Depletion of ClO<sub>3</sub> from 0.5 and 1.0mM substrate solutions. See Materials and Methods for procedures.
- Fig. 5.  ${\rm ClO_3}$  as a competitive inhibitor of  ${\rm NO_3}$  uptake. See Materials and Methods for procedures.
- Fig.6. Semilog plot of NO<sub>3</sub> efflux vs time. See Materials and Methods for procedures.
- Fig.7. Effect of  ${\rm ClO_3}$  on  ${\rm NO_3}$  efflux. See Materials and Methods for procedures.

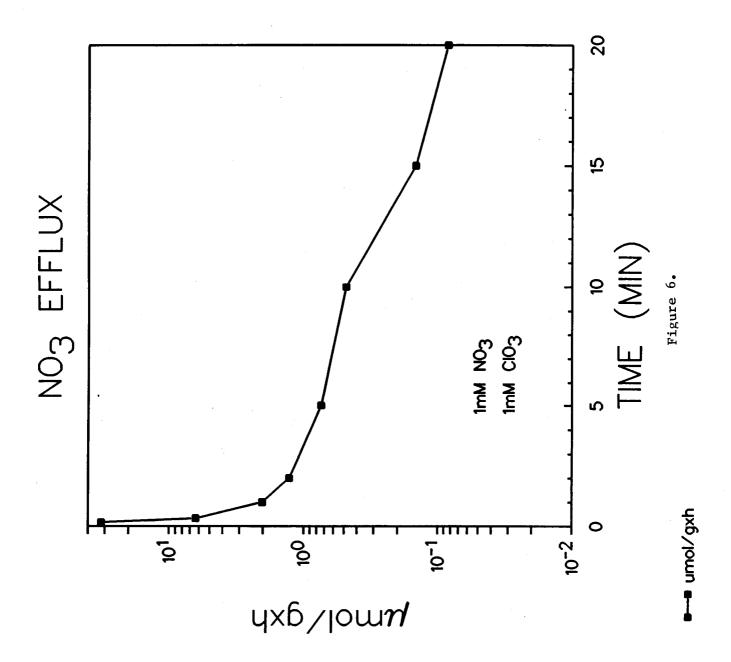


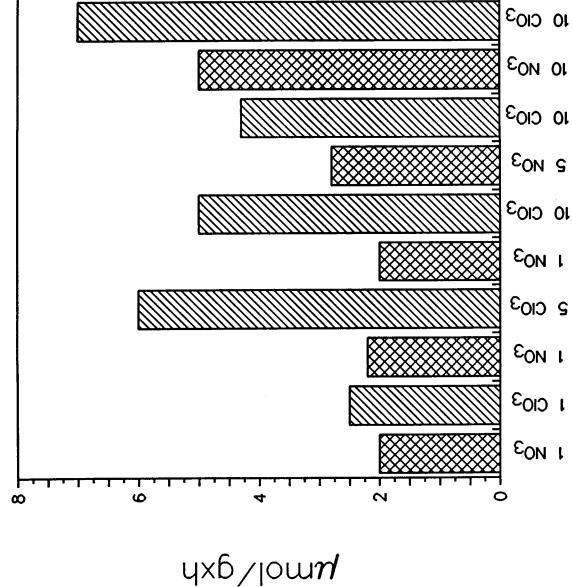












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Robert D. MacElroy, NASA-Ames Research Center Point of Contact: MS 239-4, Moffett Field, CA 94035 415-694-5573 or FTS 8-464-5573  16. Abstract Since N assimilation is one of the most costly functions of a plant, its efflux before assimilation results in a serious energy cost and loss in efficiency which could decrease yields. Efficient crop production is critical to CELSS. Our objective is to determine the extent of efflux of the N species NO <sub>2</sub> -, NH <sub>1</sub> +, NO <sub>2</sub> -, and urea after uptake, and possible means of regulation. We found that NO <sub>3</sub> - efflux became serious as its substrate level increased. Efflux/Influx (E/I) of NO <sub>2</sub> - was greater in darkness (35%) than in light (14%) and the ratio greatly increased with increased substrate NO <sub>3</sub> -, (up to 45% at 10mM). It seems advantageous to use the lowest possible nutrient concentration of NO <sub>3</sub> The feasibility of using C10 <sub>2</sub> - as a trapping agent (competitive inhibitor of NO <sub>3</sub> - uptake) for effluxed NO <sub>3</sub> <sup>2</sup> was assessed and its toxicity determined.					
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