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<th>Intra- and Interfractional Variations in Geometric Arrangement between Lung Tumours and Implanted Markers</th>
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Keywords: Lung cancer, fiducial marker, respiratory motion management, 4DCT

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Results: The intrafractional variations differed between patients. The root mean squares of standard deviations for each phase were 0.6, 0.9, and 1.5 mm in the right-left, anterior-posterior, and superior-inferior directions, respectively. The maximum difference in intrafractional variation among 10 phases was correlated with the amplitude of tumour motion in all directions and the tumour-marker distance in the anterior-posterior and superior-inferior directions. The interfractional variations were within 2.5 mm.

Conclusions: The intrafractional variations differed according to the amount of tumour motion and the tumour-marker distance. Additionally, interfractional variations of up to 2.5 mm were observed. Thus, a corresponding margin should be considered during implanted marker-based beam delivery to account for these variations.
Title

Intra- and Interfractional Variations in Geometric Arrangement between Lung Tumours and Implanted Markers

Authors

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Running header

Variation between lung tumour and markers

Keywords

Lung cancer, fiducial marker, positional variation, 4DCT.

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Abstract

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Introduction
Stereotactic body radiation therapy (SBRT) is an innovative technique that delivers high-dose radiation limited precisely to the region of the tumour [1,2]. In SBRT for targets affected by respiratory motion, such as lung tumours, appropriate motion management is recommended to reduce doses delivered to the surrounding normal tissues. Several methods of accounting for respiratory motion have been developed, including methods in which the radiation delivery is synchronised with respiration; i.e. the dynamic tumour tracking (DTT) method and the respiratory gating method [3].

With the above respiratory-synchronised methods, markers implanted either in the tumour itself or nearby are often used as the internal surrogate to localise the tumour position [4-6]. However, the position of the implanted markers does not always represent the tumour position because the tumour and markers move non-synchronously during respiration, especially in cases in which the markers were located slightly distal from the tumour [7]. This intrafractional positional difference between the tumour and markers should be incorporated into the DTT or respiratory gating irradiation treatment plan by using a wider gating window, within which the beam is delivered during the respiration cycle. Furthermore, the relative position of the tumour with respect to the markers may vary from day to day; therefore, the interfractional positional difference must be addressed. However, little about these variations is known.

The purpose of this study was to quantify the intra- and interfractional variations between the lung tumour position and the position of the implanted markers to evaluate the margin necessary to account for the associated errors during respiratory-synchronised irradiation treatment.
Materials and Methods

Patients and implanted markers

Fifteen patients who underwent SBRT for a solitary lung tumour were enrolled in this study. With the approval of the Institutional Review Board, written informed consent was obtained from all patients. One to two weeks prior to the date of the computed tomography (CT) simulation, four or five disposable gold markers (Olympus Corporation, Tokyo, Japan), spherical markers with a diameter of 1.5 mm, were implanted transbronchially. The insertion technique was similar to the one reported by Harada et al [4]. Prior to the implantation, the relative position between tumour and bronchi was evaluated on the multiplanar reformatted CT images. The markers were implanted into the peripheral surrounding bronchi near tumour under fluoroscopy guidance. A total of 66 markers were placed. The median interval between marker placement and the CT simulation was 8 (range, 2 to 16) days. Twelve markers were coughed up before CT simulation. After CT simulation, 2 markers were coughed up on the seventh and thirteenth day, and 1 marker migrated on the sixth day after insertion. The markers that coughed up or migrated after CT simulation during the treatment period were excluded from this analysis. No adverse effect associated with the implantation was observed. The characteristics of patients and tumours are shown in Table 1.

Patient set-up and four-dimensional CT data acquisition

The patients were immobilised using vacuum immobilisation devices: BodyFix system (Elekta AB, Stockholm, Sweden) or Esform (Engineering System, Nagano, Japan). After set-up with skin marks, four-dimensional CT (4DCT) data were acquired using a 16-multidetector row CT: LightSpeed RT or BrightSpeed (General Electric Healthcare, Waukesha, WI, USA) with an axial slice thickness of 2.5 mm. The cine duration time of the scan at each couch position was set to 6.0 or 7.0 s, which was more than the maximum
observed respiratory period. Simultaneously, the respiratory phase was monitored using the Varian Real-time Position Management system (Varian Medical Systems, Palo Alto, CA, USA) under free breathing without coaching. CT slices and respiratory phase data were transferred to the Advantage SIM workstation (General Electric Healthcare, Waukesha, WI, USA) and sorted into 10 respiratory phase bins. Motion phases were assigned for each respiratory phase as percentages; end-inhalation corresponded to 0% and end-exhalation to 50%. 4DCT scans were performed during the CT simulation (CT-1) and repeated once during the course of treatment (CT-2). Fifteen pairs, corresponding to a total of thirty 4DCT scans, were obtained. The median period from the day of CT-1 until the day of CT-2 was 8 days (range, 4 to 12). All 4DCT datasets were imported into a commercial radiotherapy planning system, iPlan 4.5.1 (BrainLAB AG, Fieldkirchen, Germany).

Analysis

The intrafractional variations assessed in this study were defined as the residual tumour motions relative to the markers due to respiration. In all 10 phases of the CT-1 scans, gross tumours and implanted markers were contoured manually with a pulmonary window setting (window level, -700 Hounsfield units; window width, 2000 Hounsfield units) by a single radiation oncologist. The centroid of the tumour $G_{t,n} = (x_{t,n}, y_{t,n}, z_{t,n})$ and the centroids of all 3 to 5 markers $G_{m,n} = (x_{m,n}, y_{m,n}, z_{m,n})$ were recorded at $n\%$ respiratory phase ($0 \leq n \leq 90$). The coordinates ($x$, $y$, $z$) correspond to the right-left (RL), anterior-posterior (AP), and superior-inferior (SI) directions, respectively. Along each axis, a positive value corresponds to the right, anterior, and superior directions. The relative position of the tumour and centroid of markers for each phase was represented by the vector $V_n = G_{t,n} - G_{m,n}$. Using the relative positions on 50% phase images ($V_{50}$) as a reference, the error $E_n = V_n - V_{50}$ was calculated. The mean ($M_n$) and standard deviation ($SD_n$) of $E_n$ in fifteen 4DCT CT-1 datasets were also
calculated. The mean and SD of $M_n$ ($0 \leq n \leq 90$) were calculated to evaluate systematic displacement between respiratory phases. The root mean square (RMS) of $SD_n$ ($0 \leq n \leq 90$) was calculated to evaluate interpatient variations. The range of intrafractional variations is defined as the maximum difference in $E_n$ among 10 phases for each direction. To evaluate the influence of the tumour motion amplitude and the tumour-marker distance on the ranges of intrafractional variations, a multiple linear regression analysis was performed. The tumour motion amplitude was defined as the maximal difference in the tumour centroid position among the 10 respiratory phases in each direction. The tumour-marker distance in each direction was defined as the distance between the tumour centroid and the centroid of all 3 to 5 markers in the 50% phase images.

The interfractional variations in this study represent the residual setup errors after correction based on the implanted markers. Firstly, to correct the rotational set up errors, the 50% phase images for CT-2 were rigidly registered to the 50% phase images of CT-1 based on bony structure. Then the translational errors were modified by registering those images based on the marker centroids. The interfractional variations were evaluated as the residual difference in the tumour centroids for each direction between CT-1 and CT-2 for each patient.

Results

**Tumour motion amplitude and tumour-marker distance**

The median (range) tumour motion amplitudes in CT-1 were 1.8 mm (0.4 to 5.6), 3.1 mm (0.6 to 7.8), and 8.2 mm (0.9 to 28.9) in the RL, AP, and SI directions, respectively. The median values (range) of the distance between the tumour centroid and the centroid of all markers in the 50% phase images for CT-1 were 11.9 mm (1.9 to 30.5), 8.1 mm (0.7 to 35.3), and 10.3 mm (0.3 to 30.1) in the RL, AP, and SI directions, respectively.
Intrafractional variations
The divergence in the range of intrafractional variations between patients is shown in Fig. 1, and the values of $E_n$ in the respiratory phases ($0 \leq n \leq 90$) are shown in Fig. 2. The means ± SD of $M_n$ ($0 \leq n \leq 90$) were 0.1±0.1 mm, 0.3±0.2 mm, and 0.0±0.2 mm, and the RMS of SD$_n$ were 0.6 mm, 0.9 mm, and 1.5 mm in the RL, AP, and SI directions, respectively. These results indicate that the systematic difference between respiratory phases is negligible. In addition, as shown in Fig. 2, the further towards inhale then the greater the intrafractional variations.

The tumour motion amplitude was positively correlated with the range of intrafractional variations in all directions, and the tumour-marker distances were also positively correlated in the AP and SI directions (Table 2).

Interfractional variations
The median (range) interfractional variations were -0.1 mm (-2.4 to 0.7), 0.1 mm (-2.3 to 2.4), and -0.6 mm (-1.3 to 1.6) in the RL, AP, and SI directions, respectively. As shown in Fig. 3, all interfractional variations were within 2.5 mm; the greatest variations were in the AP direction. The 95th percentiles of interfractional variations for one side of each direction were 0.6 and 2.1 mm to the right and left, 1.9 and 2.1 mm in the anterior and posterior directions, and 1.6 and 1.3 mm in the superior and inferior directions.

Discussion
Implanted markers are often used as a surrogate for the tumour position in radiation therapy for lung tumours. The transcutaneous and transbronchial approaches are the two major methods for implantation of markers in the vicinity of lung tumours [8-13]. These procedures may cause pneumothorax as a complication, which can delay radiation delivery and could be life-threatening for those with comorbidities. The incidences of all pneumothorax and of those
requiring chest tube placement after transcutaneous implantation have been reported to be 30 to 67% and 16 to 40%, respectively [8-10]. By contrast, the reported incidence of pneumothorax with the transbronchial approach is low [10-13] and in our series no complication was observed. Therefore, the transbronchial approach is preferable due to its less invasive nature. However, the placement of markers near or inside the tumour is more difficult with the transbronchial approach than with the transcutaneous approach, because in the former the markers are placed along the small bronchi near a tumour. The greater distance between the tumour and markers leads to a larger positional error [14]. This error must be considered when performing radiotherapy using markers placed outside the tumour. In the current study, we quantified the intra- and interfractional positional variations between the lung tumour and implanted markers using 4DCT scans to determine the necessary margin for respiratory-synchronised irradiation using implanted markers. Another issue about the markers implanted transbronchially is the low fixation rate. In our series, the fixation rate of implanted markers was 77.3%: 51 of 66 markers implanted markers were fixed throughout treatment. This is comparable to the reported fixation rate using the same insertion technique [11]. Due to the low fixation rate, we inserted 4 or 5 markers to avoid an additional insertion procedure and used multiple markers as a surrogate for the tumour position to address the change in geometric arrangement of markers by dislocation.

Although several authors reported the intrafractional verification of the tumour position by the kilo-voltage (kV) X-ray images during gating irradiation, they calculated the tumour positions from the detected positions of the implanted markers assuming the relative position between the tumour and markers were constant [7,15]. The planar kV X-ray imaging is superior to CT in terms of the temporal resolution but it is difficult to quantify the motion of tumor itself accurately on the projected images. To quantify the variations between the tumour and implanted markers, we used 4DCT. Two studies are available which evaluated the
geometrical difference between tumour and markers due to respiration using 4DCT. Smith et al. analysed the motion of lung tissue in 10 patients with deformable registration between exhalation and inhalation of 4DCT scans and reported stronger correlations between tumour and surrounding lung tissues in the upper lobes than in the lower lobes [16]. Finally, they concluded that the correlation between the tumour and the surrounding tissue was highly specific to the patient and lobe [16]. Since the amplitude of the tumour motion is typically smaller in the upper lobe than that in the lower lobe, then it is likely the bigger variations observed in lower lobe tumours by Smith et al. may be related to the amplitude of motion.

Yamazaki et al. evaluated the distances between tumours and the distal bronchi during respiration cycle with 4DCT for 8 patients. They showed that the distances in the mid-inhalation to end-inhalation phase images were significantly larger than the distances in the end-exhalation phase images [17]. Smith et al. and Yamazaki et al. suggested that markers that are closer to the tumour give a more accurate representation of tumour motion [16,17]. These results are consistent with our findings: the values needed to compensate for the intrafractional variations differed between patients, and depended on the amplitude of tumour motion and the tumour-marker distance.

Moreover, our results indicated that the intrafractional errors were different for each patient both in direction and in amplitude, as shown in Fig 1. A uniform isotropic margin was not adequate to cover the errors observed. Consequently, the variations must be evaluated on a per-patient basis and compensated for by addition of a patient-specific margin in DTT or gating irradiation treatment with a wider gating window. In our treatment planning process of DTT, we create an enlarged target volume which cover the intrafractional variations in the following steps. Firstly, 4DCT images from each phase are translated based on the centroid of the markers. Then, the phase images are superimposed onto the 50% phase image that is used as a reference image set. Finally, the enlarged target volume is delineated encompassing gross
tumour volumes on all fused phase image. This enlarged volume can compensate for the patient-specific intrafractional variations.

All interfractional variations in the present study were within 2.5 mm. Previous reports on interfractional variations between lung tumours and implanted markers are summarised in Supplementary Table 1 (Electronic Appendix). The reported values are larger than those in this study. This discrepancy may be attributed to two causes. One is a change in tumour-marker distance during the course of treatment. Several investigators reported tumour shrinkage and deformation after radiotherapy with conventional fractionation [5,10,14], which altered the distance. Meanwhile, Imura et al. [11] evaluated the interfractional variations in the distances between markers using orthogonal X-ray images with a median treatment time of 6 days, and showed that the variations during treatment were within 2 mm in 95% of cases. Their results support our finding that the interfractional variation between the tumour and markers was smaller in those undergoing hypofractionated treatment than in those undergoing conventional fractionation. A second interesting finding was the respiratory phase reproducibility. Van der Voort van Zyp et al. assessed marker displacement compared to the centroid of the tumour in patients that underwent SBRT; however, their results were influenced by the nonsynchronous tumour-marker motion due to divergence in the timing of breath holding [6]. Persson et al. also used the breath-hold CT with voluntary deep inspiration [18]. The interfractional variations in the current study were evaluated using end-exhale phase images under free breathing, which has high reproducibility compared with breath-holding.

Several limitations of our study should be mentioned. Firstly, the intrafractional variations evaluated with 4DCT during a few respiration cycles may not be representative of those during treatment in some patients although a single 4DCT is thought to be reliable for the tumour motion in the majority of patients [19]. Therefore, in our institution, we validate margins to compensate for the intrafractional variations, by visual verification with kV x-ray
fluoroscopy after the margins are determined based on the simulation 4DCT. Secondly, motion artefacts affected the contouring of tumours and implanted markers evaluated in binned 4DCT images. Because of this uncertainty in contouring, the intrafractional variations for tumours with larger motion may be over- or underestimated [20]. Furthermore, we evaluated the position of markers of 1.5-mm diameter using 4DCT with a 2.5-mm axial slice thickness. The use of 2.5-mm CT slice thickness would affect the accuracy of contouring the markers, with a maximum uncertainty of localising tumours and markers of 1.25 mm in the SI direction [6]. Those errors could be reduced by acquiring CT images with a thinner axial slice thickness or by the volumetric acquisition [21,22].

**Conclusions**

Intrafractional variations of the difference between tumour centroid and marker centroid position increased with both tumour motion amplitude and tumour-marker distance. Additionally interfractional variations of the distance between between tumour centroid and marker centroid position were observed up to 2.5mm. Thus, an appropriate margin to account for these variations should be considered when planning implanted-marker-based beam delivery.
References


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Conflicts of interest
Takashi Mizowaki, Masaki Kokubo, and Masahiro Hiraoka have a consultancy agreement with Mitsubishi Heavy Industries, Ltd., Japan.
Figure legends

Fig. 1. The range of intrafractional variations in the RL (a), AP (b), and SI (c) directions for each patient rearranged according to the three-dimensional tumour motion amplitude in descending order. Abbreviations: RL, right-left; AP, anterior-posterior; SI, superior-inferior

Fig. 2. $E_n$ values in the RL (a), AP (b), and SI (c) directions for each respiratory phase. $E_n$ is the error in the relative position of the tumour to the centroid of the markers on n% phase images ($0 \leq n \leq 90$), using the relative position on the 50% phase images as a reference. Other abbreviations are as in Fig. 1.

Fig. 3. Interfractional variation in the RL, AP, and SI directions. Abbreviations are as in Fig. 1.
Table 1. Characteristics of patients and tumours (n=15).

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<tr>
<td>Median</td>
<td>82</td>
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<tr>
<td>[range]</td>
<td>[54–87]</td>
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<tr>
<td><strong>Gender</strong></td>
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<tr>
<td>Male</td>
<td>12</td>
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<tr>
<td>Female</td>
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<tr>
<td><strong>Tumour size</strong></td>
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<tr>
<td>( \leq 20 \text{ mm} )</td>
<td>4</td>
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<tr>
<td>( &gt;20 \text{ to } \leq 30 \text{ mm} )</td>
<td>6</td>
</tr>
<tr>
<td>( &gt;30 \text{ to } \leq 50 \text{ mm} )</td>
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<tr>
<td><strong>Tumour location</strong></td>
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<td>Right middle lobe</td>
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</tr>
<tr>
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<td>7</td>
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<td>Left upper lobe</td>
<td>2</td>
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<tr>
<td>Left lower lobe</td>
<td>4</td>
</tr>
<tr>
<td><strong>No. of implanted markers</strong></td>
<td></td>
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<td>4</td>
<td>9</td>
</tr>
<tr>
<td>5</td>
<td>6</td>
</tr>
<tr>
<td><strong>No. of markers evaluated</strong></td>
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<td>3</td>
<td>10</td>
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Table 2. Predictive factors for the range of intrafractional variation as determined by multiple linear regression analysis.

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<tr>
<td></td>
<td></td>
<td>RL β</td>
<td>p</td>
<td>AP β</td>
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<tr>
<td>Tumour motion amplitude</td>
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<td>0.539</td>
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<td>R</td>
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*Abbreviations:* R, correlation coefficient; other abbreviations are as in Fig. 1.
Figure 2

(a) RL [mm]

(b) AP [mm]

(c) SI [mm]

Phase (%)
Click here to download Supplementary Files: Supplementary Table 1.doc