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Kasper, S., van der Meer, M.T.J., Castaneda, I.S., Tjallingii, R., Brummer, G.-J., Sinninghe Damsté, J. S. & Schouten, S. (2015). Testing the alkenone D/H ratio as a paleo indicator of sea surface salinity in a coastal ocean margin (Mozambique Channel). *Organic Geochemistry*, 78, 62-68.

Published version: [dx.doi.org/10.1016/j.orggeochem.2014.10.011](https://doi.org/10.1016/j.orggeochem.2014.10.011)

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Testing the alkenone D/H ratio as a paleo indicator of sea surface salinity  
in a coastal ocean margin (Mozambique Channel)

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1 ABSTRACT

2 Reconstructing past ocean salinity is important for assessing paleoceanographic change and  
3 therefore past climatic dynamics. Commonly, sea water salinity reconstruction is based on  
4 planktonic foraminifera oxygen isotope values combined with sea surface temperature  
5 reconstruction. However, the approach relies on multiple proxies, resulting in rather large  
6 uncertainty and, consequently, relatively low accuracy of salinity estimates. An alternative tool  
7 for past ocean salinity reconstruction is the hydrogen isotope composition of long chain (C<sub>37</sub>)  
8 alkenones ( $\delta D_{\text{alkenone}}$ ). Here, we have applied  $\delta D_{\text{alkenone}}$  to a 39 ka sedimentary record from the  
9 Eastern South African continental shelf in the Mozambique Channel, close to the Zambezi River  
10 mouth. Despite changes in global seawater  $\delta D$  related to glacial – interglacial ice volume effects,  
11 no clear changes were observed in the  $\delta D_{\text{alkenone}}$  record throughout the entire 39 ka. The BIT  
12 index record from the same core, which provides information on relative contributions of soil  
13 organic matter (OM) vs. marine input, indicates high soil OM input during the glacial and low  
14 input during the Holocene. This suggests a more pronounced freshwater influence at the core  
15 location during the glacial, resulting in alkenones depleted in D during that time, thereby  
16 explaining the lack of a clear glacial-interglacial alkenone  $\delta D$  shift. The correlation between the  
17 BIT index and  $\delta D_{\text{alkenone}}$  during the glacial period suggests that increased continental runoff  
18 potentially changed the growth conditions of the alkenone-producing haptophytes, promoting  
19 coastal haptophyte species with generally more enriched  $\delta D_{\text{alkenone}}$  values. We therefore suggest  
20 that the application of  $\delta D_{\text{alkenone}}$  for reconstructing past salinity in coastal settings may be  
21 complicated by changes in the alkenone-producing haptophyte community.

22

23 **Keywords:** stable hydrogen isotopes, alkenones, salinity, BIT index, coastal environment,  
24 Mozambique Channel, Zambezi River, glacial, interglacial.

## 25 **1. Introduction**

26 The interaction and feedback mechanisms at the interface between oceanic and atmospheric  
27 circulation on glacial – interglacial timescales are largely unknown, but are thought to play an  
28 important role in the global transmission of climate change (Manabe, 1969; Rahmstorf, 2002).  
29 Accurate reconstruction of past oceanic and atmospheric changes is crucial for a better  
30 understanding of these mechanisms and should eventually improve prediction of future climate  
31 development (Esper et al., 2005). Reconstruction of ocean circulation requires an estimate of  
32 seawater density, which is determined by salinity and temperature. Therefore, in order to  
33 understand past ocean circulation in relation to climate change it is essential to be able to  
34 reconstruct paleosalinity and temperature with reasonable accuracy.

35  
36 Ocean salinity and the oxygen isotope composition of seawater are linearly correlated (Epstein  
37 and Mayeda, 1953) and this can be utilized for reconstruction of past seawater salinity using the  
38  $\delta^{18}\text{O}$  record of carbonate shells of foraminifera (Duplessy et al., 1991; Rostek et al., 1993) under  
39 the assumption of temporal consistency of the relationship between  $\delta^{18}\text{O}$  and salinity (Rohling  
40 and Bigg, 1998; Rohling et al., 2000). However, salinity reconstruction based on  $\delta^{18}\text{O}$  values of  
41 foraminiferal carbonate requires accurate correction for the global ice volume (Rostek et al.,  
42 1993) and knowledge of the temperature during calcification (Erez and Luz, 1983). The global  
43 ice volume can be estimated from the mean ocean  $\delta^{18}\text{O}$  record (Waelbroeck et al., 2002), whereas  
44 the temperature is mostly obtained from Mg/Ca ratio values of foraminifera (Nürnberg et al.,  
45 1996; Lea et al., 1999; Mashiotta et al., 1999) or the alkenone-based  $U^{K'}_{37}$  Index (Brassell et al.,  
46 1986; Rostek et al., 1993; Nürnberg et al., 1996). However, the correlation between  $\delta^{18}\text{O}$  and  
47 salinity is affected by mixing and evaporation processes (Craig and Gordon, 1965), seasonality  
48 (Strain and Tan, 1993) and likely changes over glacial – interglacial periods (Rohling and Bigg,

49 1998). Consequently, salinity reconstruction based on  $\delta^{18}\text{O}$  values of foraminiferal carbonate is  
50 subject to uncertainty and is difficult to calibrate (Rohling and Bigg, 1998; Rohling, 2000).

51  
52 In addition to the correlation between oxygen isotopes and salinity, there is also a strong  
53 correlation between the hydrogen isotope composition of seawater ( $\delta\text{D}_{\text{sw}}$ ) and salinity (Craig,  
54 1961; Friedman et al., 1964).  $\delta\text{D}_{\text{sw}}$  is recorded in the non-exchangeable hydrogen of organic  
55 matter (OM; Sessions et al., 1999; Sauer et al., 2001; Sachse et al., 2012) and can potentially be  
56 used for reconstructing past  $\delta\text{D}_{\text{sw}}$ . The hydrogen isotope composition of long chain alkenones, in  
57 particular  $\text{C}_{37}$  alkenones ( $\delta\text{D}_{\text{alkenone}}$ ) appears well suited for reconstructing past  $\delta\text{D}$  values of  
58 seawater (Englebrecht and Sachs, 2005; Schouten et al., 2006). The isotopic composition of  
59 covalently bound hydrogen is thought to be preserved over long geological timescales (Sessions  
60 et al., 2004). Culture studies have shown that  $\delta\text{D}_{\text{alkenone}}$  of two common marine haptophytes,  
61 *Emiliania huxleyi* and *Gephyrocapsa oceanica*, depends mainly on salinity and the hydrogen  
62 isotope composition of the growth medium and, to a lesser extent, growth rate (Schouten et al.,  
63 2006). Attempts to reconstruct past salinity change based on  $\delta\text{D}_{\text{alkenone}}$  has led to reasonable  
64 results for the eastern Mediterranean Sea and Black Sea (van der Meer et al., 2007, 2008; Coolen  
65 et al., 2013) and the Panama Basin (Pahnke et al., 2007). Recently, the approach has also been  
66 applied to open marine sediments to evaluate relative salinity changes in the Agulhas Leakage  
67 area at the southern tip of the African continent over two glacial termination periods (Kasper et  
68 al., 2014). However, the lack of a clear correlation between  $\delta\text{D}_{\text{alkenone}}$  and seawater salinity in the  
69 Chesapeake Bay estuary suggests that the interpretation of this salinity proxy may become  
70 problematic for coastal environments (Schwab and Sachs, 2011). A possible explanation for the  
71 absence of a clear correlation relates to change in species composition since different haptophytes  
72 may have specific D/H fractionation characteristics. Comparison of the coastal haptophyte alga

73 *Isochrysis galbana* with the open ocean species *E. huxleyi* in a culture experiment (M'Boule et  
74 al., 2014) revealed a strong fractionation difference between the two species. *I. galbana*  
75 fractionated to an extent of ca. 90‰ less than *E. huxleyi*, although the rate of change in D/H  
76 fractionation per salinity unit remained similar for both species (M'Boule et al., 2014, Chivall et  
77 al., 2014). This raises the question as to which extent  $\delta D_{\text{alkenone}}$  is applicable as a paleosalinity  
78 proxy in coastal settings, where differences in haptophyte communities are to be expected  
79 (Marlowe et al., 1990; Noble et al., 2003; Örnólfssdóttir et al., 2004; Schwab and Sachs, 2011).

80  
81 To investigate the  $\delta D_{\text{alkenone}}$ -based sea surface salinity proxy in a freshwater runoff-influenced  
82 continental margin, we analyzed a 39,000 year (39 ka) long record of  $\delta D_{\text{alkenone}}$  from a marine  
83 sediment core (64PE304-80) from the Mozambique Channel near the Zambezi River mouth  
84 inside the Zambezi River fan (Fig. 1; Beiersdorf et al., 1980). The core site is off the continental  
85 shelf in the channel, which is traversed by the Mozambique Current, a branch of the South  
86 Equatorial Current (SEC; Fig. 1). The Mozambique Current is dominated by large anti-cyclonic  
87 eddies passing through the Mozambique Channel, with an annual net flow rate of ca. 15 Sv (de  
88 Ruijter et al., 2002; Schouten et al., 2003; Ullgren et al., 2014). However, interannual net flow  
89 rate varies substantially by up to 9 Sv in volume transport (Ridderinkhof et al., 2010).  
90 Additionally, there is a substantial input of continental freshwater to the Mozambique Channel  
91 (Siddorn et al., 2001). A main contributor is the Zambezi River, with a runoff of ca. 224 km<sup>3</sup>/yr  
92 (7.1 x 10<sup>-3</sup>Sv; Meybeck, 1982). Due to the location of the core site within the Zambezi River fan  
93 and independent reconstruction of high freshwater outflow by the BIT index, we would expect  
94 variation in freshwater runoff and therefore in salinity at the core site over time, suggesting it as a  
95 promising record for testing  $\delta D_{\text{alkenone}}$  as a paleo sea surface salinity proxy at a river dominated  
96 ocean margin.

97

## 98 **2. Material and methods**

99

### 100 *2.1. Core and chronology*

101

102 Samples were taken from core 64PE304-80 (18° 14' 26.6274"S; 37° 52' 8.6874" E, 1329 m water  
103 depth), located north of the Zambezi River Delta at the Mozambique shelf (Fig. 1). The core was  
104 taken during the RV Pelagia cruise "64PE304, INATEX - GEO" (Brummer et al., 2009). The  
105 chronology was established from <sup>14</sup>C dating of 20 samples consisting of ca. 10 mg mixed surface-  
106 dwelling planktonic foraminifera (*Globigerinoides acculifer*, *G. trilobus* and *G. ruber*). These  
107 samples were collected from the washed and sieved > 250 µm sediment fraction by selecting  
108 individual foraminifera specimens under a microscope (Fig. 2). The age model for the upper 6 m,  
109 spanning the last 20 ka, was constrained by 11 <sup>14</sup>C dates and by stratigraphic correlation with a  
110 parallel core (GIK16160-3; Wang et al., 2013b; Van der Lubbe et al., 2014). The age model  
111 between 20 and 39.5 ka BP was constrained by 9 additional <sup>14</sup>C dating points. The <sup>14</sup>C age values  
112 were converted to calendar age using the MARINE09 calibration curve that applies a standard  
113 reservoir correction of ca. 400 yr (Reimer et al., 2011).

114

### 115 *2.2. Extraction*

116

117 Sediment samples were freeze dried and homogenized with a mortar and pestle. The  
118 homogenized material was extracted using accelerated solvent extraction (ASE) with  
119 dichloromethane (DCM):MeOH 9:1 (v/v) and a pressure of 6895 kPa in 3 extraction cycles. The

120 extract was separated over an Al<sub>2</sub>O<sub>3</sub> column into apolar, ketone and polar fractions using  
121 hexane:DCM 9:1 (v/v), hexane:DCM 1:1 (v/v) and DCM:MeOH 1:1 (v/v), respectively. Column  
122 chromatography was carried out using about 4 cm activated Al<sub>2</sub>O<sub>3</sub> (MPO Chemicals; technical  
123 quality, basic A) as stationary phase in a Pasteur pipette . Fractions were eluted with 3 column  
124 volumes of eluent.

125

### 126 *2.3. Branched isoprenoid tetraether (BIT) index*

127

128 The polar fraction was analyzed for the BIT index, a proxy originally thought to reflect the input  
129 of soil OM (Hopmans et al., 2004) but which has recently been shown to be affected by in situ  
130 production of branched glycerol dibiphytanyl glycerol tetraethers (br GDGTs) in rivers (Zell et  
131 al., 2013; De Jonge et al., 2014). Analysis was performed using high performance liquid  
132 chromatography-mass spectrometry (HPLC-MS) with an Agilent 1100 series instrument  
133 equipped with an auto-injector. Separation was achieved with an Alltech Prevail Cyano column  
134 (2.1 x 150 mm, 3µm), maintained at 30 °C. GDGTs were eluted with 99% hexane and 1%  
135 propanol for 5 min, followed by a linear gradient to 1.8% propanol in 45 min, followed by back-  
136 flushing with hexane/propanol (9:1, v/v) at 0.2ml/min for 10 min. Detection was achieved using  
137 atmospheric pressure positive ion chemical ionization MS (APCI-MS). APCI-MS settings were:  
138 nebulizer 414 kPa, vaporizer 400 °C, drying gas (N<sub>2</sub>) at 6 l/min and 200 °C, capillary voltage -3  
139 kV and a corona 5 µA (ca. 3.2 kV). GDGTs were detected using selected ion monitoring (SIM) of  
140 the [M+H]<sup>+</sup> ions (*m/z* 1292 for crenarchaeol and *m/z* 1050, 1036 and 1022 for br GDGTs,  
141 respectively, dwell time 234 ms; Schouten et al., 2007) and integrated peak areas were used for  
142 quantification. BIT values were calculated according to the formula of Hopmans et al. (2004).



143

144 *2.4. C<sub>37</sub>/C<sub>38</sub> alkenone ratio*

145

146 The ketone fraction, containing long chain alkenones, was analyzed using an Agilent 6890 gas  
147 chromatography (GC) instrument with a flame ionization detection (FID) and an Agilent CP Sil-5  
148 fused silica column (50 m x 0.32 mm, film thickness 0.12 µm) with He as carrier gas. The GC  
149 oven was programmed from 70 to 200 °C at 20 °C/min and then at 3°C/min to 320 °C (held 25  
150 min). Peak integration for the calculation of ratio values for the total abundance of the C<sub>37</sub>/C<sub>38</sub>  
151 alkenones was performed using the Atlas 8.2 Chromatography Data System software from  
152 Thermo Electron Cooperation.

153

154 *2.5. Alkenone δD analysis*

155

156 The ketone fraction was analyzed using GC-high temperature conversion-isotope ratio MS (GC-  
157 TC-irMS) to determine the combined hydrogen isotope composition of the di- and triunsaturated  
158 C<sub>37</sub> alkenones (van der Meer et al., 2013). Isotope analyses were carried out with a Thermo –  
159 Scientific DELTA V GC-TC-irMS instrument. The GC temperature conditions were: 70 to 145 °C  
160 at 20 °C/min, then heated at 8 °C/min to 200 °C and to 320 °C (held 20 min) at 4°C/min. An  
161 Agilent CP Sil-5 column (25 m x 0.32 mm) with a film thickness of 0.4 µm was used with He as  
162 carrier gas at 1 ml/min (constant flow). The high temperature conversion reactor was at 1420 °C.  
163 The H<sub>3</sub><sup>+</sup> correction factor was determined daily and was constant at 5.6 ± 0.5. The δD<sub>alkenone</sub>  
164 values were calculated with the Isodat software relative to pulses of H<sub>2</sub> reference gas. A set of  
165 standard *n*-alkanes with known isotopic composition (Mix B prepared by A.Schimmelmann,  
166 University of Indiana) was analyzed daily prior to analyzing samples in order to monitor system

167 performance. Samples were only analyzed when the alkanes in Mix B had an average deviation  
168 from their off-line determined value of  $< 5\text{‰}$ . Squalane was co-injected as internal standard with  
169 each sample to monitor the accuracy and precision of alkenone isotope values. The standard had  
170 an average  $\delta D$  value of  $-165 \pm 3.6\text{‰}$ , which compared favorably with that of  $-170\text{‰}$  determined  
171 offline. The alkenone fraction was analyzed in duplicate if sufficient amount of material was  
172 available. Standard deviation of replicate analyses varied from  $\pm 0.1\text{‰}$  to  $\pm 5.9\text{‰}$ .

173

### 174 **3. Results**

175

#### 176 *3.1. BIT index*

177 To trace the relative importance of the river outflow at the core location we determined the BIT  
178 index, indicating riverine transported soil OM (Hopmans et al., 2004; Kim et al., 2007; Walsh et  
179 al., 2008; Schouten et al., 2013) as well as in situ produced crenarcheol from the river system  
180 (Zell et al., 2013; De Jonge et al., 2014). Values ranged between 0 and 1- a value near 0 indicates  
181 marine dominated OM and values near 1 a dominance of soil/river OM. Values were relatively  
182 high at ca. 0.50 between 39 and 38 ka and decreased to ca. 0.3 at 37 ka (Fig. 2). Between 37 and  
183 16 ka they increased continuously to a maximum of ca. 0.7 during Heinrich Event 1 (HE1).  
184 Subsequently, they decreased to a plateau of ca. 0.25 between 15 and 12 ka in the Younger Dryas  
185 (YD). At ca. 12 ka they decreased to the Holocene level of ca. 0.1.

186

#### 187 *3.2. $\delta D_{alkenone}$*

188 The  $\delta D_{alkenone}$  values were measured as the combined signal from  $C_{37:2}$  and  $C_{37:3}$ , as this has  
189 been suggested to yield a more robust water  $\delta D$  and salinity signal and to reduce additional  
190 biosynthetic effects related to the synthesis of  $C_{37:3}$  alkenone from the  $C_{37:2}$  alkenone (van der

191 Meer et al., 2013). The total variability in the  $\delta D_{\text{alkenone}}$  record was ca. 23‰, with absolute values  
192 ranging between -181 and -204‰. They varied around a mean of  $-191\text{‰} \pm 5\text{‰}$  ( $1\sigma$  standard  
193 deviation) over the entire record (Fig. 2). No trend from glacial Marine Isotope Stage (MIS) 3,  
194 through the Last Glacial Maximum (LGM) to the Holocene, was observed. Instead, the record  
195 was highly variable, showing a minor tendency of ca. 5‰ to increasingly D-depleted values with  
196 decreasing age during the Holocene. The record showed several periods of relatively D-enriched  
197 values of ca. -185‰ to -180‰ at 31 ka, 23 ka and during HE1, and more D-depleted values of ca.  
198 -200‰ at 37 ka, 27 ka and the LGM.

199

### 200 3.3. $C_{37}/C_{38}$ alkenone ratio

201 The ratio ranged between 0.8 and 1.7, with an average of  $1.3 \pm 0.2$  over the entire record (Fig. 2).  
202 During the period between 39– 21ka, it was  $1.3 \pm 0.1$ , and after 21 ka it increased to a maximum  
203 of 1.7 at 19 ka, followed by a decrease to 1.3 at 10 ka. After this, it increased slightly to 1.4 at 9  
204 ka and then rapidly decreased to the minimum value of ca. 0.8 at 8 ka. It was generally lower at  
205  $1.1 \pm 0.1$  during the Holocene compared with the glacial period.

206

## 207 4. Discussion

208

209 The transition from glacial to interglacial periods is marked by a significant decrease in the stable  
210 oxygen isotope ratio ( $\delta^{18}\text{O}_{\text{sw}}$ ) and the stable hydrogen isotope ratio ( $\delta D_{\text{sw}}$ ) of seawater as a  
211 consequence of melting of a large ice volume accumulated during the glacial (Shackelton, 1987).  
212 Global change in  $\delta^{18}\text{O}_{\text{sw}}$  was obtained from a mean global benthic isotope record (Waelbroeck et  
213 al., 2002) and indicates a gradual increase in  $\delta^{18}\text{O}_{\text{sw}}$  values from 0.53 to 1.02‰ between 39 and

214 20 ka BP (Fig. 2e). The values decrease linearly between 18 and 8 ka by almost 1‰. Based on a  
215 meteoric water line for the Indian Ocean (Srivastava et al., 2010), this record suggests an increase  
216 in the mean global  $\delta D_{sw}$  of ca. 4‰ during the last glacial and a decrease of nearly 8‰ during the  
217 deglaciation. In contrast, the  $\delta D_{alkenone}$  record from the upstream Mozambique slope showed  
218 fairly constant average values of  $-191 \pm 5\text{‰}$ , with no trend that could be attributed to the inferred  
219 change related to the glacial – interglacial transition (Fig. 2b). The absence of a clear glacial-  
220 interglacial change related to the changing global ice volume is unlike that for a  $\delta D_{alkenone}$  record  
221 obtained for open ocean sediments in the Agulhas Leakage area (Kasper et al., 2014). This  
222 suggests that additional environmental factors affect the  $\delta D$  values of alkenones in the  
223 Mozambique Channel. This could be related to the change in salinity, in combination with the  
224 input of relatively D-depleted freshwater via the Zambezi River outflow (Schefuß et al., 2011).  
225 An impact of temperature induced change in the  $U^{K'}_{37}$  ratio on the fractionation of hydrogen  
226 isotopes seems unlikely as this effect has not been observed in marine haptophyte algae  
227 (Schouten et al., 2006; van der Meer et al., 2013). Furthermore, the temperature change in this  
228 region has found to be relatively small (3.5 °C; Wang et al., 2013b) and we did not find any  
229 correlation between  $\delta D_{alkenone}$  and local  $U^{K'}_{37}$  sea surface temperature (SST; data not shown).  
230

231 In order to assess changes in freshwater input, we examined the BIT index for the same core  
232 (64PE304-80). The values indicate a relative increase in continental OM input during MIS2/3,  
233 with a relative maximum at ca. 17 ka during HE1 and at ca. 12 ka during the Younger Dryas,  
234 suggesting an increased influence of river outflow (Fig. 2; Van der Lubbe et al., 2014). These  
235 events were followed by a rapid decrease in the index during the deglaciation phase between 15-  
236 12ka and the period between 11-8ka, showing reduced input of continent-derived OM. This could  
237 indicate a strongly reduced river outflow, consistent with the rising sea level at the time and the

238 subsequent transgression of the shelf (Walford et al., 2005; März et al., 2008), moving the  
239 Zambezi River mouth further away from the core site. Based on  $\delta D$  values of leaf wax alkanes,  
240 Schefuß et al. (2011) demonstrated that, during northern hemisphere cold events, such as the  
241 Younger Dryas and HE1, rainfall was enhanced in the Zambezi catchment area in response to a  
242 southwards forcing of the intertropical convergence zone (ITCZ). In contrast, a northward shift of  
243 the ITCZ in the Holocene would have resulted in reduced rainfall and decreasing river discharge  
244 as compared with glacial conditions (Schefuß et al., 2011; Wang et al., 2013a). The BIT record  
245 covering the last 17 ka from the Zambezi River mouth presented by Schefuß et al. (2011) is in  
246 good agreement with the BIT record here. The constant high BIT values during the glacial  
247 indicate an increased impact of freshwater input at the study site compared with the Holocene  
248 when BIT values were low. Increased freshwater input, either via closer proximity to the coast or  
249 increased precipitation or a combination of both, would lead to a relative depletion in  $\delta D_{sw}$  at the  
250 core site and subsequently to a decrease in  $\delta D_{alkenone}$  during the glacial. The combination of lower  
251 salinity, resulting in increased biosynthetic hydrogen isotope fractionation (Schouten et al., 2006)  
252 and the more negative  $\delta D_{sw}$  values from the freshwater runoff may have resulted in production of  
253 relatively D-depleted alkenones during the glacial; this may have counteracted the global ice  
254 volume-induced increase in  $\delta D_{sw}$ , potentially leading to relatively constant values of alkenone  $\delta D$   
255 during the glacial-interglacial transition.

256

257 During MIS 2/3, positive excursions in  $\delta D_{alkenone}$  of up to 10‰ correspond with elevated BIT  
258 values (Fig. 2), such as between 34 and 31 ka, between 26 and 24 ka, and during HE1 (19-16 ka)  
259 (Fig. 2). In fact, a crossplot of BIT values vs.  $\delta D_{alkenone}$  showed (Fig. 3) that, during the glacial  
260 period (39.4 to 16.2 ka), BIT index and  $\delta D_{alkenone}$  correlated significantly ( $R^2$  0.209, p value  
261 0.0001), whilst during the Holocene no statistically significant relationship was observed ( $R^2$

262 0.0186, p value 0.5249). This suggests that variation in freshwater input had a significant impact  
263 on the variability in  $\delta D_{\text{alkenone}}$  during the glacial. However, a negative correlation between BIT  
264 and  $\delta D_{\text{alkenone}}$  would be expected, as an increasing freshwater input would lead to an input of D-  
265 depleted water, as well as increasing fractionation due to a decrease in salinity. Thus, the trend  
266 cannot be explained by a salinity effect or by an increasing influx of D-depleted water.

267 Alternative factors, which could lead to the apparent positive correlation between BIT values and  
268  $\delta D$  of alkenones during the glacial period, are changes in growth rate or in haptophyte species  
269 composition (Schouten et al., 2006; Wolhowe et al., 2009; M'Boule et al., 2014). The reason why  
270 this effect is apparent in the glacial and not in the interglacial, may be because the core site was in  
271 much closer proximity to the coast during glacial sea level low stands (Walford et al., 2005; März  
272 et al., 2008; van der Lubbe et al., 2014), thereby allowing a potential proliferation of coastal  
273 haptophyte species as well as an increased impact of river-transported nutrients on growth rate.

274 Regarding the latter hypothesis, although high freshwater input might have provided nutrients to  
275 the core site, thereby promoting algal growth (Verity et al., 1993; Pedersen and Borum, 1996;  
276 Gillanders and Kingsford, 2002), increased growth rate would be expected to lead to a depletion  
277 in  $\delta D_{\text{alkenone}}$  (Schouten et al., 2006) and so cannot explain the positive excursions in  $\delta D_{\text{alkenone}}$ .

278 Since coastal haptophytes produce alkenones 90‰ enriched in D compared with open ocean  
279 haptophytes (M'Boule et al., 2014; Chivall et al., 2014), changes in marine vs. coastal  
280 haptophytes in the alkenone producing community could explain the positive excursions of ca.  
281 10‰ in hydrogen isotopic composition during the glacial period. The relative amount of the  $C_{37:4}$   
282 alkenone has been suggested as a marker for low salinity haptophyte algae (Schulz et al., 2000;  
283 Liu et al., 2008; 2011), but its abundance was very low in the sediment record (data not shown).

284 However, the hypothesis of a changing contribution to the alkenone pool is supported by the  
285 observed change in the ratio between  $C_{37}$  and  $C_{38}$  alkenones (Fig. 2). This ratio has been reported

286 to be more elevated for coastal haptophytes than for open ocean species (Marlowe et al., 1984;  
287 Prahl et al., 1988; Schulz et al., 2000; Liu et al., 2011). No significant positive anomalies in the  
288  $C_{37}/C_{38}$  ratio were apparent between 39 and 22 ka. Nevertheless,  $C_{37}/C_{38}$  ratio indicated elevated  
289 values during the glacial, with the highest at ca. 18.5 ka (Fig. 2), which coincides with relatively  
290 more enriched  $\delta D_{\text{alkenone}}$  values and elevated BIT values (Fig. 2). After 18.5 ka the ratio generally  
291 decrease to values of ca. 1.1 throughout the Holocene. This suggests that, during these periods of  
292 elevated  $C_{37}/C_{38}$  alkenone ratio, coastal haptophyte species may have contributed more to the  
293 alkenone pool and consequently shifted  $\delta D_{\text{alkenone}}$  towards more positive values at times of  
294 increased freshwater input. Changing haptophyte assemblages in coastal settings in response to  
295 changing environmental conditions appear, therefore, to have a strong effect on  $\delta D_{\text{alkenone}}$ . This  
296 highlights the care that has to be taken when using the  $\delta D_{\text{alkenone}}$  as a proxy for freshwater input in  
297 coastal settings or other environments where changes in the dominant alkenone producer(s) are  
298 likely to have occurred.

299

## 300 **5. Conclusion**

301 Despite changes in global seawater  $\delta D$  related to glacial – interglacial ice volume effects, no long  
302 term change in the 39 ka record of  $\delta D_{\text{alkenone}}$  off the Mozambique shelf was observed. We suggest  
303 that glacial – interglacial changes in the  $\delta D_{\text{alkenone}}$  were masked by changing freshwater input to  
304 the core site, as evidenced by the BIT index. During the glacial, the site was in closer proximity  
305 to the coast, when there was likely also larger river outflow. These conditions probably led to  
306 locally more D-depleted water and increased fractionation of hydrogen isotopes during alkenone  
307 biosynthesis due to decreased salinity. The correlation between periods of enriched  $\delta D_{\text{alkenone}}$  and  
308 elevated BIT values during the glacial is possibly related to an increased contribution from  
309 coastal alkenone-producing haptophyte species coinciding with elevated river outflow. This

310 complicates the application of  $\delta D_{\text{alkenone}}$  as a proxy for freshwater input in coastal settings.

311 However, the technique appears promising in more open ocean settings.

312

313

#### 314 **Acknowledgments**

315 We thank the two anonymous reviewers for their comments, which improved the manuscript. D. Chivall (Royal

316 NIOZ) is thanked for input to the manuscript. We acknowledge financial support from The Seventh Framework

317 Programme PEOPLE Work Programme, Grant 238512 (Marie Curie Initial Training Network 'GATEWAYS'). The

318 Netherlands Organization for Scientific Research (NWO) is acknowledged for funding M.v.d.M (VIDI), S.S. (VICI),

319 I.S.C. (NEBROC II) and R.T. (INATEX program, G.-J. B.; project number 839.08.434). R.T. acknowledges the

320 SCAN2 program on advanced instrumentation. A. Mets is thanked for laboratory assistance. We thank the Captain

321 and crew of the R/V Pelagia and participants of the INATEX cruise for collecting core 64PE304-80. Y. Wang at Kiel

322 University is acknowledged for extracting some of the 64PE304-80. Data presented here can be found at Pangaea.de;

323 doi:10.1594/PANGAEA.836271.

324 *Associate Editor – S. Wakeham*

325

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546 **Fig. 1.** Map of oceanic currents in the study region and core location. (a) Main ocean current  
547 systems in the Western Indian Ocean (white arrows), showing the South Equatorial Current  
548 (SEC), the Eastern Madagascar Current (EMC), the Mozambique Current (MC) and the greater  
549 Agulhas Current (Beal et al., 2011). The location of core 64PE304-80 is given by the red circle in  
550 the Mozambique Channel. Fig. (b) Shows an enlargement of the Mozambique Channel and the  
551 location of the Zambezi River mouth; the large gray arrow indicates the general current direction  
552 of the Mozambique Current, small gray arrows coastal circulation cells, and white shaded area  
553 represents the sediment distribution of Zambezi River fan (Sætre and Da Silva, 1984).

554

555 **Fig. 2.**(a) Temperature variation over Greenland indicated by the  $\delta^{18}\text{O}$  record of GRIP ice core  
556 (blue; Greenland Ice-core Project, 1993); (b)  $\delta\text{D}_{\text{alkenone}}$  record (red line) with analytical error ( $1\sigma$ ),  
557 (c)  $\text{C}_{37}/\text{C}_{38}$  alkenone ratio (black line) and (d) BIT index (green) for core 64PE304-80; (e) mean  
558 global benthic  $\delta^{18}\text{O}_{\text{sw}}$  curve (Waelbroeck et al., 2002). Gray shaded bars indicate Younger Dryas  
559 (YD) (ca. 12.5-11 ka), Heinrich Event 1 (HE1) (ca. 19-14.6 ka), Last Glacial Maximum (LGM;  
560 ca. 23-19 ka) and Heinrich Event 2 (HE2; ca. 24 ka). Radiocarbon samples are indicated as black  
561 triangles.

562

563 **Fig. 3.** Cross plot of  $\delta\text{D}_{\text{alkenone}}$  vs. BIT index. Values for Holocene (ca. 12.7 – 0.5 ka) are indicated  
564 by closed gray circles, regression II (dashed line). Values for pre-LGM (ca. 39.4 – 16.1 ka) are  
565 indicated by open squares, regression I (solid line).

566