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The role of Mg in the crystallisation of monohydrocalcite.

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ABSTRACT

Monohydrocalcite is a member of the carbonate family which forms in Mg-rich environments at a wide range of Mg/Ca ratios ($Mg^{2+}_{aq}/Ca^{2+}_{aq} \geq 0.17 < 65$). Although found in modern sedimentary deposits and as a product of biomineralization, there is a lack of information about its formation mechanisms and about the role of Mg during its crystallization. In this work we have quantitatively assessed the mechanism of crystallization of monohydrocalcite through in situ synchrotron-based small and wide angle X-ray scattering (SAXS/WAXS) and off-line spectroscopic, microscopic and wet chemical analyses. Monohydrocalcite crystallizes via a 4-stage process beginning with highly supersaturated solutions from which a Mg-bearing, amorphous calcium carbonate (ACC) precursor precipitates. This precursor crystallizes to monohydrocalcite via a nucleation-controlled reaction in stage two, while in stage three it is further aged through Ostwald-ripening at a rate of 1.8 ± 0.1 nm/h$^{1/2}$. In stage four, a secondary Ostwald ripening process (66.3 ± 4.3 nm/h$^{1/2}$) coincides with the release of Mg from the monohydrocalcite structure and the concomitant formation of minor hydromagnesite. Our data reveal that monohydrocalcite can accommodate significant amounts of Mg in its structure (χ_{MgCO3}=0.26) and that its Mg content and dehydration temperature are directly proportional to the saturation index for monohydrocalcite (SI_{MHC}) immediately after mixing the stock solutions. However, its crystallite and particle size are inversely proportional to these parameters. At high

35 supersaturations (SI_{MHC}=3.89) nanometer-sized single crystals of monohydrocalcite form,

36 while at low values (SI_{MHC}=2.43) the process leads to low-angle branching spherulites. Many

37 carbonates produced during biomineralization form at similar conditions to most synthetic

38 monohydrocalcites, and thus we hypothesize that some calcite or aragonite deposits found in

39 the geologic record that have formed at high Mg/Ca ratios could be secondary in origin and

40 may have originally formed via a metastable monohydrocalcite intermediate.

41

42 **Keywords**: monohydrocalcite, carbonates, magnesium, synchrotron, SAXS, WAXS,

43 scattering.

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45

46 **1. INTRODUCTION**

47

48   A variety of calcium carbonate minerals are stable at Earth surface conditions. These

49 include the common polymorphs calcite, vaterite and aragonite, and the less common and

50 hydrated phases monohydrocalcite ($CaCO_3 \cdot H_2O$) and ikaite ($CaCO_3 \cdot 6H_2O$).

51 Monohydrocalcite forms in a variety of modern natural environments including saline spring

52 waters (Ito, 1993), marine polar systems (e.g., Antarctic lakes, or Ikka Fjord, Greenland; Bird

53 et al., 1991; Dahl and Buchardt, 2006), basaltic caves (Broughton, 1972; Onac, 2000;

54 Léveillé et al., 2000), cold/humid mine galleries and lacustrine deposits. Representative of the

55 latter environment is for example, the oldest known monohydrocalcite, 800 ka. (Solotchina et

56 al. (2009), that was found in deep sediments at Lake Hovsgol, NW Mongolia. Other

57 lacustrine examples include Lake Kivu in Africa, or Lake Fellmongery and Lake Butler in S.

58 Australia (Stoffers and Fischbeck, 1974 and Taylor, 1975). Monohydrocalcite is usually

59 found in association with other carbonates like Mg-calcite, aragonite, Ca-rich dolomite

60 ($CaMg(CO_3)_2$) or Mg carbonates like hydromagnesite ($Mg_5(CO_3)_4(OH)_2 \cdot 4H_2O$) and

61 nesquehonite ($MgCO_3 \cdot 3H_2O$) (Fischbeck and Mueller, 1971; Broughton, 1972; Nishiyama et

62 al., 2013). Monohydrocalcite is also formed as a product of biomineralization by certain

63 molluscs (Lowenstam, 1981), flatworms (calcareous corpuscles of Platyhelminthes; Señorale-

64 Pose et al., 2008), vertebrates (otoliths; Carlström, 1963), guinea pigs (bladder stones;

65 Skinner et al., 1977) or is even found as a decay product in Saguaro cacti (Garvie, 2003,

66 2006). It has also been described associated with algae (Taylor, 1975) or halo bacilli

67 (Rivadeneyra et al., 2004). However, despite its occurrence in a wide variety of systems little

68 is known about the mechanism and kinetics of its formation pathway.

69   At Earth surface temperatures and pressures, monohydrocalcite is thermodynamically

70 unstable relative to anhydrous calcite and aragonite. In the presence of a Mg-free aqueous

71 fluid it will slowly (>2 days at 100 °C, several days/weeks at ambient temperature) transform

72 to calcite (Stoffers and Fischbeck 1974; Taylor, 1975). Yet, even minor concentrations of

73 aqueous magnesium will lead to its transformation to aragonite over 25 days at ambient

74 temperatures (Brooks et al., 1950; Kamiya et al., 1977; Dahl and Buchardt, 2006; Munemoto

75 and Fukushi, 2008; Fukushi et al., 2011). This effect is believed to be due to the inhibiting

76 effect of Mg^{2+} on calcite crystallization (Chen et al., 2004; Mucci and Morse, 1983, Davis et

77 al., 2000; Bots et al., 2011). The instability of monohydrocalcite with respect to calcite and

78 aragonite explains the relatively low abundance of this phase within modern environmental

79 systems and the geological record.

80   The mechanism of monohydrocalcite crystallisation in many systems is unknown, but

81 some studies indicate that it can form from an amorphous precursor (Kamiya et al., 1977;

82 Loste et al., 2003; Fukushi et al., 2011; Nishiyama et al., 2013). Such a crystallization

83 pathway has been shown to be common for many Ca-Mg carbonates (e.g., vaterite, aragonite,

84 dolomite; Bots et al., 2012; Rodriguez-Blanco et al., 2012; Sand et al., 2012; Rodriguez-

85 Blanco et al., 2013; Ihli et al., 2012), and other carbonate and phosphate systems (Roncal-

86 Herrero et al., 2009; Roncal-Herrero et al., 2011; Vallina et al., 2013), but this amorphous to

87 crystalline transition is not ubiquitous (Van Driessche et al., 2012). In the carbonate system,

88 these studies have shown that the transformation of the amorphous calcium carbonate (ACC)

89 precursor to stable crystalline phases (i.e., vaterite, dolomite) occurs via a nucleation

90 dominated (spherulitic) growth mechanism. Despite its presence in a variety of natural

91 settings and biominerals, a quantitative evaluation of the kinetics and mechanisms of

92 monohydrocalcite formation from an amorphous precursor is lacking.

93   Monohydrocalcite has a hexagonal structure with space group $P3_112$ and an atomic

94 structure consisting of irregular 8-folded Ca-O polyhedra, with a central Ca^{2+} ion surrounded

95 by carbonate groups and water molecules (Effenberger, 1981; Neumann and Epple, 2007;

96 Swainson, 2008). The presence of the water makes its structure more open and less dense

97 compared to anhydrous $CaCO_3$ (e.g., calcite or aragonite). Interestingly, monohydrocalcite is

98 described as a pure 'calcium' carbonate phase in mineralogical databases, but a number of

99 studies have shown that Mg is ubiquitous in all synthetic and natural monohydrocalcite

100 samples (Mg/Ca= ~ 0.01-0.53; Hull and Turnbull, 1973; Taylor, 1975; Neumann and Epple,

2007; Nebel et al., 2008; Fukushi et al., 2011; Nishiyama et al., 2013). The Mg content of monohydrocalcite is likely to be linked to the overall fluid chemistry and in particular to the Mg contents of the fluid in which it forms. In both natural and most synthetic monohydrocalcites the presence of high concentrations of Mg relative to Ca in the precipitating solution ($Mg^{2+}_{aq}/Ca^{2+}_{aq} \geq 0.17 < 65$) is a prerequisite for its formation (e.g., Munemoto and Fukushi, 2008; Neumann and Epple, 2007; Last et al., 2010; Kimura and Koga, 2011). Recently, Nishiyama et al (2013) have also shown that the crystallinity and particle size of synthetic monohydrocalcite decrease with Mg content. This clearly shows that Mg is a key component in monohydrocalcite, and must be present in the fluids in which it crystallizes. However, the exact mechanisms by which Mg controls the formation and stability of monohydrocalcite are unknown.

This study aims to determine the mechanism of monohydrocalcite crystallisation in solution via an ACC precursor. We evaluate the role of Mg during this process and test the hypothesis that a nucleation-dominated process controls the formation of monohydrocalcite from the Mg-containing precursor ACC. This has been done through a series of experiments where the nucleation and crystallization of monohydrocalcite from Mg doped-ACC was followed in situ and in real time with synchrotron-based X-ray scattering combined with microscopic and wet chemical characterization of the solids and solutions throughout the reaction. Our results demonstrate that the formation of monohydrocalcite takes place via a 4-stage process that starts with the precipitation of an Mg-rich ACC precursor. This precursor crystallizes in stage two to monohydrocalcite through a nucleation-controlled dissolution-reprecipitation reaction. In stage three Ostwald-ripening dominates, while in stage four, a secondary monohydrocalcite crystallization, also through Ostwald ripening, occurs concomitantly with the loss of some of its structural Mg. We also show that, depending on the aqueous Mg/Ca ratio and supersaturation, monohydrocalcite can accommodate large amounts of Mg in its structure (up to ~25% Ca replaced by Mg) and that the aqueous chemistry of the starting solution controls the crystallite size, unit-cell dimensions and particle size/shape of the forming monohydrocalcite. Based on these findings we discuss the implications of this monohydrocalcite crystallization pathway, on its occurrence in various natural settings and as a biomineral.

2. EXPERIMENTAL

135   Experiments were carried out at 21 °C by rapid addition (under constant and vigorous

136   stirring) of a CaCl$_2$/MgCl$_2$ solution (700 mM CaCl$_2$; 300 mM MgCl$_2$; Ca/Mg ratio of 7/3) to

137   a 1000mM Na$_2$CO$_3$ solution to achieve a (Ca+Mg)/CO$_3$ ratio of 1/1. In all cases immediately

138   upon mixing a white gel precipitated. The crystallization of this white gel was followed on-

139   line using in situ and time resolved small and wide angle X-ray Scattering (SAXS/WAXS) on

140   beamline I22 (Diamond Light Source Ltd., UK). Solutions were mixed using a stopped-flow

141   apparatus (Bio-Logic, Paris, France) and the resulting white suspensions were continuously

142   circulated with a peristaltic pump through a capillary cell in line with the synchrotron beam.

143   Simultaneous SAXS (RAPID detector; Marchal et al., 2009) and WAXS (HOTWAXS

144   detector; Bateman et al., 2007) patterns were collected over 12 hours at 1 min/frame. The

145   time resolved scattering patterns were detector-response corrected, and background

146   subtracted using a scattering pattern from the starting Na$_2$CO$_3$ solution. Individual WAXS

147   patterns were fit using XFIT (Cheary and Coelho, 1992) and the areas under the Bragg peaks

148   normalized to values from 0 to 1 to express the degree of reactions (α) as a function of time

149   (Rodriguez-Blanco et al., 2011). Crystallite size was calculated from the Bragg peak full-

150   width half-maximum (FWHM) using the Scherrer equation (Scherrer, 1918) and the unit cell

151   parameters were determined with TOPAS (Coelho, 2006), using the Swainson (2008) model

152   for the monohydrocalcite structure.

153   The variations in the scattering peak position in the SAXS patterns were used to

154   derive the changes in the particle sizes of the solids throughout the experiments via the

155   expression:

156

157   d = 2π/q   [1]

158

159   where d (nm) is particle diameter and q (nm^{-1}) is the scattering vector (de Moor et al, 1999;

160   de Moor et al, 1999b). These particle size results were also cross-validated by whole SAXS

161   pattern fitting using GNOM (Svergun, 1992).

162   Experiments equivalent to the online SAXS/WAXS experiments (i.e., 1000mM

163   Na$_2$CO$_3$ solution mixed with a 700 mM CaCl$_2$ and 300 mM MgCl$_2$ solution) were performed

164   to follow the reaction process via time-dependent solid characterization and solution

165   analyses. At specific time steps aliquots of the reacting suspension were removed and

166   immediately vacuum filtered (0.2 μm polycarbonate membranes). The resulting solids were

167   washed with water and isopropanol following Rodriguez-Blanco et al (2008). An aliquot of

168   each solid sample was dissolved in 0.1 M HCl and analysed together with the corresponding

169   solution samples for aqueous Ca and Mg concentrations ($[Ca^{2+}_{aq}]$ and $[Mg^{2+}_{aq}]$) using ion

170   chromatography (Dionex LC 220) following Bots et al (2011). Throughout all experiments

171   the pH was recorded at a 10 second time resolution using an Orion pH meter and WinWedge

172   v3.4.1 software (TAL technologies) with a precision of 0.001. Finally, the total carbonate

173   concentration in solution was determine by assuming that all Ca and Mg precipitated formed

174   Ca/Mg carbonate, therefore the moles of Ca + Mg removed form solution must equal the total

175   moles of carbonate removed. The total carbonate was then calculated from the difference

176   between the initial concentration and the amount removed at each time point. From these

177   solution data the saturation indexes (SI) with respect to different Ca and Mg-bearing

178   carbonates were calculated using PHREEQC (Parkhurst, 1995) and are presented in Table 1.

179   This was done using the Pitzer activity coefficient models (Pitzer, 1979) and two solubility

180   products, $K_{sp(MHC)}$, for monohydrocalcite ($10^{-7.60}$; Hull and Turnbull, 1973) and ($10^{-7.05}$; Krajl

181   and Brečević, 1995). The saturation index of monohydrocalcite is defined as:

182    $$SI_{MHC} = \log \frac{a_{Ca^{2+}} \times a_{CO_3^2}}{K_{sp(MHC)}}$$ [2]

183

184   To determine the effect of initial supersaturation (i.e. saturation index of the aqueous

185   solution for monohydrocalcite immediately after mixing the stock solutions) on the

186   crystallization reaction, experiments were also performed following the same procedure but

187   starting at different initial supersaturations with respect to monohydrocalcite, yet keeping a

188   1/1 (Ca+Mg)/CO$_3$ ratio (Table 2) in all cases. Using the $[Ca^{2+}_{aq}]$ and $[Mg^{2+}_{aq}]$ concentrations

189   obtained from the analyses of the dissolved solids, the molar fraction of MgCO$_3$ in each solid

190   sample, X_{MgCO_3}, was calculated following:

191    $$_{MgCO_3} = \frac{M_{MgCO_3}}{M_{MgCO_3} + M_{CaCO_3}}$$ [3]

192

193   where M_{MgCO3} and M_{CaCO3} correspond to the molar concentration of MgCO$_3$ and CaCO$_3$,

194   respectively.

195

196   The mineralogy of the solids from off-line experiments was characterized by powder

197   X-ray diffraction (PXRD) using a Bruker D8 X-ray Diffractometer (CuKα1, 5-90° two theta,

198 0.001°/step; 0.1 or 1 sec/step). The PXRD patterns were also used to derive the crystallite

199 size using the Scherrer equation (Scherrer, 1918). All samples were imaged by high-

200 resolution transmission electron microscopy (HR-TEM; Philips CM200 field emission gun

201 TEM equipped with a Gatan Imaging Filter, GIF-200 and a Gatan camera for selected area

202 electron diffraction, SAED, pattern collection) or scanning electron microscopy (SEM; LEO

203 1530 Gemini). Finally, the water content of the samples was determined using

204 thermogravimetric analyses (TGA, Mettler Toledo, 25-800 °C in air, 10°C/min). The total

205 water content for the initial white precipitate was calculated from the difference in weight

206 between 25 and 550°C in accordance with the method of Radha et al. (2012) for determining

207 the water content of ACC. For crystalline samples the difference in weight between 100 and

208 550°C was used, in accordance with the method of Hull and Turnbull (1973) for determining

209 the water content of monohydrocalcite.

210

211

212 **3. RESULTS AND DISCUSSION**

213

214 Our data revealed that over the 12 hours of our experiments monohydrocalcite formed

215 through a multi-stage crystallisation pathway. The first stage of the reaction was

216 characterized by the formation of a Mg-bearing amorphous calcium carbonate phase. In stage

217 two, this amorphous phase crystallised to nanoparticulate monohydrocalcite. The rate of

218 crystallization slowed considerably during stage three, coincident with only minor changes

219 in the solution composition (e.g., pH). Finally, in the fourth stage of the reaction secondary

220 growth of the nanoparticulate monohydrocalcite was observed. This was accompanied by the

221 simultaneous decrease in $[Mg^{2+}_{aq}]$ and the formation of minor amounts of hydromagnesite

222 $(Mg_5(CO_3)_4(OH)_2 \cdot 4H_2O)$. Each of these four stages is described in detail below:

223 In **Stage I** of the reaction a white gel precipitated immediately upon solution mixing.

224 The PXRD patterns of this gel (Fig. 1a, lower pattern) showed no Bragg peaks and only three

225 humps located at 2θ (λ=1.54 Å) 17, 31 and 45°, indicating the solid is poorly-ordered. These

226 three broad humps (asterisks in Fig. 1a), are located at the same positions as those observed

227 for Mg-free amorphous calcium carbonate (ACC; Rodriguez-Blanco et al., 2008). Based on

228 the chemical analyses of this amorphous phase (Table 1, 0.83 hour sample) and the TGA

229 results, the formula for this compound was determined to be $Ca_{0.74}Mg_{0.26}CO_3 \cdot 1.18H_2O$. In

230 line with previous studies where the incorporation of Mg into ACC has been quantified

231 (Nebel and Epple, 2008; Wang et al., 2009; Rodriguez-Blanco et al., 2012), we will term this
232 poorly-ordered phase, Mg-ACC, following the notation of Günther et al (2005). Throughout
233 this first stage, Mg-ACC was the only solid phase present. In both the on-line and off-line
234 experiments and regardless of conditions, the Mg-ACC started to crystallize after
235 approximately 50 minutes to 1 hour, initiating **Stage II**. During this stage the background
236 intensity in the PXRD patterns decreased with time, and Bragg peaks for monohydrocalcite
237 formed, grew and sharpened (Fig. 1a). The on-line time resolved WAXS data (Fig. 1b) shows
238 the simultaneous growth of the monohydrocalcite (110), (111), (300) and (301) peaks, after ~
239 1 hour. The whole reaction is clearly illustrated when the change in area for a specific Bragg
240 peak, expressed as the degree of reactions (α), is plotted as a function of time (Fig. 2). Stage
241 II of the reaction lasted for about 1 hour and was characterized by the rapid increase in
242 monohydrocalcite peak area. After the fast growth in stage II, a much slower growth phase,
243 between ~ 2 and 8 hours followed (**stage III**). During this stage, only minor changes in Bragg
244 peak intensities / areas were observed (Fig. 1b and 2). The final stage of the reaction -**stage
245 IV**- started after about 8.3 hours. During this stage a secondary growth in the
246 monohydrocalcite Bragg peak areas was observed. This growth phase lasted about 2-3 hours
247 and the peak areas reaching a stable maximum after ~ 11 hours. This same evolution was
248 observed for all other monohydrocalcite Bragg peaks. An PXRD examination of off-line
249 experimental solids sampled during stage IV revealed the presence of a second, but minor
250 phase of hydromagnesite (ICDD PDF 01-070-1177; Fig. S1). Quantitative X-ray refinement
251 with TOPAS (Coelho, 2006) determined that its abundance was ~4.5%. The on-line time
252 resolved WAXS patterns (Fig. 3) also revealed small hydromagnesite (110) and (011) Bragg
253 peaks that began to grow at the onset of stage IV, in parallel with the second stage of
254 monohydrocalcite crystallization.

255   Analysis of the SAXS data (Fig. 4) indicates that the Mg-ACC that formed in stage I
256 consisted of particles with a diameter of between 40 and 60 nm in size, a size that remained
257 constant throughout this first stage. This is consistent with particle sizes derived from the
258 TEM imaging of the Mg-ACC particles, which indicted an average diameter of ~ 40 nm
259 (n=100; Fig. 5a; Table 1). However, the Mg-ACC particle diameters varied between 15 and
260 210 nm revealing a high degree of polydispersity. The corresponding SAED patterns revealed
261 only diffuse rings for the Mg-ACC, confirming its poorly-ordered nature (Fig. 5d). During
262 stage II, a distinct scattering peak appeared in the SAXS patterns (arrows in Fig. 4a)
263 indicating that the crystals formed in the experiment were relatively monodispersed. The
264 position of the scattering peak in the time resolved SAXS patterns moved towards lower q

with time indicting particle growth (Fig. 4a). From the position of this peak and using equation [1] the particle size of the monohydrocalcite crystals was determined (Fig. 4b). At the beginning of stage II, when the first Bragg peaks for monohydrocalcite appeared in the WAXS data (Fig. 1b and Fig. 2), the corresponding SAXS data revealed an average diameter of ~9 nm for the newly forming monohydrocalcite crystals. During the rest of stages II and III these monohydrocalcite nanocrystals grew only slightly and very slowly reaching a maximum diameter of ~15 nm after 8 hours. This same trend in average particles sizes was confirmed (open symbols in Fig. 4b) through selected full pattern evaluation with GNOM (Svergun, 1992). TEM microphotographs of the monohydrocalcite that formed in stage II and persisted throughout stage III (Fig. 5b) revealed that the monohydrocalcite consisted of nanocrystals, which exhibited sub-spherical to slightly elongated morphologies. Their average dimensions were 28±7 (L) x 20±5 (W) nm, with the minimum size being around 14x10 nm (n=100). The corresponding SAED pattern showed diffraction rings with only poorly developed spots (Fig. 5e), evidencing the presence of very small but crystalline nanocrystals. At the onset of stage IV, and simultaneously with the secondary growth in Bragg peak areas for monohydrocalcite, the SAXS data also revealed a steady increase in the monohydrocalcite nanocrystal diameters, which reached a maximum of ~45 nm at the end of the reaction. Photomicrographs of a sample obtained after 10 hr (stage IV; Fig. 5c) showed euhedral or subhedral single monohydrocalcite crystals with sizes of 77±20 (L) x 47±10 (W) nm. Again large variations in dimensions were observed, with the largest and smallest crystals being 155x80 and 40x30 nm (n=100). SAED pattern from these crystals exhibited clearly defined spots within the diffraction rings (Fig. 5f). HR-TEM images of the sample taken from the end of stage IV (Fig. 5c, white arrow) revealed the presence of hydromagnesite as a minor phase consisting of ~100x5 nm platy crystals, confirming the WAXS and PXRD observations (Fig. 3 and Fig. S1).

Evaluating the corresponding WAXS patterns for the Scherrer crystallite size (Fig. 2) showed a reasonable agreement with the particle size determined from the SAXS and TEM data, and also confirmed that the forming monohydrocalcite crystals were anisotropic. For example, based on the Bragg peak (110) the data reveals that the monohydrocalcite crystals formed during stage II had a crystallite size of ~15 nm, increasing to ~ 20 nm during stage III, which is in agreement with the particle sizes derived from the SAXS and TEM data. Once stage IV of the crystallization was initiated, the monohydrocalcite nanocrystallite size steadily increased to > 80 nm, which is almost double the average particle size from SAXS,

but closer to that derived from the TEM data. However, although this general trend was the same in all other Bragg peaks, the crystallite sizes in all stages were dramatically different between the different Bragg peaks. For example, the final crystallite size from the (141) peak at the end of stage 3 was 16 nm, half that of the (110) peak, and at the end of the reaction it only reached a value of 28 nm (Fig. 2), almost three times smaller than the final crystallite size from the (110) peak, indicating a clear anisotropic growth of the monohydrocalcite nanocrystals. This is consistent with the elongated particle morphology observed by TEM.

Analysis of the WAXS data also revealed changes in unit cell dimensions during the formation of monohydrocalcite (Fig. 6). Once formed, the unit cell volume (721.6 Å^3) remained virtually constant during stages II-III with a slight decrease during stage IV ($\Delta V \approx -0.2$ Å^3; Fig. 6a). However, more substantive changes were observed in the a and c axis dimensions (Fig. 6b and c, respectively). Their evolution mirrored each other, with the a-axis decreasing and the c-axis increasing in length ($\Delta c = -\Delta a \approx 0.02$ Å). These changes were small during stages II and III, but became more significant during the secondary monohydrocalcite crystallization (stage IV).

The changes in solution and solid phase chemistry during the formation and crystallisation of monohydrocalcite showed a clear link between the evolution of pH, $[Ca^{2+}_{aq}]$, $[Mg^{2+}_{aq}]$ and molar fraction of Mg in the solids (Fig. 7) over the four stage of the reaction. The reaction started with a pH of 11.60 (the pH of the initial Na_2CO_3 solution), which upon mixing with the Ca/Mg solution rapidly dropped to 9.17. During stage I the pH increased slightly reaching a value of 9.24, while the formation of monohydrocalcite in stage II did not change the pH. In stage III only a small decrease in pH to a value of 9.16 was evident. However, at the end of stage III and during stage IV the pH dropped dramatically to 8.65. The $[Ca^{2+}_{aq}]$ was 1.25 mM during stage I and decreased to ≈ 0.6 mM by stage III. It then decreased further reaching a concentration < 0.1 mM at the end of stage IV (Fig. 7). $[Mg^{2+}_{aq}]$ was 28 mM during stages I and II, but increased by ~ 20 % to 35 mM during stage III. During stage IV it decreased by 40% to 13 mM. These changes in $[Mg^{2+}_{aq}]$ were mirrored throughout the whole reaction by the change in χ_{MgCO3} in the solids (Fig. 7). Furthermore, the timing of these changes in aqueous composition were all coupled with the increase in monohydrocalcite particle size (Fig. 4 and 5) and formation of hydromagnesite (Fig. 3).

From the χ_{MgCO3} values and the TGA results, the formula of the stage II monohydrocalcite was derived to be $Ca_{0.74}Mg_{0.26}CO_3 \cdot 0.99H_2O$. However, although the total χ_{MgCO3} increased during stage IV (Fig. 7), this does not represent the Mg in monohydrocalcite

but primarily the contribution from the minor (~ 4.5%) hydromagnesite ($Mg_5(CO_3)_4(OH)_2$ $4H_2O$). Using the quantitative XRD results and the composition of the final solid obtained from chemical analysis (χ_{MgCO3} = 0.26, Table 1) we calculated that the molar fraction of Mg in monohydrocalcite at the end of stage IV was only ~ 0.065 moles, which is a 75% decrease compared to the Mg content of the stage II monohydrocalcite (0.26 moles). The remaining Mg was transferred into the newly formed hydromagnesite.

One last line of evidence that supports the observations above is the evolution of the saturation states during the reaction. PHREEQC modelling of the solution during the reaction showed that immediately after the precipitation of Mg-ACC upon mixing, the aqueous solution was supersaturated with respect to both monohydrocalcite and hydromagnesite (Fig. 8) and that the SI for both phases varied little during stages I to III ($SI_{MHC} \approx 0.5$; $SI_{HMgS} \approx 6$). The biggest change in both SI occurred at the onset of stage IV, where SI_{MHC} became undersaturated ($\approx$ -0.85) while SI_{HMgS} dropped to ≈ 0.2. This change in SI was concomitant with the removal of Mg from solution and the associated drop in pH, due to the formation of hydromagnesite. There are however, two factors that need to be taken into account when SI_{MHC} is calculated. Firstly, two solubility products for monohydrocalcite are available in the literature: $10^{-7.60}$ (Hull and Turnbull, 1973), and $10^{-7.05}$ (Krajl and Brečević, 1995) and this results in a ~0.55 difference in the calculated SI_{MHC} (Fig. 8). Secondly, the total ionic strength in our experiments was high ($\leq$1 M) and thus even an error of 1% in the ionic strength of the starting solution could also result in variations up to ~0.5 units in the calculated SI_{MHC}.
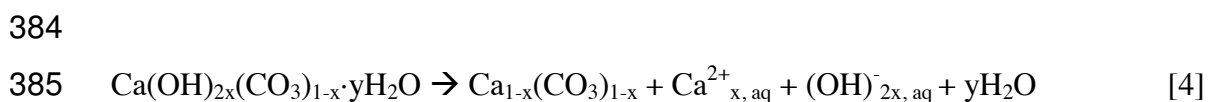

3.1. Stability of the precursor Mg-ACC and the mechanism of monohydrocalcite nucleation and growth

The formation of monohydrocalcite proceeded, in all our experiments, through a poorly-ordered Mg-ACC precursor, which precipitated from solutions with an initial $[Ca^{2+}_{aq}]$:$[Mg^{2+}_{aq}]$ of 7:3. The resulting phase was considerably more stable (crystallization started at ~50 minutes) compared to pure ACC (no Mg; crystallization started <2 minutes; Ogino et al., 1987; Rodriguez-Blanco et al., 2011; Rodriguez-Blanco et al., 2012, Bots et al., 2012), or Mg-ACC formed in a solution with $[Ca^{2+}_{aq}]$:$[Mg^{2+}_{aq}]$ of 9 to 1 (crystallized to calcite within ~ 14 minutes; Rodriguez-Blanco et al., 2012). However, the Mg-ACC in the current study was less stable than Mg-ACC produced from solutions where 50% of the

calcium was replaced with magnesium (i.e., $[Ca^{2+}_{aq}]:[Mg^{2+}_{aq}] = 1:1$), which did not crystallize at ambient temperatures even after longer reaction times (>1 day; Rodriguez-Blanco et al., 2013). The increasing stability of ACC with increasing Mg contents is not unexpected, and confirms both abiotic (Rodriguez-Blanco et al., 2011; Rodriguez-Blanco et al., 2013; Ajikumar et al., 2005) and biotic (Politi et al,. 2010; Loste et al., 2003; Raz et al., 2003) data that demonstrate the importance of Mg in stabilizing amorphous precursors and delaying crystallization. The most likely cause of these effects is the high dehydration energy of the Mg-aquo ion compared to the Ca ion (di Tommaso and de Leeuw, 2010). Hydrated and poorly-ordered ACC is less thermodynamically stable than the dehydrated and more-ordered ACC. This dehydration and local ordering in the amorphous precursor precedes its crystallization (Radha et al., 2010; Bots et al., 2012). Hydrated Mg located within the nanoporous structure of ACC (Goodwin et al., 2010) would retard its dehydration and breakdown, slowing its transformation to crystalline phases.

In stage I and prior to the formation of monohydrocalcite, the pH increased by a small amount (0.07 units) (Fig. 7), which likely corresponds to the release of OH$^-$ due to the onset of Mg-ACC dissolution. Kojima et al (1993) showed that ACC incorporates minor amounts of OH$^-$ ions into its structure when it forms at a basic starting pH. The Mg-ACC in our on-line experiment started forming at a pH of 11.5 (the pH of the carbonate starting solution; Fig. 7) and thus minor OH$^-$ in our Mg-ACC is not unexpected. Its dissolution would release OH$^-$ ions, thus explaining the slight pH increase following:

$$Ca(OH)_{2x}(CO_3)_{1-x}\cdot yH_2O \rightarrow Ca_{1-x}(CO_3)_{1-x} + Ca^{2+}_{x,\,aq} + (OH)^-_{2x,\,aq} + yH_2O \qquad [4]$$

The first monohydrocalcite crystals formed during stage II were significantly smaller than the initial Mg-ACC nanoparticles (Fig. 4b), yet once monohydrocalcite started forming, its diameter remained virtually constant throughout stage II and III. Analysis of the WAXS data shows that the amount of monohydrocalcite crystallising increased at a constant rate throughout stage II of the reaction (Fig. 2). However, this increase was not due to particle growth (Fig. 4b) and we suggest that the formation of monohydrocalcite during stage II proceeds via the nucleation of new crystalline solids, i.e., via a nucleation-controlled reaction. The constant rate of monohydrocalcite crystallization with time (i.e., linear increase in the amount of monohydrocalcite) during most of stage II indicates a zeroth order reaction, and a constant rate of nucleation. Furthermore, the pH remained virtually constant during stage II,

397   indicating a constant dissolution of Mg-ACC coupled with the constant nucleation of

398   monohydrocalcite. Taking all this into account, we propose that the transformation form Mg-

399   ACC to monohydrocalcite occurs via a simultaneous dissolution of Mg-ACC and nucleation

400   of monohydrocalcite at a constant rate. Nucleation controlled growth has been shown for

401   several other crystalline carbonate phases formed from amorphous Ca-Mg carbonates (e.g.,

402   vaterite and proto-dolomite; Bots et al., 2012; Rodriguez-Blanco et al., 2013). These studies

403   suggested that the large difference in solubility between the amorphous precursor and the

404   crystalline phases as well as the high supersaturations with respect to all crystalline phases,

405   promotes nucleation. This nucleation process will continue until all of the amorphous

406   precursor has been consumed, leading to a constant nucleation rate.

407   Stage IV of the reaction is key to understanding the effect of Mg on the formation of

408   monohydrocalcite. The SAXS data (Fig. 4b) shows a dramatic increase in particle size.

409   During stage IV, this increase can be fitted to a straight line when plotted as a function of $t^{1/2}$,

410   indicating a surface-controlled Ostwald-ripening mechanism (Wagner, 1961; Tobler et al.,

411   2009). TEM images and both WAXS and PXRD data confirm that larger and better

412   developed monohydrocalcite crystals formed during this stage of the reaction, supporting the

413   Ostwald-ripening mechanism. The photomicrographs of monohydrocalcites from stages III

414   and IV (Fig. 5 b and c) show an unambiguous increase in the nanocrystal sizes and a

415   corresponding change in the SAED patterns from rings with faint spots to discrete spots,

416   indicting an increase in particle size.

417   Ostwald ripening usually occurs during the later stages of crystallisation reactions and

418   involves particle growth without the formation of new material (Wagner, 1961). All

419   nanoparticles formed in stages III are undoubtedly crystalline, but some of them are too small

420   (<10 nm) to produce a significant amount of diffraction. Therefore scattering from these

421   crystal is observed in the background of the PXRD/WAXS patterns during stages II and III

422   (e.g. Fig. 1 after 1 hour of reaction). This behaviour is similar to that observed for ultra-small

423   iron oxides crystallites which are too small to diffract singly (Machala et al., 2007; Ahmed et

424   al., 2010). The intensity of the background in the XRD/WAXS patterns decreased during

425   stages III and IV, while the Bragg peaks increased in intensity and became sharper. These

426   changes in background and Bragg peak intensity are not due to the formation of more

427   monohydrocalcite, but reflect the growth of the monohydrocalcite nanoparticles to a size

428   where they diffract strongly, due to the Ostwald ripening process. The linear fits to the

429   particle growth data with the surface-controlled growth model Fig. 4b show different rates

430   (slopes) for the Ostwald-ripening process during stages III (slow; 1.8 ± 0.1 nm/h$^{1/2}$) and IV

431   (fast; 66.3 ± 4.3 nm/h$^{1/2}$).

432   The rapid change in monohydrocalcite ripening rate is coincident with the formation

433   of hydromagnesite; its formation lead to the fast removal of Mg from solution and triggered

434   the rapid increase in monohydrocalcite growth rate. We suggest that structural Mg from

435   monohydrocalcite during the ripening process was released into the solution leading to the

436   observed increase in $[Mg^{2+}_{aq}]$ during stage III (Fig. 7). This ultimately drove the

437   monohydrocalcite composition toward a pure $CaCO_3 \cdot H_2O$ end-member. The high

438   concentration of Mg in solution during stage II suppressed the ripening rate, and produced a

439   feedback effect, which decreased the rate of ripening even more (decrease in the slope of the

440   degree of reaction plot during stage III; Fig. 2). The presence of Mg in solution is known to

441   suppress the dissolution and growth of calcium carbonates (e.g., calcite; Chen et al., 2004;

442   Mucci and Morse, 1983; Davis et al., 2000). Thus, is not unexpected that the high

443   concentration of $\{Mg^{2+}_{aq}]$ present during stage III significantly reduced the dissolution and

444   reprecipitation process during the Ostwald ripening. Although the aqueous solution was

445   supersaturated with respect to hydromagnesite throughout the reaction, this progressive

446   increase in $[Mg^{2+}_{aq}]$ eventually triggered the nucleation of hydromagnesite at the end of stage

447   III (Fig. 3). Its formation removed $[Mg^{2+}_{aq}]$ from solution (Fig. 7) and induced a dramatic

448   drop in pH following:

449

450    $4Mg^{2+} + 3HCO_3^- + 6H_2O \rightarrow Mg_4(CO_3)_3(OH)_2 \cdot 4H_2O + 5H^+$ [5]

451   (hydromagnesite)
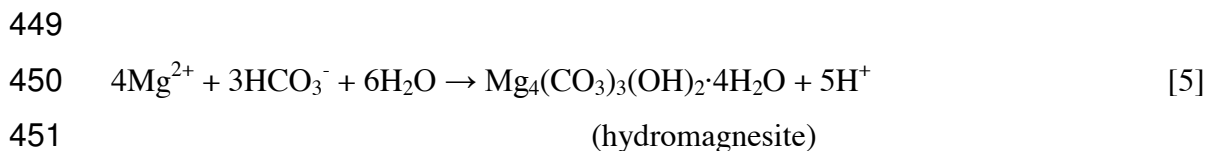
452

453   The main consequence of this Mg removal from the aqueous solution was an acceleration of

454   the monohydrocalcite ripening reaction, which was translated into a rapid growth in particle

455   size and the formation of highly crystalline low-Mg monohydrocalcite.

456   PHREEQC calculations (Table 1) also revealed that the aqueous solution was in

457   equilibrium or slightly supersaturated with respect to nesquehonite ($Mg(HCO_3)(OH) \cdot 2(H_2O)$)

458   during stages II and II, but became undersaturated in this mineral at stage IV. Nesquehonite

459   was never detected using conventional XRD, synchrotron-based WAXS or HR-TEM

460   imaging. However, Nishiyama et al (2013) suggested that the formation of monohydrocalcite

461   would require the paragenesis of a hydrous Mg-bearing carbonate e.g., nesquehonite, that

462   would transform to hydromagnesite after longer reaction times. Despite the lack of

463   experimental evidences, we can not completely discard the possibility that small amounts of

464 nanocrystalline nesquehonite were present during stages II and III, and that these eventually
465 transformed to hydromagnesite during stage IV.

466   The inhibiting effect of Mg has been described previously for other Ca-Mg carbonates
467 (Bischoff, 1968; Berner, 1975; Reddy and Wang, 1980; Mucci and Morse, 1983; Davis et al.,
468 2000). For example, Zhang and Dawe (2000) have suggested that the calcite growth rate is
469 inversely proportional to the Mg concentration in solution. This effect has been attributed to
470 the stronger hydration shell of Mg in comparison to Ca (di Tommaso and de Leeuw, 2010;
471 Moomaw and Maguire, 2008). The higher energy, which Mg needs to dehydrate before
472 incorporating into a carbonate structure controls the kinetics of crystal growth (Mucci and
473 Morse, 1983; Nancollas and Purdie, 1964; De Boer, 1977). Therefore, monohydrocalcite
474 growth would be favoured at lower Mg concentrations.

475

476 **3.2. Monohydrocalcite chemical and structural variability**

477

478   The molar fraction of Mg in monohydrocalcite (χ_{MgCO3}) was ~0.25 during stages II
479 and III (i.e., before the formation of hydromagnesite). This is close to the maximum values in
480 the literature for natural or synthetic monohydrocalcites (χ_{MgCO3}= ~0.01-0.34; Brooks et al,
481 1950; Sapozhnikov and Tsvetkov, 1959; Marschner, 1969; Hull and Turnbull, 1973; Taylor,
482 1975; Skinner, 1977; Neumann and Epple, 2007; Nebel et al., 2008; Munemoto and Fukushi,
483 2008; Nishiyama et al., 2013). Many of these studies do not include information about the
484 exact conditions at which monohydrocalcite formed (temperature, solution composition,
485 synthesis method, etc.), therefore the factors controlling χ_{MgCO3} in monohydrocalcite are
486 difficult to evaluate. However, we suggest that the high level of Mg in the monohydrocalcite
487 crystallised from Mg-ACC in this study (Fig. 7) is likely due to the high supersaturation of
488 the initial solution with respect to all carbonate phases.

489   To test the effect of supersaturation on the incorporation of Mg into the resulting
490 monohydrocalcite, a set of batch experiments were performed at different initial
491 supersaturations (Table 2). Regardless of supersaturation, immediately after mixing the initial
492 solutions an amorphous precursor formed. Experiments with initial SI_{MHC}>2.43 resulted in
493 the crystallization of monohydrocalcite, while only Mg-calcite was obtained at lower
494 supersaturations (Table 2 and Fig. S2). No other Ca/Mg-bearing phases were observed.
495 PXRD analyses of the monohydrocalcite revealed that its crystallite size was inversely
496 proportional to the starting supersaturation (Table 2), varying between 35 nm at SI_{MHC} = 2.43
497 to 16 nm at SI_{MHC} = 3.89. Furthermore, the χ_{MgCO3} of the monohydrocalcite increased with

498   initial supersaturation, from 0.017 (SI_{MHC} = 2.43) to 0.164 (SI_{MHC} = 3.89, Fig. 9a). This data

499   is consistent with previously published values for χ_{MgCO3} of synthetic monohydrocalcite

500   produced using different initial Mg content in solution and different supersaturations (Fig. 9a;

501   Neumann and Epple (2007); Munemoto and Fukushi (2008), however the original χ_{MgCO3} in

502   the later reference may be higher than reported due to sample treatment). Recently,

503   Nishiyama et al. (2013) synthesized a series of monohydrocalcites using highly variable

504   starting $[Mg^{2+}_{aq}]/[Ca^{2+}_{aq}]/[CO_{3,aq}]$ ratios and most often an excess of carbonate, which lead to

505   a much higher crystallization pH (9.8-11.4) compared to ours (< 9.25). Their data shows that

506   the χ_{MgCO3} in monohydrocalcite is proportional to their starting $[Mg^{2+}_{aq}]/[Ca^{2+}_{aq}]$ ratio, but

507   these is no clear link between supersaturation and Mg contents (χ_{MgCO3}) (Fig. 9a). In addition,

508   Nishiyama et al. (2013) used a different solution mixing approach (addition of the Na_2CO_3

509   solution to the $CaCl_2/MgCl_2$ solution) compared to ours (addition of the $CaCl_2/MgCl_2$

510   solution to the Na_2CO_3 solution). In an earlier study (Rodriguez-Blanco et al., 2012) we

511   showed that, in the pure carbonate system, different mixing approaches lead to different

512   initial synthesis pH, which in turn dramatically affected the crystallization pathway. Taken as

513   a whole, both the initial $[Mg^{2+}_{aq}]/[Ca^{2+}_{aq}]/[CO_{3,aq}]$ ratios (Nishiyama et al., 2013) and initial

514   supersaturation (this study) are key factors in controlling the Mg content of monohydrocalcite

515   (Fig. 9a). This is also consistent with Loste et al. (2003) who showed that in other carbonate

516   systems, precipitation at high supersaturation levels is so rapid that it results in less

517   distinction between Ca and Mg ions. Therefore we infer that the Mg content of the solid is

518   proportional to the initial supersaturation level and the Mg content in the initial amorphous

519   precursor phase.

520   TGA analysis of the monohydrocalcite samples showed that the H_2O content (0.93 –

521   1.03 per formula unit) was similar in all solids and proportional to initial supersaturation.

522   However, the release of structural water during heating was different depending on the molar

523   fraction of Mg (χ_{MgCO3}) (Fig. 9b). In samples with χ_{MgCO3}<0.06 all water was released at low

524   temperature (150-200°C), while in the Mg-rich monohydrocalcite samples (χ_{MgCO3}>0.06) the

525   retention of water was greater, and it was progressively released during heating to T>450 °C.

526   The final weight loss starting at 550 °C was due to carbonate decomposition. The retention of

527   water to higher temperatures in samples with χ_{MgCO3}>0.06 is related to the greater

528   dehydration energy (~20% higher) of Mg^{2+} in comparison to Ca^{2+} (Nancollas and Purdie,

529   1964; Lippmann, 1973; De Boer, 1977). Ca^{2+} and Mg^{2+} ion dehydration temperature ranges

530   have been defined from 100 - 300 °C, and from 300 - 550 °C respectively, as the release of

531   water associated with Mg^{2+} in the monohydrocalcite structure requires a higher temperature

532   (300-550 °C) compared to water associated with Ca^{2+}. A detailed quantitative analysis of the

533   TGA data (Fig. 9b, inset) shows that the amount of water released at higher temperatures (>

534   300 °C) is directly proportional to the molar fraction of Mg in the solid.

535   Photomicrographs of high or low Mg-containing monohydrocalcite, reveals two

536   distinct morphologies and sizes. Monohydrocalcite χ_{MgCO3}>0.06 consists of individual,

537   nanometer-sized crystals that are <35 nm in size (Fig. 9c). In contrast, monohydrocalcite with

538   χ_{MgCO3}<0.06 consists also of nanoparticulate crystallites (<35nm), but these are aggregated to

539   form elongated particles >5 µm in length. The morphology of these low Mg-

540   monohydrocalcites (Fig. 9d) is identical to "type 2" spherulites described by Granasy et al.

541   (2005), who demonstrated that spherulitic growth occurs via growth front nucleation. This

542   mechanism consists of the continuous nucleation of misaligned equivalent structural units on

543   the surface of growing spherulites, and proceeds via unidirectional growth with low angle

544   branching (Fig. 9d). This process is nucleation controlled with very little, if any, growth of

545   the mineral phase. Spherulites occur in many systems (i.e., organic and inorganic), and

546   usually require high levels of supersaturation in order drive the nucleation controlled process

547   (Shtukenberg et al., 2012; Andreassen, 2005; Andreassen et al., 2010; Beck and Andreassen,

548   2010). These observations are also consistent previous studies who observed

549   monohydrocalcite crystals with spherulitic morphologies (Kimura and Koga, 2011; Neumann

550   and Epple, 2007; Dahl and Buchardt, 2006, Dejehet et al., 1999; Ferguson et al., 1978;

551   Duedall and Buckley, 1971; Nishiyama et al., 2013).

552   Therefore, taking into account its crystal chemistry, crystallite size, thermal behaviour

553   and morphology, we propose a new categorization for monohydrocalcite:

554   - High-Mg monohydrocalcite (χ_{MgCO3}>0.06) consists of individual nanometer-sized

555   crystals (<35 nm) (Fig. 9c) with a significant part (6-25%) of its structural H_2O being

556   associated with the Mg ion, therefore displaying a progressive dehydration during heating to

557   >500°C (Fig. 9b). Such high-Mg monohydrocalcites are uncommon in nature, but can be

558   synthesised in the laboratory at high initial supersaturation levels (SI>3.25).

559   - Low-Mg monohydrocalcite (χ_{MgCO3}<0.06) which forms a "type 2" spherulite

560   morphology. Less than 6% of the structural water in the low-Mg monohydrocalcite is bonded

561   to Mg, so it fully dehydrates at low temperatures (150-200°C). They have the same

562   composition as natural monohydrocalcites reported in the literature, and can be synthesized in

563   the laboratory at lower supersaturation levels (SI<3.25).

564   These observations indicate that despite their different morphologies (single

565   nanometer sized crystals and low-angle branching spherulites, respectively; Fig. 9c and 9d)

566 and levels of supersaturations at which they form, high- and low-Mg monohydrocalcite both
567 crystallize via a nucleation-dominated growth process. The difference in particle size and
568 morphology is likely controlled by the aqueous Mg concentration. At high concentrations,
569 Mg poisons the formation of spherulites but still allows direct nucleation in solution,
570 producing the non-aggregated, individual high-Mg monohydrocalcite crystals. At low
571 supersaturations, the Mg concentration is low and monohydrocalcite forms via growth-front
572 nucleation permitting the development of the low angle branching "type 2" spherulites.

573 Combining the mechanistic results described above with chemical data from our on-
574 line experiment, and data from other studies (Fig. S3) reveals interesting relationships.
575 Firstly, our on-line experiment shows an increase in nanocrystal sizes during the secondary
576 crystallisation of monohydrocalcite, which is coupled with a significant decrease in χ_{MgCO3}
577 (from ~0.26 to ~0.065). This corresponds to the transition from high- to low-Mg
578 monohydrocalcite, suggesting that the former would be metastable and rapidly transforming
579 to the latter, possibly triggered by the removal of Mg from aqueous solution. Secondly, Davis
580 et al. (2000) determined that the solubility of Mg-calcite ($Ca_{1-x}Mg_xCO_3$; x=0-0.20) varies by
581 approximately half an order of magnitude depending on the Mg content of the solid ($K_{sp} = 10^{-8.0}$
582 $^{8.0}$ - $10^{-8.5}$). A similar behaviour should be expected for monohydrocalcite. The saturation
583 indexes calculated for monohydrocalcite using the available solubility products from Hull
584 and Turnbull (1973) and Krajl and Brečević (1995) are negative and show a difference of
585 ~0.55 (Fig. 8). We suggest that may be due to difference in the Mg contents of the
586 monohydrocalcites used in their respective studies. This hypothesis is supported by the recent
587 findings of Nishiyama et al. (2013), who reported that the solubility of synthetic
588 monohydrocalcite increases with higher Mg/Ca ratios in the solid. They showed that the K_{sp}
589 of monohydrocalcite can reach maximum values of $K_{sp} = 10^{-6.77}$, which is almost one order of
590 magnitude higher than the value of Hull and Turnbull (1973). Furthermore, Nishiyama et al.
591 (2013) also reported an decrease in monohydrocalcite crystallite size (broadening of Bragg
592 peaks in PXRD) with increasing solid Mg/Ca ratios, which again support our interpretation of
593 the transition from high- to low-Mg monohydrocalcite.

594 Although an in-depth study of the structural changes in monohydrocalcite as a
595 function of Mg content are outside of the scope of this study, the changes in unit cell
596 parameters during crystallisation (Fig. 6) may be better understood when the χ_{MgCO3} of the
597 monohydrocalcite is taken into account. Regardless of Mg content, the unit cell volume
598 remained virtually constant during stages II-III and only decreased slightly during stage IV
599 ($\Delta V \approx -0.2$ Å^3), in parallel with the decrease in monohydrocalcite χ_{MgCO3}. This small decrease

600 in volume is a consequence of a mirrored change in a-axis and c-axis dimensions, which may
601 be explained by the change in the monohydrocalcite internal structure during the loss of Mg
602 and the transition from the high to low-Mg type (stage III to IV). The structure of
603 monohydrocalcite is less dense and more open than calcite or aragonite (Effenberger, 1981;
604 Neumann and Epple, 2007; Swainson, 2008), therefore it may more easily adapt to a
605 changing Mg content.

606
607

608 **4. SUMMARY AND IMPLICATIONS**

609

610   Results show that under the conditions of our study monohydrocalcite forms via a
611 Mg-rich ACC precursor. The Mg-ACC forms rapidly then transforms to monohydrocalcite
612 via dissolution and reprecipitation, with monohydrocalcite forming via a nucleation-
613 controlled reaction. This crystallization path is consistent with those observed for the
614 formation of vaterite, calcite and dolomite from (Mg-)ACC. The crystalline phase which
615 forms from the (Mg-)ACC is controlled by the magnesium content of the precursor: pure
616 ACC crystallises to vaterite (Bots et al., 2012); 10% Mg-ACC crystallises to calcite
617 (Rodriguez-Blanco et al., 2012); ~ 30% Mg-ACC crystallises to monohydrocalcite (this study
618 and Nishiyama et al., 2013), and 50% Mg-ACC crystallises to protodolomite/dolomite at
619 higher temperatures (Rodriguez-Blanco et al., 2013). The nucleation controlled growth of
620 these phases from ACC is thought to be due to the high solubility of (Mg-)ACC. Any
621 solution in equilibrium with the amorphous phase will be highly supersaturated with respect
622 to the crystalline phase, leading to nucleation controlled growth. This highlights the universal
623 nature of the nucleation-controlled formation during crystallization from ACC.

624   Our results highlight the importance of Mg in the formation of monohydrocalcite. The
625 mechanisms by which Mg controls phase stability are complex, but are related to 3 key
626 factors. Firstly, the presence of Mg in solution is known to inhibit the formation of vaterite
627 and calcite (Bischoff, 1968; Kitano, 1966; Bischoff and Fyfe, 1968; Berner, 1975; Mucci and
628 Morse, 1983; Deleuze and Brantley, 1997; Chen et al., 2004; Bots et al., 2011). We suggest
629 that the Mg levels in the current study (i.e. Mg/(Mg + Ca) = 0.3) were high enough to inhibit
630 the formation of both calcite and vaterite, and favoured the formation of monohydrocalcite.
631 Secondly, the hydration shell of Mg is more strongly bound than the hydration shell of Ca (di
632 Tommaso and de Leeuw, 2010; Moomaw and Maguire, 2008). The energy required to
633 desolvate Mg is higher than that of Ca, a fact which explains the higher temperature required

to crystallize anhydrous Ca-Mg carbonates, e.g. dolomite or magnesite (Christ and Hostetler 1970; Lipmann, 1973; Kittrick and Peryea, 1986; Rodriguez-Blanco et al., 2013). The hydrated nature of monohydrocalcite means that full dehydration of Mg is not required before incorporation into the crystal lattice, therefore it will more likely form than the anhydrous calcium carbonate phases. The TGA analysis of the monohydrocalcite samples support this, as it shows increasing water content with Mg (Fig. 9). Thirdly, monohydrocalcite has a high capacity to accommodate Mg within its structure favouring its formation at higher Mg/Ca ratios. Thus although the initial solutions used in our work were more supersaturated with respect to all anhydrous $CaCO_3$ polymorphs, the formation of monohydrocalcite was favoured.

We have shown that the crystal chemistry and growth morphology of monohydrocalcite (Stoffers and Fischbeck, 1974; Taylor, 1975; Munemoto and Fukushi, 2008) can be controlled by the initial supersaturation of the aqueous solution. We showed that with increasing supersaturation the Mg content in the monohydrocalcite increases up to $\chi_{MgCO3} = \sim 0.26$, similar to the Mg-contents in Mg-calcites reported previously (Lenders et al., 2012; Goldschmidt et al., 1955; Goldschmidt and Graf, 1958). This has led to us defining 2 types of monohydrocalcite i.e. high-Mg and low Mg monohydrocalcite, classified according to their Mg content and morphology. All natural monohydrocalcites, which have been analysed to date, are low Mg monohydrocalites (Mg content $\sim 0.01 > \chi_{MgCO3} < 0.075$, Hull and Turnbull, 1973; Taylor, 1975; Fukushi et al., 2011 and references therein). We suggest that the lack of high-Mg monohydrocalcite in natural environment is due to 2 factors. Firstly, for high-Mg monohydrocalcite to form, initial solutions must be extremely highly supersaturated ($SI_{MHC} \sim 3.5 - 4$; see Table 1 and Table 2). Such extremely high supersaturation levels are difficult to achieve through natural processes. Secondly, our analysis of the monohydrocalcite crystallization pathway shows that high-Mg monohydrocalcite breaks down to low-Mg monohydrocalcite within hours of its formation, therefore high-Mg monohydrocalcite if it forms at all is likely a transient phase. Furthermore, despite monohydrocalcite not being as abundant as calcite and or aragonite in the geological record, it may still have a potential as a palaeoenvironmental indicator (Solotchina et al., 2009). The isotopic composition of the hydration water in natural low-Mg monohydrocalcites may be useful as a $\delta^{18}O$ record of the palaeoenvironmental conditions of the water from which it crystallized. Such an approach has been successfully applied to other hydrated carbonates like ikaite, which is even scarcer in nature and more unstable than monohydrocalcite (Rickaby et al, 2011; Lu et al., 2012).

668   Nevertheless, our experiments documented a simple methodology to synthesize
669   monohydrocalcite with specific Mg/Ca ratios, specific particle sizes and shapes (i.e., reactive
670   surface), which may have applications in a number of fields. For example, synthetic
671   monohydrocalcite have been reported to have a high sorption capacity for PO_4^{3-} and AsO_4^{3-}
672   (Yagi and Fukushi, 2012; Yagi and Fukushi, 2011; Fukushi et al., 2011). Thus, by producing
673   monohydrocalcite with specific compositions, particle sizes and shapes the effectivity of
674   these remediation materials could be improved.

675   Finally, the formation of monohydrocalcite and other CaMg carbonates as a
676   consequence of biological activity has been reported in many systems (Lowenstam, 1981;
677   Sánchez-Román et al., 2011). Microorganisms often promote the formation of highly
678   supersaturated microenvironments which facilitate the precipitation of carbonates (e.g.,
679   bacterial cells which absorb Ca, Mg or other metallic cations and act as nucleation sites),
680   even when the conditions for supersaturation have not been reached in the surrounding
681   environment (Vasconcelos and Mckenzie, 1997; Párraga et al., 1998). Systems characterized
682   by high initial aqueous Mg/Mg+Ca values (e.g., ≥ 0.3) will lead to the formation of a Mg-rich
683   ACC precursor, which in turn will crystallize to monohydrocalcite. We suggest that this may
684   be the primary crystallization pathway during biomineralization of monohydrocalcite.

685   It is interesting to note that monohydrocalcite forms at a broad range conditions in the
686   laboratory and natural environment (i.e. a wide range of solution compositions and Mg/Ca
687   ratios) (Nishiyama et al., 2013; Wood, 2012; Fukushi et al., 2011 and references therein).
688   Yet, monohydrocalcite is scarce in nature, a fact that can be explained by (1) its metastability
689   with respect to aragonite and calcite, and (2) the high supersaturation required to form it
690   compared to other calcium carbonates. However, the chemistry of many carbonates formed
691   during biomineralization processes indicates that they have formed from high Mg/Ca ratios
692   seawater solutions (Davis et al., 2000; Ries, 2010; Ries et al, 2008; Ries, 2004; Reeder et al.,
693   1983), and in some cases at high supersaturations (e.g., Señorale-Pose et al., 2008; Hood et
694   al., 2011; Hood and Wallace, 2012). Furthermore, the paragenesis of some mineral deposits
695   which now contain aragonite also include a variety of Ca-Mg-bearing carbonates like
696   dolomite, hydromagnesite or nesquehonite (Fischbeck and Mueller, 1971; Broughton, 1972).
697   Contrary to monohydrocalcite, aragonite does not necessarily require the presence of Mg to
698   form (e.g., Ogino et al., 1987; Sand et al., 2012), but it is well known that Mg and SO_4^{2-} are
699   the key ions responsible for the switch from calcite to aragonite seas (Sandberg, 1983; Bots et
700   al., 2011). During geological timescales the mineralogy of most calcifying organisms was
701   influenced by the Mg/Ca ratio of the oceans in which they evolved (Stanley, 2006; Porter,

702   2007). For that reason we consider that some aragonite and (Mg)-calcite deposits that are

703   known to have formed at high Mg/Ca ratios (e.g., coralline red algal beds; rhodoliths, Schäfer

704   et al., 2011; ooids, Brehm et al., 2006; Davies et al., 1978; Ferguson et al., 1978 or

705   microbialites, Burne and Moore, 1987) could be secondary in origin and may have originally

706   been a metastable monohydrocalcite intermediate which recrystallized. This secondary

707   crystallization via monohydrocalcite could be the reason why the skeletal mineralogy and

708   chemistry of calcareous organisms that recorded the Mg/Ca ratio of the seawater in which

709   they grow varied so dramatically during the Phanerozoic (Ries, 2004) and this in turn may

710   affect Mg/Ca ratio – based paleoenvironmental reconstructions. This may also explain why

711   some early Cambrian fossils may have preferentially formed low-cost, high-Mg calcite, while

712   later the increasing physiological cost of biomineralization lead to low Mg-calcite skeletons

713   (Wood, 2011). Whether monohydrocalcite played a role in these early biomineralization

714   processes is still an open question, although if monohydrocalcite was an intermediate phase

715   during these biomineralization reactions it may have big implications for seawater chemistry

716   reconstructions and other paleoproxies and would require further study.

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718   5. **ACKNOWLEDGEMENTS**

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728   **6. REFERENCES**

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Figures
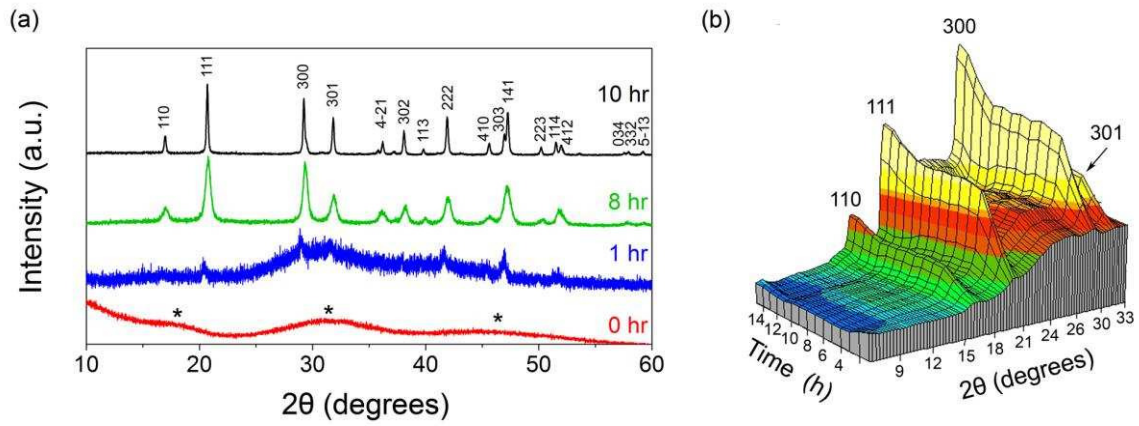
1039 **Fig. 1.** (a) PXRD patterns from off-line crystallization experiments with samples removed
1040 from each run after 0, 1, 8 and 10 hr. (b) 3D WAXS plot showing the time resolved formation
1041 of monohydrocalcite from the on-line experiment with only some major peaks shown.

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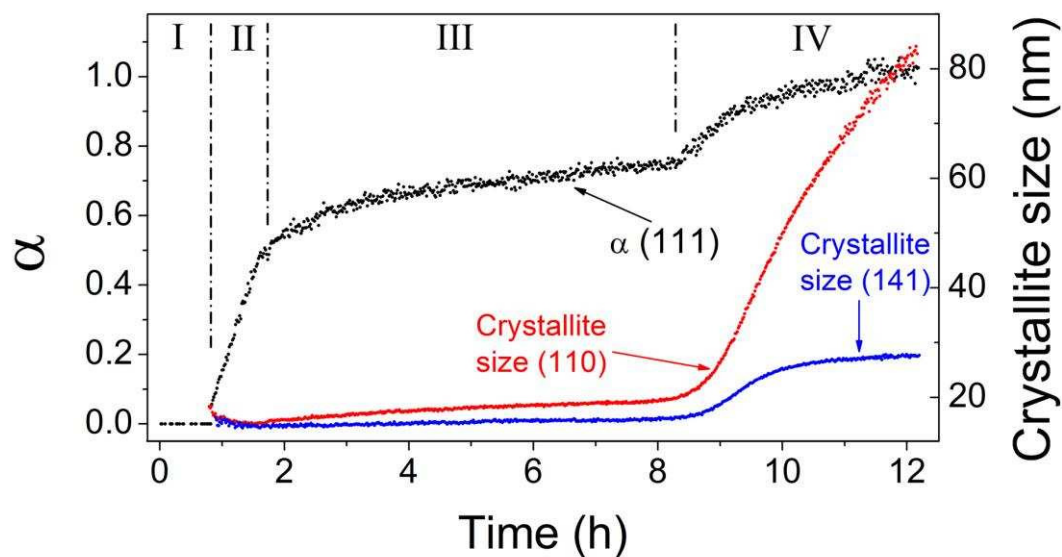
Fig. 2. Evolution of the degree of reaction (α) of the monohydrocalcite (111) Bragg peak (black symbols). The same growth profiles are true for all other Bragg peaks. Shown also with the right-hand y-axis are Scherrer crystallite sizes for the (110) and (141) peaks of monohydrocalcite as a function of time as derived from the WAXS data. I to IV correspond to the crystallization stages described in the text.
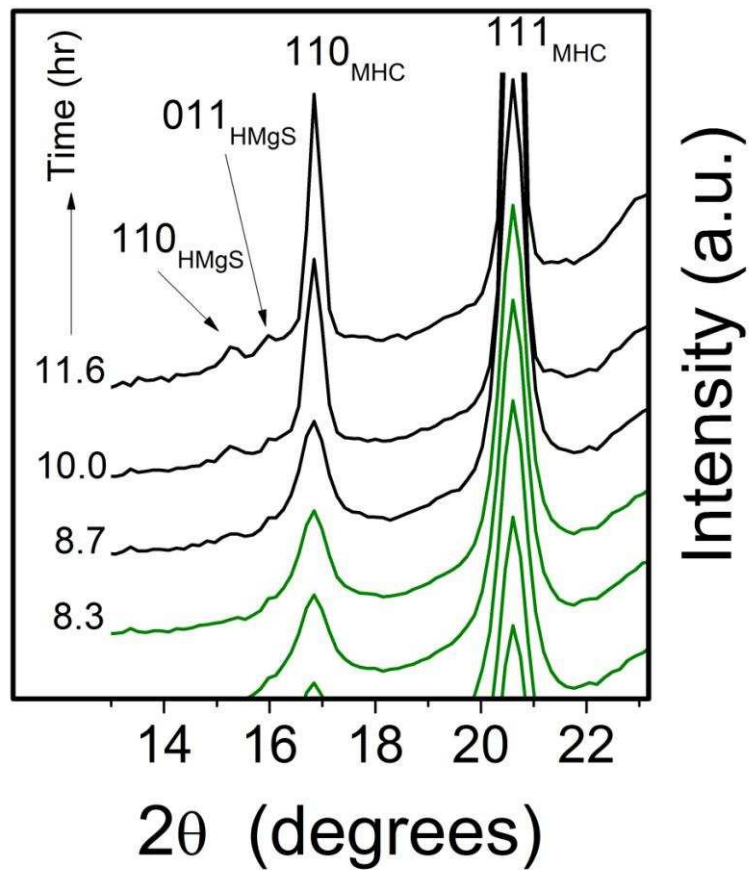
Fig. 3. Selected WAXS patterns from the on-line experiment showing the growth of the monohydrocalcite (MHC) 110 and 111 Bragg peaks and the formation of the small hydromagnesite (HMgS) 110 and 011 Bragg peaks during stages III (green patterns) and IV (black patterns) of the reaction.
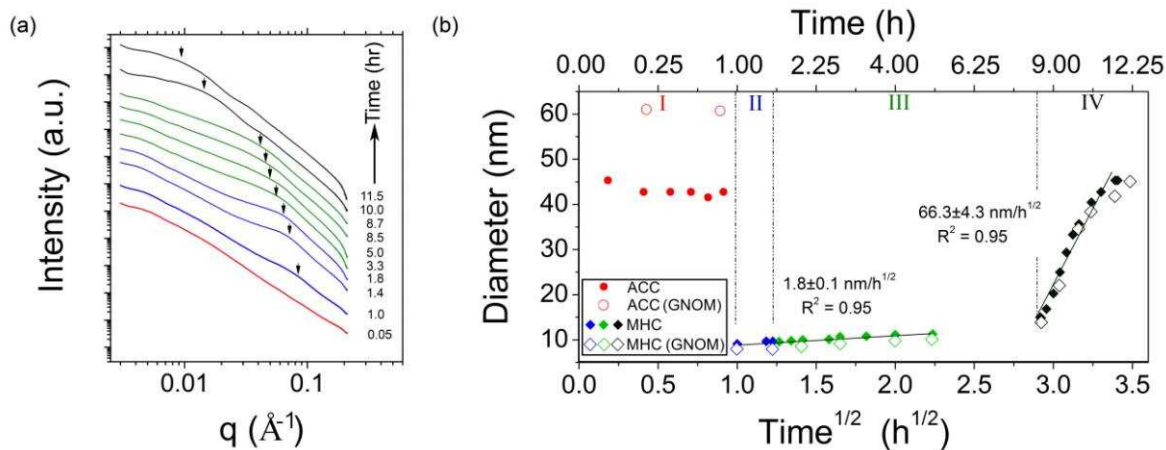
Fig. 4. (a) Double logarithmic plot showing selected SAXS patterns from the on-line in situ experiment with arrows indicating a changing position of the scattering peak, reflecting the growth of monohydrocalcite particles with time; colours of the patterns matches the colour code of the four stages of reaction shown in (b) Time evolution of the particle diameters for the initial amorphous phase (Mg-ACC stage I, red symbols) and for monohydrocalcite (MHC stages II to IV, blue, green and black symbols respectively) as derived from the SAXS data. Empty symbols correspond to GNOM code (Svergun, 1992). The differences in growth rates of the monohydrocalcite particles are evidenced by the fit in stages II-III and IV.
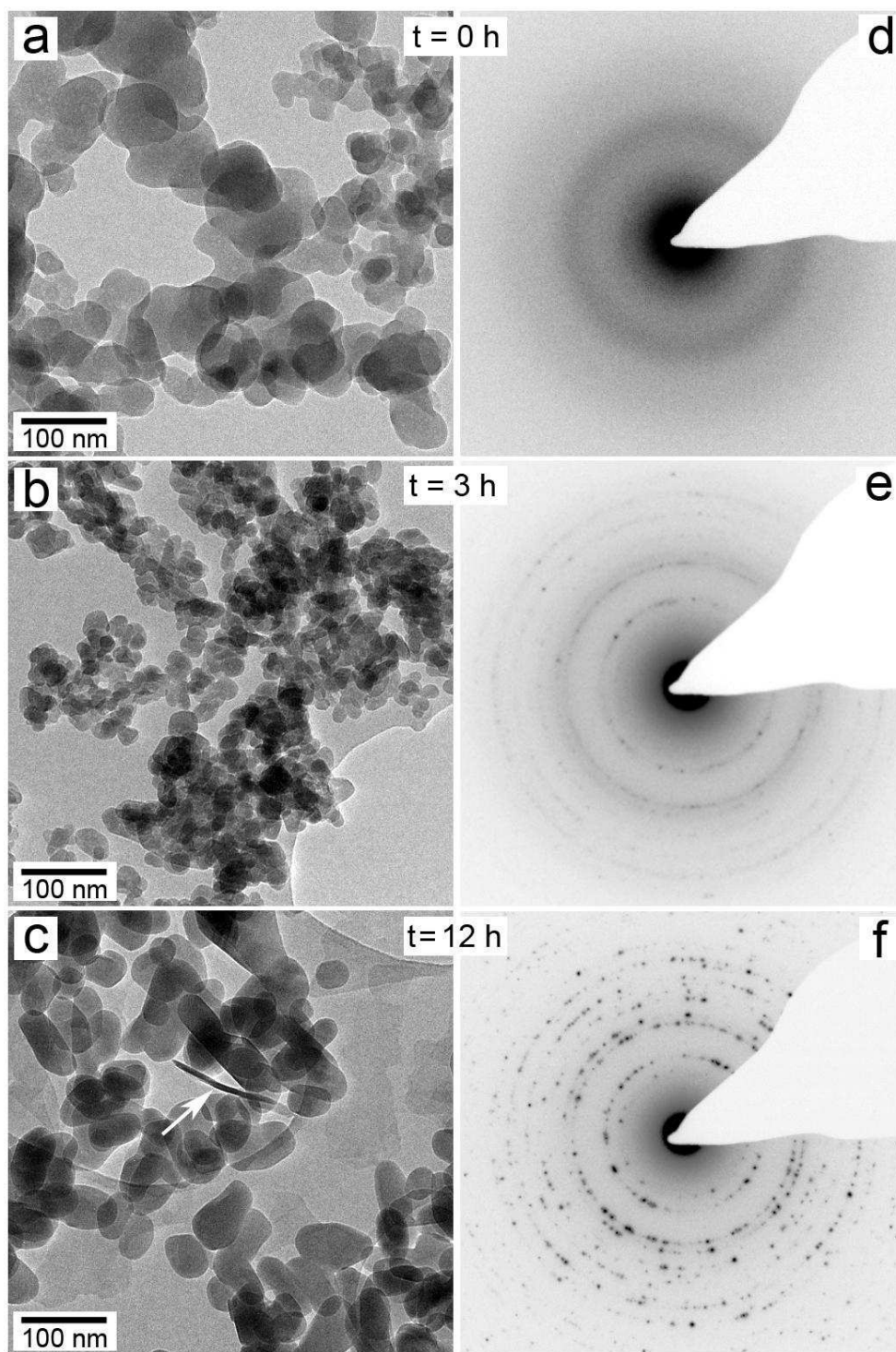
Fig. 5. HR-TEM microphotographs and corresponding SAED patterns of solids during the crystallization reaction. (a, d) Mg-ACC precursor; stage I); (b, e) Monohydrocalcite nanocrystals from stage III; (c, f) Bigger monohydrocalcite particles and a platy hydromagnesite crystal (white arrow) from the end of stage IV.
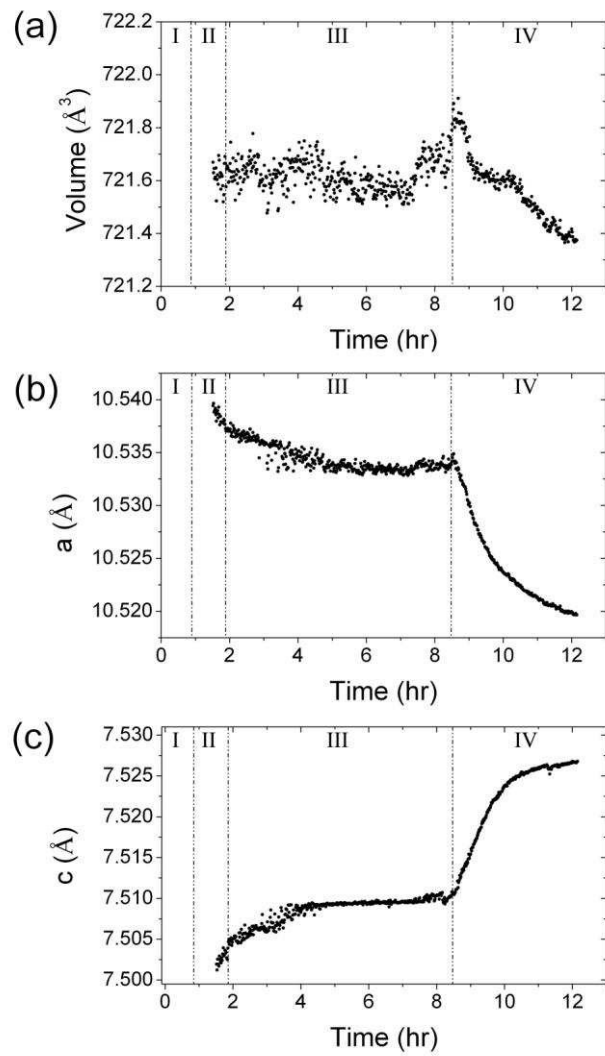
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1086   **Fig. 6.** Evolution of the unit cell volume (a) and of the a and c parameters (panels b and c) for
1087   monohydrocalcite over time as derived from the on-line WAXS data for stages II to IV.
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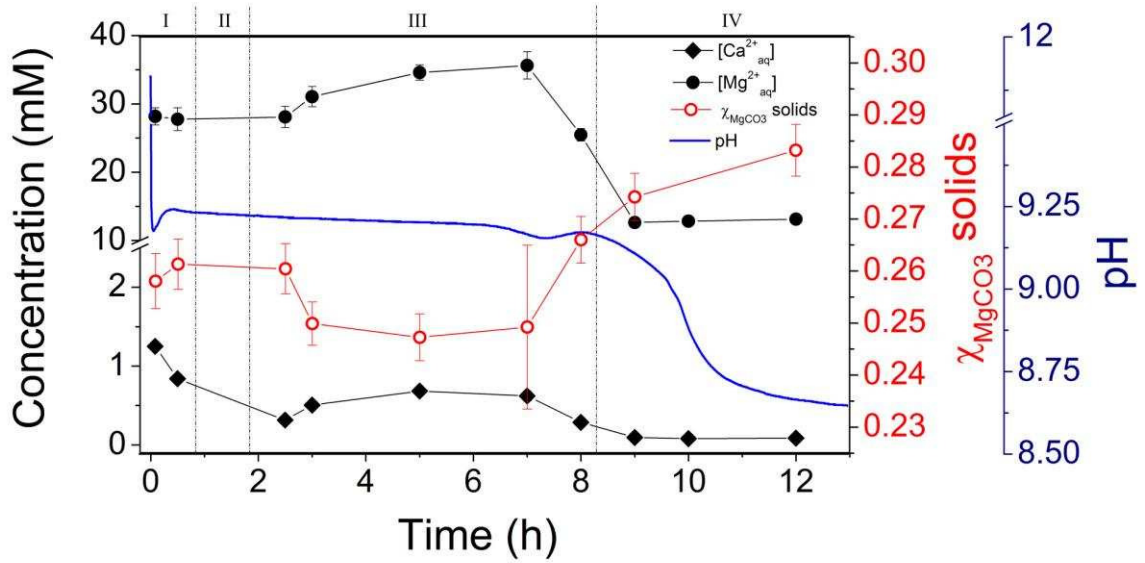
Fig. 7. Evolution of the pH, the concentrations of $[Ca^{2+}_{aq}]$, $[Mg^{2+}_{aq}]$ and of the Mg molar fraction (χ_{MgCO3}) in the solids over the course of the four stages of the crystallization reaction. Error bars correspond to the standard deviation of three measurements each.

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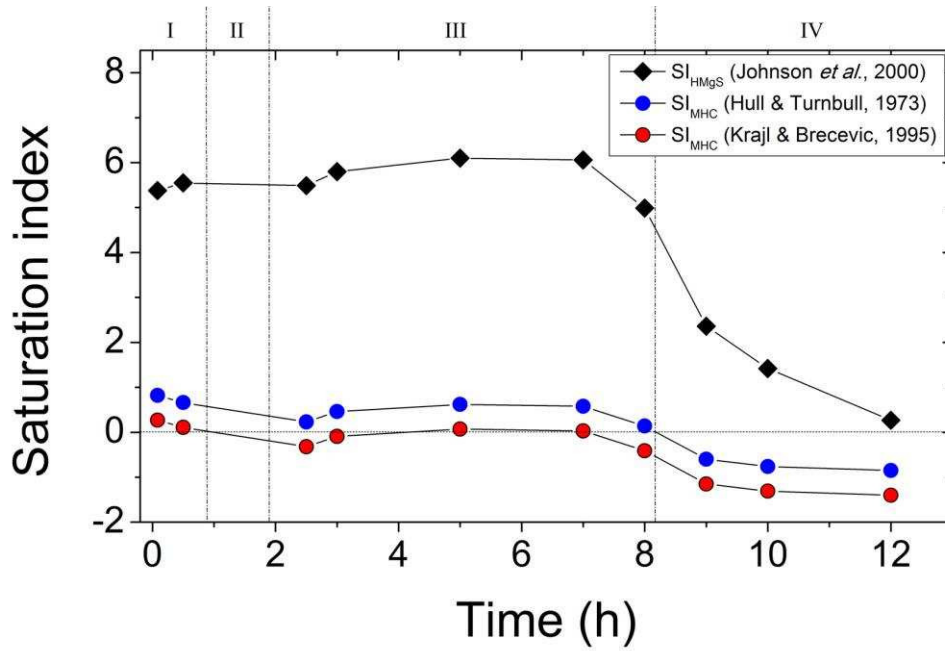
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1100   **Fig. 8.** Evolution of the saturation indexes for monohydrocalcite (SI_{MHC}) and hydromagnesite
1101   (SI_{HMgS}) calculated from the data shown in Fig. 7 and Table 1.
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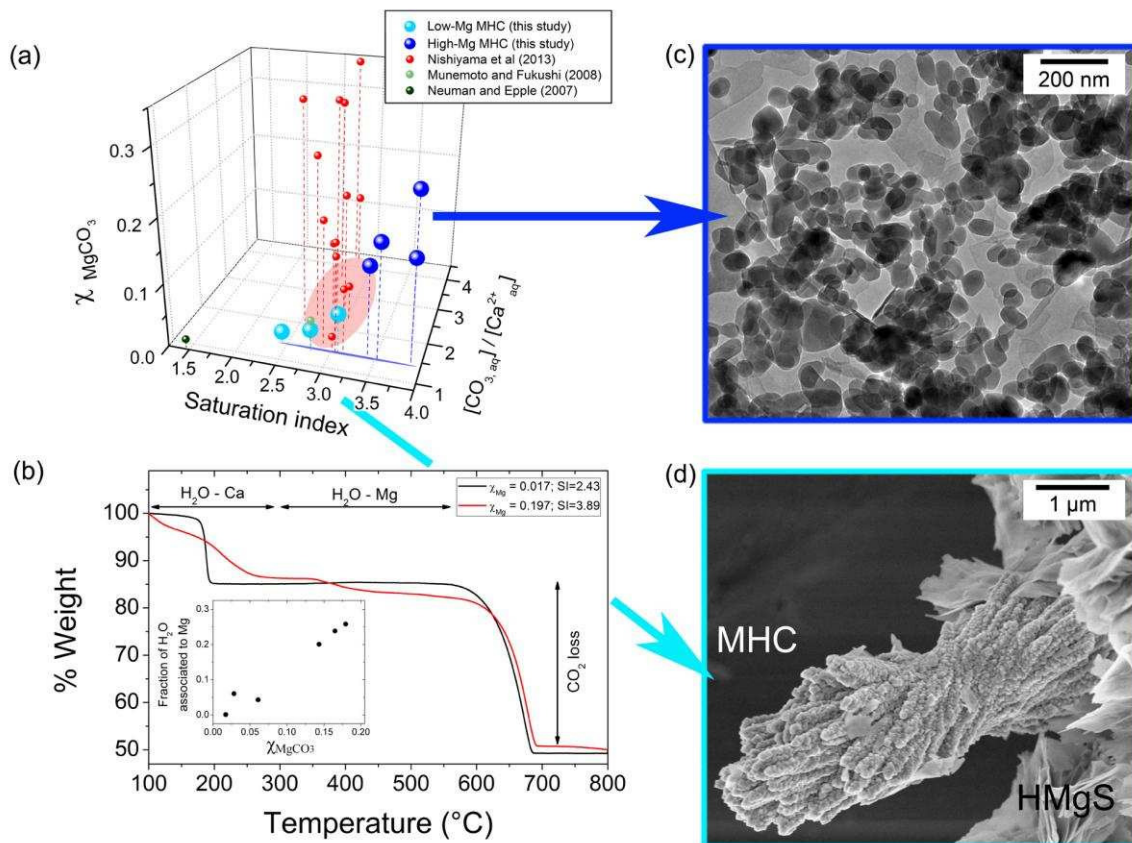
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1107 **Fig. 9.** (a) Three dimensional plot showing the relationship between the molar fraction of Mg
1108 (χ_{MgCO3}) in monohydrocalcite as a function of its initial saturation index (SI_{MHC}) and starting
1109 $[CO_{3,aq}]/[Ca^{2+}_{aq}]$ ratios. Blue and turquoise larger spheres are our off-line experimental data
1110 (Table 2) that were all obtained at the same starting $[CO_{3,aq}]/[Ca^{2+}_{aq}]$ ratio; the smaller, red,
1111 green and black spheres correspond to literature data. (b) Comparative TGA graphs for
1112 monohydrocalcite synthesised at high (SI=3.89) and low (SI=2.43) initial supersaturations
1113 (Table 2) with the inset showing the fraction of Mg-bonded structural water in
1114 monohydrocalcite vs. χ_{MgCO3}. Images (c) and (d) illustrate the differences in morphology,
1115 particle sizes and formation modes for our high- and low-Mg monohydrocalcite (MHC),
1116 respectively. Hydromagnesite (HMgS) crystals are also present in image (d).

1117

Table 1.

Evolution of the aqueous solution composition, the magnesium content in the solids (χ_{MgCO3}) and the solid mineralogy during the course of a typical experiment. Saturation indexes for monohydrocalcite, hydromagnesite and nesquehonite (SI_{MHC} and SI_{HMgS}, SI_{nesq}) were calculated from the measured pH and $[Ca^{2+}_{aq}]$ and $[Mg^{2+}_{aq}]$ values combined with the solubility products for monohydrocalcite reported by (1) Hull and Turnbull (1973), and (2) Krajl and Brečević (1995) and those for hydromagnesite and nesquehonite from Johnson et al. (2000). Average particle sizes were evaluated from the SAXS data or from photomicrographs.

Stage	Time (h)	pH	$[Ca^{2+}_{aq}]$ (mM)	$[Mg^{2+}_{aq}]$ (mM)	χ_{MgCO3} solid	Mineralogy	Particle sizes from SAXS / Crystallite sizes from Scherrer (nm)	Particle sizes from TEM imaging (nm)	SI_{MHC} (1)	SI_{MHC} (2)	SI_{HMgS}	SI_{nesq}
Pre-mixing	0	-	350	150	-	-	40-60 / -		3.89	3.34	13.06	1.17
I	0.083	9.239	1.25 ± 0.05	28.17 ± 1.26	0.258 ± 0.005	Mg-ACC	40-60 / -	~ 40	0.82	0.27	5.38	0.03
	0.5	9.217	0.84 ± 0.04	27.75 ± 1.68	0.261 ± 0.005		43 / -		0.66	0.11	5.55	0.04
II	-	-	-	-	-	-	-	-	-	-	-	-
III	2.5	9.214	0.32 ± 0.02	28.08 ± 1.57	0.260 ± 0.005	MHC	10 / 15	28±7 (L) x 20±5 (W)	0.23	-0.32	5.49	0.04
	3	9.203	0.51 ± 0.03	31.05 ± 1.48	0.250 ± 0.004		11 / 15		0.46	-0.09	5.8	0.11
	5	9.167	0.68 ± 0.03	34.59 ± 1.14	0.247 ± 0.005		11 / 16		0.62	0.07	6.1	0.18
	7	9.171	0.62 ± 0.06	35.62 ± 2.00	0.249 ± 0.016		- / 17		0.58	0.03	6.06	0.19
	8	9.111	0.29 ± 0.01	25.47 ± 0.93	0.266 ± 0.004		- / 17.5		0.14	-0.41	4.99	-0.05
IV	9	8.893	0.09 ± 0.01	12.64 ± 0.75	0.274 ± 0.005	MHC+HMgS	15 / 22.5		-0.6	-1.15	2.36	-0.60
	10	8.646	0.08 ± 0.00	12.83 ± 0.39	-		35 / 37.5	155±80 (L) x 40±30 (W)	-0.76	-1.31	1.42	-0.70
	12	8.609	0.09 ± 0.00	13.12 ± 0.54	0.283 ± 0.005		45 / 45		-0.85	-1.4	0.27	-0.85

Table 2.

Initial stock solution concentrations, and corresponding calculated monohydrocalcite saturation index (SI_{MHC}) for the solution immediately after mixing for each experiment. The first line represents the conditions used in the on-line SAXS/WAXS experiments. The mineralogy, Scherrer crystallite size and Mg (χ_{MgCO3}), and H_2O content of the solid products after 2 h of reaction are also presented.. The saturation index of monohydrocalcite was calculated using the solubility product from Hull and Turnbull (1973).

$Na_2CO_{3(ini)}$ (mM)	$[Ca^{2+}_{aq}]_{(ini)}$ (mM)	$[Mg^{2+}_{aq}]_{(ini)}$ (mM)	SI_{MHC} (t = 0 sec)	Mineralogy from PXRD	Scherrer crystallite size (average) (nm)	χ_{MgCO3}* solid	H_2O content
500	350	150	3.89	MHC	16	0.164	0.99
250	175	75	3.52	MHC	17	0.180	1.03
200	140	60	3.41	MHC	-	0.143	1.00
100	70	30	3.08	MHC	-	0.061	0.97
50	35	15	2.76	MHC	35	0.028	0.95
25	17.5	7.5	2.43	MHC	35	0.017	0.93
12.5	8.75	3.75	2.10	Mg-Calcite	43	0.073	0
5	3.5	1.5	1.63	Mg-Calcite	131	0.032	0

* Errors for χ_{MgCO3} of solids are in all cases lower than 0.002
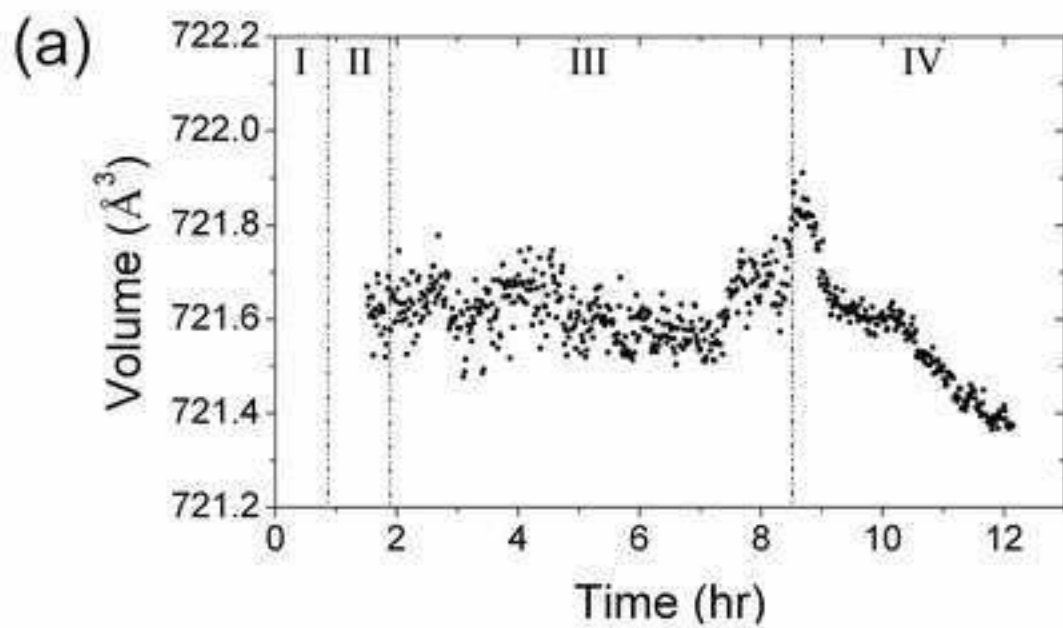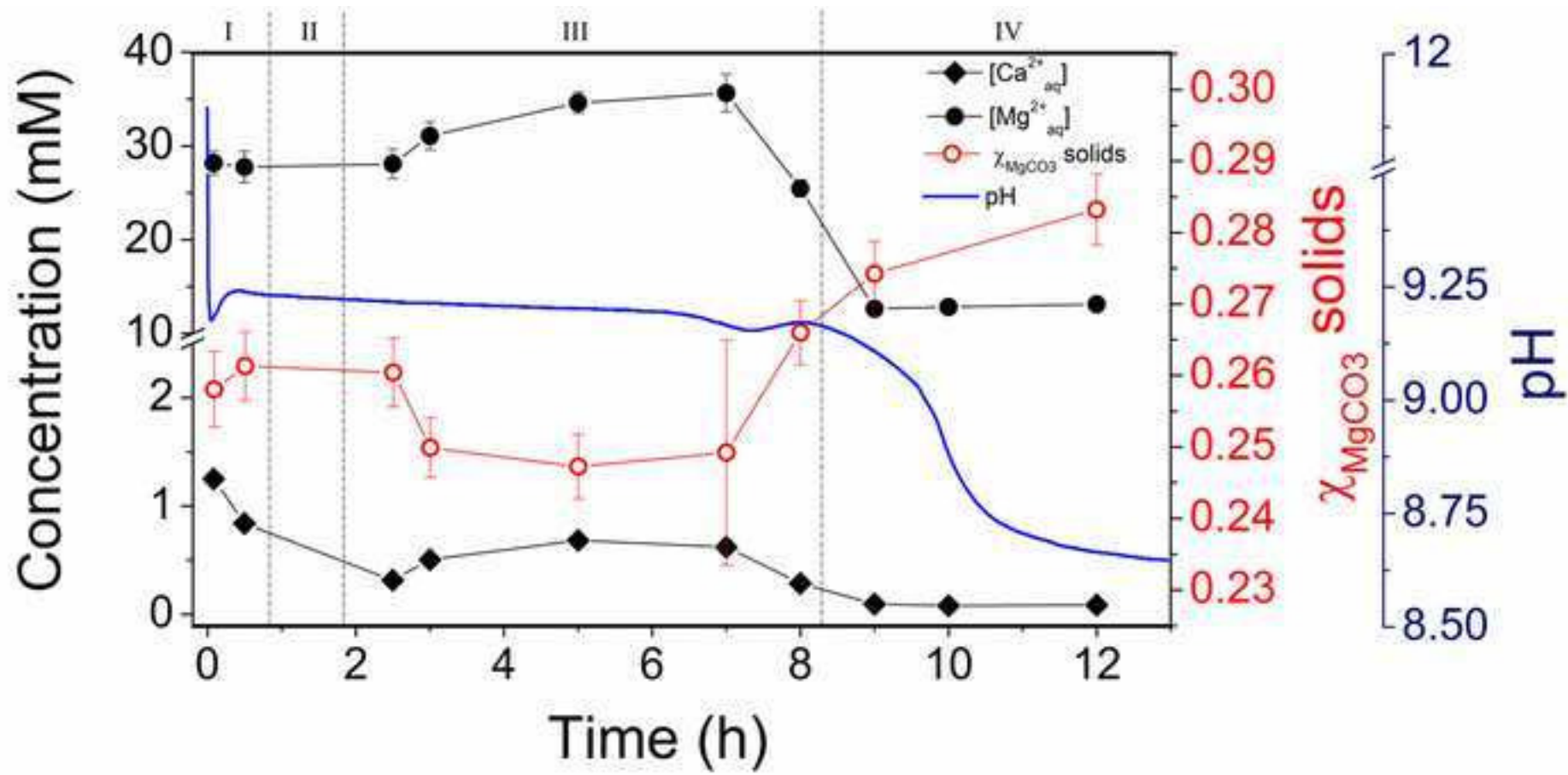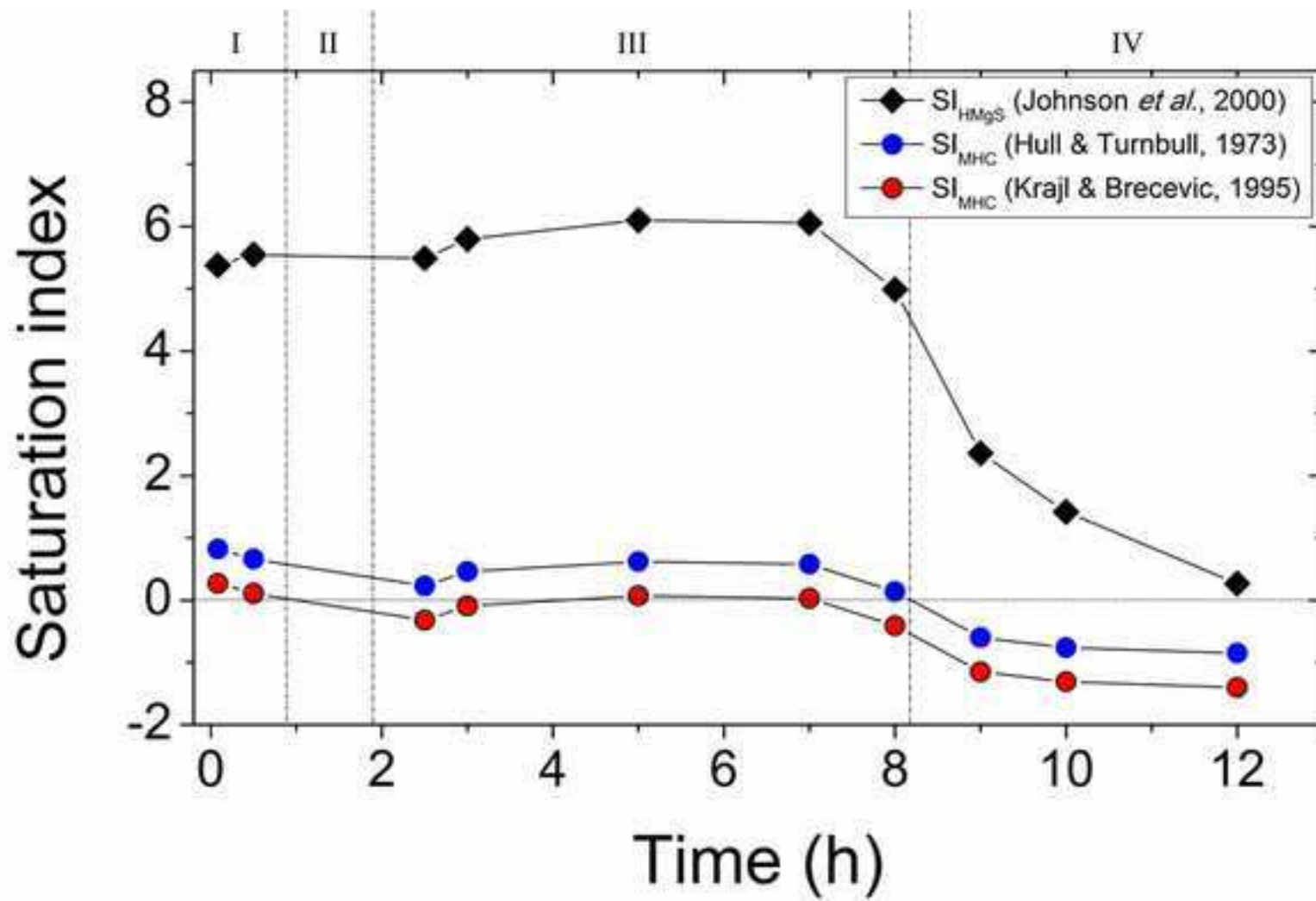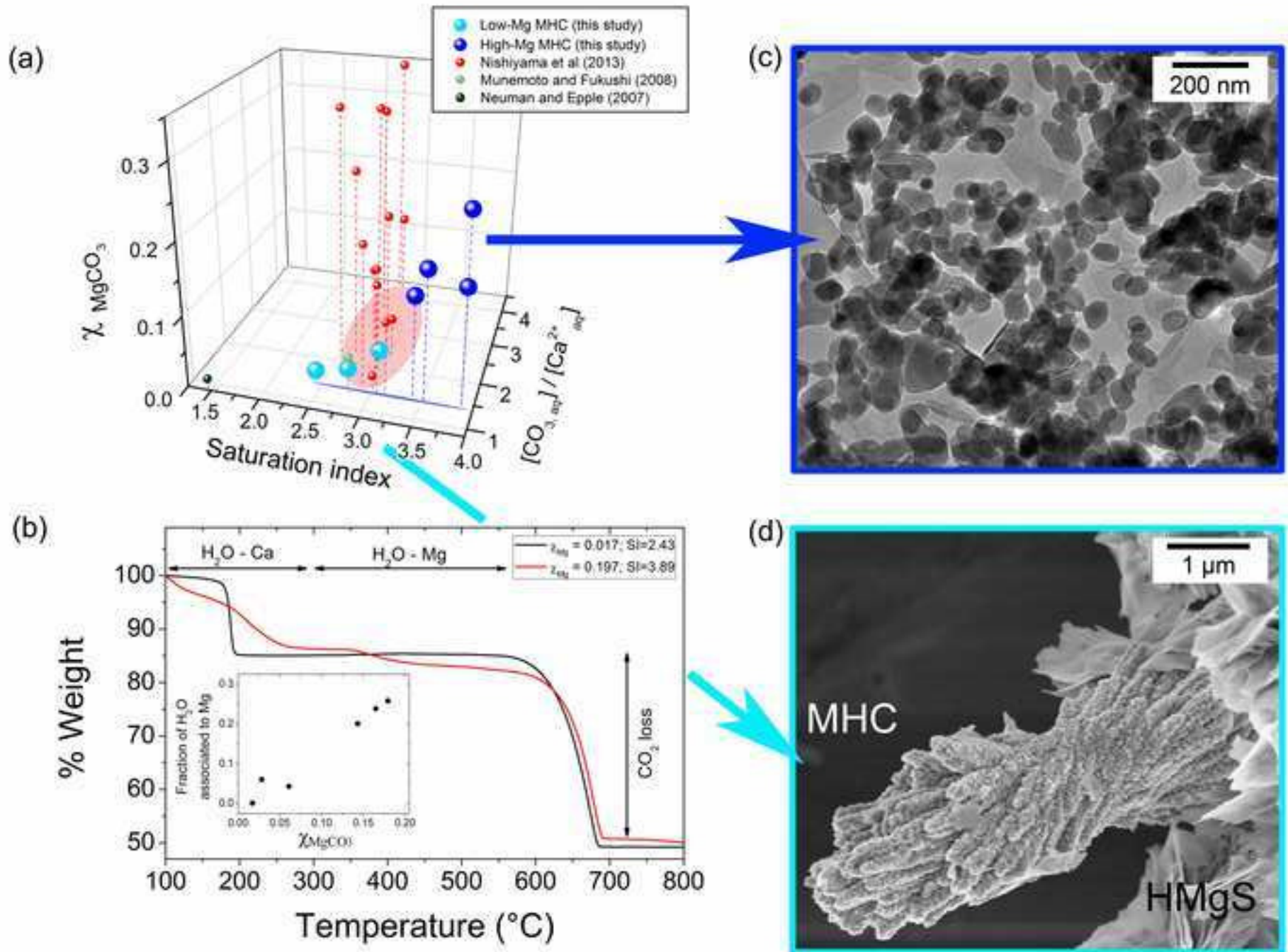
Fig 9.jpg
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Supplementary Information

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Supplementary information

for manuscript:

The role of Mg in the crystallisation of monohydrocalcite.

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The following diagrams constitute the supplementary information for the manuscript entitled '*The role of Mg in the crystallization of monohydrocalcite*', which details the formation mechanisms for monohydrocalcite from solution via a poorly ordered Mg-rich carbonate precursor.
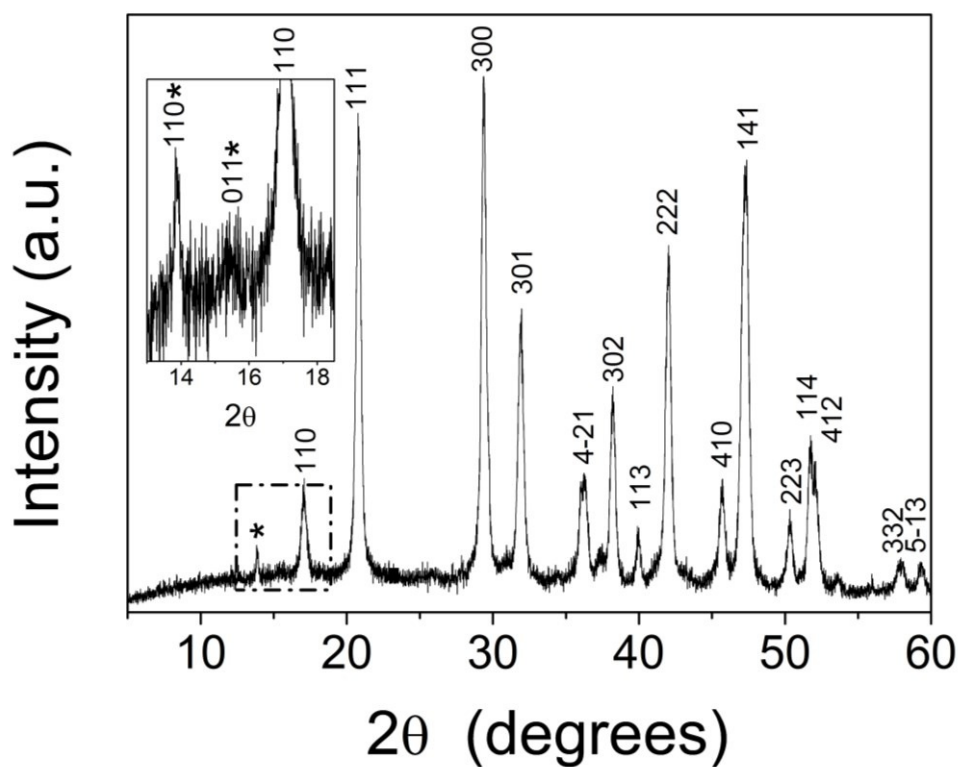
Figure S1: Off-line PXRD pattern of the solids from stage IV; main image shows a pattern of monohydrocalcite with its Bragg peaks marked and with only a tiny peak (at 2 theta of ~ 14; marked with a *) indicating hydromagnesite; in the inset the minor (<5%) contribution from the hydromagnesite with its (110) and (011) Bragg peaks (marked with *) are shown.
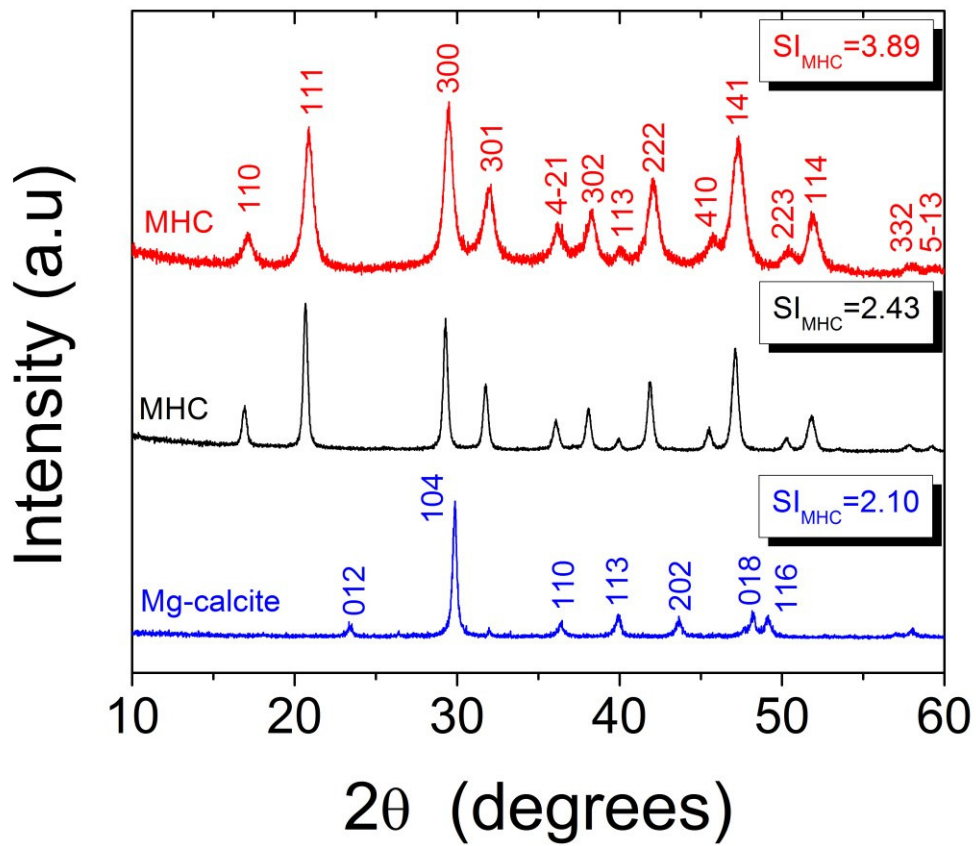
Figure S2: Off-line PXRD patterns of solids obtained after 2 hours at three different initial (prior to mixing) saturation indexes (SI_{MHC}) showing the increase in crystallinity and change in mineralogy from pure monohydrocalcite to Mg-calcite with decreasing initial SI. See Table 2 of manuscript for details.
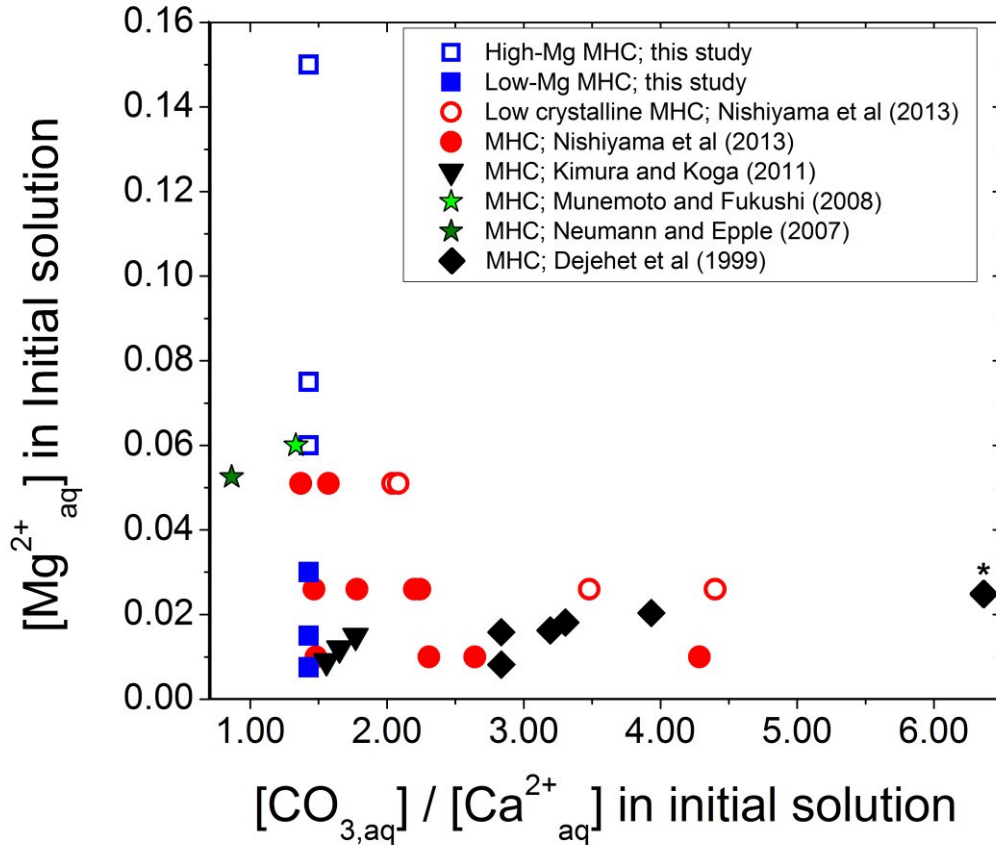
Figure S3: Compositions of monohydrocalcites (MHC) produced in the current study and compared with the literature plotted as a function of initial $[CO_{3,aq}]/[Ca^{2+}_{aq}]$ and $[Mg^{2+}_{aq}]$ concentrations. Filled and empty blue squares correspond to our low-Mg ($\chi_{MgCO3}<0.06$) and high-Mg ($\chi_{MgCO3}>0.06$) monohydrocalcite samples, respectively. Empty circles correspond to 'low crystalline monohydrocalcite' described by Nishiyama *et al*. (2013), which is structurally and chemically similar to the high-Mg monohydrocalcite described in this study. One of the samples from Dejehet *et al* (1999; marked with an *) has a χ_{MgCO3} equivalent to our high-Mg ($\chi_{MgCO3}>0.06$) monohydrocalcite and was produced at very high initial carbonate concentrations.

References:

Nishiyama, R., Munemoto, T., Fukushi, K. (2013) Formation condition of monohydrocalcite from CaCl$_2$–MgCl$_2$–Na$_2$CO$_3$ solutions. *Geochimica et Cosmochimica Acta*, **100**, 217-231.