1	A Type 2 A/C ₂ plasmid carrying the <i>aacC4</i> apramycin resistance gene and the
2	erm(42) erythromycin resistance gene recovered from two Salmonella enterica
3	serovars
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26 Abstract

27 **Objective:** To determine the relationships between RepA/C_2 plasmids carrying several 28 antibiotic resistance genes found in isolates of Salmonella enterica serovars Ohio and 29 Senftenberg from pigs. 30 Methods: Illumina HiSeq was used to sequence seven S. enterica isolates. BLAST 31 searches identified relevant A/C_2 plasmid contigs, and contigs were assembled using 32 PCR. 33 Results: Two serovar Ohio isolates were ST329 and the five Senftenberg isolates were 34 ST210. The A/C₂ plasmids recovered from the seven isolates belong to Type 2 and 35 contain two resistance islands. Their backbones were closely related, differing by five or 36 fewer single nucleotide polymorphisms. The sul2-containing resistance island ARI-B is 37 19.9 kb and also contains the kanamycin and neomycin resistance gene aphA1, the 38 tetracycline resistance gene tetA(D), and an erythromycin resistance gene, erm(42), not 39 previously seen in A/C_2 plasmids. A second 30.3 kb resistance island, RI-119, is in a 40 unique location in the A/C₂ backbone 8.2 kb downstream of rhs. RI-119 contained 41 genes conferring resistance to apramycin, netilmicin, tobramycin (*aacC4*), hygromycin 42 (hph), sulphonamides (sul1) and spectinomycin and streptomycin (aadA2). In one of the 43 seven plasmids, this resistance region contained two IS26-mediated deletions. A discrete 44 5.7 kb segment containing the *aacC4* and *hph* genes and bounded by IS26 on one side 45 and the IR of Tn5393 on the other was identified. 46 **Conclusions:** The presence of almost identical A/C_2 plasmids in two serovars indicates 47 a common origin. Type 2 A/C₂ plasmids continue to evolve via addition of new 48 resistance regions such as RI-119 and evolution of existing ones.

49

51 Introduction

52 Plasmids of the incompatibility groups A and C (later combined as A/C) were among the 53 earliest plasmids to be associated with antibiotic resistance in Gram-negative bacteria. 54 However, the sequence of the *repA* gene in the reference plasmid RA1 differs 55 significantly from that of most of the sequenced A/C plasmids, and they are now designated A/C₁ (RA1) or A/C₂ based on the *repA* gene sequence.¹ A/C₂ plasmids 56 57 contribute to multiple antibiotic resistance in Salmonella enterica, a major cause of 58 foodborne illness,²⁻⁶ and several A/C₂ plasmids from *S. enterica* serovars Newport, Heidelberg and Typhimurium have been sequenced.^{3-5, 7, 8} A/C₂ plasmids also mobilize 59 60 S. enterica genomic island 1 (SGI1) carrying genes conferring resistance to multiple antibiotics.9, 10 61 62 Recently, the A/C_2 group was subdivided into two types, Type 1 and Type 2

(Figure 1a), that diverged a long time ago.¹¹ They have accumulated extensive SNPs and differ by two insertions or deletions (i1 and i2 in Figure 1a) within the backbone and two regions where part of a large gene has been replaced (R1 and R2 in Figure 1a).¹¹ The replacements give rise to two versions of the *rhs* gene (*rhs1* and *rhs2*) and open reading frames between *traA* and *traC* that predict proteins of 1832 aa (orf1832) in Type 1 and 1847 aa (orf1847) in Type 2.

We previously reported an unusual class 1 integron-associated gene cassette configuration in seven *S. enterica* isolates, two of serovar Ohio and five of serovar Senftenberg sourced in Australia from pigs. These isolates all carried an A/C₂ plasmid, and also shared resistance to apramycin, gentamicin, kanamycin, neomycin, streptomycin, spectinomycin, sulfamethoxazole and tetracycline.⁶ They carry the *aacC4* gene⁶, which confers resistance to apramycin, netilmicin and tobramycin.¹² However, conjugative transfer of the shared resistance genes and the A/C₂ plasmids could not be 76 detected.⁶ Here, we have sequenced the seven isolates and completed the sequence of 77 the A/C_2 plasmids.

78

79 Materials and methods

80 DNA sequencing and sequence analysis

81 The *S. enterica* isolates examined in this study are described elsewhere.⁶ Genomic DNA

82 was isolated as described previously¹³ and was sequenced using an Illumina HiSeq

83 platform at the Australian Genome Research Facility. Paired-end reads of 100 bp were

- 84 assembled using Velvet,¹⁴ yielding between 100-200 contigs with an average read depth
- 85 of 47- to 60-fold. Contigs carrying parts of the A/C_2 backbone defined previously¹¹ were
- 86 recovered using standalone BLAST (<u>www.ncbi.nlm.nih.gov/books/NBK52640</u>). Contigs
- 87 containing resistance genes were identified using ResFinder 2.1.¹⁵ All junctions between
- 88 contigs were confirmed by PCR using 20 ng of genomic DNA. Amplicons were
- resolved by electrophoresis and sequenced as described previously.¹⁶ Sequencher 5.2.3
- 90 (Gene Codes Corporation, Ann Arbor, MI, USA) was used for the final sequence
- 91 assembly. Reading frames not annotated previously¹¹ were predicted using ORF Finder
- 92 (<u>www.ncbi.nlm.nih.gov/projects/gorf/</u>) and annotated manually. The sequence type (ST)
- 93 of each strain was determined using the Warwick MLST scheme
- 94 (http://mlst.warwick.ac.uk/mlst/dbs/Senterica).
- 95

96 Nucleotide sequence accession number

- 97 The complete sequence of a representative plasmid, pSRC119-A/C, is deposited in
- 98 GenBank under accession number KM670336.
- 99
- 100

101 **Results**

102 *pSRC119-A/C*

103 The sequence of pSRC119-A/C, the A/C_2 plasmid from isolate SRC119, was assembled 104 from five contigs by using PCR to link across four copies of IS26 (GenBank Accession 105 Number KM670336). pSRC119-A/C is 174,068 bp long, and the backbone (Figure 1b) 106 includes the i1 and i2 insertions together with R1-2 (orf1847) and R2-2 (rhs2), the Type 107 2 versions of R1 and R2, making it a Type 2 A/C_2 plasmid. It contains two resistance 108 regions at the locations shown in Figure 1b. The first resistance island contains the *sul2* gene, and islands at this position were recently named ARI-B.¹¹ In pSRC119-A/C a 109 110 deletion has removed 4477 bp from the adjacent plasmid backbone (corresponding to 111 bases 25590 - 30066 in pRMH760, GenBank accession number KF976462). The second 112 resistance island was named RI-119 and is 30,345 bp. We have recently shown that 113 additional resistance islands in Type 2 A/ C_2 plasmids are found in several different positions, mostly clustered within or around the *rhs* gene.¹¹ RI-119 is located further 114 115 away at a new position, 8.2 kb downstream of the *rhs* stop codon. It replaces an 891 bp 116 segment of the A/C₂ backbone (corresponding to bases 157537 - 158427 in pRMH760) 117 that includes part of the *uvrD* and *kfrA* genes.

118 ARI-B in pSRC119-A/C

119 ARI-B is 19.9 kb long. The boundary at the *sul2* (RH) end, defined by comparison to 120 pRMH760 that lacks an ARI-B island, is the same as in all other A/C₂ plasmids with 121 ARI-B. The left hand boundary is identical to that found in the plasmids pIP1202 and 122 pP99-018 (GenBank Accession Numbers NC_009141 and NC_008612, respectively) 123 both of which are A/C₂ Type 2. These three plasmids have a segment from one end of 124 GI*sul2*¹⁷ at one end and a 4.4 kb segment sharing 98.9% nucleotide identity with the 125 IncN plasmid R46 (GenBank Accession Number AY046276) at the other end (Figure 126 1c). In addition to sul2 (sulphonamide resistance), the GIsul2 fragment includes a 127 complete copy of the small mobile element CR2. In pSRC119-A/C, 4286 bp at the right end are identical to the *sul2* end of GI*sul2*¹⁷ (bases 3902209-3906494 in GenBank 128 129 Accession Number CP001918) and in pIP1202 and pP99-018 4920 bp of GIsul2 are 130 present (Figure 1c). Indeed, it appears that the island was originally formed via the 131 integration of GIsul2. However, the internal composition of ARI-B differs. ARI-B in 132 pSRC119-A/C also carries genes conferring resistance to kanamycin and neomycin 133 (aphA1b), tetracycline (tetA(D)) and a novel gene, erm(42), that confers resistance to erythromycin, tilmicosin and clindamycin¹⁸ and to gamithromycin and tildipirosin.¹⁹ The 134 erm(42) gene was only recently identified in Pasteurella multocida (GenBank Accession 135 Numbers FR734406 and CP003022),¹⁸ where it is part of an integrative, conjugative 136 element (ICE) that can transfer across species and genus boundaries.²⁰ In GenBank there 137 138 is only one other example of this resistance gene and it is found in *Photobacterium* 139 damselae (GenBank Accession Number AB601890). Comparison of the three 140 sequences revealed a shared boundary upstream of the gene but different divergence 141 points downstream (Figure 1d).

142 *RI-119*

The 30345 bp RI-119 (Figure 2a) is a mosaic that contains adjacent genes conferring resistance to apramycin, netilmicin and tobramycin (*aacC4*) and to hygromycin (*hph*), as well as the unusual class 1 integron configuration previously shown to carry an *aadA2* cassette conferring resistance to streptomycin and spectinomycin and the *sul1* gene conferring resistance to sulphonamides.⁶ IRt of the 6988 bp integron is at the right-hand boundary of the resistance island and IRi is separated from an IS26 by 20 bp of sequencederived from Tn21.

150	The remainder of RI-119 is 23357 bp, and at the left-hand boundary of the island
151	there are 364 bp from the <i>tnpA</i> end of Tn1721, with the IR adjacent to the A/C ₂
152	backbone sequence. The <i>aacC4</i> and <i>hph</i> genes are in a 5693 bp structure (bases 138374
153	to 144066 in GenBank Accession Number KM670336) bounded by IS26 at one end and
154	IR_{tnp} of Tn5393 at the other end (Figure 2b). This configuration is present in three other
155	plasmids, p9134, pPWD4_103, and pK1HV (GenBank Accession Numbers KF705205,
156	HQ114284 and HF545434, respectively) (Figure. 2c). Though the structure is within a
157	resistance island in each of these plasmids, the surrounding sequence is different,
158	suggesting that this may be a mobile unit that can spread between plasmids.
159	The IS26- <i>aacC4-hph-ΔtnpA5393 structure in pSRC119-A/C separates two parts</i>
159 160	The IS26- <i>aacC4-hph-ΔtnpA5393 structure</i> in pSRC119-A/C separates two parts of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft
160	of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft
160 161	of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft genome of <i>Klebsiella pneumonia</i> strain KPNIH18 (AKA10100038). The genes in this
160 161 162	of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft genome of <i>Klebsiella pneumonia</i> strain KPNIH18 (AKA10100038). The genes in this segment encode proteins predicted to be associated with plasmid replication or
160 161 162 163	of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft genome of <i>Klebsiella pneumonia</i> strain KPNIH18 (AKA10100038). The genes in this segment encode proteins predicted to be associated with plasmid replication or conjugative transfer. In RI-119, part of this segment has been inverted, probably due to
160 161 162 163 164	of a 13.5 kb segment sharing 99.8% nucleotide identity with one found in the draft genome of <i>Klebsiella pneumonia</i> strain KPNIH18 (AKA10100038). The genes in this segment encode proteins predicted to be associated with plasmid replication or conjugative transfer. In RI-119, part of this segment has been inverted, probably due to inversion of the segment between the two oppositely-oriented IS26 (Figure 2a). An

168 Closely related A/C₂ plasmids in different serovars

169 Closure of the A/C₂ plasmids from four additional Senftenberg isolates, SRC69, SRC91,

- 170 SRC102 and SRC103, and two Ohio isolates, SRC22, SRC74, revealed that the A/C_2
- 171 plasmids they carried were almost identical to pSRC119-A/C. The resistance islands

172 were in the same locations in all seven plasmids, and the backbones differed from

173 pSRC119-A/C by only 2-5 SNPs. Three SNPs were shared by the two Ohio isolates, and

a single SNP was shared by two of the four remaining Senftenberg isolates. However,

the plasmid from SRC102 had two IS26-mediated deletions relative to RI-119, one of

176 which has truncated the *hph* gene. These deletions have removed 5453 bp (bases

177 132921-138373 in GenBank Accession Number KM670336) and 10608 bp (bases

178 141081-151688 in GenBank Accession Number KM670336).

179 The five Senftenberg isolates were all ST210 (*aroC* 74, *dnaN* 6, *hemD* 70, *hisD*

180 8, *purE* 7, *sucA* 79, *thrA* 13) and the two Ohio isolates were ST329 (82-38-26-12-115-

181 78-70). ST329 and ST210 differ at all seven alleles, and though conjugation was not

182 detected,⁶ the A/C₂ plasmid has clearly transferred between the two *S. enterica* strains.

183

184 **Discussion**

185 The Type 2 A/C₂ plasmids recovered in this study have acquired three resistance genes

186 (*erm*(42), *aacC4* and *hph*) not previously seen in A/C_2 plasmids. These genes

187 complement genes conferring resistance to kanamycin, neomycin, tetracycline,

tobramycin, sulphonamides, spectinomycin and streptomycin. The *aacC4* and *hph* genes

are located in a discrete structure that appears to have moved as a unit into three

190 different plasmids.

191 The ARI-B island in pSRC119-A/C belongs to a specific sub-group of Type 2

192 plasmids that include a fragment from the IncN plasmid R46 in ARI-B and the *erm*(42)

193 gene has been incorporated into ARI-B. One member of this group (pP99-018) includes

194 only an ARI-B island whereas pSRC119-A/C and pIP1202 have each acquired an

additional resistance island.

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204	
205	Transparency declaration
206	None to declare.
207	
208	References
209	1. Carattoli A, Miriagou V, Bertini A et al. Replicon typing of plasmids encoding
210	resistance to newer beta-lactams. Emerg Infect Dis 2006; 12: 1145-8.
211	2. Poole TL, Edrington TS, Brichta-Harhay DM <i>et al.</i> Conjugative transferability of
212	the A/C plasmids from Salmonella enterica isolates that possess or lack bla_{CMY} in the
213	A/C plasmid backbone. Foodborne Pathog Dis 2009; 6: 1185-94.
214	3. Call DR, Singer RS, Meng D <i>et al</i> . <i>bla</i> _{CMY-2} -positive IncA/C plasmids from
215	Escherichia coli and Salmonella enterica are a distinct component of a larger lineage of
216	plasmids. Antimicrob Agents Chemother 2010; 54: 590-6.
217	4. Han J, Lynne AM, David DE <i>et al.</i> DNA sequence analysis of plasmids from
218	multidrug resistant Salmonella enterica serotype Heidelberg isolates. PLoS One 2012; 7:
219	e51160.

- 5. Han J, Lynne AM, David DE et al. Sequencing of plasmids from a multi-
- 221 antimicrobial resistant Salmonella enterica serovar Dublin strain. Food Research
- 222 International 2012; **45**: 931-4.
- 223 6. Evershed NJ, Levings RS, Wilson NL et al. Unusual class 1 integron-associated
- 224 gene cassette configuration found in IncA/C plasmids from Salmonella enterica.
- 225 Antimicrob Agents Chemother 2009; **53**: 2640-2.
- 226 7. Welch TJ, Fricke WF, McDermott PF *et al*. Multiple antimicrobial resistance in
- 227 plague: an emerging public health risk. *PLoS One* 2007; **2**: e309.
- 8. Hoffmann M, Muruvanda T, Allard MW et al. Complete Genome Sequence of a
- 229 Multidrug-Resistant Salmonella enterica Serovar Typhimurium var. 5- Strain Isolated
- from Chicken Breast. *Genome Announc* 2013; 1: e01068-13.
- 231 9. Doublet B, Boyd D, Mulvey MR *et al*. The *Salmonella* genomic island 1 is an
- integrative mobilizable element. *Mol Microbiol* 2005; **55**: 1911-24.
- 233 10. Douard G, Praud K, Cloeckaert A et al. The Salmonella genomic island 1 is
- specifically mobilized in trans by the IncA/C multidrug resistance plasmid family. *PLoS*
- 235 *One* 2010; **5**: e15302.
- 236 11. Harmer CJ, Hall RM. pRMH760, a Precursor of A/C Plasmids Carrying blacmy
- and bla_{NDM} genes. *Microb Drug Resist* 2014; **20**: 416-23.
- 238 12. Brau B, Pilz U, Piepersberg W. Genes for gentamicin-(3)-N-acetyltransferases
- 239 III and IV: I. Nucleotide sequence of the AAC(3)-IV gene and possible involvement of
- an IS140 element in its expression. *Mol Gen Genet* 1984; **193**: 179-87.
- 241 13. Wilson K. Preparation of genomic DNA from bacteria. In: *Current Protocols in*
- 242 *Molecular Biology*. Hoboken: John Wiley & Sons Inc, 2001, 2.4.1-2.4.5.
- 243 14. Zerbino DR, Birney E. Velvet: algorithms for de novo short read assembly using
- 244 de Bruijn graphs. *Genome Res* 2008; **18**: 821-9.

245 15. Zankari E, Hasman H, Cosentino S *et al*. Identification of acquired antimicrobial
246 resistance genes. *J Antimicrob Chemother* 2012; **67**: 2640-4.

247 16. Cain AK, Liu X, Djordjevic SP *et al.* Transposons related to Tn¹⁶⁹⁶ in IncHI2

248 plasmids in multiply antibiotic resistant Salmonella enterica serovar Typhimurium from

Australian animals. *Microb Drug Resist* 2010; **16**: 197-202.

250 17. Nigro SJ, Hall RM. GIsul2, a genomic island carrying the sul2 sulphonamide

resistance gene and the small mobile element CR2 found in the *Enterobacter cloacae*

subspecies cloacae type strain ATCC 13047 from 1890, Shigella flexneri ATCC 700930

from 1954 and Acinetobacter baumannii ATCC 17978 from 1951. J Antimicrob

254 *Chemother* 2011; **66**: 2175-6.

255 18. Kadlec K, Brenner Michael G, Sweeney MT et al. Molecular basis of macrolide,

triamilide, and lincosamide resistance in *Pasteurella multocida* from bovine respiratory

disease. Antimicrob Agents Chemother 2011; 55: 2475-7.

258 19. Michael GB, Eidam C, Kadlec K et al. Increased MICs of gamithromycin and

tildipirosin in the presence of the genes erm(42) and msr(E)-mph(E) for bovine

260 Pasteurella multocida and Mannheimia haemolytica. J Antimicrob Chemother 2012; 67:

261 1555-7.

262 20. Michael GB, Kadlec K, Sweeney MT et al. ICEPmu1, an integrative conjugative

263 element (ICE) of *Pasteurella multocida*: analysis of the regions that comprise 12

antimicrobial resistance genes. *J Antimicrob Chemother* 2012; **67**: 84-90.

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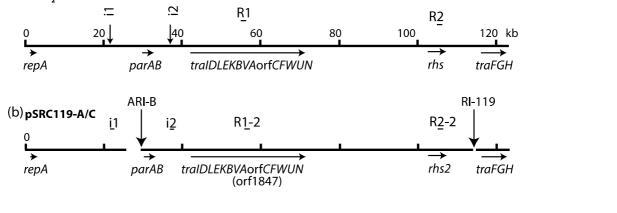
271 Figure Legends

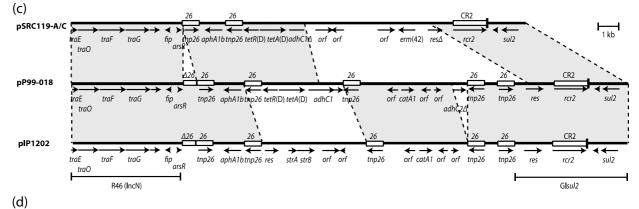
272	Figure 1. Comparison of pSRC119-A/C to other A/C ₂ plasmids. (a) A/C ₂ backbone
273	structure showing difference between Type 1 and Type 2 groups as defined
274	previously ¹¹ and (b) backbone structure of Type 2 plasmid pSRC119-A/C. Regions
275	containing genes involved in plasmid replication (<i>rep</i>), partitioning (<i>par</i>) and
276	conjugative transfer (<i>tra</i>), and the <i>rhs</i> gene are indicated by arrows below. The
277	remaining genes and open reading frames are annotated in GenBank Accession
278	Number <mark>KM670336</mark> . The location of insertions i1 and i2 found in Type 2, and two
279	regions of replacement R1 (in the orf) and R2 (in <i>rhs</i>) are shown above. In
280	pSRC119-A/C, vertical arrows indicate the location of the ARI-B and RI-119
281	resistance islands. Gaps in the horizontal line indicate segments of sequence that
282	are missing relative to the A/C_2 backbone. Figure is drawn to scale with lengths in
283	kb shown above the line in (a). (c) pSRC119-A/C ARI-B and related ARI-B
284	structures. Regions are drawn to scale from GenBank Accession Numbers
285	KM670336 (pSRC119-A/C), NC_008612 (pP99-018), and NC_009141 (pIP1202).
286	(d) Regions surrounding the <i>erm</i> (42) gene. Structures were drawn to scale from
287	GenBank Accession Numbers CP003022 (P. multocida), and AB601890 (P.
288	damselae). In (c) and (d), shared regions are indicated by shading. IS and the small
289	mobile element CR2 are shown as open boxes with a vertical bar indicating the ori
290	end of CR2. IS numbers or names are indicated above. Genes and open reading
291	frames (orfs) are shown below the line as named arrows indicating the direction of
292	transcription. Segments derived from the IncN plasmid R46 or GIsul2 are indicated
293	below.

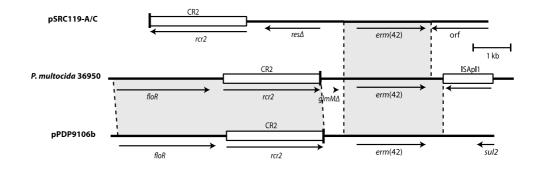
295	Figure 2. RI-119. (a) Structure of RI-119. Flanking backbone is shown as a dashed
296	line. The arrangement of genes in KPNIH18 (drawn to scale from AKA10100038) is
297	shown below. (b) Regions surrounding the IS26-aacC4-hph-IR $_{5393}$ unit. Regions are
298	drawn to scale from GenBank Accession Numbers KM670336 (pSRC119-A/C),
299	KF705205 (p9134), GQ114284 (pPWD4_103) and HF545434 (pK1HV). Shading
300	indicates shared regions. Inverted repeats (IR) are shown as thick vertical lines with
301	the origin named above, and IS are shown as open boxes with names above. Genes
302	and open reading frames (orfs) are shown below the line as named arrows
303	indicating the direction of transcription. The origins of individual segments are
304	indicated above. In (b), horizontal lines with different patterns denote different genetic
305	contexts and the direct repeats surrounding the Tn5393-derived portion of p9134 are
306	shown below.
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320 Figure 1









(a)

