REVIEW ARTICLE

Etiology of the Protein-Energy Wasting Syndrome in Chronic Kidney Disease: A Consensus Statement From the International Society of Renal Nutrition and Metabolism (ISRNM)

Juan Jesús Carrero, PhD,* Peter Stenvinkel, MD, PhD,* Lilian Cuppari, PhD,† T. Alp Ikizler, MD, PhD,‡ Kamyar Kalantar-Zadeh, MD, PhD,§ George Kaysen, MD, PhD,¶ William E. Mitch, MD, PhD,** S. Russ Price, PhD,††‡‡ Christoph Wanner, MD, PhD,§§ Angela Y. M. Wang, MD, PhD,¶¶ Pieter ter Wee, MD, PhD,*** and Harold A. Franch, MD††‡‡

Protein-energy wasting (PEW), a term proposed by the International Society of Renal Nutrition and Metabolism (ISRNM), refers to the multiple nutritional and catabolic alterations that occur in chronic kidney disease (CKD) and associate with morbidity and mortality. To increase awareness, identify research needs, and provide the basis for future work to understand therapies and consequences of PEW, ISRNM provides this consensus statement of current knowledge on the etiology of PEW syndrome in CKD. Although insufficient food intake (true undernutrition) due to poor appetite and dietary restrictions contribute, other highly prevalent factors are required for the full syndrome to develop. These include uremia-induced alterations such as increased energy expenditure, persistent inflammation, acidosis, and multiple endocrine disorders that render a state of hypermetabolism leading to excess catabolism of muscle and fat. In addition, comorbid conditions associated with CKD, poor physical activity, frailty, and the dialysis procedure per se further contribute to PEW.

Published by Elsevier Inc. on behalf of the National Kidney Foundation, Inc.

This article has an online CPE activity available at www.kidney.org/professionals/CRN/ceuMain.cfm

Introduction

A SYNDROME OF adverse changes in nutrition and body composition is highly prevalent in patients with chronic kidney disease (CKD), especially in those undergoing dialysis, and it is associated with high morbidity and mortality. A summary of the mechanisms involved in these alterations is provided in Figure 1. Although insufficient food intake (true undernutrition) due to poor appetite and dietary restrictions contributes to these problems, there are features of the syndrome that cannot be explained by undernutrition alone. Many contributing causes are directly related to kidney

*Division of Renal Medicine, Department of Clinical Science, Intervention and Technology, Karolinska Institutet, Solna, Sweden.

[†]Division of Nephrology, Department of Medicine, Federal University of Sao Paulo, Sao Paulo, Brazil.

[‡]Department of Medicine, Division of Nephrology, Vanderbilt University School of Medicine, Nashville, Tennessee.

[§]Harold Simmons Center, Division of Nephrology and Hypertension, University of California Irvine Medical Center, Orange, California. [¶]Departments of Internal Medicine and Biochemistry and Molecular Medicine,

¹Departments of Internal Medicine and Biochemistry and Molecular Medicine, University of California–Davis, Davis, California.

** Nephrology Division, Department of Medicine, Baylor College of Medicine, Houston, Texas.

^{††}Renal Division, Department of Medicine, Emory University School of Medicine, Atlanta, Georgia.

^{‡‡}Research Service, Atlanta Department of Veterans Affairs Medical Center, Decatur, Georgia.

^{§§}Division of Nephrology, Department of Internal Medicine, University of Würzburg, Würzburg, Germany.

[¶]Department of Medicine, Queen Mary Hospital, University of Hong Kong, Hong Kong, Hong Kong. *** Department of Nephrology, VU Medical Center, Amsterdam, The Netherlands.

Financial Disclosure: J.J.C. has received funding from Abbott Nutrition, Shire (honoraria); P.S. has been a consultant for Abbott Renal Care, Bayer, Baxter Renal, Diaverum, Amgen, Takeda, Gambro; T.A.I. has been a consultant for Abbott Renal Care, Abbott Nutrition, DSI, Inc., Baxter Renal, Amgen, Affymax, Inc., Fresenius Medical Care North America, Fresenius-Kabi, Satellite Healthcare; K.K-Z. has been funded by Abbot Renal Nutrition (honoraria); G.K. has been a consultant for Merck, Renal Research Institute; A.Y.M.W has received monetary compensation to speak from Fresenius Kabi, Sanofi Renal, Baxter Healthcare (honoraria), Abbvie, Sanofi Renal, and grants funding from Baxter Healthcare. All other authors declare that they have no relevant financial interests.

Address correspondence to Harold A. Franch, MD, Renal Division, Emory University School of Medicine, WMRB #3301, 1639 Pierce Drive, Atlanta, GA 30306. E-mail: hfranch@emory.edu

Published by Elsevier Inc. on behalf of the National Kidney Foundation, Inc. 1051-2276/\$36.00

http://dx.doi.org/10.1053/j.jrn.2013.01.001

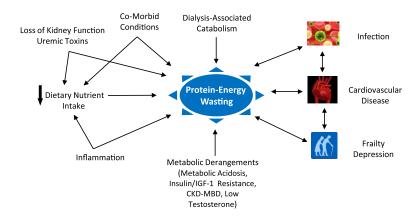


Figure 1. A conceptual model for etiology of PEW in CKD and direct clinical implications. PEW is the result of multiple mechanisms inherent to CKD, including undernutrition, systemic inflammation, comorbidities, hormonal derangements, the dialysis procedure, and other consequences of uremic toxicity. PEW may cause infection, CVD, frailty, and depression, but these complications may also increase the extent of PEW.

disease, including increased resting energy expenditure (REE), persistent inflammation, acidosis, multiple endocrine disorders, and the dialysis procedure itself. However, this syndrome shares etiologic factors that contribute to cachexia in non-CKD populations, including comorbid conditions associated with cachexia, decreased physical activity, frailty, and aging. The CKD and end-stage renal disease (ESRD) populations are unique in the constant surveillance that facilitates the

Table 1. Causes of PEW in CKD Patients

- 1. Decreased protein and energy intake
 - a. Anorexia
 - i. Dysregulation in circulating appetite mediators
 - ii. Hypothalamic amino acid sensing
 - iii. Nitrogen-based uremic toxins
 - b. Dietary restrictions
 - c. Alterations in organs involved in nutrient intake
 - d. Depression
 - e. Inability to obtain or prepare food
- 2. Hypermetabolism
 - a. Increased energy expenditure
 - i. Inflammation
 - ii. Increased circulating proinflammatory cytokines
 - iii. Insulin resistance secondary to obesity
 - iv. Altered adiponectin and resistin metabolism
 - b. Hormonal disorders
 - i. Insulin resistance of CKD
 - ii. Increased glucocorticoid activity
- 3. Metabolic acidosis
- 4. Decreased physical activity
- 5. Decreased anabolism
 - a. Decreased nutrient intake
 - b. Resistance to GH/IGF-1
 - c. Testosterone deficiency
 - d. Low thyroid hormone levels
- 6. Comorbidities and lifestyle
 - a. Comorbidities (diabetes mellitus, CHF, depression, coronary artery disease, peripheral vascular disease)
- 7. Dialysis
 - a. Nutrient losses into dialysate
 - b. Dialysis-related inflammation
 - c. Dialysis-related hypermetabolism
 - d. Loss of residual renal function

diagnosis of wasting before frank cachexia begins. Given the unique features of the syndrome, the International Society of Renal Nutrition and Metabolism (ISRNM) proposed a common nomenclature and diagnostic criteria for these alterations in the context of CKD.¹ Protein-energy wasting (PEW) was proposed to denote concurrent losses in protein and energy stores, with cachexia being regarded as only the end stage. ISRNM's intention was to begin creating a framework to identify and understand disorders that promote PEW.² To further this process, the ISRNM now provides a consensus review of current knowledge on the etiology of PEW in kidney disease (Table 1) to provide a basis for future advances in diagnosis and therapy and to identify gaps in knowledge for future research.

Undernutrition and Anorexia

Low energy and/or protein intake associates with a significant decline of nutritional parameters (including hypoalbuminemia) and increased risk of morbidity and mortality in patients with advanced CKD.^{3,4} In most of these studies, dietary energy and protein intakes are lower than recommended for patients undergoing either hemodialysis (HD)^{3,5,6} or peritoneal dialysis (PD).^{7,8} However, dietary recalls underestimate dietary intake,⁹⁻¹¹ and improving accuracy of dietary monitoring is needed. There is presently limited information correlating dietary composition, including micro/macronutrient intake, with outcomes.¹²⁻¹⁴ In one study that was based on foodfrequency questionnaires, HD patients consume significantly lower amounts of potassium, dietary fiber, vitamin C, and certain cardioprotective carotenoids.¹⁵ Data from the Third National Health and Nutrition Examination Survey showed that high dietary total fiber intake was associated with lower risk of inflammation and mortality in CKD patients.¹⁶ Many of the restrictions in renal diets contradict current recommendations for healthy eating. Although limiting dietary sodium, phosphate, potassium, and fluid intake prevents important patient complications, problems arise when these restrictions are not accompanied

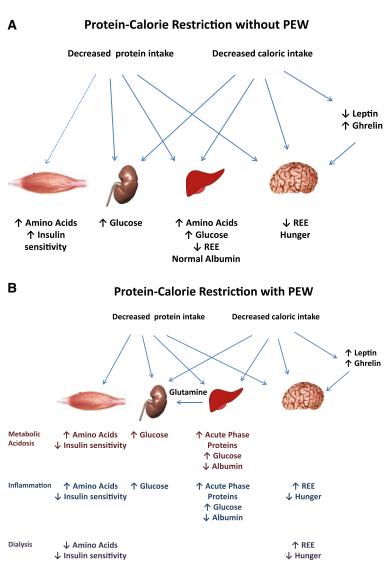
with appropriate counseling on alternative food choices and/or strategies to ensure adequate nutrient intake.^{17,18}

Anorexia often drives inadequate protein and energy intake and directly contributes to poor quality of life.^{6,19-21} The prevalence of anorexia has been reported at 35% to 50% of ESRD patients.^{22,23} Although few studies exist in CKD stages 1-3, a progressive, spontaneous decrease in food intake occurs with greater loss of kidney function, which correlates with accumulation of nitrogen-derived uremic toxins.²⁴⁻²⁶ The factors influencing food intake involve not only metabolic signals but also anomalies in the digestive system and psychological and acquired aspects, including a desire for pleasure, social behavior, and customs.²⁷ Anorexia may be mediated by circulating appetite regulators, such as gastric mediators (such as cholecystokinin,²⁸ peptide YY,²⁹ ghrelin,^{30,31} or obestatin), adipokines (such as leptin³² and visfatin³³), or cytokines

Figure 2. Response to reduced dietary protein and energy intake. (A) Normal response. Reduced dietary protein and energy drive an increase in hunger and a fall in REE, loss of protein preferentially from the visceral organs, and increased insulin sensitivity of muscle. The liver and kidney provide glucose, and serum albumin is maintained at a normal level. (B) Response with PEW. During PEW, the adaptations to increase hunger and lower REE are blunted in part by an increased half-life of leptin and ghrelin and in part by inflammation and dialysis. The loss of protein occurs preferentially from muscle because of the effects of metabolic acidosis, glucocorticoids, and inflammation, leading to increased insulin resistance. Dialysis results in the loss of amino acids, stimulating muscle protein breakdown. Under the influence of inflammation and metabolic acidosis, the liver makes glutamine for deamination in the kidney, increases acute-phase reactants, and reduces serum albumin. The kidney increases glucose production from glutamine under the influence of metabolic acidosis.

(such as tumor necrosis factor [TNF], interleukin [IL]-6, and IL-1 $\beta^{19,20}$), but these mediators need additional research in the uremic milieu.³⁴ Signaling by hypothalamic neurons that sense the ratio of essential to nonessential amino acids³⁵ may be influenced by the fall in branchedchain amino acid levels with uremia or dialysis, creating the so-called brain hyperserotoninergic-like syndrome. The role of other complications of uremia on anorexia need to be further explored, including dental and oral problems (such as palatability problems or incidence of periodontitis³⁶), gastric alterations³⁷ (motility disorders,³⁸ dyspepsia,³⁹ or bacterial infections in the intestine⁴⁰), and depression.^{21,41}

Although reduced intake of food or poor absorption of nutrients plays a critical role in most cases of PEW,^{42,43} the science of starvation suggests that additional mechanisms are needed for PEW to occur (Fig. 2). Decreased



energy intake reduces insulin secretion and stimulates the production of sugar from glycogen and increased mobilization of fatty acids.⁴⁴ Activation of these systems contributes to a reduction in basal metabolic rate and mobilization of free fatty acids and amino acids.^{44,45} Muscle proteolysis only transiently increases in early starvation, but muscle release of amino acids declines over the first 2 weeks of starvation and visceral organ proteins are used preferentially to muscle.⁴⁶ Muscle and visceral proteins can be preserved to some extent because of heightened insulin sensitivity, and diets with as little as 0.55 g/kg/day of balanced protein may be well tolerated.⁴⁷ Below that level, the loss of visceral protein and increases in lipolysis lead to fatty infiltration of the liver and decreased plasma protein synthesis.⁴⁸ However, plasma proteins, particularly prealbumin and S-albumin, have increased half-life and do not change in concentration with moderate calorie or protein restriction alone.^{49,50} Generally, other factors in addition to starvation (especially inflammation and acidosis) are required for accelerated muscle loss and hypoalbuminemia. However, depletion of visceral protein stores caused by prolonged decreased energy intake or frequent intermittent starvation causes disruption of certain protective mechanisms. Heavy ketone body formation marks a transition in metabolism to more severe starvation and causes a loss of the adaptation that prevented hypoalbuminemia and limited muscle wasting earlier in starvation.^{44,51} The acid and the ketone bodies in severe starvation appear to be critical in making protein loss from muscle greater than from other organs and making amino acids a critical source of glucose.⁴⁶

Hypermetabolism Increased Energy Expenditure

In simple starvation, the body reduces energy expenditure to conserve energy needs. REE is usually normal in stable maintenance dialysis or CKD patients. In contrast, REE increases from 12% to 20% in CKD patients during the HD procedure⁵² or in the presence of comorbidities such as cardiovascular disease (CVD),⁵³ severe hyperparathyroidism,⁵⁴ poorly controlled diabetes,⁵⁵ inflammation, PEW,^{53,56,57} and loss of residual kidney function.⁵³ In PD patients, PEW was more frequent among patients in the highest tertile of REE when compared with those in the lower tertile.⁵³ Because protein catabolism and inflammation result in elevated energy expenditure,⁵⁸ higher energy intake alone should not correct increased REE under these circumstances (although this has not been rigorously tested). Increased REE is frequently mitigated by decreased physical activity, leading to a reduction, rather than an increase, in total energy expenditure in some studies.^{59,60}

Persistent Inflammation

Inflammation overcomes the adaptive responses protecting muscle and reducing REE during decreased protein and energy intake. Inflammation activates intracellular NADPH oxidases, creating signals that induce muscle insulin resistance.⁶¹ The inflammatory response is associated with a rise in REE, which can be so severe that starvation responses are activated in well-fed individuals.^{44,61} Inflammation is associated with a decline in albumin concentration and reduces the synthesis and half-life of albumin.⁶² Inflammation explains the requirement for infection to promote edema and hypoalbuminemia in kwashiorkor.⁴⁸ Protein, DNA, and lipid oxidation occur in severe starvation as a result of depletion of dietary antioxidants, exhaustion of autophagy, depletion of protein stores, and/or from inflammation.^{50,63} It is interesting to note that increased oxidative signaling is associated with muscle insulin resistance, muscle wasting, and atherosclerotic disease.^{51,64} Thus, inflammation causes increased REE, preferential muscle loss, and oxidation.

Inflammatory markers are increased in most conditions associated with loss of muscle mass, including CKD,65-68 cancer, congestive heart failure (CHF), chronic pulmonary disease, acquired immune deficiency syndrome, and aging.⁶⁹ Muscle loss due to inflammation has been ascribed to inflammatory cytokines.^{65,70} Animal studies show that infusion of TNF, IL-1, and IL-6 causes an increase in muscle protein breakdown, resulting in muscle atrophy.⁶⁹ Proinflammatory cytokines also act on the central nervous system to decrease appetite⁷¹ and increase REE.⁷² Proinflammatory cytokines impair insulin/insulin-like growth factor (IGF)-1 signaling by augmenting the level of glucocorticoids (see Impairment of Insulin/IGF-1) and by directly inducing insulin and IGF-1 resistance in skeletal muscle.⁷³ Multiple studies show that high circulating levels of IL-6, a prominent biomarker of inflammation, contribute to inflammatory muscle protein losses.⁷⁴ In part, these losses are triggered by alteration of IL-6 signaling due to interaction with acute-phase proteins, including serum amyloid A, to impair insulin/IGF-1 signaling via the activator of transcription 3 and suppressor of cytokine signaling 3.⁷⁴ Ineffective utilization of exogenous amino acids for muscle protein synthesis during HD has been linked to increased skeletal muscle expression of IL-6.75 In uremic skeletal muscle, IL-6 has also been linked to increased caspase-3 activity (an initial step resulting in loss of muscle protein).76

Myostatin, a member of the transforming growth factor (TGF)- β superfamily of proteins, is induced by CKD in mouse models via cytokine-activated pathways, and down-regulating the myostatin receptor improved IGF-1 signaling, enhanced satellite cell function, and suppressed inflammatory cytokines.⁷⁷ Significantly, inflammation-induced increase in muscle protein degradation in CKD can be blocked by a humanized antibody inhibiting the function of myostatin, leading to increased muscle growth, suppression of the levels of inflammatory cytokines, and improvement in insulin/IGF-1 resistance.⁷⁸ Consistent with a role of myostatin in PEW, its endogenous inhibitor,

follistatin, is induced by exercise,⁷⁹ one of the few interventions that increases muscle strength and mass in CKD.⁸⁰ However, in CKD patients, follistatin is positively correlated with inflammation and PEW resistance to its action.⁸¹ Thus, it is possible that inflammation or CKD alters the balance between follistatin, and myostatin to regulate muscle mass in uremia and that intervening in myostatin signaling might preserve muscle and/or reduce inflammation.⁸²⁻⁸⁴

TNF-related weak inducer of apoptosis (TWEAK), a member of the TNF superfamily⁸⁵ binds to its receptor (Fn14) linked to signaling pathways involved in the regulation of nuclear factor kappa light-chain enhancer of activated B cells (NF- κ B), myogenesis, and apoptotic cascades.⁸⁶ TWEAK-Fn14 expression is induced in animal models of tissue injury and inflammation, and biomarker studies show a significant interaction between soluble TWEAK and IL-6 in the prediction of mortality and reduced muscle strength in HD patients.⁸⁵ Recently, alternative pathways of $NF-\kappa B$ activation have been identified that regulate distinct forms of NF-*k*B and its effectors.⁸⁷ Finally, complex regulation of IL-15, an immunoregulatory cytokine with proinflammatory activity but also paradoxical anabolic functions, may play a role in insulin/IGF-1 resistance.88

Abdominal Obesity and Adipokines

Observational studies indicate improved survival in obese patients undergoing HD. Thus, it is hypothesized that dialysis patients at high risk of PEW are protected by excess weight. However, obesity does not necessarily imply good nutritional status, and muscle wasting occurring despite fat accumulation in the general population has been termed "obese sarcopenia".⁸⁹ Furthermore, the regional fat distribution has metabolic implications. Abdominal subcutaneous tissue in otherwise healthy subjects is proinflammatory and CKD patients have increased expression of proinflammatory cytokines and adipokines in abdominal subcutaneous tissue compared with healthy controls.⁹⁰⁻⁹² Observational studies in CKD patients link abdominal fat with inflammation, insulin resistance, hyperadipokinemia, dyslipidemia and oxidative stress, ^{91,93-98} and cardiovascular events. ⁹⁹ In a large cohort of prevalent HD patients, each kilogram of body mass index (BMI) increase reduced the risk of dying whereas, concomitantly, each centimeter increase of waist circumference raised mortality risk.¹⁰⁰ Thus, although a high BMI in the setting of CKD may signal health and better nutritional status, abnormal deposition of abdominal fat may be detrimental because of metabolic derangements. This concept was demonstrated in a study of disproportional fat mass accumulation in HD patients by modeling the body as a bicone centered on the waist.¹⁰¹ In addition, the recent observations that waist circumference modifies the mortality risk associated with circulating triglycerides,¹⁰² leptin, and adiponectin¹⁰³ underscores the overall effect that abdominal obesity has on PEW.

Although leptin inhibits food intake and increases energy consumption via the hypothalamic melanocortin system,³ evidence is lacking that the markedly elevated circulating leptin level in uremia contributes clinically to anorexia and PEW.⁹⁰ In fact, the positive association between circulating leptin levels and improved nutrition in CKD suggests that uremia is a state of leptin resistance.^{27,104,105} Although early reports in CKD showed that higher adiponectin levels are linked to better outcomes,¹⁰⁶ recent studies showed the opposite.¹⁰⁷ Lower fat mass in PEW increases circulating adiponectin, causing adiponectin to lose its association with mortality after adjustment for BMI in diseases such as CHF.¹⁰⁸ Adiponectin has anti-inflammatory, antiatherogenic, and insulin sensitizing actions, and increased adiponectin has been suggested to be a "reparatory response" to the microvascular insults in uremia,¹⁰⁹ but experimental data suggest that adiponectin also promotes weight loss via increased energy expenditure.¹¹⁰ Therefore, although adiponectin is a biomarker of PEW, its role in pathogenesis remains to be determined.

Visfatin is expressed in human atherosclerotic plaques and is associated with plaque destabilization, independently predicting coronary artery disease in humans.¹¹¹ Although studies are limited, visfatin in CKD is positively associated with endothelial dysfunction and inflammation and negatively associated with HDL cholesterol.^{90,112} Visfatin may also be involved in appetite regulation and nutrient homeostasis,⁹⁰ and elevated visfatin levels were associated with loss of appetite and low fasting serum amino acids in dialysis patients.³³ The inconsistency in PEW is that plasma visfatin in normal individuals is related positively to fat mass. However, Hallschmid et al.¹¹³ found that visfatin in human cerebrospinal fluid was negatively correlated with fat mass, suggesting that central nervous system visfatin insufficiency and/or resistance drives higher plasma levels.

Hormonal Disorders

Impairment of Insulin/IGF-1

As a direct consequence of the kidneys' role as modulator of endocrine function, kidney disease causes abnormalities in the excretion, synthesis, and action of many hormones. Resistance to insulin, growth hormone (GH), and IGF-1 are implicated in loss of muscle mass in adult CKD patients. Insulin or IGF-1 bind distinct cell surface receptors to activate similar downstream signaling pathways, which act to prevent loss of muscle protein.¹¹⁴ When muscle is lost, large multinucleated myofibers decrease in size rather than decrease in number. Regenerative systems that involve the fusion of muscle cell precursor cells (i.e., satellite or stem cells) with myofibers are also inhibited. Although current evidence suggests that myofiber shrinkage due to accelerated protein degradation is the predominant mechanism for loss of muscle mass, myofiber shrinkage and satellite cell fusion are regulated by insulin and IGFs. This has led to the hypothesis that the integrated outputs of these insulin/IGFactivated signaling pathways determine the balance between protein accretion and loss, determining overall changes in muscle mass.

The effect of low insulin on muscle is clear: Uncontrolled type 1 diabetes mellitus leads to negative nitrogen balance, lean tissue atrophy, and hyperaminoacidemia easily reversed through the provision of insulin.¹¹⁵ The net protein anabolic effect of insulin involves a blunting of proteolvsis rather than enhanced protein synthesis. Alterations in insulin function in uremia were reported as early as 1951,¹¹⁶ and the alterations in glucose metabolism in the face of hyperinsulinemia and diminished tissue sensitivity to insulin are partially correctable by HD.^{117,118} In insulin-deprived animals, muscle protein breakdown is significantly increased, a process that is mediated by the proteasome-ubiquitin pathway.^{110,111} Enhanced protein catabolism applies to insulin-deficient and insulinresistant states. HD patients with suboptimally controlled type 2 diabetes have a higher rate of muscle protein loss than HD patients without diabetes.¹¹⁹ Altered insulin sensitivity is primarily due to a postreceptor defect altering primarily skeletal muscle, rather than hepatic glucose uptake. Furthermore, the extent of insulin resistance correlates with muscle protein breakdown in HD patients who are not diagnosed with diabetes mellitus.¹²⁰ Individual uremic toxins removed by dialysis (such as P-cresol, the byproduct of tyrosine metabolism) have been shown to induce insulin resistance.¹²¹ Insulin resistance represents a major target for intervention in PEW. For example, treatment with an insulin sensitizer (PPAR γ agonist, rosiglitazone) suppressed muscle proteolysis in insulin-resistant mice.¹²² It is not surprising that the use of rosiglitazone treatment was associated with significantly lower all-cause mortality and higher Salbumin among insulin-free, but not insulin-requiring, diabetic HD patients.¹²³

Uremia, inflammatory cytokines, acidosis, glucocorticoids, and angiotensin (ANG) II share a common mechanism of muscle wasting: impairment of insulin/IGF-1 actions by altering signaling through the phosphatidylinositol 3-kinase (PI3-kinase)/Akt pathway.^{124,125} Although the precise signals causing insulin/IGF-1 resistance in CKD are unknown, several steps in PI3-kinase/Akt signaling contribute to the impairment, including activation of the FoxO family transcription factors that induce the expression of several atrophy-inducing genes involved in the ubiquitin-proteasome and autophagic proteolytic systems.¹²⁶ Dysfunctional PI3-kinase/Akt activity can also result in activation of caspase-3, an apoptotic protease that also degrades actin in actomyosin complexes.¹²⁷ A byproduct of this proteolytic reaction is a characteristic actin fragment that was shown to serve as a biomarker of muscle wasting in HD patients and others with conditions associated with muscle wasting.⁷⁶

Testosterone Deficiency and Low Thyroid Hormone Levels

Prolactin retention in CKD impairs the production of gonadotropic hormones in men and women.¹²⁸ In men, this translates into a high prevalence of testosterone deficiency (hypogonadism).¹²⁹⁻¹³² Testosterone levels are also abnormally low among women. Testosterone is an anabolic hormone that induces skeletal muscle hypertrophy by promoting nitrogen retention, stimulating fractional muscle protein synthesis, inducing myoblast differentiation, and augmenting the efficiency of amino acid reuse by skeletal muscle. Testosterone suppresses myostatin expression, inhibits apoptosis, induces muscle IGF-I mRNA expression, and affects the differentiation of mesenchymal-derived pluripotent stem cells into myocytes.¹³³ In dialysis and predialysis patients, low testosterone levels were associated with increased mortality risk, ¹³⁴⁻¹³⁶ and the observation that adjustment for serum creatinine levels (a surrogate marker of muscle mass) abrogated this mortality prediction¹³⁴ may indirectly support this pathophysiological mechanism. In CKD stage 2-4, endogenous testosterone was an independent determinant of bioelectrical impedance analysis-estimated muscle mass and muscle strength (handgrip).¹³⁷ Randomized intervention studies with androgen therapy in CKD patients (alone or in combination with resistance training) have shown significant improvement of muscle mass and nutritional status.^{138,139}

Available data cannot distinguish if low thyroid hormone levels in CKD patients with PEW are an adaptation that reduces energy expenditure and minimizes protein catabolism or a maladaptation participating in the wasting syndrome.¹⁴⁰ Low triiodothyronine levels in CKD stage 5 patients correlate with systemic inflammatory markers, endothelial dysfunction, and all-cause as well as cardiovascular mortality.¹⁴¹⁻¹⁴⁴ Correction of metabolic acidosis in dialysis patients improves these hormonal derangements.^{145,146} The correlation of triiodothyronine with mortality prediction was abrogated after adjustment for C-reactive protein and albumin as surrogates of PEW.¹⁴⁷ Thus, even if low thyroid hormone participates in the PEW process and is not adaptive, then changes in thyroid hormones may act as an intermediate link among inflammation, acidosis, PEW, and mortality and not a primary cause.

Metabolic Acidosis and Glucocorticoids

Metabolic acidosis is a key mechanism in the starvation response, inducing release of branched-chain amino acids from muscle during ketosis. It also causes insulin resistance leading to loss of muscle mass. Acidosis does not alter insulin/IGF-1 receptor binding, but rather it inhibits intracellular signaling. In a rat model of CKD-induced acidosis, intracellular pH of myofibers was not changed,¹⁴⁸ but an acidic extracellular pH is sufficient to reduce postreceptor signaling through insulin/IGF-1 pathways in cultured muscle cells.¹⁴⁹ However, a decline in extracellular pH by itself is not sufficient to induce muscle wasting in a rat CKD model.^{73,114,122,148,150,151} Metabolic acidosis induces increased adrenal glucocorticoid production, and adrenalectomized rats have markedly reduced muscle wasting that is restored by replacement of glucocorticoids. Glucocorticoids induce insulin/IGF-1 resistance in skeletal muscle by altering the same signaling pathways that are affected by acidosis, but they act on slightly different signaling molecules within the pathway.¹²⁶ Notably, prevailing evidence from other comorbidities of CKD, including ANG II and inflammation, indicates that insulin/IGF-1 resistance and elevated glucocorticoids are the common physiological responses that are causative for the increase in protein and amino acid catabolism as well as the suppression of protein synthesis.74,152,153 Understanding this coordinated response may provide additional clues in how insulin/ IGF1 signaling controls muscle wasting.

Correction of acidosis has salutary effects on nutritional parameters. For example, sodium bicarbonate stimulates the growth of premature infants and that of children with renal tubular acidosis.^{138,154} Likewise, treatment with sodium bicarbonate and potassium bicarbonate improves the nitrogen balance of elderly women with mild metabolic acidosis.^{155,156} In normal adults, induction of metabolic acidosis not only decreases serum albumin concentration but also stimulates nitrogen losses due to accelerated degradation of protein and essential amino acids.^{157,158} Similar adverse consequences occur in CKD patients who develop metabolic acidosis: Acidosis decreases serum levels of essential branched-chain amino acids in muscle although muscle protein degradation is accelerated. Both abnormalities correct when metabolic acidosis is treated.¹⁵⁹ Likewise, in patients being treated by HD or PD, treatment of metabolic acidosis reduced the excessive rate of protein degradation.^{160,161} Longterm clinical trials show similar results in PD and CKD.^{162,163} Thus, acidosis contributes to PEW, and correction of acidosis ameliorates it.

Comorbidities and Lifestyle Comorbidities

Typical comorbidities associated with CKD or ESRD contribute to a catabolic milieu and the development of PEW. As Table 2 demonstrates, these factors share common etiologic mechanisms with PEW. Given its high prevalence in CKD patients, diabetes may be the most important single comorbidity. Pupim et al.¹⁶⁴ showed that diabetes mellitus is an important predictor of lean body mass loss in dialysis patients, and reduced insulin signaling by insulin absence or resistance results in increased muscle protein breakdown.¹¹⁹ Diabetes also causes CVD and neuropathy that contribute to infection, muscle atrophy, and diabetic gastroparesis (with resultant food intake impairment). One out of three long-term diabetic dialysis patients no longer require hypoglycemic therapy. The poor outcomes in this subgroup with "burnt-out diabetes" may be the result of PEW.¹⁶⁵

Another common comorbid state in CKD patients is CVD, in particular CHE.¹⁶⁶ Cardiac cachexia is in many ways indistinguishable from uremic PEW and shares important mechanisms. Inadequate cardiac output drives neurohumeral responses associated with PEW including glucocorticoids, increased ANG II, and sympathetic nerve activity. Right ventricular heart failure with passive

Table 2. Typical Comorbidities in CKD Patients that Contribute to PEW

Comorbitiy	Possible Effects Related to Etiology of PEW
Diabetes/metabolic syndrome	Gastroparesis, inflammation/oxidation, insulin deprivation (type I), insulin resistance, pain
CVD/heart failure	Cachexia, inflammation, glucocorticoid release, sympathetic nerve overactivity, increased circulating ANG II, insulin resistance, decreased activity, pain
Peripheral vascular disease	Reduced activity, ulcers, inflammation, pain
Fluid overload	Inflammatory cytokine release, gut edema, leg ulcers, decreased physical activity, pain
Hyperparathyrodism, CKD-MBD	Increased energy expenditure, glucose intolerance, hypovitaminosis D, muscle wasting, gastric ulcers, heart disease
Anemia	Frailty, decreased activity, iron-deficiency, high output heart failure
Autoimmune/rheumatologic disorders	Inflammation, intercurrent infections, joint involvement reduces activity, glucocorticoids, pain
Gastrointestinal disorders	Anorexia, swallowing disorders, nutrient malabsorption, acidosis (diarrhea, drains, and ostomies), inflammation, infections, pain
Chronic lung diseases	Increased work of breathing (raises REE), decreased activity, intercurrent infections, inflammation, glucocorticoids, fluid overload, ANG II
Liver disease	Hypoalbuminemia, volume overload, ANG II, infections, inflammation, acidosis (bowel therapies), anorexia, pain
Infections	Inflammation, reduced appetite, increased REE, pain
Pain	Anorexia, constipation (narcotics), inflammation
Psychiatric disorders/depression/dementia	Unwillingness to eat, anorexia, inability to obtain/prepare food, inflammation, decreased activity
Neurologic disorders	Reduced activity, anorexia, swallowing disorders, pain
Malignant diseases	Cancer cachexia, inflammation, increased REE, decreased activity, pain

congestion of the liver and gut wall edema can be associated with alterations in nutrient absorption, anorexia, and gut mucosal barrier function.^{166,167}

CKD-metabolic bone disease (CKD-MBD) is a comorbid condition associated with PEW. PEW can contribute to CKD-MBD because loss of body weight, inflammation, and physical inactivity lead to bone loss. Certain conditions associated with CKD such as protein losses and anorexia can predispose these patients to hypovitaminosis D. What is more controversial is the extent to which CKD-MBD contributes to the etiology of PEW. Low circulating levels of vitamin D, a decrease in klotho, and a rise in fibroblast growth factor-23 can increase parathyroid hormone synthesis, thereby contributing to the development of secondary hyperparathyroidism.¹⁶⁸ Vitamin D and/or parathyroid hormone have long been considered contributors to PEW, and vitamin D appears to be involved in some key molecular pathways for PEW and muscle regulation.¹⁶⁹ Experimental animal models of kidney failure and humans show improvements in muscle size and strength, markers of muscle metabolism, and/or serum albumin with replacement of 1,25-dihydroxyvitamin D.^{170,171} Finally, there is a positive association between 25-hydroxyvitamin D and testosterone levels, suggesting an additional mechanism in which vitamin D may regulate muscle mass.¹⁷²

Symptoms of depression are more frequent with reduction in kidney function and relate to poor outcome and mortality.¹⁷³⁻¹⁷⁵ Cytokines are thought to be important mediators of brain immune connection and may play an important role in the pathogenesis of depression because of their effect on neurotransmitters and neurohormones.^{173,176} In dialysis patients, symptoms of depression are associated with increased IL-6 levels,^{177,178} and 8 weeks of fluoxetine treatment resulted in decreased serum IL-1 β levels.¹⁷⁹ Depression may also lead to fatigue¹⁸⁰ and unwillingness to eat,²³ contributing in a vicious circle to anorexia, physical inactivity, and overall PEW.

Poor Physical Activity, Frailty

Decreased physical activity likely plays a major role in the etiology of PEW and the associated cardiovascular mortality. This is in part because individuals with low physical activity have increased risk of CKD secondary to obesity, diabetes, hypertension, and heart disease, and common comorbidities in CKD patients, especially heart disease and aging, are partly associated with decreased ability to exercise. Furthermore, complications of CKD including anemia, volume overload, and muscle wasting limit exercise ability. Subjects with stages 3-5 CKD have lower median peak oxygen consumption, and in some individuals this limits exercise enough to prevent activities of daily living.¹⁸¹ Muscle weakness is common in stage 5 CKD by measures including grip strength, rising from a chair, and maximum gait speed.¹⁸¹ In the geriatric population, IL-6 level is associated with current physical performance and

predicts a future decline in physical activity.¹⁸² Lack of exercise can increase inflammatory markers, and this change, rather than the decreased muscle mass, may dominate the association with mortality.¹⁸³

In CKD, just as in the diabetic patients, exercise-induced benefits in muscle are likely due to improvement in insulin/ IGF-1 sensitivity of skeletal muscle.¹⁸⁴ Endurance exercise decreased the rate of protein degradation in the muscle of CKD rats,¹⁸⁵ and aerobic exercise reduced the amount of caspase-3-generated 14-kDa actin biomarker in muscle from HD patients.¹⁸⁵ Resistance exercise also has beneficial effects on muscle in CKD patients by increasing mitochondrial biogenesis.¹⁸⁶ Because these anabolic actions are attributed to insulin/IGF-1 signaling, it is possible that so called "exercise mimetics" (e.g., resveratrol) will prove equally effective at slowing muscle loss in CKD patients.¹⁸⁷ Regardless of mechanism, a Cochrane review of trials to improve physical activity in CKD concluded that the interventions improved markers of PEW and traditional cardiovascular risk factors.¹⁸⁸

Dialysis Procedure

As discussed above, dialysis may contribute to PEW through infectious, inflammatory, and volume-related complications. Recent studies have improved our understanding of how dialysis treatment per se affects protein and energy homeostasis. Amino acid and protein losses during the dialysis session, together with low nutrient intake, promote low nutrient availability for muscle synthesis.¹⁸⁹⁻¹⁹² In a study comparing high (1.4 g/kg/day) and low (0.5 g/kg/day) protein diets, Borah et al.¹⁹³ found negative nitrogen balance on every day on the low-protein diet, but only on HD days with high protein intake. Several more recent studies show that the catabolic effects of HD, especially on the protein homeostasis, are profound, affecting whole-body and skeletal muscle protein homeostasis. All of these careful metabolic studies consistently show a decrease in protein synthesis at the whole-body level whereas one specific study showed an additional increase in whole-body protein breakdown.¹⁹⁴ Moreover, two separate studies showed a significant increase in net skeletal muscle protein breakdown; in one study, these undesirable effects persisted for at least 2 hours after the completion of HD.^{58,195} Mechanistically, the net protein breakdown has been related to (1) an absolute decline in amino acid levels due to dialysis losses-a study in normal swine showed that a reduction in plasma amino acid concentrations similar to the extent seen during dialysis signals an inhibition of muscle protein synthesis and that corresponding changes in eIF2B activity suggest a possible role in mediating the response¹⁹⁶; (2) imbalances in amino acid levels; and (3) activation of the inflammatory cascade.58,197,198 Fortunately, in HD and PD patients, concurrent amino acid supplementation can prevent

or reverse these adverse effects,¹⁹⁹⁻²⁰¹ providing an opportunity for treatment of PEW.

Inadequate dialysis is well known to cause PEW,^{4,5} and it has been shown in PD patients that loss of residual kidney function contributes to PEW.²⁰² Loss of residual kidney function is independently associated with reduced dietary energy, protein,²⁰³ and micronutrient intake.²⁰⁴ It was also associated with increased inflammation²⁰⁵ and increased resting energy metabolism.⁵³

Conclusion

This ISRNM working group concludes that advances in understanding how inflammation, insulin resistance, oxidative stress, glucocorticoids, and acidosis modify the response to reduced protein and energy intake provide a strong model framework to understand the pathophysiology of PEW. PEW naturally develops with the progression of CKD and is an inherent component of advanced disease. Although dialysis reverses uremia, residual metabolic derangements, inflammation, comorbid conditions, and the dialysis procedure itself may allow PEW to develop or worsen. As new mediators are discovered, integration of those mediators into the model and refinement of hypotheses are needed. Regarding clinical outcomes, the ability to separate the effects of nutrition, aging, and comorbidities is critical for understanding etiology, and, perhaps more importantly, for the design of future therapeutic clinical trials including anti-inflammatory and anabolic treatment strategies.

References

1. Fouque D, Kalantar-Zadeh K, Kopple J, Cano N, Chauveau P, Cuppari L, et al. A proposed nomenclature and diagnostic criteria for protein-energy wasting in acute and chronic kidney disease. *Kidney Int.* 2008;73:391-398.

2. Hoffer LJ. The need for consistent criteria for identifying malnutrition. *Nestle Nutr Workshop Ser Clin Perform Programme*. 2009;12:41-52.

3. Araujo IC, Kamimura MA, Draibe SA, Canziani ME, Manfredi SR, Avesani CM, et al. Nutritional parameters and mortality in incident hemodialysis patients. *J Ren Nutr.* 2006;16:27-35.

4. Kalantar-Zadeh K, Supasyndh O, Lehn RS, McAllister CJ, Kopple JD. Normalized protein nitrogen appearance is correlated with hospitalization and mortality in hemodialysis patients with kt/v greater than 1.20. *J Ren Nutr.* 2003;13:15-25.

5. Bossola M, Muscaritoli M, Tazza L, Panocchia N, Liberatori M, Giungi S, et al. Variables associated with reduced dietary intake in hemodialysis patients. *J Ren Nutr.* 2005;15:244-252.

6. Burrowes JD, Larive B, Chertow GM, Cockram DB, Dwyer JT, Greene T, et al. Self-reported appetite, hospitalization and death in haemodialysis patients: findings from the hemodialysis (HEMO) study. *Nephrol Dial Transpl.* 2005;20:2765-2774.

7. Bazanelli AP, Kamimura MA, da Silva CB, Avesani CM, Lopes MG, Manfredi SR, et al. Resting energy expenditure in peritoneal dialysis patients. *Perit Dial Int.* 2006;26:697-704.

8. Wang AY, Sea MM, Ng K, Kwan M, Lui SF, Woo J. Nutrient intake during peritoneal dialysis at the Prince of Wales Hospital in Hong Kong. *Am J Kidney Dis.* 2007;49:682-692. 9. Kloppenburg WD, de Jong PE, Huisman RM. The contradiction of stable body mass despite low reported dietary energy intake in chronic haemodialysis patients. *Nephrol Dial Transpl.* 2002;17:1628-1633.

10. Bazanelli AP, Kamimura MA, Vasselai P, Draibe SA, Cuppari L. Underreporting of energy intake in peritoneal dialysis patients. *J Ren Nutr.* 2010;20:263–269.

11. Huang X, Sjogren P, Cederholm T, Arnlov J, Lindholm B, Riserus U, et al. Serum and adipose tissue fatty acid composition as biomarkers of habitual dietary fat intake in elderly men with chronic kidney disease. *Nephrol Dial Transpl.* 2012. Available at: http://ndt.oxfordjournals.org/content/early/2012/12/09/ndt.gfs478.long. Accessed January 29, 2013.

12. Huang X, Stenvinkel P, Qureshi AR, Cederholm T, Barany P, Heimburger O, et al. Clinical determinants and mortality predictability of stearoyl-coa desaturase-1 activity indices in dialysis patients. *J Intern Med.* 2012 Aug 17; http://dx.doi.org/10.1111/j.1365-2796.2012.02573.x [Epub ahead of print].

13. Huang X, Stenvinkel P, Qureshi AR, Riserus U, Cederholm T, Barany P, et al. Essential polyunsaturated fatty acids, inflammation and mortality in dialysis patients. *Nephrol Dial Transpl.* 2012;27:3615-3620.

14. Jankowska M, Marszall M, Debska-Slizien A, Carrero JJ, Lindholm B, Czarnowski W, et al. Vitamin B6 and the immunity in kidney transplant recipients. *J Ren Nutr.* 2013;23:57-64.

15. Kalantar-Zadeh K, Kopple JD, Deepak S, Block D, Block G. Food intake characteristics of hemodialysis patients as obtained by food frequency questionnaire. *J Ren Nutr.* 2002;12:17-31.

16. Krishnamurthy VM, Wei G, Baird BC, Murtaugh M, Chonchol MB, Raphael KL, et al. High dietary fiber intake is associated with decreased in-flammation and all-cause mortality in patients with chronic kidney disease. *Kidney Int.* 2012;81:300-306.

17. Hollingdale R, Sutton D, Hart K. Facilitating dietary change in renal disease: investigating patients' perspectives. *J Ren Care*. 2008;34: 136-142.

18. Paes-Barreto JG, Barreto Silva MI, Qureshi AR, Bregman R, Cervante VF, Carrero JJ, et al. Can renal nutrition education improve adherence to a low-protein diet in patients with stages 3 to 5 chronic kidney disease? *J Ren Nutr.* 2012. Available at: http://www.jrnjournal.org/article/ \$1051-2276(12)00203-8/abstract. Accessed January 29, 2013.

19. Carrero JJ, Qureshi AR, Axelsson J, Avesani CM, Suliman ME, Kato S, et al. Comparison of nutritional and inflammatory markers in dialysis patients with reduced appetite. *Am J Clin Nutr.* 2007;85:695-701.

20. Kalantar-Zadeh K, Block G, McAllister CJ, Humphreys MH, Kopple JD. Appetite and inflammation, nutrition, anemia, and clinical outcome in hemodialysis patients. *Am J Clin Nutr.* 2004;80:299-307.

21. Lopes AA, Elder SJ, Ginsberg N, Andreucci VE, Cruz JM, Fukuhara S, et al. Lack of appetite in haemodialysis patients—associations with patient characteristics, indicators of nutritional status and outcomes in the international DOPPS. *Nephrol Dial Transpl.* 2007;22:3538-3546.

22. Bossola M, Tazza L, Giungi S, Luciani G. Anorexia in hemodialysis patients: an update. *Kidney Int.* 2006;70:417-422.

23. Carrero JJ. Identification of patients with eating disorders: clinical and biochemical signs of appetite loss in dialysis patients. *J Ren Nutr.* 2009;19: 10-15.

24. Kopple JD, Berg R, Houser H, Steinman TI, Teschan P. Nutritional status of patients with different levels of chronic renal insufficiency. Modification of diet in renal disease (MDRD) study group. *Kidney Int Suppl.* 1989;27:S184-S194.

25. Duenhas MR, Draibe SA, Avesani CM, Sesso R, Cuppari L. Influence of renal function on spontaneous dietary intake and on nutritional status of chronic renal insufficiency patients. *Eur J Clin Nutr.* 2003;57:1473-1478.

26. Ikizler TA, Greene JH, Wingard RL, Parker RA, Hakim RM. Spontaneous dietary protein intake during progression of chronic renal failure. *J Am Soc Nephrol.* 1995;6:1386-1391.

27. Carrero JJ. Mechanisms of altered regulation of food intake in chronic kidney disease. J Ren Nutr. 2011;21:7-11.

28. Wright M, Woodrow G, O'Brien S, Armstrong E, King N, Dye L, et al. Cholecystokinin and leptin: their influence upon the eating behaviour and nutrient intake of dialysis patients. *Nephrol Dial Transpl.* 2004;19:133–140.

29. Perez-Fontan M, Cordido F, Rodriguez-Carmona A, Penin M, Diaz-Cambre H, Lopez-Muniz A, et al. Short-term regulation of peptide YY secretion by a mixed meal or peritoneal glucose-based dialysate in patients with chronic renal failure. *Nephrol Dial Transpl.* 2008;23:3696-3703.

30. Muscaritoli M, Molfino A, Chiappini MG, Laviano A, Ammann T, Spinsanti P, et al. Anorexia in hemodialysis patients: the possible role of des-acyl ghrelin. *Am J Nephrol.* 2007;27:360–365.

31. Carrero JJ, Nakashima A, Qureshi AR, Lindholm B, Heimburger O, Barany P, et al. Protein-energy wasting modifies the association of ghrelin with inflammation, leptin, and mortality in hemodialysis patients. *Kidney Int.* 2011;79:749-756.

32. Cheung W, Yu PX, Little BM, Cone RD, Marks DL, Mak RH. Role of leptin and melanocortin signaling in uremia-associated cachexia. *J Clin Invest.* 2005;115:1659-1665.

33. Carrero JJ, Witasp A, Stenvinkel P, Qureshi AR, Heimburger O, Barany P, et al. Visfatin is increased in chronic kidney disease patients with poor appetite and correlates negatively with fasting serum amino acids and triglyceride levels. *Nephrol Dial Transpl.* 2010;25:901-906.

34. Carrero JJ, Stenvinkel P. Nutrition: Can ghrelin improve appetite in uremic wasting? *Nat Rev Nephrol.* 2009;5:672-673.

35. Bossola M, Scribano D, Colacicco L, Tavazzi B, Giungi S, Zuppi C, et al. Anorexia and plasma levels of free tryptophan, branched chain amino acids, and ghrelin in hemodialysis patients. *J Ren Nutr.* 2009;19:248-255.

36. Akar H, Akar GC, Carrero JJ, Stenvinkel P, Lindholm B. Systemic consequences of poor oral health in chronic kidney disease patients. *Clin J Am Soc Nephrol.* 2011;6:218–226.

37. Bossola M, Luciani G, Rosa F, Tazza L. Appetite and gastrointestinal symptoms in chronic hemodialysis patients. *J Ren Nutr.* 2011;21: 448-454.

38. Aguilera A, Bajo MA, Espinoza M, Olveira A, Paiva AM, Codoceo R, et al. Gastrointestinal and pancreatic function in peritoneal dialysis patients: their relationship with malnutrition and peritoneal membrane abnormalities. *Am J Kidney Dis.* 2003;42:787-796.

39. Van Vlem B, Schoonjans R, Vanholder R, De Vos M, Vandamme W, Van Laecke S, et al. Delayed gastric emptying in dyspeptic chronic hemodialysis patients. *Am J Kidney Dis.* 2000;36:962–968.

40. Aguilera A, Gonzalez-Espinoza L, Codoceo R, Jara Mdel C, Pavone M, Bajo MA, et al. Bowel bacterial overgrowth as another cause of malnutrition, inflammation, and atherosclerosis syndrome in peritoneal dialysis patients. *Adv Perit Dial.* 2010;26:130-136.

41. Hung KC, Wu CC, Chen HS, Ma WY, Tseng CF, Yang LK, et al. Serum IL-6, albumin and co-morbidities are closely correlated with symptoms of depression in patients on maintenance haemodialysis. *Nephrol Dial Transpl.* 2011;26:658-664.

42. Evans WJ, Morley JE, Argiles J, Bales C, Baracos V, Guttridge D, et al. Cachexia: a new definition. *Clin Nutr (Edinburgh, Scotland)*. 2008;27:793-799.

43. Morley JE, Thomas DR. Cachexia: new advances in the management of wasting diseases. J Am Med Dir Assoc. 2008;9:205-210.

44. Shetty PS. Adaptation to low energy intakes: the responses and limits to low intakes in infants, children and adults. *Eur J Clin Nutr.* 1999;53(suppl 1): S14–S33.

45. Emery PW. Metabolic changes in malnutrition. *Eye.* 2005;19:1029-1034.

46. Waterlow JC, Garlick PJ, Millward DJ. Protein turnover in mammalian tissues and in the whole body. Amsterdam, The Netherlands: North Holland; 1978.

47. Franch HA, Mitch WE. Navigating between the scylla and charybdis of prescribing dietary protein for chronic kidney diseases. *Annu Rev Nutr.* 2009;29:341-364.

48. Golden MH. The development of concepts of malnutrition. J Nutr. 2002;132:2117S-2122S.

49. Don BR, Kaysen G. Serum albumin: relationship to inflammation and nutrition. *Semin Dial.* 2004;17:432-437.

50. Myron Johnson A, Merlini G, Sheldon J, Ichihara K. Clinical indications for plasma protein assays: transthyretin (prealbumin) in inflammation and malnutrition. *Clin Chem Lab Med.* 2007;45:419-426.

51. Finn PF, Dice JF. Proteolytic and lipolytic responses to starvation. *Nutrition*. 2006;22:830–844.

52. Neyra R, Chen KY, Sun M, Shyr Y, Hakim RM, Ikizler TA. Increased resting energy expenditure in patients with end-stage renal disease. *JPEN J Parenter Enteral Nutr.* 2003;27:36–42.

53. Wang AY, Sea MM, Tang N, Sanderson JE, Lui SF, Li PK, et al. Resting energy expenditure and subsequent mortality risk in peritoneal dialysis patients. J Am Soc Nephrol. 2004;15:3134–3143.

54. Cuppari L, de Carvalho AB, Avesani CM, Kamimura MA, Dos Santos Lobao RR, Draibe SA. Increased resting energy expenditure in hemodialysis patients with severe hyperparathyroidism. *J Am Soc Nephrol.* 2004;15: 2933-2939.

55. Avesani CM, Cuppari L, Silva AC, Sigulem DM, Cendoroglo M, Sesso R, et al. Resting energy expenditure in pre-dialysis diabetic patients. *Nephrol Dial Transpl.* 2001;16:556-565.

56. Utaka S, Avesani CM, Draibe SA, Kamimura MA, Andreoni S, Cuppari L. Inflammation is associated with increased energy expenditure in patients with chronic kidney disease. *Am J Clin Nutr.* 2005;82: 801-805.

57. Kamimura MA, Draibe SA, Dalboni MA, Cendoroglo M, Avesani CM, Manfredi SR, et al. Serum and cellular interleukin-6 in haemodialysis patients: relationship with energy expenditure. *Nephrol Dial Transpl.* 2007;22:839-844.

58. Ikizler TA, Pupim LB, Brouillette JR, Levenhagen DK, Farmer K, Hakim RM, et al. Hemodialysis stimulates muscle and whole body protein loss and alters substrate oxidation. *Am J Physiol Endocrinol Metab.* 2002;282:E107-E116.

59. Mafra D, Deleaval P, Teta D, Cleaud C, Arkouche W, Jolivot A, et al. Influence of inflammation on total energy expenditure in hemodialysis patients. *J Ren Nutr.* 2011;21:387-393.

60. Avesani CM, Trolonge S, Deleaval P, Baria F, Mafra D, Faxen-Irving G, et al. Physical activity and energy expenditure in haemodialysis patients: an international survey. *Nephrol Dial Transpl.* 2012;27:2430-2434.

61. Spindler SR. Caloric restriction: from soup to nuts. Ageing Res Rev. 2010;9:324-353.

62. Kaysen GA, Greene T, Daugirdas JT, Kimmel PL, Schulman GW, Toto RD, et al. Longitudinal and cross-sectional effects of C-reactive protein, equilibrated normalized protein catabolic rate, and serum bicarbonate on creatinine and albumin levels in dialysis patients. *Am J Kidney Dis.* 2003;42:1200-1211.

63. Tomkins AM, Garlick PJ, Schofield WN, Waterlow JC. The combined effects of infection and malnutrition on protein metabolism in children. *Clin Sci.* 1983;65:313-324.

64. Keusch GT. The history of nutrition: malnutrition, infection and immunity. J Nutr. 2003;133:336S-340S.

65. Carrero JJ, Stenvinkel P. Inflammation in end-stage renal disease-what have we learned in 10 years? *Semin Dial*. 2010;23:498-509.

66. Stenvinkel P, Ketteler M, Johnson RJ, Lindholm B, Pecoits-Filho R, Riella M, et al. IL-10, IL-6, and TNFF-alpha: central factors in the altered cytokine network of uremia—the good, the bad, and the ugly. *Kidney Int.* 2005;67:1216-1233.

67. Carrero JJ, Chmielewski M, Axelsson J, Snaedal S, Heimburger O, Barany P, et al. Muscle atrophy, inflammation and clinical outcome in incident and prevalent dialysis patients. *Clin Nutr (Edinburgh, Scotland)*. 2008;27:557-564.

68. Miyamoto T, Rashid Qureshi A, Heimburger O, Barany P, Carrero K, Sjoberg B, et al. Inverse relationship between the inflammatory marker pentraxin-3, fat body mass, and abdominal obesity in end-stage renal disease. *Clin J Am Soc Nephrol.* 2011;6:2785-2791.

69. Delano MJ, Moldawer LL. The origins of cachexia in acute and chronic inflammatory diseases. *Nutr Clin Pract.* 2006;21:68-81.

70. Meuwese CL, Stenvinkel P, Dekker FW, Carrero JJ. Monitoring of inflammation in patients on dialysis: forewarned is forearmed. *Nat Rev Nephrol.* 2011;7:166-176.

71. Carrero JJ, Aguilera A, Stenvinkel P, Gil F, Selgas R, Lindholm B. Appetite disorders in uremia. J Ren Nutr. 2008;18:107-113.

72. Avesani CM, Carrero JJ, Axelsson J, Qureshi AR, Lindholm B, Stenvinkel P. Inflammation and wasting in chronic kidney disease: partners in crime. *Kidney Int Suppl.* 2006;70:S8–S13.

73. Hu Z, Wang H, Lee IH, Du J, Mitch WE. Endogenous glucocorticoids and impaired insulin signaling are both required to stimulate muscle wasting under pathophysiological conditions in mice. *J Clin Invest.* 2009;119:3059-3069.

74. Zhang L, Du J, Hu Z, Han G, Delafontaine P, Garcia G, et al. IL-6 and serum amyloid a synergy mediates angiotensin II-induced muscle wasting. *J Am Soc Nephrol.* 2009;20:604–612.

75. Raj DS, Moseley P, Dominic EA, Onime A, Tzamaloukas AH, Boyd A, et al. Interleukin-6 modulates hepatic and muscle protein synthesis during hemodialysis. *Kidney Int.* 2008;73:1054-1061.

76. Boivin MA, Battah SI, Dominic EA, Kalantar-Zadeh K, Ferrando A, Tzamaloukas AH, et al. Activation of caspase-3 in the skeletal muscle during haemodialysis. *Eur J Clin Invest.* 2010;40:903-910.

77. Wang XH, Hu Z, Klein JD, Zhang L, Fang F, Mitch WE. Decreased mir-29 suppresses myogenesis in CKD. J Am Soc Nephrol. 2011;22:2068-2076.

78. Zhang L, Rajan V, Lin E, Hu Z, Han HQ, Zhou X, et al. Pharmacological inhibition of myostatin suppresses systemic inflammation and muscle atrophy in mice with chronic kidney disease. *FASEB J.* 2011;25:1653-1663.

79. Hansen J, Brandt C, Nielsen AR, Hojman P, Whitham M, Febbraio MA, et al. Exercise induces a marked increase in plasma follistatin: evidence that follistatin is a contraction-induced hepatokine. *Endocrinology*. 2011;152:164-171.

80. Leehey DJ, Moinuddin I, Bast JP, Qureshi S, Jelinek CS, Cooper C, et al. Aerobic exercise in obese diabetic patients with chronic kidney disease: a randomized and controlled pilot study. *Cardiovasc Diabetol.* 2009;8:62.

81. Miyamoto T, Carrero JJ, Qureshi AR, Anderstam B, Heimburger O, Barany P, et al. Circulating follistatin in patients with chronic kidney disease: implications for muscle strength, bone mineral density, inflammation, and survival. *Clin J Am Soc Nephrol.* 2011;6:1001-1008.

82. Lazarus DD, Moldawer LL, Lowry SF. Insulin-like growth factor-1 activity is inhibited by interleukin-1 alpha, tumor necrosis factor-alpha, and interleukin-6. *Lymphokine Cytokine Res.* 1993;12:219-223.

83. Axelsson J, Qureshi AR, Divino-Filho JC, Barany P, Heimburger O, Lindholm B, et al. Are insulin-like growth factor and its binding proteins 1 and 3 clinically useful as markers of malnutrition, sarcopenia and inflammation in end-stage renal disease? *Eur J Clin Nutr.* 2006;60:718-726.

84. Mak RH, Rotwein P. Myostatin and insulin-like growth factors in uremic sarcopenia: the yin and yang in muscle mass regulation. *Kidney Int.* 2006;70:410-412.

85. Carrero JJ, Ortiz A, Qureshi AR, Martin-Ventura JL, Barany P, Heimburger O, et al. Additive effects of soluble tweak and inflammation on mortality in hemodialysis patients. *Clin J Am Soc Nephrol.* 2009;4:110-118.

86. Dogra C, Changotra H, Wedhas N, Qin X, Wergedal JE, Kumar A. TNF-related weak inducer of apoptosis (TWEAK) is a potent skeletal muscle-wasting cytokine. *FASEB J.* 2007;21:1857-1869.

87. Sun SC. The noncanonical NF-kappab pathway. Immunol Rev. 2012;246:125-140.

88. Argiles JM, Lopez-Soriano FJ, Busquets S. Therapeutic potential of interleukin-15: a myokine involved in muscle wasting and adiposity. *Drug Discov Today.* 2009;14:208-213.

89. Honda H, Qureshi AR, Axelsson J, Heimburger O, Suliman ME, Barany P, et al. Obese sarcopenia in patients with end-stage renal disease is associated with inflammation and increased mortality. *Am J Clin Nutr.* 2007;86:633–638.

90. Carrero JJ, Cordeiro AC, Lindholm B, Stenvinkel P. The emerging pleiotrophic role of adipokines in the uremic phenotype. *Curr Opin Nephrol Hypertens*. 2010;19:37-42.

91. Witasp A, Carrero JJ, Heimburger O, Lindholm B, Hammarqvist F, Stenvinkel P, et al. Increased expression of pro-inflammatory genes in abdominal subcutaneous fat in advanced chronic kidney disease patients. *J Intern Med.* 2011;269:410-419.

92. Carvalho LK, Barreto Silva MI, da Silva Vale B, Bregman R, Martucci RB, Carrero JJ, et al. Annual variation in body fat is associated with systemic inflammation in chronic kidney disease patients stages 3 and 4: a longitudinal study. *Nephrol Dial Transpl.* 2012;27:1423-1428.

93. Axelsson J, Rashid Qureshi A, Suliman ME, Honda H, Pecoits-Filho R, Heimburger O, et al. Truncal fat mass as a contributor to inflammation in end-stage renal disease. *Am J Clin Nutr.* 2004;80:1222-1229.

94. Gohda T, Gotoh H, Tanimoto M, Sato M, Io H, Kaneko K, et al. Relationship between abdominal fat accumulation and insulin resistance in hemodialysis patients. *Hypertens Res.* 2008;31:83-88.

95. Odamaki M, Furuya R, Ohkawa S, Yoneyama T, Nishikino M, Hishida A, et al. Altered abdominal fat distribution and its association with the serum lipid profile in non-diabetic haemodialysis patients. *Nephrol Dial Transpl.* 1999;14:2427-2432.

96. Odamaki M, Furuya R, Kinumura Y, Ikegaya N, Kumagai H. Association between plasma adiponectin concentration and visceral fat accumulation in hemodialysis patients. *Nephron Clin Pract.* 2006;102:c8-c13.

97. Ramos LF, Shintani A, Ikizler TA, Himmelfarb J. Oxidative stress and inflammation are associated with adiposity in moderate to severe CKD. *J Am Soc Nephrol.* 2008;19:593–599.

98. Witasp A, Carrero JJ, Hammarqvist F, Qureshi AR, Heimburger O, Schalling M, et al. Expression of osteoprotegerin in human fat tissue; implications for chronic kidney disease. *Eur J Clin Invest.* 2011;41:498-506.

99. Kamimura MA, Carrero JJ, Canziani ME, Watanabe R, Lemos MM, Cuppari L. Visceral obesity assessed by computed tomography predicts cardiovascular events in chronic kidney disease patients. *Nutr Metab Cardiovasc Dis.* 2012. Available at: http://www.nmcd-journal.com/article/S0939-4753(12)00155-X/abstract. Accessed January 29, 2013.

100. Postorino M, Marino C, Tripepi G, Zoccali C. Abdominal obesity and all-cause and cardiovascular mortality in end-stage renal disease. J Am Coll Cardiol. 2009;53:1265-1272.

101. Cordeiro AC, Qureshi AR, Stenvinkel P, Heimbürger O, Axelsson J, Bárány P, et al. Abdominal fat deposition is associated with increased inflammation, protein-energy wasting and worse outcome in patients undergoing haemodialysis. *Nephrol Dial Transplant*. 2010;25:562-568.

102. Postorino M, Marino C, Tripepi G, Zoccali C. Abdominal obesity modifies the risk of hypertriglyceridemia for all-cause and cardiovascular mortality in hemodialysis patients. *Kidney Int.* 2011;79:765-772.

103. Zoccali C, Postorino M, Marino C, Pizzini P, Cutrupi S, Tripepi G. Waist circumference modifies the relationship between the adipose tissue cytokines leptin and adiponectin and all-cause and cardiovascular mortality in haemodialysis patients. *J Intern Med.* 2011;269:172-181.

104. Yamamoto T, Carrero JJ, Lindholm B, Stenvinkel P, Axelsson J. Leptin and uremic protein-energy wasting-the axis of eating. *Semin Dial*. 2009;22:387-390.

105. Bossola M, Muscaritoli M, Valenza V, Panocchia N, Tazza L, Cascino A, et al. Anorexia and serum leptin levels in hemodialysis patients. *Nephron Clin Pract.* 2004;97:c76-c82.

106. Park SH, Carrero JJ, Lindholm B, Stenvinkel P. Adiponectin in chronic kidney disease has an opposite impact on protein-energy wasting and cardiovascular risk: two sides of the same coin. *Clin Nephrol.* 2009;72:87-96.

107. Carrero JJ, Park SH, Axelsson J, Lindholm B, Stenvinkel P. Cytokines, atherogenesis, and hypercatabolism in chronic kidney disease: a dreadful triad. *Semin Dial*. 2009;22:381-386.

108. Kistorp C, Faber J, Galatius S, Gustafsson F, Frystyk J, Flyvbjerg A, et al. Plasma adiponectin, body mass index, and mortality in patients with chronic heart failure. *Circulation*. 2005;112:1756-1762.

109. Drechsler C, Krane V, Winkler K, Dekker FW, Wanner C. Changes in adiponectin and the risk of sudden death, stroke, myocardial infarction, and mortality in hemodialysis patients. *Kidney Int.* 2009;76: 567-575.

110. Qi Y, Takahashi N, Hileman SM, Patel HR, Berg AH, Pajvani UB, et al. Adiponectin acts in the brain to decrease body weight. *Nat Med.* 2004;10:524-529.

111. Liu SW, Qiao SB, Yuan JS, Liu DQ. Association of plasma visfatin levels with inflammation, atherosclerosis and acute coronary syndromes (ACS) in humans. *Clin Endocrinol.* 2009;71:202–207.

112. Yilmaz MI, Saglam M, Qureshi AR, Carrero JJ, Caglar K, Eyileten T, et al. Endothelial dysfunction in type-2 diabetics with early diabetic nephropathy is associated with low circulating adiponectin. *Nephrol Dial Transpl.* 2008;23:1621-1627.

113. Hallschmid M, Randeva H, Tan BK, Kern W, Lehnert H. Relationship between cerebrospinal fluid visfatin (PBEF/NAMPT) levels and adiposity in humans. *Diabetes*. 2009;58:637-640.

114. Price SR, Gooch JL, Donaldson SK, Roberts-Wilson TK. Muscle atrophy in chronic kidney disease results from abnormalities in insulin signaling. *J Ren Nutr.* 2010;20:S24-S28.

115. Flakoll PJ, Carlson M, Cherrington AC. Physiological action of insulin. In: Leroith DTS, Olefsky J, eds. *Diabetes Mellitus: a Fundamental and Clinical Text.* Philadelphia, PA: Williams & Wilkins; 2000: 148-161.

116. Runyan JW Jr, Hurwitz D, Robbins SL. Effect of Kimmelstiel-Wilson syndrome on insulin requirements in diabetes. *N Engl J Med.* 1955;252:388-391.

117. DeFronzo RA, Alvestrand A, Smith D, Hendler R, Hendler E, Wahren J. Insulin resistance in uremia. J Clin Invest. 1981;67:563-568.

118. DeFronzo RA, Smith D, Alvestrand A. Insulin action in uremia. *Kidney Int Suppl.* 1983;16:S102-S114.

119. Pupim LB, Flakoll PJ, Majchrzak KM, Aftab Guy DL, Stenvinkel P, Ikizler TA. Increased muscle protein breakdown in chronic hemodialysis patients with type 2 diabetes mellitus. *Kidney Int.* 2005;68:1857-1865.

120. Siew ED, Pupim LB, Majchrzak KM, Shintani A, Flakoll PJ, Ikizler TA. Insulin resistance is associated with skeletal muscle protein breakdown in non-diabetic chronic hemodialysis patients. *Kidney Int.* 2007;71: 146-152.

121. Koppe L, Pillon NJ, Vella RE, Croze ML, Pelletier CC, Chambert S, et al. P-cresyl sulfate promotes insulin resistance associated with CKD. J Am Soc Nephrol. 2013;24:88-99.

122. Wang X, Hu Z, Hu J, Du J, Mitch WE. Insulin resistance accelerates muscle protein degradation: Activation of the ubiquitin-proteasome pathway by defects in muscle cell signaling. *Endocrinology*. 2006;147:4160–4168.

123. Brunelli SM, Thadhani R, Ikizler TA, Feldman HI. Thiazolidinedione use is associated with better survival in hemodialysis patients with non-insulin dependent diabetes. *Kidney Int.* 2009;75:961-968.

124. Mitch WE, Du J, Bailey JL, Price SR. Mechanisms causing muscle proteolysis in uremia: the influence of insulin and cytokines. *Miner Electrolyte Metab.* 1999;25:216–219.

125. Ding H, Gao XL, Hirschberg R, Vadgama JV, Kopple JD. Impaired actions of insulin-like growth factor 1 on protein synthesis and degradation in skeletal muscle of rats with chronic renal failure. Evidence for a postreceptor defect. *J Clin Invest*. 1996;97:1064–1075.

126. Zheng B, Ohkawa S, Li H, Roberts-Wilson TK, Price SR. Foxo3a mediates signaling crosstalk that coordinates ubiquitin and atrogin-1/mafbx expression during glucocorticoid-induced skeletal muscle atrophy. *FASEB J.* 2010;24:2660-2669.

127. Du J, Wang X, Miereles C, Bailey JL, Debigare R, Zheng B, et al. Activation of caspase-3 is an initial step triggering accelerated muscle proteolysis in catabolic conditions. *J Clin Invest*. 2004;113:115-123.

128. Carrero JJ, Kyriazis J, Sonmez A, Tzanakis I, Qureshi AR, Stenvinkel P, et al. Prolactin levels, endothelial dysfunction, and the risk of cardiovascular events and mortality in patients with CKD. *Clin J Am Soc Nephrol.* 2012;7:207-215.

129. Carrero JJ, Qureshi AR, Nakashima A, Arver S, Parini P, Lindholm B, et al. Prevalence and clinical implications of testosterone deficiency in men with end-stage renal disease. *Nephrol Dial Transpl.* 2011;26:184–190.

130. Gungor O, Kircelli F, Carrero JJ, Asci G, Toz H, Tatar E, et al. Endogenous testosterone and mortality in male hemodialysis patients: is it the result of aging? *Clin J Am Soc Nephrol.* 2010;5:2018-2023.

131. Yilmaz MI, Sonmez A, Qureshi AR, Saglam M, Stenvinkel P, Yaman H, et al. Endogenous testosterone, endothelial dysfunction, and cardiovascular events in men with nondialysis chronic kidney disease. *Clin J Am Soc Nephrol.* 2011;6:1617-1625.

132. Carrero JJ, Barany P, Yilmaz MI, Qureshi AR, Sonmez A, Heimburger O, et al. Testosterone deficiency is a cause of anaemia and reduced responsiveness to erythropoiesis-stimulating agents in men with chronic kidney disease. *Nephrol Dial Transpl.* 2012;27:709-715.

133. Carrero JJ, Stenvinkel P. The vulnerable man: impact of testosterone deficiency on the uraemic phenotype. *Nephrol Dial Transpl.* 2012;27:4030-4041.

134. Carrero JJ, Qureshi AR, Parini P, Arver S, Lindholm B, Barany P, et al. Low serum testosterone increases mortality risk among male dialysis patients. J Am Soc Nephrol. 2009;20:613-620.

135. Kyriazis J, Tzanakis I, Stylianou K, Katsipi I, Moisiadis D, Papadaki A, et al. Low serum testosterone, arterial stiffness and mortality in male haemodialysis patients. *Nephrol Dial Transpl.* 2011;26:2971-2977.

136. Haring R, Nauck M, Volzke H, Endlich K, Lendeckel U, Friedrich N, et al. Low serum testosterone is associated with increased mortality in men with stage 3 or greater nephropathy. *Am J Nephrol.* 2011;33: 209-217.

137. Cigarran S, Pousa M, Castro MJ, Gonzalez B, Martinez A, Barril G, et al. Endogenous testosterone, muscle strength, and fat-free mass in men with chronic kidney disease. *J Ren Nutr.* 2012; pii:S1051-2276.

138. Johansen KL, Mulligan K, Schambelan M. Anabolic effects of nandrolone decanoate in patients receiving dialysis: a randomized controlled trial. *JAMA*. 1999;281:1275–1281.

139. Johansen KL, Painter PL, Sakkas GK, Gordon P, Doyle J, Shubert T. Effects of resistance exercise training and nandrolone decanoate on body composition and muscle function among patients who receive hemodialysis: a randomized, controlled trial. J Am Soc Nephrol. 2006;17:2307-2314.

140. Vanhorebeek I, Langouche L, Van den Berghe G. Endocrine aspects of acute and prolonged critical illness. *Nat Clin Pract.* 2006;2:20-31.

141. Carrero JJ, Qureshi AR, Axelsson J, Yilmaz MI, Rehnmark S, Witt MR, et al. Clinical and biochemical implications of low thyroid hormone levels (total and free forms) in euthyroid patients with chronic kidney disease. J Intern Med. 2007;262:690-701.

142. Zoccali C, Mallamaci F, Tripepi G, Cutrupi S, Pizzini P. Low triiodothyronine and survival in end-stage renal disease. *Kidney Int.* 2006;70: 523-528.

143. Yilmaz MI, Sonmez A, Karaman M, Ay SA, Saglam M, Yaman H, et al. Low triiodothyronine alters flow-mediated vasodilatation in advanced nondiabetic kidney disease. *Am J Nephrol.* 2011;33:25-32.

144. Meuwese CL, Dekker FW, Lindholm B, Qureshi AR, Heimburger O, Barany P, et al. Baseline levels and trimestral variation of triiodothyronine and thyroxine and their association with mortality in maintenance hemodialysis patients. *Clin J Am Soc Nephrol.* 2012;7:131-138.

145. Wiederkehr MR, Kalogiros J, Krapf R. Correction of metabolic acidosis improves thyroid and growth hormone axes in haemodialysis patients. *Nephrol Dial Transpl.* 2004;19:1190-1197.

146. Disthabanchong S, Treeruttanawanich A. Oral sodium bicarbonate improves thyroid function in predialysis chronic kidney disease. *Am J Nephrol.* 2010;32:549–556.

147. Ozen KP, Asci G, Gungor O, Carrero JJ, Kircelli F, Tatar E, et al. Nutritional state alters the association between free triiodothyronine levels and mortality in hemodialysis patients. *Am J Nephrol.* 2011;33:305-312.

148. Bailey JL, England BK, Long RC Jr, Weissman J, Mitch WE. Experimental acidemia and muscle cell ph in chronic acidosis and renal failure. *Am J Physiol.* 1995;269:C706-C712. 149. Franch HA, Raissi S, Wang X, Zheng B, Bailey JL, Price SR. Acidosis impairs insulin receptor substrate-1-associated phosphoinositide 3-kinase signaling in muscle cells: consequences on proteolysis. *Am J Physiol Ren Physiol.* 2004;287:F700-F706.

150. Lee SW, Dai G, Hu Z, Wang X, Du J, Mitch WE. Regulation of muscle protein degradation: coordinated control of apoptotic and ubiquitinproteasome systems by phosphatidylinositol 3 kinase. *J Am Soc Nephrol.* 2004;15:1537-1545.

151. May RC, Kelly RA, Mitch WE. Metabolic acidosis stimulates protein degradation in rat muscle by a glucocorticoid-dependent mechanism. *J Clin Invest.* 1986;77:614–621.

152. Song YH, Li Y, Du J, Mitch WE, Rosenthal N, Delafontaine P. Muscle-specific expression of igf-1 blocks angiotensin II-induced skeletal muscle wasting. *J Clin Invest*. 2005;115:451-458.

153. Lecker SH, Jagoe RT, Gilbert A, Gomes M, Baracos V, Bailey J, et al. Multiple types of skeletal muscle atrophy involve a common program of changes in gene expression. *FASEB J.* 2004;18:39–51.

154. Kalhoff H, Diekmann L, Kunz C, Stock GJ, Manz F. Alkali therapy versus sodium chloride supplement in low birthweight infants with incipient late metabolic acidosis. *Acta Paediatr.* 1997;86:96-101.

155. McSherry E, Morris RC Jr. Attainment and maintenance of normal stature with alkali therapy in infants and children with classic renal tubular acidosis. *J Clin Invest.* 1978;61:509-527.

156. Frassetto L, Morris RC Jr, Sebastian A. Potassium bicarbonate reduces urinary nitrogen excretion in postmenopausal women. *J Clin Endocrinol Metab.* 1997;82:254–259.

157. Ballmer PE, McNurlan MA, Hulter HN, Anderson SE, Garlick PJ, Krapf R. Chronic metabolic acidosis decreases albumin synthesis and induces negative nitrogen balance in humans. *J Clin Invest*. 1995;95:39-45.

158. Reaich D, Channon SM, Scrimgeour CM, Goodship TH. Ammonium chloride-induced acidosis increases protein breakdown and amino acid oxidation in humans. *Am J Physiol*. 1992;263:E735-E739.

159. Reaich D, Channon SM, Scrimgeour CM, Daley SE, Wilkinson R, Goodship TH. Correction of acidosis in humans with CRF decreases protein degradation and amino acid oxidation. *Am J Physiol.* 1993;265:E230-E235.

160. Graham KA, Reaich D, Channon SM, Downie S, Goodship TH. Correction of acidosis in hemodialysis decreases whole-body protein degradation. *J Am Soc Nephrol.* 1997;8:632-637.

161. Graham KA, Reaich D, Channon SM, Downie S, Gilmour E, Passlick-Deetjen J, et al. Correction of acidosis in CAPD decreases whole body protein degradation. *Kidney Int.* 1996;49:1396-1400.

162. Stein A, Moorhouse J, Iles-Smith H, Baker F, Johnstone J, James G, et al. Role of an improvement in acid-base status and nutrition in CAPD patients. *Kidney Int.* 1997;52:1089-1095.

163. de Brito-Ashurst I, Varagunam M, Raftery MJ, Yaqoob MM. Bicarbonate supplementation slows progression of CKD and improves nutritional status. J Am Soc Nephrol. 2009;20:2075-2084.

164. Pupim LB, Heimburger O, Qureshi AR, Ikizler TA, Stenvinkel P. Accelerated lean body mass loss in incident chronic dialysis patients with diabetes mellitus. *Kidney Int.* 2005;68:2368–2374.

165. Kalantar-Zadeh K, Derose SF, Nicholas S, Benner D, Sharma K, Kovesdy CP. Burnt-out diabetes: impact of chronic kidney disease progression on the natural course of diabetes mellitus. *J Ren Nutr.* 2009;19:33-37.

166. von Haehling S, Lainscak M, Springer J, Anker SD. Cardiac cachexia: a systematic overview. *Pharmacol Ther.* 2009;121:227-252.

167. Wang AY, Sea MM, Tang N, Lam CW, Chan IH, Lui SF, et al. Energy intake and expenditure profile in chronic peritoneal dialysis patients complicated with circulatory congestion. *Am J Clin Nutr.* 2009;90:1179-1184.

168. Cuppari L, Garcia-Lopes MG. Hypovitaminosis D in chronic kidney disease patients: prevalence and treatment. J Ren Nutr. 2009;19:38-43.

169. Garcia LA, King KK, Ferrini MG, Norris KC, Artaza JN. 1,25(OH) 2vitamin D3 stimulates myogenic differentiation by inhibiting cell proliferation and modulating the expression of promyogenic growth factors and myostatin in C2C12 skeletal muscle cells. *Endocrinology*. 2011;152:2976-2986. 170. Harter HR, Birge SJ, Martin KJ, Klahr S, Karl IE. Effects of vitamin D metabolites on protein catabolism of muscle from uremic rats. *Kidney Int.* 1983;23:465-472.

171. Gordon PL, Sakkas GK, Doyle JW, Shubert T, Johansen KL. Relationship between vitamin d and muscle size and strength in patients on hemodialysis. *J Ren Nutr.* 2007;17:397-407.

172. Lee DM, Tajar A, Pye SR, Boonen S, Vanderschueren D, Bouillon R, et al. Association of hypogonadism with vitamin D status: the European male ageing study. *Eur J Endocrinol.* 2012;166:77-85.

173. Kimmel PL, Peterson RA, Weihs KL, Simmens SJ, Alleyne S, Cruz I, et al. Multiple measurements of depression predict mortality in a longitudinal study of chronic hemodialysis outpatients. *Kidney Int.* 2000;57:2093-2098.

174. Riezebos RK, Nauta KJ, Honig A, Dekker FW, Siegert CE. The association of depressive symptoms with survival in a Dutch cohort of patients with end-stage renal disease. *Nephrol Dial Transpl.* 2010;25:231-236.

175. Chilcot J, Wellsted D, Vilar E, Farrington K. An association between residual renal function and depression symptoms in haemodialysis patients. *Nephron Clin Pract.* 2009;113:c117-c124.

176. Schiepers OJ, Wichers MC, Maes M. Cytokines and major depression. Prog Neuropsychopharmacol Biol Psychiatry. 2005;29:201-217.

177. Sonikian M, Metaxaki P, Papavasileiou D, Boufidou F, Nikolaou C, Vlassopoulos D, et al. Effects of interleukin-6 on depression risk in dialysis patients. *Am J Nephrol.* 2010;31:303-308.

178. Montinaro V, Iaffaldano GP, Granata S, Porcelli P, Todarello O, Schena FP, et al. Emotional symptoms, quality of life and cytokine profile in hemodialysis patients. *Clin Nephrol.* 2010;73:36-43.

179. Lee SK, Lee HS, Lee TB, Kim DH, Koo JR, Kim YK, et al. The effects of antidepressant treatment on serum cytokines and nutritional status in hemodialysis patients. *J Korean Med Sci.* 2004;19:384–389.

180. Jhamb M, Argyropoulos C, Steel JL, Plantinga L, Wu AW, Fink NE, et al. Correlates and outcomes of fatigue among incident dialysis patients. *Clin J Am Soc Nephrol.* 2009;4:1779-1786.

181. Johansen KL, Painter P. Exercise in individuals with CKD. Am J Kidney Dis. 2012;59:126–134.

182. Ferrucci L, Penninx BW, Volpato S, Harris TB, Bandeen-Roche K, Balfour J, et al. Change in muscle strength explains accelerated decline of physical function in older women with high interleukin-6 serum levels. *J Am Geriatr Soc.* 2002;50:1947-1954.

183. Johansen KL, Chertow GM, Jin C, Kutner NG. Significance of frailty among dialysis patients. J Am Soc Nephrol. 2007;18:2960-2967.

184. LeBrasseur NK, Walsh K, Arany Z. Metabolic benefits of resistance training and fast glycolytic skeletal muscle. *Am J Physiol.* 2011;300: E3-E10.

185. Workeneh BT, Rondon-Berrios H, Zhang L, Hu Z, Ayehu G, Ferrando A, et al. Development of a diagnostic method for detecting increased muscle protein degradation in patients with catabolic conditions. *J Am Soc Nephrol.* 2006;17:3233-3239.

186. Rao M, Li L, Demello C, Guo D, Jaber BL, Pereira BJ, et al. Mitochondrial DNA injury and mortality in hemodialysis patients. *J Am Soc Nephrol.* 2009;20:189-196.

187. Dolinsky VW, Jones KE, Sidhu RS, Haykowsky M, Czubryt MP, Gordon T, et al. Improvements in skeletal muscle strength and cardiac function induced by resveratrol contribute to enhanced exercise performance in rats. *J Physiol*. 2012;590:2783–2799.

188. Heiwe S, Jacobson SH. Exercise training for adults with chronic kidney disease. *Cochrane Database Syst Rev.* 2011:CD003236.

189. Lofberg E, Essen P, McNurlan M, Wernerman J, Garlick P, Anderstam B, et al. Effect of hemodialysis on protein synthesis. *Clin Nephrol.* 2000;54:284–294.

190. Mokrzycki MH, Kaplan AA. Protein losses in continuous renal replacement therapies. J Am Soc Nephrol. 1996;7:2259-2263.

191. Davies SP, Reaveley DA, Brown EA, Kox WJ. Amino acid clearances and daily losses in patients with acute renal failure treated by continuous arteriovenous hemodialysis. *Crit Care Med.* 1991;19:1510-1515. 192. Schepky AG, Bensch KW, Schulz-Knappe P, Forssmann WG. Human hemofiltrate as a source of circulating bioactive peptides: determination of amino acids, peptides and proteins. *Biomed Chromatogr.* 1994;8:90-94.

193. Borah M, Schoenfeld P, Gotch F, Sargent J, Wolfsen M, Humphreys M. Nitrogen balance during intermittent dialysis therapy of uremia. *Kidney Int.* 1978;14:491-500.

194. Lim VS, Ikizler TA, Raj DS, Flanigan MJ. Does hemodialysis increase protein breakdown? Dissociation between whole-body amino acid turnover and regional muscle kinetics. *J Am Soc Nephrol.* 2005;16:862–868.

195. Raj DS, Zager P, Shah VO, Dominic EA, Adeniyi O, Blandon P, et al. Protein turnover and amino acid transport kinetics in end-stage renal disease. *Am J Physiol.* 2004;286:E136-E143.

196. Kobayashi H, Borsheim E, Anthony TG, Traber DL, Badalamenti J, Kimball SR, et al. Reduced amino acid availability inhibits muscle protein synthesis and decreases activity of initiation factor eIF2B. *Am J Physiol.* 2003;284:E488-E498.

197. Caglar K, Peng Y, Pupim LB, Flakoll PJ, Levenhagen D, Hakim RM, et al. Inflammatory signals associated with hemodialysis. *Kidney Int.* 2002;62:1408–1416.

198. Ikizler TA, Flakoll PJ, Parker RA, Hakim RM. Amino acid and albumin losses during hemodialysis. *Kidney Int.* 1994;46:830-837.

199. Pupim LB, Majchrzak KM, Flakoll PJ, Ikizler TA. Intradialytic oral nutrition improves protein homeostasis in chronic hemodialysis patients with deranged nutritional status. *J Am Soc Nephrol.* 2006;17:3149-3157.

200. Veeneman JM, Kingma HA, Boer TS, Stellaard F, De Jong PE, Reijngoud DJ, et al. Protein intake during hemodialysis maintains a positive whole body protein balance in chronic hemodialysis patients. *Am J Physiol.* 2003;284:E954-E965.

201. Tjiong HL, van den Berg JW, Wattimena JL, Rietveld T, van Dijk LJ, van der Wiel AM, et al. Dialysate as food: combined amino acid and glucose dialysate improves protein anabolism in renal failure patients on automated peritoneal dialysis. J Am Soc Nephrol. 2005;16:1486–1493.

202. Wang AY, Woo J, Wang M, Sea MM, Sanderson JE, Lui SF, et al. Important differentiation of factors that predict outcome in peritoneal dialysis patients with different degrees of residual renal function. *Nephrol Dial Transpl.* 2005;20:396–403.

203. Wang AY, Sea MM, Ip R, Law MC, Chow KM, Lui SF, et al. Independent effects of residual renal function and dialysis adequacy on actual dietary protein, calorie, and other nutrient intake in patients on continuous ambulatory peritoneal dialysis. *J Am Soc Nephrol.* 2001;12:2450-2457.

204. Wang AY, Sea MM, Ip R, Law MC, Chow KM, Lui SF, et al. Independent effects of residual renal function and dialysis adequacy on dietary micronutrient intakes in patients receiving continuous ambulatory peritoneal dialysis. *Am J Clin Nutr.* 2002;76:569-576.

205. Wang AY, Sanderson J, Sea MM, Wang M, Lam CW, Li PK, et al. Important factors other than dialysis adequacy associated with inadequate dietary protein and energy intakes in patients receiving maintenance peritoneal dialysis. *Am J Clin Nutr.* 2003;77:834–841.