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Statistical Design in Isothermal Aging of Polyimide Resins

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SYNOPSIS

Recent developments in research on polyimides for high temperature applications have led to the synthesis of many new polymers. Among the criteria that determines their thermal oxidative stability, isothermal aging is one of the most important. Isothermal aging studies require that many experimental factors are controlled to provide accurate results. In this article we describe a statistical plan that compares the isothermal stability of several polyimide resins, while minimizing the variations inherent in high-temperature aging studies.

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INTRODUCTION

The NASA Lewis Research Center HITEMP program in polymer matrix composites (PMC) focuses on the development of high-temperature polymers for advanced aircraft engine fan and compressor applications. One of this program's main goals is to produce polymers that can withstand engine operating temperatures up to 425°C (800°F). Therefore, an important feature of these polymers is their thermal oxidative stability at high temperatures.

Several polymer matrix resins have been developed at NASA Lewis to meet these goals. PMR-II-50 and VCAP-75 are two matrix resins that were investigated in engine component development programs.^{1,2} Additional resins developed at Lewis for high-temperature applications are N-CYCAP, 12F-VCAP, 3F, and MARVimides. DuPont Company and TRW, Inc. have also developed resins for high-temperature applications. These resins are Avimid-N[®] and AFR700B.

We report the findings of a study that we designed and analyzed with formal statistical methodologies. Specifically, the study was designed to compare, in a valid quantifiable manner, the respective weight losses among a selected set of five different resins after 400 h of exposure to high temperature (371°C). The resins included in our investigation were

Avimid-N, AFR700B, PMR-II-50, VCAP-75, and N-CYCAP (Fig. 1).

Implementation of a statistically designed study allowed us to address the following two questions. Can we determine weight loss differences between these polymers? If so, how accurate are the estimated differences? The design for this study accounted for a location effect within the oven, and a molding powder or disk effect within a polymer. Two molding powders were available for PMR-II-50, VCAP-75, and N-CYCAP. One disk was produced from each of these powders. We obtained two disks from each resin developed by DuPont and TRW (2 Avimid-N disks and 2 AFR700B disks, respectively). Although the disks of Avimid-N came from a single batch of molding powder as did the disks of AFR700B, the molding powder-molding powder variability within a polymer was treated in the same way as the disk-disk variability within a polymer.

EXPERIMENTAL

Instrumentation

All infrared spectra were recorded with a Nicolet model 510P Fourier Transform Spectrometer in KBr pellets. All thermal analyses were recorded on a PL Thermal Sciences Thermal Analysis System. Thermogravimetric analyses were done with a Perkin-Elmer TGS-2 on postcured neat resin samples under air using a scan rate of 10°C/min. Thermomechan-

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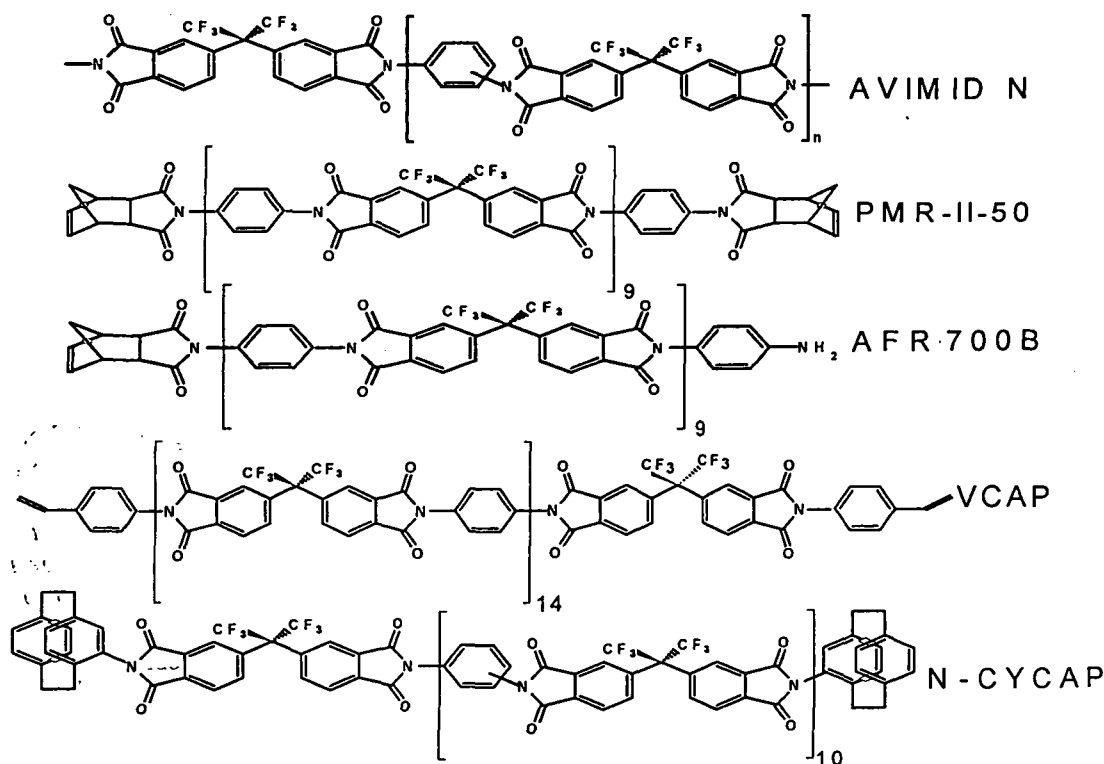


Figure 1 Polyimides aged at 371°C.

ical analyses were performed with a DuPont Instruments 943 TMA that employed a scan rate of 10°C/min on neat resin disks. Differential scanning calorimetry was carried out on imidized molding powders in a DuPont Instruments Model 910 DSC cell with a scan rate of 10°C/min under ambient air. Paired-ion HPLC analyses were done with a Beckman System Gold chromatograph using a Hamilton PRP-1 column and a gradient mixture of acetonitrile (with water as the solvent system). The water for HPLC analysis was purified with a Millipore Milli-Q system and subsequently treated with tetrabutylammonium perchlorate as a paired-ion reagent. Hardened steel dies were purchased from F. Carver. A Wabash press equipped with a 30.5 × 30.5 cm high-temperature platens was used to compression mold the resin disks. A Blue M Model #POM-6680E-3 was used to age the disks in air.

Materials

Avimid-N resin samples, *p*-phenylene diamine (*p*-PDA), and *m*-phenylenediamine (*m*-PDA) were supplied by DuPont Company Advanced Materials. AFR700B molding powder (lot nos. 308801 and 217501) were purchased from HyCOMP Inc. 4-Amino [2.2] paracyclophane was synthesized ac-

ording to an improved method.³ The following chemicals were used as received: reagent grade glacial acetic acid (Fisher), reagent grade acetic anhydride (Aldrich), 4,4'-(hexafluoroisopropylidene) diphthalic anhydride (HFDA) (Hoechst Celanese), tetrabutylammonium perchlorate (Kodak), monomethyl *cis*-5-norbornene-endo-2,3-dicarboxylate (Pharm-Eco Laboratories), *p*-aminostyrene (Polysciences Inc.), mono-Coat E63FF release agent (Chem-trend). 4,4'-(Hexafluoroisopropylidene) diphthalic anhydride dimethyl ester diacid (HFDE) was prepared by refluxing HFDA in methanol for 2 h under a nitrogen atmosphere such that the final concentration of HFDE was 50 wt %. The purity of HFDE was determined by HPLC analysis.

Synthesis of PMR-II-50, VCAP-75, and N-CYCAP Molding Powders

The number of moles of monomeric reactants in PMR-II-50 was governed by the ratio $n : n + 1 : 2$, where n , $n + 1$, and 2 are the number of moles of dianhydride (or diacid diester), diamine, and end cap, respectively, although the number of moles of monomeric reactants in VCAP-75 and N-CYCAP was governed by the ratio $n : n + 1 : 2$, where n , $n + 1$, and 2 are the number of moles of diamine, dian-

hydride (or diacid diester), and end cap, respectively. Both PMR-II-50 and VCAP-75 use HFDE as the diacid diester. N-CYCAP uses HFDA as the dianhydride. The n values for PMR-II-50, VCAP-75, and N-CYCAP are 9, 14, and 10, respectively.

PMR-II-50

To a solution of HFDE (4.53 g, 8.9 mmol) in methanol we added *p*-PDA (1.07 g, 9.9 mmol) and nadic acid ester (0.389 g, 1.98 mmol). This mixture was gently heated to reduce the methanol content and resulted in a viscous gum that was further heated in an air oven at 210°C for 2 h.

VCAP-75

To a solution of HFDE (4.84 g, 9.5 mmol) in methanol was added *p*-PDA (0.92 g, 8.5 mmol) and *p*-aminostyrene (0.15 g, 1.26 mmol). This mixture was gently heated to reduce the methanol content and resulted in a viscous gum that was further heated in an air oven at 210°C.

N-CYCAP

To a finely dispersed mixture of HFDA (4.06 g, 9.15 mmol) and glacial acetic acid (50 mL) was added *p*-PDA (0.88 g, 8.15 mmol) and 4-amino [2.2] paracyclophane (0.37 g, 1.6 mmol). The reaction mixture was refluxed. Initially, the reaction mixture separated into two phases. However, after 30 min the mixture became a clear amber solution. The homogeneous solution persisted for approximately 15 min before forming a precipitate. Refluxing was continued for an additional 12–16 h. Acetic anhydride (6 mL) was added to the slurry and stirred for an additional 2–3 h while refluxing. The reaction was cooled to room temperature and poured into a beaker containing ice water (150 mL) and stirred for 30 min. The mixture was filtered, and the molding powder was washed with cold water (3×150 mL) and air dried. The prepolymer was then vacuum dried at 100°C overnight. Typical yields ranged from 95 to 100%.

Processing Polyimide Molding Powders and Aging of Neat Resins

A cylindrical 2.54-cm hardened-steel mold equipped with a thermocouple was sprayed with a high-temperature release agent (monocoat E63FF) and dried at room temperature for 20 min. Sheets of Kapton (0.5-mm thick) were sprayed with release agent and cut to fit in the mold. The Kapton was placed in the

mold on the top and bottom of the molding powder. The mold was charged with approximately 1.3 g of prepolymer and placed in a preheated (380°C) press. When the temperature of the mold reached approximately 230°C, 6.9 MPa was applied. The temperature was increased from 230 to 370°C over 7 min and held at 370°C. After 2 h, the mold was cooled to 200°C, then the pressure was released, and finally, the resin disk was removed. No sample weight loss was observed during the molding process. All resins were postcured at 371°C in air for 16 h. Samples were predried at 135°C in air for 1 h and then placed in a desiccator. Once the postcured resins were cooled to 25°C, they were weighed and then placed in the air oven. The samples were aged for 400 h at 371°C. All of the resins remained in the oven for the 400-h period, that is, weight losses were not recorded at periodic intervals. After aging, the samples were placed in a desiccator and cooled to 25°C. The final weight loss values were calculated as follows: the weight of resins after postcuring minus the weight of resin after 400 h aging equals the weight of resin remaining.

RESULTS AND DISCUSSION

Each disk from each polymer was split into eight equally sized samples. We partitioned the tray into four locations to provide a valid comparison of percent weight losses among the resins that experienced similar aging conditions. Each location was 3 slots by 7 slots, that is, 21 slots. For a given polymer-disk combination, two samples were randomly assigned to each of the four rectangular locations. This procedure produced 20 samples per location (5 polymers \times 2 disks/polymers \times 2 samples/disks). The resulting 20 samples in a given location were randomly allocated to the 21 slots (Fig. 2). In our analysis, the percent weight losses were consistently different, for a given resin pair, from location to location (Table I).

Least squares was used to fit the following model:

$$y_{ijkl} = \mu + R_i + L_j + RL_{ij} + B_{k(i)} + LB_{jk(i)} + \epsilon_{l(ijk)} \quad (1)$$

where y_{ijkl} is % weight loss; μ is overall constant; R_i is fixed polymer effect; L_j is fixed location effect; $B_{k(i)}$ is random molding powder within polymer effect; RL_{ij} is fixed resin by location interaction effect; $LB_{jk(i)}$ is random location by molding powder interaction effect within polymer; $\epsilon_{l(ijk)}$ is random error

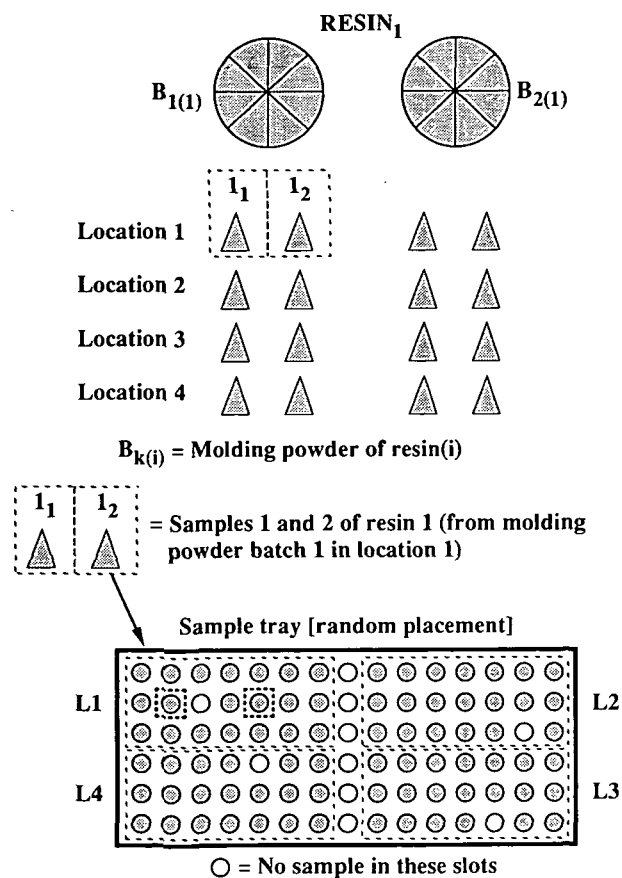


Figure 2 Experimental layout.

term; in which subscripts $i = 1, \dots, 5$ is polymer number; $j = 1, \dots, 4$ location number; $k = 1, 2$ is molding powder number; and $l = 1, 2$ sample number.

In this model, molding powders 1 and 2 from polymer 1 are different from molding powders 1 and 2 from polymer 2, etc. Sample numbers 1 and 2 from a given polymer, molding powder, and location combination are different from samples 1 and 2 from a different polymer, molding powder, and location combination. The variables $\epsilon_{l(ijk)}$, $B_{k(i)}$ and $LB_{jk(i)}$

were treated as normal random variables with zero means and variances σ_{ϵ}^2 , σ_B^2 , and σ_{LB}^2 .

The normal probability plot (used to evaluate our normal theory assumption) and residual plot (used to evaluate the constant variance assumption for every y_{ijkl}) both detected violations of the corresponding assumptions (Fig. 3). Hence, a natural logarithm transformation was applied to the y_{ijkl} 's, and least squares was used to fit the model given by eq. (2). The resulting normal probability plot showed a negligible departure from the assumed normal theory, and the residual vs. the predicted plot revealed a constant variance (Fig. 4). Thus, our analyses are applicable on the natural log scale (ln).

Table II contains development of the expected mean squares (MS) for the model

$$\ln(y_{ijkl}) = \mu + R_i + L_j + RL_{ij} + B_{k(i)} + LB_{jk(i)} + \epsilon_{l(ijk)}. \quad (2)$$

Table II is derived from the rules Neter et al.⁴ described, and indicates how to form test statistics applicable to the questions motivating this article. Table III presents an analysis of variance (ANOVA) obtained using the least squares methodology. We see that the ANOVA given in Table III contains estimates of the corresponding expected MS given in Table II. Thus, to test the null hypothesis [H_0 : Each of the 10 polymer differences are the same for each location ($RL_{ij} = 0$ for every ij)] against the alternate hypothesis [H_A : There exists at least one difference among polymers that is not the same from location to location ($RL_{ij} \neq 0$ for at least one ij combination)], the appropriate test statistic (F) is indicated by Table II to be

$$F_{12,15} = \frac{\text{MS location} \times \text{polymer}}{\text{MS location} \times \text{powder (polymer)}}. \quad (3)$$

For example,

Table I Raw Percent Weight Loss after 371°C Isothermal Aging

Location	Avimid-N	AFR700B	VCAP-75	N-CYCAP	PMR-II-50
1	9.7	26.7	26.2	25.2	28.8
2	9.8	23.6	23.1	24.7	25.2
3	8.9	21.9	22.7	22.2	24.3
4	10.7	23.5	24.5	26.3	30.8
Average (1-4)	9.8	23.9	24.2	24.6	27.2

Means averaged across locations. Aging performed in a Blue M, Model CW160E-MP with a temperature capability up to 704°C. Environment for 371°C testing: 1 atm air.

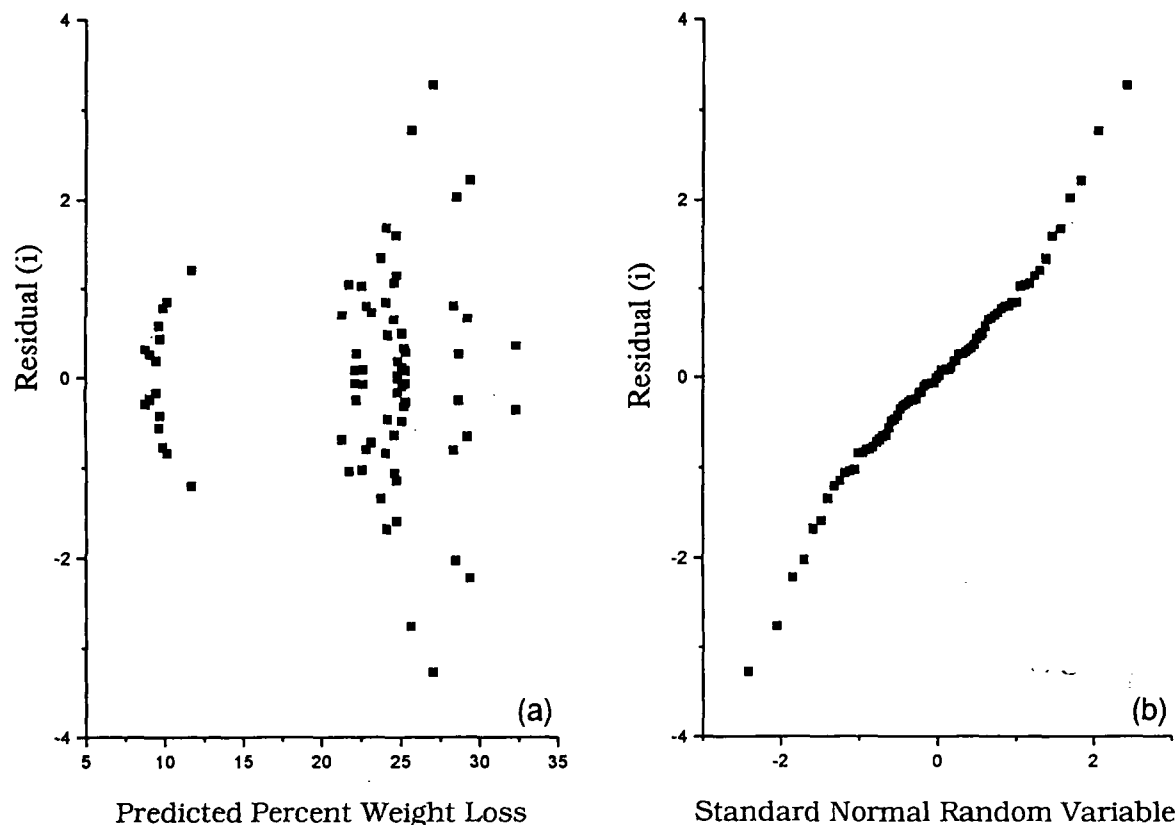


Figure 3 (a) Residual and (b) normal probability plots using the untransformed data.

$$F_{12,15,calc} = \frac{0.00919039}{0.00521} = 1.7639. \quad (4)$$

For a level of significance $\alpha = 0.05$, we require $F_{(12,15,\alpha=0.05)} = 2.48$. Because $F_{(12,15,calc)} < 2.48$, we do not have strong evidence to conclude that there exists at least one difference among polymers that is not the same from location to location (H_A). Instead, we conclude that there is no interaction of location with polymer type (H_0). In fact, the p value (observed significance level) > 0.10 .

Because polymer differences are consistent from location to location, it is valid to average the polymer log percent weight losses across locations before comparing them to one another. Therefore, to test this null hypothesis (H_0 : All polymers are the same with respect to average log percent weight loss.) against an alternate hypothesis (H_A : At least one polymer is different from one or more polymers with respect to average log percent weight loss.), we use Table II to identify the appropriate statistic as

$$F_{4,5} = \frac{\text{MS polymer}}{\text{MS powder (polymer)}}. \quad (5)$$

For example,

$$F_{4,5,calc} = \frac{2.842411}{0.00924} = 307.6202. \quad (6)$$

For a level of significance $\alpha = 0.05$, we require $F_{(4,5,\alpha=0.05)} = 5.19$. Because $F_{(4,5,calc)} > 5.19$, we have strong evidence to conclude that there exists at least one difference among the true average log percent weight losses for the respective polymers (H_A). The p value $\ll 0.01$.

Fisher's protected least significant difference (LSD)⁵ and Tukey's honest significant difference (HSD)⁴ are the two multiple comparison methods we applied to evaluate the differences in percent weight loss from polymer to polymer (on the ln scale) (Table IV). A 0.05 significance level was chosen for each method. Differences in the mean of the logs among the five polymers are displayed with material footnoted Table IV. Fisher's protected LSD grouped the polymers as indicated by footnote a. With the more conservative Tukey's HSD, we observed the grouping illustrated with footnote b.

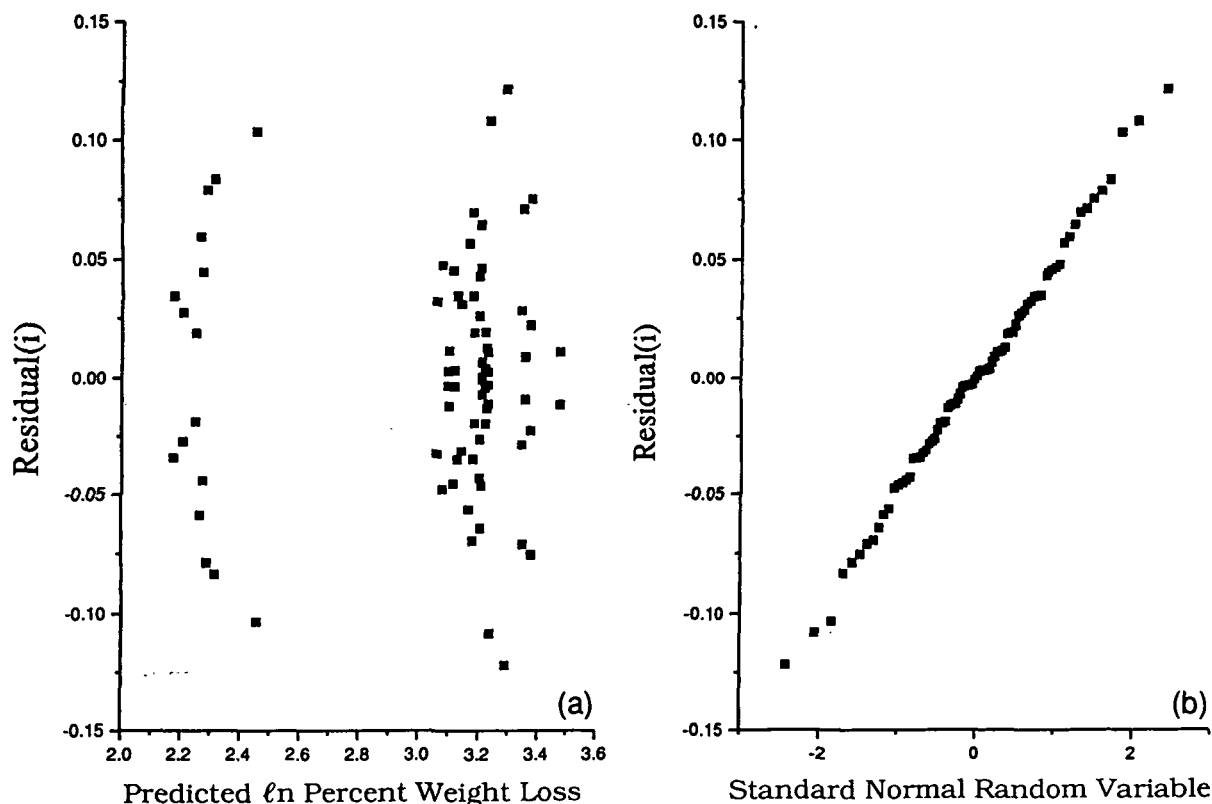


Figure 4 (a) Residual and (b) normal probability plots using the transformed data.

Resins are grouped together when there is no significant difference in the weight loss among them on the natural log scale according to the method used. We conclude that there is a 5% chance that we have made at least one mistake in the seven significant differences detected using the LSD method, and in the four significant differences using the HSD method (footnotes a and b in Table IV).

The only difference in Tukey's HSD and Fisher's protected LSD is that PMR-II-50 is judged to be in

the same group as AFR700B, VCAP-75, and N-CY-CAP with Tukey's HSD (footnote b, Table IV), whereas the LSD method identified that an important difference existed between PMR-II-50 and the other polymers (footnote a, Table IV). Another graphic representation of Fisher's protected LSD and Tukey's HSD methods can be seen in Figures 5 and 6, respectively. The conclusions drawn from Figures 5 and 6 are mathematically equivalent to those drawn from Table IV(b). This corresponds to

Table II Expected Mean Squares for Stated Model [Eq. (2)]

Source	Degrees of Freedom	Expected Mean Square
Corrected total	79	
Polymer	4	$(\frac{16}{4}) \sum_{i=1}^5 R_i^2 + 8\sigma_B^2 + \sigma^2$
Location	3	$(\frac{20}{3}) \sum_{j=1}^4 L_j^2 + 2\sigma_{LB}^2 + \sigma^2$
Location \times polymer	12	$(\frac{4}{12}) \sum_{i=1}^5 \sum_{j=1}^4 RL_{ij}^2 + 2\sigma_{LB}^2 + \sigma^2$
Powder (polymer)	5	$8\sigma_B^2 + \sigma^2$
Location \times powder (polymer)	15	$2\sigma_{LB}^2 + \sigma^2$
Error	40	σ^2

Table III Analysis of Variance for Fitted Model Using $\ln(y)$

Source	Degrees of Freedom	Sum of Squares	Mean Squares
Corrected total	79	12.096	
Polymer	4	11.369	2.8424
Location	3	0.29665	0.09888
Location \times polymer	12	0.11028	0.00919
Powder (polymer)	15	0.04623	0.00924
Location \times powder (polymer)	1	0.07816	0.00521
Error	40	0.19519	0.00488

the underscoring in Table IV(a). If the bounds do not overlap in Figures 5 and 6, significant differences exist above and beyond that due to randomness among the means of the natural log percent weight loss for each of the resins.

Tukey's HSD method is more conservative because its interpretation implies that not only is there at most a 5% chance that we have made at least one mistake in the four identified differences, but that we have a 95% chance or confidence that all the $\binom{5}{2} = 10$ intervals for the differences in means of the log percent weight losses actually include the true differences. This cannot be said using Fisher's protected LSD. The raw mean of percent weight loss for each of the polymers averaged across locations and for each location are shown in Table I.

The 10 HSD 95% intervals mentioned in the previous paragraph take on the form

$$\hat{D} \pm T \cdot s\{\hat{D}\}. \quad (7)$$

\hat{D} is the estimated difference in the average log per-

cent weight loss, after averaging across locations, for a given pair of polymers. The estimated standard deviation of \hat{D} is

$$s\{\hat{D}\} = \left[\frac{2}{16} (\text{MS powder (polymer)}) \right]^{1/2}. \quad (8)$$

The constant T is available from statistical tables. (See Neter et al.⁴ for details of Tukey's HSD multiple comparison methods.) Table V contains the 10 intervals constructed using Tukey's HSD 95% confidence interval methodology.

Now, one might ask about the interpretation of our results in terms of the raw data (percent weight loss). We gain insight into this question by back-transforming with the exponential transformation. Because our normal probability plot revealed logs of the percent weight losses to be normally distributed, we are dealing with the lognormal distribution; that is, $\ln(y_{ijkl})$ is normally distributed. Denoting δ as the true mean of a lognormally distributed random variable, δ is also the median of the lognormally distributed random variable because of symmetry.

Table IV Comparison of Fisher's Protected LSD Test and Tukey's Honest Significant Difference Test

(a) Mean of \ln Averaged Over Locations For Each Resin ^{a,b}					
Resin	Avimid-N	AFR700B	VCAP-75	N-CYCAP	PMR-II-50
Mean of \ln	2.278	3.172	3.185	3.198	3.301
(b) Differences of Means ^c					
Avimid-N		0.893 ^{a,b}	0.906 ^{a,b}	0.920 ^{a,b}	1.022 ^{a,b}
AFR700B			0.013	0.026	0.129 ^a
VCAP-75				0.014	0.116 ^a
N-CYCAP					0.103 ^a
PMR-II-50					

^a Break in underscoring indicates an important statistical difference has been determined using Fisher's protected LSD: $\text{LSD}_{0.05} = 0.087$.

^b Break in underscoring indicates an important statistical difference has been determined using Tukey's HSD: $\text{HSD}_{0.05} = 0.188$.

^c After transformation.

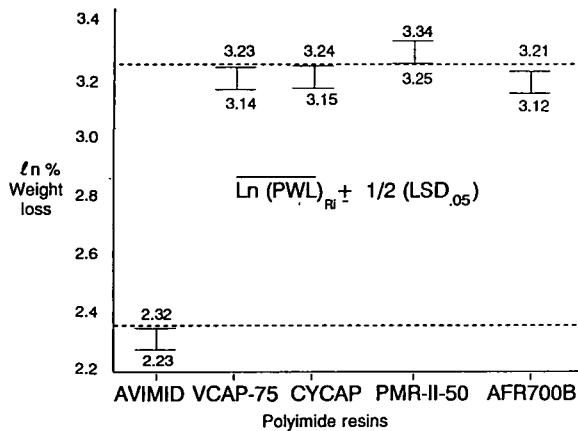


Figure 5 Fisher's protected least significant difference test.

Furthermore, the \ln transformation is monotonic, that implies that if the median of the original variable (y) is, for example, m , the median of the transformed variable ($\ln y$) is $\ln(m)$. Hence, $\delta = \ln(m)$. Back-transforming with the exponential function gives $\exp(\delta) = m$, the median of the untransformed (or original) variable. To continue, if we let $\delta_i = \ln(m_i)$, where δ_i is the mean of the i^{th} lognormally distributed population and m_i is the median of the respective untransformed population, we have

$$\delta_i - \delta_j = \ln(m_i) - \ln(m_j), \quad i \neq j. \quad (9)$$

Thus, using the exponential transformation, we obtain

$$\begin{aligned} \exp(\delta_i - \delta_j) &= \exp[\ln(m_i) - \ln(m_j)] \\ \text{or } \exp(\delta_i - \delta_j) &= m_i/m_j. \end{aligned} \quad (10)$$

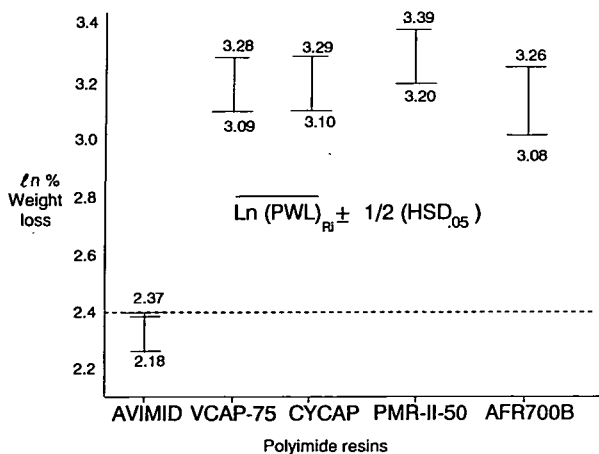


Figure 6 Tukey's honest significant difference test.

Table V HSD 95% Family Confidence Intervals for True Difference in Average Log Percent Weight Losses^a

Polymer Difference ^b	Confidence Interval
5-1	(0.7051, 1.082)
2-1	(0.7179, 1.094)
3-1	(0.7317, 1.108)
4-1	(0.8343, 1.211)
2-5	(-0.1754, 0.2011)
3-5	(-0.1617, 0.2148)
4-5	(-0.0590, 0.3175)
3-2	(-0.1745, 0.2020)
4-2	(-0.0718, 0.3046)
4-3	(-0.0856, 0.2909)

^a $\delta_i - \delta_j$, corresponds to the respective $(\frac{5}{2})_{-10}$ polymer differences.

^b Polymers: Avimid-N = 1, VCAP-75 = 2, N-CYCAP = 3, PMR-II-50 = 4, AFR700B = 5.

Now, m_i/m_j is the ratio of medians from respective populations in terms of the untransformed data. Turning to Table V, we use the exponential transformation of the end points from each of the respective intervals. The resulting confidence intervals estimate the ratio of the corresponding polymer median percent weight losses (with the same group or family confidence level equal to 95%). These intervals are given in Table VI.

There is one final yet important note regarding our findings. Deming⁶⁻⁸ and, more recently, Hahn and Meeker⁹ have detailed the important distinction between an enumerative and an analytic study. Our study is considered analytic. Hence, our results require the critical assumption that molding powders

Table VI HSD 95% Family Confidence Intervals for Ratio of Median Percent Weight Losses^a

Polymer Ratio ^b	Confidence Interval
5/1	(2.024, 2.951)
2/1	(2.050, 2.986)
3/1	(2.079, 3.028)
4/1	(2.303, 3.357)
2/5	(0.8391, 1.223)
3/5	(0.8507, 1.240)
4/5	(0.9427, 1.374)
3/2	(0.8399, 1.224)
4/2	(0.9307, 1.356)
4/3	(0.9180, 1.338)

^a m_i/m_j , corresponding to the respective $(\frac{5}{2})_{-10}$ polymer ratios.

^b Polymers: Avimid-N = 1, VCAP-75 = 2, N-CYCAP = 3, PMR-II-50 = 4, AFR700B = 5.

made in the future vary in the same manner as those selected for this study and that the procedure for making disks varies in the same manner in the future as for this study. This complies with the requirement noted by Hahn and Meeker¹⁰ that "analytic studies require the critical added assumption that the process about which one wishes to make inferences is statistically identical to that from which the sample was selected."

SUMMARY AND CONCLUSIONS

A method was developed to reliably determine if differences exist in the thermal oxidative stability of resins. In addition to quantifying these differences by percent weight loss, the design of the experiment enabled us to correctly adjust for temperature gradients in the oven and the variability that exists within a resin type (i.e., molding powder-molding powder, disk-disk, and within a given disk). Finally, although two methods were used to determine differences in thermal oxidative stability, the only resin that was noticeably different (lowest weight loss) in both analyses was Avimid-N. We believe that this difference can be attributed to two factors. (1) Although all of the other polyimides in this study, which were addition-curing polyimides, have end caps, Avimid-N does not. These end caps have aliphatic carbons that can oxidize and contribute toward thermal oxidative weight loss. (2) Of the five polymers studied, Avimid-N was the only polymer not processed in our laboratories and we have no basis for comparing the processing conditions because those for Avimid-N are proprietary. However, it is well known that processing conditions contribute to the thermal oxidative stability of polymers.¹¹

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