

Effect of bacteria density and accumulated inert solids on the effluent pollutant concentrations predicted by a Constructed Wetlands model

Roger Samsó^a, Jordi Blázquez^a, Núria Agulló^a, Joan Grau^b, Ricardo Torres^b, Joan García^a

^a*GEMMA - Group of Environmental Engineering and Microbiology, Department of Hydraulic, Maritime and Environmental Engineering, Universitat Politècnica de Catalunya-BarcelonaTech, c/ Jordi Girona, 1-3, Building D1, E-08034, Barcelona, Spain.*

^b*Fluid Mechanics Department, Universitat Politècnica de Catalunya-BarcelonaTech, c/ Urgell 187. E-08036, Barcelona, Spain.*

Abstract

Constructed wetlands are a widely adopted technology for the treatment of wastewater in small communities. The understanding of their internal functioning has increased at an unprecedented pace over recent years, in part thanks to the use of mathematical models. BIO_PORE model is one of the most recent models developed for constructed wetlands. This model was built in the COMSOL MultiphysicsTM software and implements the biokinetic expressions of Constructed Wetlands Model 1 (CWM1) to describe the fate and transport of organic matter, nitrogen and sulphur in horizontal subsurface-flow constructed wetlands. In previous studies, CWM1 was extended with the inclusion of two empirical parameters (M_{bio_max} and M_{cap}) that proved to be essential to provide realistic bacteria growth rates and dynamics. The aim of the current work was to determine the effect of these two parameters on the effluent pollutant concentrations predicted by the model. To that end, nine simulations, each with a different M_{bio_max} - M_{cap} pair, were launched on

a high-end multi-processor computer and the effluent COD and ammonia nitrogen concentrations obtained on each simulation were qualitatively compared among them. Prior to this study, a finite element mesh optimization procedure was carried out to reduce computational cost. Results of the mesh optimization procedure indicated that among the 5 tested meshes of different element size, the mesh utilized for this model in previous studies represented a fair compromise between output accuracy and computation time. Results of the sensitivity analysis showed that the value of M_{cap} has a dramatic effect on the simulated effluent concentrations of COD and ammonia nitrogen, which clearly decreased for increasing values of this parameter. On the other hand, $M_{bio.max}$ was also sensitive, but its effects on the model output were less important and no clear relation could be established between its value and the simulated effluent concentration of COD and ammonia nitrogen.

Keywords: Local sensitivity, mesh optimization, bacteria, growth, parallel computing, batch

1. Introduction

Constructed Wetlands (CWs) are wastewater treatment systems usually applied for communities of less than 2000PE. This technology provides comparable treatment efficiencies with significantly lower energy and maintenance requirements than conventional technologies (García et al., 2010; Puigagut et al., 2007).

However, and due to the diversity and complexity of the physic-chemical and biological processes occurring within CWs, their functioning is far less well understood than that of activated sludge systems. To bridge this knowl-

10 edge gap, several mathematical models have been developed in recent years
11 to simulate CWs functioning (Meyer et al., 2014; Samsó et al., 2014b).

12 The BIO_PORE model is one of such models and was developed in COM-
13 SOL MultiphysicsTM, a commercial finite elements (FE) simulation platform
14 (Meyer et al., 2014; Samsó and García, 2014a; Samsó et al., 2014b; Samsó and
15 García, 2013a,b). This model aims at describing the hydraulics and hydrody-
16 namics of CWs, as well as the removal of the most common pollutants found
17 in wastewater. To that end, it implements the biokinetic model Constructed
18 Wetlands Model 1 (CWM1) (Langergraber et al., 2009), which describes the
19 fate of organic mater, nitrogen and sulphur. This biokinetic model is based
20 on the formulation of the well-known Activated Sludge Model series (ASMs)
21 for aerobic and anoxic processes (Henze et al., 2000) and on the Anaerobic
22 Digestion Model 1 (ADM1) to describe anaerobic processes (Batstone et al.,
23 2002).

24 In BIO_PORE two logistic functions are added to the original formula-
25 tion of CWM1, which involve two new empirical parameters $M_{bio.max}$ and
26 M_{cap} (Samsó and García, 2013a). These two parameters represent, respec-
27 tively, the maximum microbial biomass (carrying capacity) and the maximum
28 amount of particulate solids that can be maintained in a representative vol-
29 ume of granular material. The function involving $M_{bio.max}$ has already been
30 used in several bioclogging studies (Brovelli et al., 2009) and adds a negative
31 feedback term to the growth of all bacteria groups to prevent their unlim-
32 ited growth in areas where substrates concentrations are high. On the other
33 hand, the expression involving parameter M_{cap} also adds a negative feedback
34 term to the growth equations, but in this case it decreases the growth rate

35 of bacteria due to the progressive accumulation of inert solids in the pore
36 space of the granular media (Samsó and García, 2014a). Our previous stud-
37 ies proved the importance of these two functions in order to obtain realistic
38 bacteria concentrations within the granular media (Samsó and García, 2014a;
39 Samsó and García, 2013a). As bacterial communities play a major role on
40 the treatment of wastewater in CWs, these two functions also improved the
41 model predictions regarding effluent pollutant concentrations.

42 However, in these previous studies a sensitivity analysis of parameters
43 $M_{bio,max}$ and M_{cap} was not carried out and so their effect on the model output
44 could not be evaluated. A parameter with high sensitivity is one for which
45 small changes in its value produce large variation in a certain output of the
46 model. On the contrary, low sensitivity parameters are those which do not
47 affect model outputs even for large changes on their value. In this context, the
48 main objective of the current work was to evaluate the sensitivity of $M_{bio,max}$
49 and M_{cap} on the effluent pollutant concentrations of COD and ammonia and
50 ammonium nitrogen predicted by the model. To that end, the BIO_PORE
51 model was used with the same domain, parameter values and initial and
52 boundary conditions than in our previous paper in which the model was
53 calibrated (Samsó and García, 2013a). Due to the large computational cost
54 associated with solving the model for a simulated period of an entire year
55 of operation of a wetland (up to 16 hours for dense finite elements (FE)
56 meshes with a current desktop computer), and due to the large number of
57 simulations needed for the current and for further studies, a previous mesh
58 optimization procedure was carried out. The objective of this part of the
59 study was to find the FE mesh which would provide the best compromise

60 between numerical solutions accuracy and computational cost.

61 The two empirical parameters discussed in this work are essential to ob-
62 tain realistic bacteria concentrations when simulating CWs and this study
63 shows how they affect the effluent pollutant concentrations predicted by the
64 BIO_PORE model. In this work we also exploited the batch and paral-
65 lel computation functionalities of COMSOL MultiphysicsTM on a high-end
66 multi-processor computer which is easily justified by the large number of
67 simulations performed.

68 2. Methods

69 The local parameter sensitivity analysis and the mesh optimization pro-
70 cedure were performed using the exact same domain, parameter values and
71 boundary and initial conditions as in Samsó and García (2013a). For this
72 reason, only the basic equations of the BIO_PORE model are described in
73 this section. For an in-depth description of all model equations the reader is
74 referred to the original source. All simulations performed in this study were
75 run for the entire first year of operation of a pilot wetland.

76 2.1. BIO_PORE model description

77 2.1.1. Governing equations

78 In BIO_PORE model, the saturated porous media flow is described using
79 the Darcy equation (Eq. 1).

$$q_i = -K_{ij} \frac{\partial H}{\partial x_j} \quad (1)$$

80 Where, q_i is the specific discharge [LT^{-1}], K_{ij} is the saturated hydraulic
81 conductivity tensor [LT^{-1}], and $\frac{\partial H}{\partial x_j}$ the hydraulic gradient vector (unitless).

82 Since in CWs both saturated and unsaturated conditions coexist, the *De-*
 83 *formed Geometry* node of COMSOL MultiphysicsTM was used to dynamically
 84 adjust the top boundary of the model domain to the simulated shape and
 85 location of the water table.

86 The fate and transport of the aqueous phase (mobile) wastewater com-
 87 ponents of CWM1 (Table 1) are described with reactive transport equa-
 88 tions, one for each component, in which the reactive term accounts for the
 89 production/consumption of the substrate through microbial activity (Eq.
 90 2)(Clement et al., 1998).

$$\frac{\partial C_k}{\partial t} = \frac{\partial}{\partial x_i} \left(D_{ij} \frac{\partial C_k}{\partial x_j} \right) - \frac{\partial}{\partial x_i} (q_i C_k) + r_r - r_{att} + r_{det} + s_s \quad (2)$$

91 Where $k = 1, 2..m$

92 Where, m is the total number of aqueous phase species (dissolved and
 93 particulate, see Table 1), C_k [ML^{-3}] is the concentration of the k^{th} aqueous
 94 phase species, D_{ij} [L^2T^{-1}] is the hydrodynamic dispersion tensor. q_i [LT^{-1}]
 95 is the specific discharge and acts as the coupling variable between equations
 96 1 and 2. r_r [$ML^{-3}T^{-1}$] is the reaction rate of the k^{th} species in the aqueous
 97 phase. r_{att} [$ML^{-3}T^{-1}$] and r_{det} [$ML^{-3}T^{-1}$] are attachment and detachment
 98 rates, respectively, and are used to simulate mass exchanges between the
 99 aqueous and the solid phases of particulate components X_S and X_I . s_s
 100 [$ML^{-3}T^{-1}$] is the source/sink term, which represents external sources or
 101 sinks of species C_k . This last term is only used to simulate oxygen release
 102 and nutrients uptake through plant roots (see Samsó and García (2013a) for
 103 more details).

104 On the other hand, Eq. (3) describes the fate of the solid phase (immobile)

105 species (Table 1):

$$\frac{d\check{C}_l}{dt} = \check{r}_r + r_{att} - r_{det} \quad (3)$$

106 Where $l = 1, 2, ..n$

107 Where, n is the total number of solid phase species (particulate only),
108 $\check{C}_l[ML^{-3}]$ is the concentration of the l^{th} species and $\check{r}_r[ML^{-3}T^{-1}]$ is the
109 reaction rate of the l^{th} species on the solid phase.

110 The growth and decay rates of each bacteria group included in CWM1
111 (Table 2) are described using Monod expressions (Monod, 1949), to which
112 the product of two logistic expressions was added (Eq. 4):

$$f_{GL} = \left(1 - \frac{M_{bio}}{M_{bio,max}}\right) \left(1 - \frac{M_{X_{If}}}{M_{cap}}\right) \quad (4)$$

113 Where, $M_{bio,max}$ and M_{cap} [M] are two empirical parameters represent-
114 ing, respectively, the maximum microbial biomass (carrying capacity) and
115 the maximum amount of particulate solids that can be maintained in a rep-
116 resentative volume of granular material. On the other hand, M_{bio} and $M_{X_{If}}$
117 [M] are, respectively, the sum of the total microbial biomass and the actual
118 mass of immobile X_I present in the representative volume.

119 Table 3 shows the biokinetic processes rates of the BIO_PORE model
120 resulting from the inclusion of Eq. 4 to the original formulation of CWM1.

121 Notice that all kinetic parameters of CWM1 are interpolated to account
122 for water temperature variations.

123 *2.1.2. Model domain*

124 The model domain corresponds to a longitudinal section of wetland *C2*
125 of the pilot system described in García et al. (2004a,b) (Figure 1). This
126 wetland was 10.3 *m* long and 5.3 *m* wide, with a bottom slope of 1%. The
127 granular media consisted of fine granitic gravel ($D_{60} = 3.5$ *mm*, $C_u = 1.7$,
128 initial porosity $n = 40\%$) with a depth of approximately 0.6 *m* at the inlet
129 and 0.7 *m* at the outlet.

130 *2.1.3. Initial and boundary conditions*

131 Experimentally measured flow-rates, ranging from 1.1 to 2.45 $m^3 \cdot d^{-1}$ were
132 imposed at boundary 3 (inlet) and a hydraulic head of 0.5 *m* at boundary 5
133 (outlet). An hydraulic head of 0.5 *m* was set as the initial condition for the
134 Darcy equation.

135 For the transport equations, inflow concentrations of the components
136 listed in Table 1, which were obtained from field measurements (see Section
137 2.1.4), were imposed at boundary 3. An outflow boundary condition was
138 imposed at boundary 5. The initial concentrations of all substrates within
139 the wetland were set to 10 $mg \cdot L^{-1}$.

140 The initial concentrations of the different bacteria groups within the wet-
141 land were set to 1 $mg \cdot L^{-1}$ to recreate start-up conditions.

142 *2.1.4. Experimental data and parameter values*

143 The experimental data measured along the first year of operation of the
144 pilot wetland and used to feed the model consisted of: 39 values of flow
145 rate, 32 values of water temperature, 31 values of inflow COD and 33 values
146 of inflow $NH_4 - N$. The fractioning of the inflow COD was made using

147 recommended values for primary effluents in ASMs (Henze et al., 2000): 15%
148 S_F , 50% X_{Sm} (0% X_{Sf}), 20% S_A , 5% S_I and 10% X_{Im} (0% X_{If}). 28 values
149 of COD and 34 of $NH_4 - N$ were measured at the outlet of the pilot wetland
150 during the same period of time.

151 The inflow concentrations of the rest of components of CWM1 ($0 \text{ mg} \cdot \text{L}^{-1}$
152 for S_{NO} and S_{H_2S} , and $72 \text{ mg} \cdot \text{L}^{-1}$ for S_{SO_4}) correspond to mean values
153 measured from different samples extracted from the same pilot wetland by
154 García et al. (2004b). Inflow oxygen concentration was set to zero, since DO
155 concentration in primary treated wastewater is usually very small (Tyroller
156 et al., 2010).

157 Values of the hydraulic and hydrodynamic parameters obtained by Samsó
158 and García (2013a) and utilised for all simulations are shown in Table 4.

159 2.2. Mesh Optimization

160 After a previous detailed study with simplified versions of the model
161 (progressively increasing the number of functional bacterial groups)(results
162 not shown), 5 triangular meshes of different elements densities (Table 5)
163 were chosen to perform the mesh optimization of the complete model (with
164 all bacteria groups listed in Table 2). Among those meshes, $M_{0.1}$ was the
165 coarsest, $M_{0.025}$ the most dense and $M_{BIO-PORE}$ was the one used by Samsó
166 and García (2013a,b). $M_{BIO-PORE}$ was the only mesh with a predefined
167 numbers of elements at boundaries 3 (20 elements), 4 (550 elements) and
168 5 (7 elements), which were reckoned as the most critical ones numerically
169 (large concentration gradients).

170 Simulated effluent concentrations of COD (sum of S_F , S_A , S_I , X_{Sm} and
171 X_{Im}) and S_{NH} , as well as the simulation time were recorded for all different

172 meshes. Although the simulated effluent concentrations of the rest of model
173 components could have also been studied, only COD and S_{NH} were used for
174 the sake of brevity and because these are the two most widely used water
175 quality indicators. The Sum of Squared Errors (SSE) for the effluent COD
176 and S_{NH} curves for all different meshes were calculated using the coarser
177 mesh ($M_{0.1}$) as a reference, to showcase the progressive accuracy gains with
178 increasing mesh densities. The optimal mesh corresponds to that after which
179 any further increments on the number of elements does not produce notable
180 improvements on the numerical accuracy of the solution (SSE remains fairly
181 constant). Moreover, for evident practical reasons, the optimal mesh is also
182 that with the shortest computational cost/time.

183 *2.3. Parameter sensitivity*

184 The sensitivity of M_{bio_max} and M_{cap} was studied by giving three different
185 values to each of the two parameters (Table 6) and running a different sim-
186 ulation for each different pair (9 simulations in total) (Table 7). The reason
187 for selection the values of Table 6 are discussed later in the text. Notice that
188 the range of variability of M_{cap} was smaller than that of M_{bio_max} . In the first
189 case, the highest value of M_{cap} was 3 times the smallest, whereas for M_{bio_max}
190 the highest was 5 times the smallest.

191 The sensitivity of the two parameters was determined qualitatively by
192 comparing the effluent concentrations of COD among them with the 9 dif-
193 ferent parameter pairs. The same is done for the simulated effluent concen-
194 trations of S_{NH} . A qualitative comparison was made between the effluent
195 concentrations of COD and S_{NH} obtained with each parameter pair.

196 The mesh used to execute all these processes was the optimum mesh
197 obtained in the previous step (Section 2.2).

198 *2.4. Launching simulations and hardware specifications*

199 In this work two different computers were used. For the mesh optimiza-
200 tion procedure, a desktop PC was used. This computer features an Intel[®]
201 Xenon[®] E5-1620 processor with 4 cores (8 threads) running at a frequency
202 of 3600GHz and 16 GB of RAM memory. The Linux kernel and COMSOL
203 Multiphysics[™] versions installed on this computer were 3.2.0-56 and v4.3b,
204 respectively.

205 On the other hand, for the sensitivity analysis the cluster functionalities
206 of COMSOL Multiphysics[™] were used to run several simulations in parallel
207 on a high-end multi-processor computer. This computer consisted of 4 CPUs
208 *AMD Opteron[™] 6140* with 8 cores each (2.6 GHz), a total of 64 GB of RAM
209 memory and run Linux Kernel 2.6.38. The COMSOL Multiphysics[™] version
210 installed in this machine was v4.2a. Since this machine was shared with
211 other researchers, only 3 parallel simulations (using 4 CPU cores each) were
212 launched at a time (see Figure 2). Therefore only 12 cores, out of the 32
213 available, were utilized. A bash script was used to automatically launch each
214 different batch of 3 parallel simulations without any intervention.

215 **3. Results and discussion**

216 *3.1. Mesh optimization*

217 In the current study the focus was not on how well or bad simulated ef-
218 fluent concentrations fit experimental data, since that discussion was already

219 made in Samsó and García (2013a), but rather on the comparison of the
220 simulation results obtained with different meshes. However, note that the
221 poor fitting of the simulated effluent COD and S_{NH} with experimental data
222 at the beginning of all simulations (Figures 3 and 4), was due to the fact that
223 initial bacteria and accumulated solids concentrations were underestimated.
224 However, after around 70 days of simulated time, the fitting improved.

225 Figures 3 and 4 show that the effluent pollutant concentrations of COD
226 and S_{NH} obtained with the different meshes (Table 5) are visually different
227 in some cases.

228 Finer FE meshes provide more accurate numerical results. Thus in our
229 study, mesh $M_{0.025}$, with a maximum element size of 2,5 *cm* and a total
230 number of 28884 elements is the one giving more accurate results. Despite
231 even better results could have been obtained by further refining the mesh, the
232 total simulation time of $M_{0.025}$ (16 hours and 18 minutes) was already seen as
233 too large for practical reasons. Moreover, refining the mesh to such an extent
234 would only make sense if field data, which is given as model input and later
235 used to compare with simulated effluent concentrations, had been gathered
236 in higher frequency. In fact, Figure 4 clearly shows that almost identical
237 results were obtained for simulated effluent S_{NH} concentrations with meshes
238 $M_{BIO-PORE}$ and $M_{0.03}$ which account for c.a. 30% less elements than $M_{0.025}$.
239 That is also confirmed with the tendency of the SSE for S_{NH} (Figure 6),
240 which shows clear signs of stabilization already with meshes $M_{BIO-PORE}$ and
241 $M_{0.03}$. Therefore, further mesh refinements would not improve the description
242 of the effluent S_{NH} concentrations. In the case of COD (Figure 3), although
243 the differences between the curves obtained with different meshes were higher

244 than for S_{NH} , and the SSE still did not show signs of stabilization (Figure
245 5), the maximum difference of effluent COD concentrations obtained with
246 meshes $M_{BIO-PORE}$ and $M_{0.025}$ was lower than $15 \text{ mgCOD} \cdot \text{L}^{-1}$, which was
247 only around 8% the maximum effluent COD concentration simulated with
248 mesh $M_{0.025}$. Moreover, note that the reference mesh $M_{0.1}$ was already fine
249 (1860 elements) and thus reaching SSE stability is more difficult than if a
250 coarser mesh had been used as a reference to calculate SSE.

251 Table 8 shows that, in general, the simulation time increased with in-
252 creasing mesh densities. $M_{BIO-PORE}$ was the exception, and although it had
253 213 less elements than $M_{0.03}$ the former took 25 *minutes* more than the later
254 to reach the final solution (see Table 8). The most likely reason for that is
255 that the mesh element quality of $M_{BIO-PORE}$ was lower than that of $M_{0.03}$
256 and thus the solver algorithm required a few more iterations at every time
257 step to reach a solution. In fact, $M_{BIO-PORE}$ was the one with the second
258 largest maximum element size (0.05 *cm*), only after $M_{0.1}$, but in contrast it
259 was the mesh with the highest elements densities in boundaries 3, 4 and 5,
260 which were the ones accounting for the highest concentration gradients. The
261 relation between number of elements and simulated time can also be observed
262 in Figures 5 and 6, and shows that a linear relationship ($R^2 = 0.97$) exists
263 between the two.

264 According to these results, the mesh with a better compromise between
265 numerical accuracy and simulation time was $M_{0.03}$. The results obtained
266 with mesh $M_{BIO-PORE}$ were almost as good as those obtained with $M_{0.03}$
267 (see Figures 3, 4, 5 and 6), and since mesh $M_{BIO-PORE}$ had already been
268 used successfully in a previous work (Samsó and García, 2013a), it was chosen

269 as the one to be used for the sensitivity analysis.

270 3.2. Parameter sensitivity

271 Despite BIO_PORE includes more than 50 parameters, only the sensitiv-
272 ity of M_{bio_max} and M_{cap} was analysed because they are two new additions to
273 the formulation of CWM1. Moreover, the sensitivity of the different parame-
274 ters of CWM1 has already been studied in other works (Mburu et al., 2012).
275 Note that the type of analysis carried out in this work is a local sensitiv-
276 ity analysis, which only addresses sensitivity relative to the point estimates
277 chosen and not for the entire parameter distribution.

278 The first parenthesis of Eq. 4 (involving M_{bio_max}) limits the maximum
279 concentration of bacteria that each pore of the granular media can hold
280 (carrying capacity) by stopping the growth of bacteria once M_{bio} reaches the
281 value of M_{bio_max} . The second parenthesis works in the same way, but M_{cap}
282 corresponds to the maximum amount of particulate solids (X_{Sf} and X_{If})
283 porosity can hold, and bacterial growth stops once $M_{X_{If}} = M_{cap}$.

284 The values given to parameters M_{bio_max} were chosen based on our pre-
285 vious experiences with the BIO_PORE model, since no literature values for
286 these parameters exist for CWs. In fact the intermediate value of this pa-
287 rameter used in the current work was that obtained from the calibration of
288 the model in Samsó and García (2013a), and the other two were chosen to
289 be at a sound distance from the first. On the other hand, the amount of
290 accumulated solids in horizontal subsurface flow CWs presents a great vari-
291 ability depending on the COD and TSS loading rates and on the turn-over
292 rates. Measurements carried out by Caselles-Osorio et al. (2007) in 6 full-
293 scale horizontal subsurface flow CWs showed that accumulated solids ranged

294 from as low as $2.3 \text{ kgVS} \cdot \text{m}^{-2}$ up to $57.3 \text{ kgVS} \cdot \text{m}^{-2}$ (between around 6 and
295 $162 \text{ kgCOD} \cdot \text{m}^{-3}$, considering an average wetland depth of 0.5 m and that
296 $1 \text{ gVS} \approx 1.42 \text{ gCOD}$ (Samsó and García, 2014a)). In this study we selected
297 the values of M_{cap} to be in the lower part of that range, since the gravel size
298 of the pilot system was quite fine ($D_{60} = 3.5 \text{ mm}$ and $C_u = 1.7$).

299 Results indicate that M_{bio_max} and M_{cap} are both very sensitive parame-
300 ters since they had a large impact on the simulated concentrations of COD
301 (Figure 7) and S_{NH} (Figure 8). At the beginning of all simulations, effluent
302 concentrations obtained with the different pairs of M_{bio_max} and M_{cap} were
303 very similar, and it was not until around simulated day 60 that they started
304 diverging. Figures 7 and 8 show that both for COD and S_{NH} the most sen-
305 sitive parameter was M_{cap} , and the higher its value, and thus the higher the
306 capacity of porosity to retain particulate solids (X_{If} and X_{Sf}), the lower the
307 effluent concentrations of the two pollutants. A possible reasoning for this
308 behaviour is that for high values of M_{cap} the amount of slowly biodegradable
309 particulate COD (X_{Sf}) that can be reached in the granular media is much
310 higher than that the maximum bacteria biomass present in the same loca-
311 tion (which is limited by the value of M_{bio_max}) can biodegrade, and so they
312 accumulate. Therefore this accumulated organic matter, which also contains
313 a fraction of organic nitrogen, is retained within the system and does not
314 add to the concentrations of COD and S_{NH} measured at the outlet.

315 On the other hand, although perturbations of the M_{bio_max} value pro-
316 duced observable changes in the effluent COD and S_{NH} concentrations, these
317 changes were smaller than those produced by changing the value of M_{cap} . Re-
318 garding the effluent COD concentrations (Figure 7), for $M_{cap} = 15 \text{ kgVS} \cdot \text{m}^{-3}$

319 and $M_{cap} = 10 \text{ kgVS} \cdot \text{m}^{-3}$, the higher the value of $M_{bio.max}$ the higher the
 320 effluent concentrations of COD. This can be explained by the fact that the
 321 higher the maximum concentrations of biomass in a specific point of the
 322 granular media, the larger proportion of the accumulated X_{Sf} can be hy-
 323 drolised and thus released through the outlet (in the form of S_F , S_A , S_I
 324 and S_{NH}) increasing the effluent concentrations of COD and S_{NH} . On the
 325 contrary, for $M_{cap} = 5 \text{ kgVS} \cdot \text{m}^{-3}$, the tendency is different and the effluent
 326 concentrations are higher for $M_{bio.max} = 0.3 \text{ kgVS} \cdot \text{m}^{-3}$, intermediate for
 327 $M_{bio.max} = 0.1 \text{ Kg} \cdot \text{m}^{-3}$ and the lowest for $M_{bio.max} = 0.5 \text{ KgVS} \cdot \text{m}^{-3}$.
 328 Therefore no clear pattern can be extracted for $M_{bio.max}$ when the values of
 329 M_{cap} are relatively small.

330 Regarding S_{NH} (Figure 8), for $M_{cap} = 15 \text{ kgVS} \cdot \text{m}^{-3}$, the effluent
 331 concentrations of this component are almost the same regardless of the
 332 value of $M_{bio.max}$. For the intermediate value of M_{cap} ($10 \text{ kgVS} \cdot \text{m}^{-3}$),
 333 $M_{bio.max} = 0.5 \text{ kgVS} \cdot \text{m}^{-3}$ gives the highest effluent concentration, while
 334 for $M_{bio.max} = 0.3 \text{ kgVS} \cdot \text{m}^{-3}$ and $M_{bio.max} = 0.1 \text{ kgVS} \cdot \text{m}^{-3}$ the ef-
 335 fluent concentrations are almost identical. For the lowest value of M_{cap}
 336 ($5 \text{ kgVS} \cdot \text{m}^{-3}$) there are also differences between the curves, but in this
 337 case $M_{bio.max} = 0.1 \text{ kgVS} \cdot \text{m}^{-3}$ gives the lowest effluent concentrations of
 338 S_{NH} while $M_{bio.max} = 0.5 \text{ kgVS} \cdot \text{m}^{-3}$ and $M_{bio.max} = 0.3 \text{ kgVS} \cdot \text{m}^{-3}$ give
 339 almost the same results.

340 Therefore, contrarily to what happened for M_{cap} , for $M_{bio.max}$ although
 341 some patterns can be detected for the effluent COD concentrations, there
 342 is not a clear distinguishable tendency regarding the effluent concentrations
 343 of S_{NH} obtained with the different values of this parameter. However, the

344 higher the value of M_{cap} , the larger the difference between the effluent con-
345 centrations obtained with the different values o $M_{bio.max}$.

346 4. Conclusions

347 In this work we performed a mesh optimization procedure in order to
348 reduce the simulation time (while maintaining similar numerical accuracy)
349 for subsequent simulations, and we also performed a local sensitivity analysis
350 of parameters M_{cap} and $M_{bio.max}$.

351 Results of the mesh optimization procedure indicated that for homoge-
352 neous meshes, a positive linear relationship existed between the number
353 of elements and simulated time. The best compromise between numeri-
354 cal accuracy and computational cost was obtained with meshes $M_{0.03}$ and
355 $M_{BIO-PORE}$. Therefore $M_{BIO-PORE}$ was selected as the optimal mesh to
356 carry out the sensitivity analysis.

357 Despite the range of values given to M_{cap} was smaller than that given to
358 $M_{bio.max}$, the former parameter proved to be the most sensitive one, and the
359 higher its value the lower the simulated effluent concentrations of COD and
360 S_{NH} of the wetland. This was due to the fact that for larger values of M_{cap} ,
361 more slowly biodegradable solids can accumulate in a specific point, and if
362 there is not enough bacteria to hydrolyse them, they are not released and
363 thus the effluent concentrations of COD and S_{NH} does not increase.

364 On the other hand, from the values given to $M_{bio.max}$ no clear recognisable
365 pattern on the effluent concentrations of COD and S_{NH} could be observed.

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Table 1: Description of the components considered in BIO_PORE model . S_i are dissolved species (all in the aqueous phase by definition) and X_i are particulate species (either in aqueous or solid phase).

Component	Description	Unit	Phase
S_O	Dissolved oxygen	$mgCOD \cdot L^{-1}$	Aqueous
S_F	Soluble fermentable COD	$mgCOD \cdot L^{-1}$	Aqueous
S_A	Fermentation products as acetate as COD	$mgCOD \cdot L^{-1}$	Aqueous
S_I	Inert soluble COD	$mgCOD \cdot L^{-1}$	Aqueous
X_{Sm}	Aqueous slowly biodegradable particulate COD	$mgCOD \cdot L^{-1}$	Aqueous
X_{Sf}	Solid slowly biodegradable particulate COD	$mgCOD \cdot L^{-1}$	Solid
X_{Im}	Aqueous inert particulate COD	$mgCOD \cdot L^{-1}$	Aqueous
X_{If}	Solid inert particulate COD	$mgCOD \cdot L^{-1}$	Solid
S_{NO}	Nitrite and nitrate nitrogen	$mgN \cdot L^{-1}$	Aqueous
S_{NH}	Ammonium and ammonia nitrogen	$mgN \cdot L^{-1}$	Aqueous
S_{SO4}	Sulphate sulphur	$mgS \cdot L^{-1}$	Aqueous
S_{H2S}	Dihydrogensulphide sulphur	$mgS \cdot L^{-1}$	Aqueous

Table 2: Functional bacterial groups considered in BIO_PORE. Bacteria concentrations are given in units of COD ($mgCOD \cdot L^{-1}$).

Component	Description	Phase
X_H	Heterotrophic bacteria	Solid
X_A	Autotrophic nitrifying bacteria	Solid
X_{FB}	Fermenting bacteria	Solid
X_{AMB}	Acetotrophic methanogenic bacteria	Solid
X_{ASRB}	Acetotrophic sulphate reducing bacteria	Solid
X_{SOB}	Sulphide oxidising bacteria	Solid

Table 3: Processes rates in $mg \cdot d^{-1}$ (adapted from Langergraber et al. (2009)).

j	Process	Process rate ρ_j
1	Hydrolysis X_{Sf}	$k_h \left[\frac{\frac{X_{Sf}}{X_H + X_{FB}}}{K_X \left(\frac{X_{Sf}}{X_H + X_{FB}} \right)} \right] (X_H + \eta_h X_{FB})$
2	Aerobic growth of X_H on S_F	$\mu_H \cdot f_{GL} \left(\frac{S_F}{K_{SFH} + S_F} \right) \left(\frac{S_F}{S_F + S_A} \right) \left(\frac{S_O}{K_{SOH} + S_O} \right) \left(\frac{S_{NH}}{K_{SNHH} + S_{NH}} \right) \left(\frac{K_{SH2SH}}{K_{SH2SH} + S_{H2S}} \right) X_H$
3	Anoxic growth of X_H on S_F	$\eta_g \cdot \mu_H \cdot f_{GL} \left(\frac{S_F}{K_{SFH} + S_F} \right) \left(\frac{S_F}{S_F + S_A} \right) \left(\frac{K_{SOH}}{K_{SOH} + S_O} \right) \left(\frac{S_{NO}}{K_{SNOH} + S_{NO}} \right) \left(\frac{S_{NH}}{K_{SNHH} + S_{NH}} \right) \left(\frac{K_{SH2SH}}{K_{SH2SH} + S_{H2S}} \right) X_H$
4	Aerobic growth of X_H on S_A	$\mu_H \cdot f_{GL} \left(\frac{S_A}{K_{SAH} + S_A} \right) \left(\frac{S_A}{S_F + S_A} \right) \left(\frac{S_O}{K_{SOH} + S_O} \right) \left(\frac{S_{NH}}{K_{SNHH} + S_{NH}} \right) \left(\frac{K_{SH2SH}}{K_{SH2SH} + S_{H2S}} \right) X_H$
5	Anoxic growth of X_H on S_A	$\eta_g \mu_H \cdot f_{GL} \left(\frac{S_A}{K_{SAH} + S_A} \right) \left(\frac{S_A}{S_F + S_A} \right) \left(\frac{K_{SOH}}{K_{SOH} + S_O} \right) \left(\frac{S_{NO}}{K_{SNOH} + S_{NO}} \right) \left(\frac{S_{NH}}{K_{SNHH} + S_{NH}} \right) \left(\frac{K_{SH2SH}}{K_{SH2SH} + S_{H2S}} \right) X_H$
6	Lysis of X_H	$b_X X_H$
7	Aerobic growth of X_A on S_{NH}	$\mu_A \cdot f_{GL} \left(\frac{S_{NH}}{K_{SNHA} + S_{NH}} \right) \left(\frac{S_O}{K_{SOA} + S_O} \right) \left(\frac{K_{SH2SA}}{K_{SH2SA} + S_{H2S}} \right) X_A$
8	Lysis of X_A	$b_A X_A$
9	Growth of X_{FB}	$\mu_{FB} \cdot f_{GL} \left(\frac{S_F}{K_{SFFB} + S_F} \right) \left(\frac{K_{SH2SFB}}{K_{SH2SFB} + S_{H2S}} \right) \left(\frac{K_{SOFB}}{K_{SOFB} + S_O} \right) \left(\frac{K_{SNOFB}}{K_{SNOFB} + S_{NO}} \right) \left(\frac{S_{NH}}{K_{SNHFB} + S_{NH}} \right) X_{FB}$
10	Lysis of X_{FB}	$b_{FB} X_{FB}$
11	Growth of X_{AMB}	$\mu_{AMB} \cdot f_{GL} \left(\frac{S_A}{K_{SAMB} + S_A} \right) \left(\frac{K_{SH2SAMB}}{K_{SH2SAMB} + S_{H2S}} \right) \left(\frac{K_{SOAMB}}{K_{SOAMB} + S_O} \right) \left(\frac{K_{SNOAMB}}{K_{SNOAMB} + S_{NO}} \right) \left(\frac{S_{NH}}{K_{SNHAMB} + S_{NH}} \right) X_{AMB}$
12	Lysis of X_{AMB}	$b_{AMB} X_{AMB}$
13	Growth of X_{ASRB}	$\mu_{ASRB} \cdot f_{GL} \left(\frac{S_A}{K_{SAASRB} + S_A} \right) \left(\frac{S_{SO4}}{K_{SO4ASRB} + S_{SO4}} \right) \left(\frac{K_{SH2SASRB}}{K_{SH2SASRB} + S_{H2S}} \right) \left(\frac{K_{SOASRB}}{K_{SOASRB} + S_O} \right) \left(\frac{K_{SNOASRB}}{K_{SNOASRB} + S_{NO}} \right) \left(\frac{S_{NH}}{K_{SNHASRB} + S_{NH}} \right) X_{ASRB}$
14	Lysis of X_{ASRB}	$b_{ASRB} X_{ASRB}$
15	Aerobic growth of X_{SOB} on S_{H2S}	$\mu_{SOB} \cdot f_{GL} \left(\frac{S_{H2S}}{K_{SH2SSOB} + S_{H2S}} \right) \left(\frac{S_O}{K_{SOSOB} + S_O} \right) \left(\frac{S_{NH}}{K_{SNHSOB} + S_{NH}} \right) X_{SOB}$
16	Anoxic growth of X_{SOB} on S_{H2S}	$\mu_{SOB} \cdot f_{GL} \cdot \eta_{SOB} \left(\frac{S_{H2S}}{K_{SH2SSOB} + S_{H2S}} \right) \left(\frac{S_{NO}}{K_{SNOSOB} + S_{NO}} \right) \left(\frac{K_{SOSOB}}{K_{SOSOB} + S_O} \right) \left(\frac{S_{NH}}{K_{SNHSOB} + S_{NH}} \right) X_{SOB}$
17	Lysis of X_{SOB}	$b_{SOB} X_{SOB}$

Table 4: Values of the hydraulic and hydrodynamic parameters of the granular media.

Parameter	Description	Unit	Value
α_L	Longitudinal dispersivity	m	0.05
α_T	Transverse dispersivity	m	0.005
K	Hydraulic conductivity	$m \cdot d^{-1}$	50

Table 5: Meshes used in the mesh optimization procedure.

Mesh	Maximum element size (m)	Number of elements
$M_{0.1}$	0.1	1860
$M_{0.04}$	0.04	11446
$M_{BIO-PORE}$	0.05 ^a	19851
$M_{0.03}$	0.03	20064
$M_{0.025}$	0.025	28884

^aNote that $M_{BIO-PORE}$ was built with a maximum element size of 0.05 m but fixing the number of elements at boundaries 3 (20 elements), 4 (550 elements) and 5 (7 elements), and its total number of elements is very similar to that of $M_{0.03}$.

Table 6: Values for M_{cap} and $M_{bio-max}$.

Value	$M_{cap}(kgVS \cdot m^{-3})$	$M_{bio-max}(kgVS \cdot m^{-3})$
Minimum	3	0.1
Intermediate	5	0.3
Maximum	15	0.5

Table 7: Combinations of M_{cap} and M_{bio_max} values for the different simulations carried out for the local sensitivity analysis.

Parameter	$M_{cap}(kgVS \cdot m^{-3})$	$M_{bio_max}(kgVS \cdot m^{-3})$
S_1	15	0.5
S_2	15	0.3
S_3	15	0.1
S_4	10	0.5
S_5	10	0.3
S_6	10	0.1
S_7	5	0.5
S_8	5	0.3
S_9	5	0.1

Table 8: Number of elements and simulation time for each of the meshes used for mesh optimization.

Mesh	Number of triangular elements	Simulation time (hours)
$M_{0.1}$	1860	1.04
$M_{0.04}$	11446	5.41
$M_{BIO-PORE}$	19851	9.96 ^a
$M_{0.03}$	20064	9.53
$M_{0.025}$	28884	16.30

^aNotice that although $M_{BIO-PORE}$ had fewer elements than $M_{0.03}$ its simulation time was slightly higher. Notice as well that $M_{BIO-PORE}$ was the only one of the selected meshes with higher elements density in boundaries 3, 4 and 5 (see Figure 1).

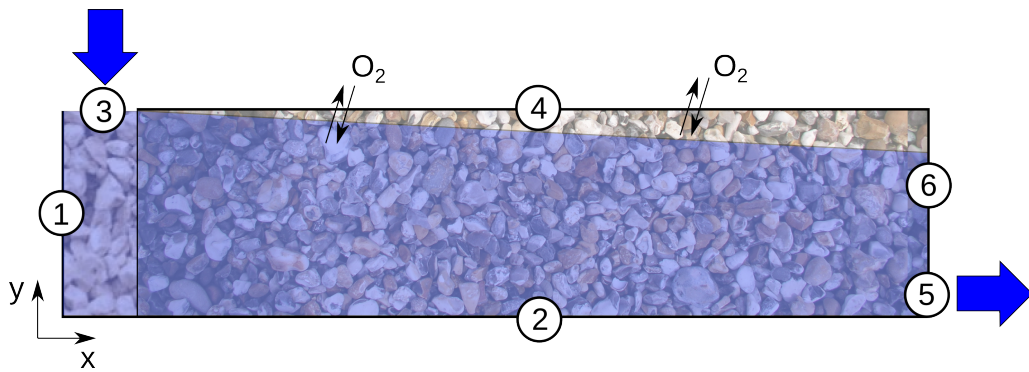


Figure 1: Model domain, representing a longitudinal section of wetland *C2* in García et al. (2004a), and numbers of the different boundaries (obtained from Samsó and García (2013a)). The numbers identify the different boundaries of the domain. Numbers 1 and 5 correspond to the inlet and outlet sections, respectively.

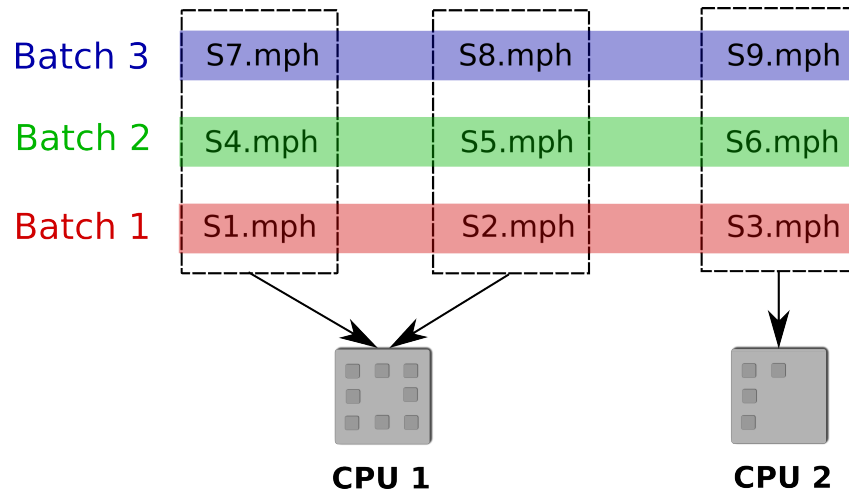


Figure 2: CPU and processor utilisation in the high-end multi-processor computer during the sensitivity analysis. Model files built in COMSOL MultiphysicsTM have *mph* extension. Three batches of 3 parallel simulations, each with a different $M_{cap} - M_{bio_max}$ pair (see Table 7), were launched. Each simulation took up only 4 processor cores. All cores of CPU1 were used, while CPU2 was only loaded to a 50%.

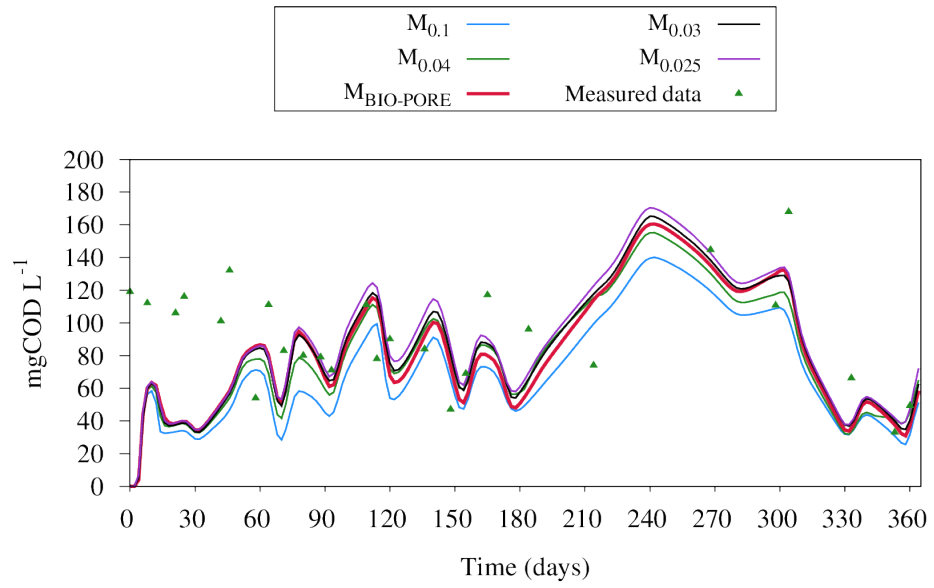


Figure 3: Simulated effluent COD concentrations obtained from the mesh optimization procedure with the meshes of Table 5.

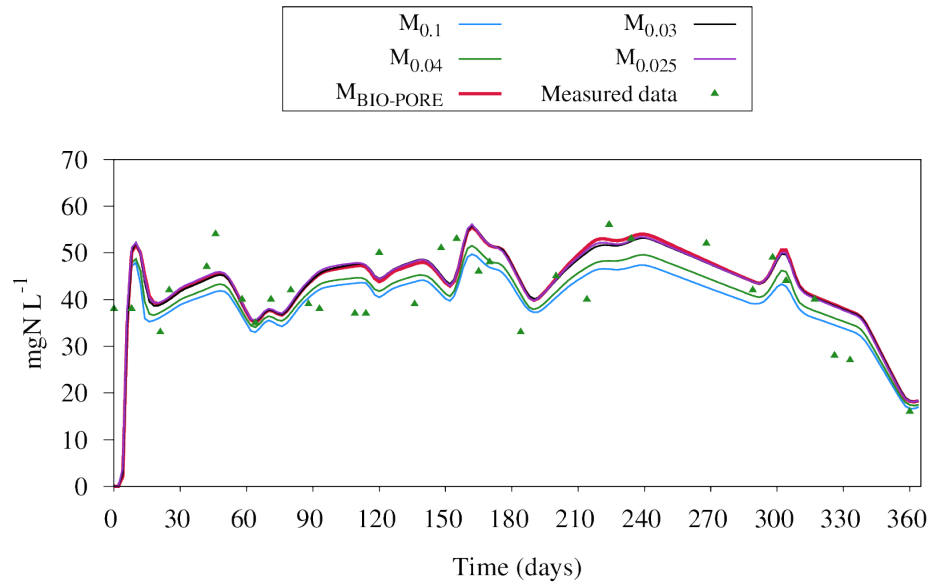


Figure 4: Simulated effluent S_{NH} concentrations obtained from the mesh optimization procedure with the meshes of Table 5.

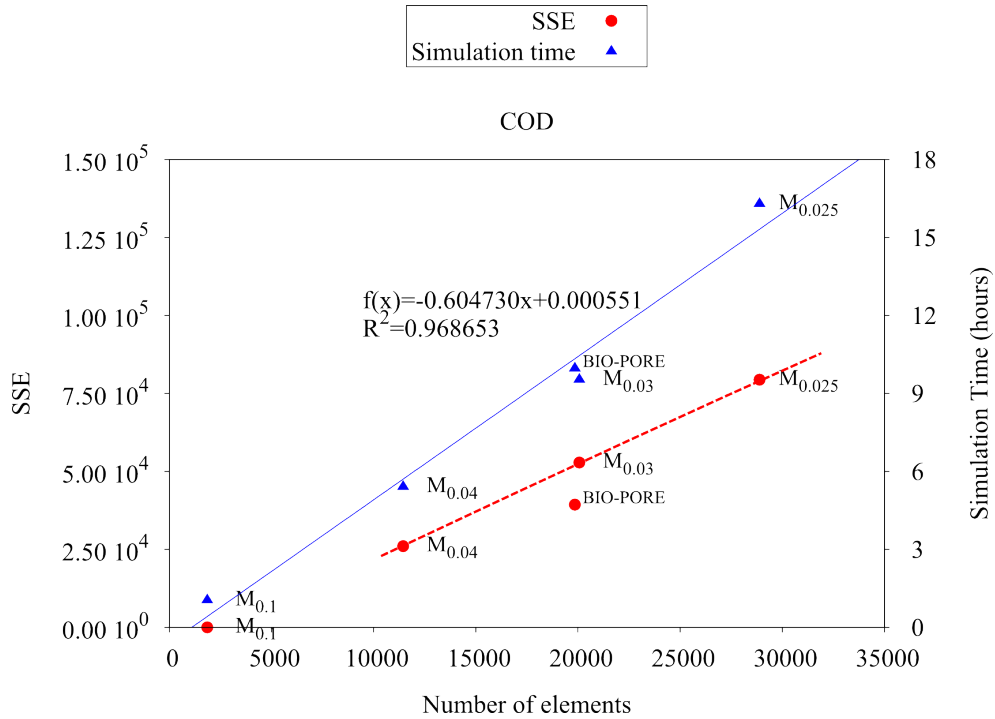


Figure 5: Sum of Squared Errors (SSE) (left y-axes) and simulation time (right y-axes) for the simulated effluent COD concentrations obtained with meshes of different elements density (see Table 6). The blue line shows the positive linear relationship ($R^2 = 0.97$) between the number of triangular elements of the mesh and the simulation time. The dotted red line was drawn to show that the SSE does not tend to a constant value with increasing number of elements. Notice that this line was drawn neglecting the SSE of $M_{BIO-PORE}$ since this mesh was built with a pre-set number of elements in specific domain boundaries. Mesh $M_{0.1}$ was also neglected, since it was the reference mesh, from which all SSE plotted in this figure were calculated.

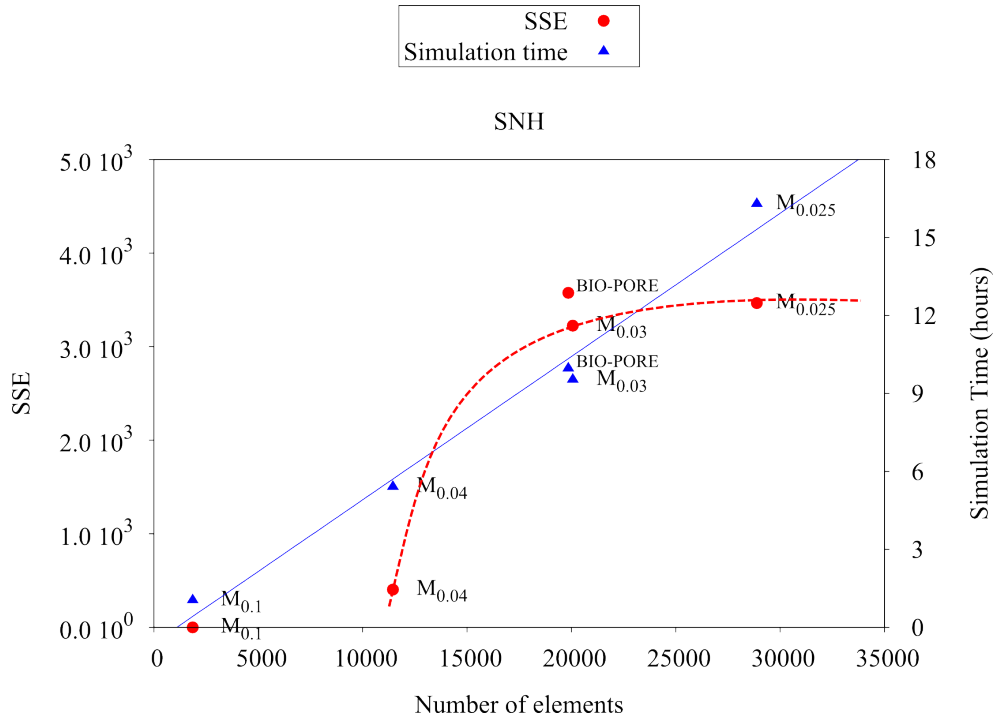


Figure 6: Sum of Squared Errors (SSE) (left y-axes) and simulation time (right y-axes) for the simulated effluent S_{NH} concentrations obtained with meshes of different elements density (see Table 6). The R^2 of the linear regression of the Simulation time is the same as in Figure 5, since all data shown in both figures was obtained from the same simulations (each focusing on different model outputs). The dotted red line was drawn to show that for S_{NH} the SSE tends to a constant value with increasing number of elements. As in the previous figure, the SSE of meshes $M_{BIO-PORE}$ and $M_{0.1}$ were neglected to draw this line.

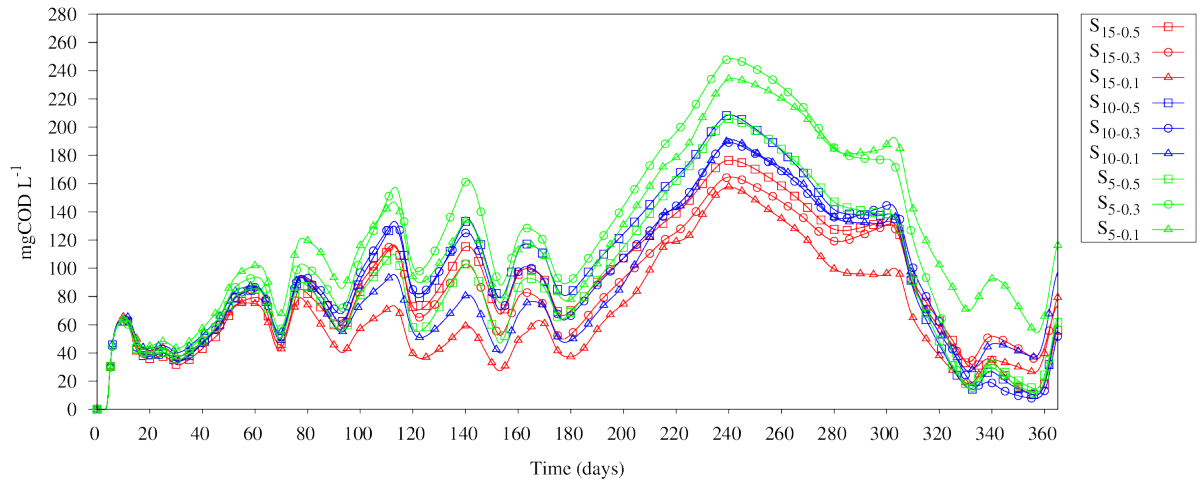


Figure 7: Effluent COD concentrations obtained with the combinations of M_{bio_max} and M_{cap} shown in Table 7.

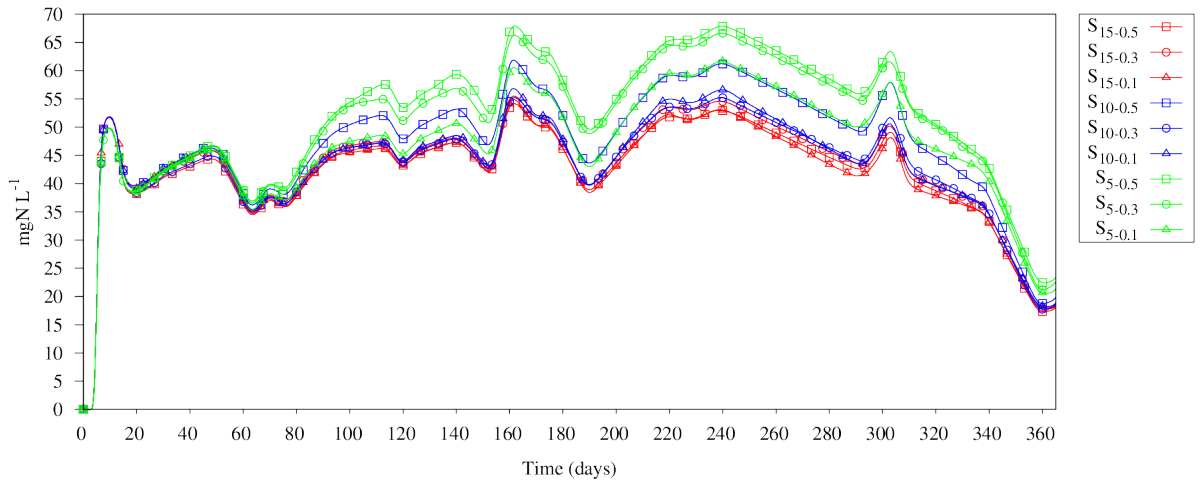


Figure 8: Effluent S_{NH} concentrations obtained with the combinations of $M_{bio-max}$ and M_{cap} shown in Table 7.