



Universidade de São Paulo

Biblioteca Digital da Produção Intelectual - BDPI

Departamento de Microbiologia - ICB/BMM

Artigos e Materiais de Revistas Científicas - ICB/BMM

2014-03-18

relA enhances the adherence of enteropathogenic *Escherichia coli*

PloS One, San Francisco, v.9, n.3, p.e91703, 2014

<http://www.producao.usp.br/handle/BDPI/44789>

Downloaded from: Biblioteca Digital da Produção Intelectual - BDPI, Universidade de São Paulo

relA Enhances the Adherence of Enteropathogenic *Escherichia coli*

Beny Spira*, Gerson Moura Ferreira, Luiz Gustavo de Almeida

Departamento de Microbiologia, Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo-SP, Brazil

Abstract

Enteropathogenic *Escherichia coli* (EPEC) is a known causative agent of diarrhea in children. In the process of colonization of the small intestine, EPEC synthesizes two types of adhesins, the bundle-forming pilus (BFP) and intimin. The BFP pilus is an adhesin associated with the initial stages of adherence of EPEC to epithelial cells, while the outer membrane protein intimin carries out the intimate adherence that takes place at the third stage of infection. BFP is encoded by the *bfp* operon located in plasmid EAF, present only in typical EPEC isolates, while *eae*, the gene that encodes intimin is situated in the LEE, a chromosomal pathogenicity island. Transcription of *bfp* and *eae* is regulated by the products of the *perABC* operon, also present in plasmid EAF. Here we show that deletion of *relA*, that encodes a guanosine penta and tetraphosphate synthetase impairs EPEC adherence to epithelial cells *in vitro*. In the absence of *relA*, the transcription of the regulatory operon *perABC* is reduced, resulting in lower levels of BFP and intimin. Bacterial adherence, BFP and intimin synthesis and *perABC* expression are restored upon complementation with the wild-type *relA* allele.

Citation: Spira B, Ferreira GM, de Almeida LG (2014) *relA* Enhances the Adherence of Enteropathogenic *Escherichia coli*. PLoS ONE 9(3): e91703. doi:10.1371/journal.pone.0091703

Editor: Axel Cloeckert, Institut National de la Recherche Agronomique, France

Received: November 23, 2013; **Accepted:** February 14, 2014; **Published:** March 18, 2014

Copyright: © 2014 Spira et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by a grant from Fundação de Amparo a Pesquisa do Estado de São Paulo (FAPESP). GMF was supported by a FAPESP scholarship. LGA was supported by CNPq and CAPES scholarships. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: benys@usp.br

Introduction

Enteropathogenic *Escherichia coli* (EPEC) is one of the causes of infant diarrhea in developing countries [1,2]. Typical EPEC cells form microcolonies on epithelial cell monolayers, a pattern known as localized adherence (LA) [3]. These strains carry a large plasmid known as EAF, which harbors two operons, *bfp* and *perABC* (or *per*), needed to confer on EPEC the LA phenotype. The *bfp* operon is formed by 14 genes that are associated with the biogenesis of the bundle-forming pilus (BFP), a type IV fimbria found in typical EPEC strains [4]. *bfpA*, the first gene of the operon encodes the main subunit of the fimbria. BFP is required in the first stage of EPEC infection, i.e., for the formation of the bacterial clusters that will ultimately result in the LA pattern [5–7]. The second stage of EPEC infection is characterized by the secretion of bacterial effector via a type III secretion system into the host cell followed by the corruption of the host signal transduction. EPEC chromosome harbors a 36 kb pathogenicity island known as the locus of enterocyte effacement, LEE [8]. Many genes associated with EPEC virulence are present in the LEE, including the ones encoding the type III secretion system, the secreted proteins and the adhesin intimin. Intimin is implicated in the third stage of infection, which involves enterocyte effacement, pedestal formation at the enterocyte membrane and intimate bacterial attachment to the host cell [9,10]. In typical EPEC, transcription of the *bfp* operon and of the LEE genes are activated, respectively, by PerA and PerC, encoded by the *perABC* operon [11,12].

The nucleotides guanosine tetra and penta-phosphate, collectively referred as ppGpp, accumulate in response to adverse environmental conditions. It was first noticed in *E. coli* starved for

amino acids [13] and later observed under other stress conditions, such as deprivation of carbon, nitrogen [14] and phosphate [15]. ppGpp accumulation is accompanied by the stringent response, a metabolic adjustment characterised by a dramatic decrease in stable RNA and ribosome synthesis, general protein inhibition and the synthesis of specific proteins, such as the alternative sigma factor RpoS [16,17]. In *E. coli* ppGpp is synthesized by two related proteins - RelA and SpoT. The ribosome-bound RelA is activated by uncharged tRNAs under conditions of amino acid limitation or by the addition of inhibitors of aminoacyl tRNA synthetases [18]. The bifunctional enzyme SpoT displays a strong hydrolase and a weak synthetase activity, owe to the presence of two functional domains [19]. SpoT senses several stress conditions, in a still unclear fashion, and usually responds by inhibiting its hydrolase activity [20]. Interestingly, though all wild-type strains respond in a similar fashion to nutrient limitation, the intrinsic levels of ppGpp are not constant throughout the *E. coli* species [21,22].

Mutations in *relA* confer on the bacterium a relaxed phenotype, which consists of a continuous accumulation of stable RNA under amino acid starvation. This mutant, however, responds normally to other stress conditions. Artificial induction of ppGpp to high levels results in growth arrest and inhibition of protein synthesis [23]. ppGpp is associated with diverse cellular functions, such as rRNA synthesis, mRNA elongation, amino acids, carbohydrate and lipid metabolism, DNA replication and virulence [15,24]. The mechanism through which ppGpp exerts so many pleiotropic effects is not entirely clear. It can be partially explained by the fact that ppGpp interacts directly with RNA polymerase and shifts the affinity of the core enzyme towards alternative sigma factors [25].

However, the effect of ppGpp on protein synthesis and other post-transcriptional events [17,26] cannot be explained by its interaction with RNA polymerase.

Here we report the effect of *relA* and ppGpp on the synthesis of EPEC virulence factors. We show that deletion of *relA* impairs bacterial adherence, reduces the synthesis of the adhesins BFP and intimin and inhibits the transcription of the *perABC* operon.

Materials and Methods

Media and growth conditions

LB medium is as described [27]. T-salts medium is a Tris-buffered minimal medium supplemented with 0.2% glucose [28] and variable concentrations of KH_2PO_4 . Amino-triazole (AT) plates were prepared as described [29]. Dulbecco's Modified Eagle's Medium (DMEM) is a medium for epithelial cells (Cultilab-Brazil). HEp-2 cells were cultured in flasks containing DMEM enriched with 10% fetal calf serum (FCS), 50 U penicillin and 50 $\mu\text{g}/\text{ml}$ streptomycin at 37°C. The antibiotics were omitted in assays where bacteria were added. For overnight growth, bacteria were usually cultivated in LB medium, for all other purposes they were grown in DMEM. Growth rate was calculated according to the formula: $\mu = \frac{\ln \frac{N}{N_0}}{t}$, where N and N_0 respectively correspond to initial and final OD_{600} of the exponential growth phase and t is time-course of the growth curve.

Bacterial strains and plasmids

The strains and plasmids used in this study are described in Table 1.

Construction of strains and plasmids

Strain GMF204 (LRT9 $\Delta\text{relA}::\text{Km}$) was obtained by P1 transduction [27] from strain CF1652. Replacement of the $\Delta\text{relA}::\text{Km}$ mutation by the *relA*⁺ allele was performed by co-transducing *relA*⁺ and *argA::Tn10* (38 kDa apart) from strain CF12489 (*argA::Tn10 relA*⁺) into strain GMF204. The transductants were selected for tetracycline resistance and kanamycin sensitivity. The $\Delta\text{relA}::\text{Km}$ mutant and the *relA*⁺ transductant were tested for ppGpp accumulation under amino acid starvation (Figure S1) and also for growth on amino 1,2,4-triazole (AT) plates (not shown).

The $\Delta\text{relA}::\text{Cm}$ mutant was constructed by λ Red recombineering as described [30]. Primers relA1-fw-P1-fw and relA1-rev-P1-rev were used to amplify the CmR gene using plasmid pKD3 as a template. The PCR product was purified from a gel and electrotransformed into strain BW25113 (pKD46). The *relA* deletion was confirmed by PCR and then transduced to strain LRT9. To confirm the ΔrelA phenotype the resulting strain BS230 (LRT9 $\Delta\text{relA}::\text{Cm}$) was tested for ppGpp accumulation under amino acid starvation and for growth on amino-triazole (not shown). LG01 (LRT9 *lacZ::Tn5*) was obtained by P1 transduction from strain MG1655 *lacZ::Tn5*.

The BfpA and intimin proteins were tagged with the sequential peptide affinity (SPA) marker to allow the detection of these proteins in immunoblots. The SPA tag is a highly immunogenic 8 kDa peptide composed of a 3XFLAG tag preceded by a calmodulin binding protein epitope, which is recognized with high affinity by the commercially available anti-FLAG M2 antibody. The small size of the SPA tag and its integration next to the carboxy-terminal amino acid makes it unlikely to disturb protein activity [31]. Introduction of the SPA tag into *bfpA* and *eae* was performed essentially as described [31]. Intimin is a 94 kDa protein whose N-terminal is conserved and the C-terminal, which

bind to receptors on epithelial cells is highly polymorphic [32]. Thus, to insert the SPA tag in the C terminus of intimin in strain LRT9, we first sequenced the 3' end of LRT9 *eae* ORF. A 400 bp fragment encompassing the 3'-end of *eae* was amplified by PCR and sequenced. Alignment with the *eae* sequences published in the Genbank database showed that LRT9 carries a class β -1 intimin. To promote DNA recombination in LRT9 the $\Delta\text{recBCD}::\text{P}tac\text{-gam-bet-exo}$ (inducible λ -Red system)-*cat* genes were transduced from strain KM32 into LRT9. DNA hybrid primers *eae*3127(F)-SPA and *eae*3207(R)-SPA each containing at its 5'-end 40 nt of a sequence immediately upstream or immediately downstream of the *eae* stop codon and the adjacent 20 nt on each primer corresponding to the ends of the SPA-KmR sequence were used to amplify a 1.7 kb DNA fragment with plasmid pJL148 as a template. The PCR product was purified from an agarose gel and electrotransformed into LRT9, giving rise to strain LRT9 *eae::SPA-Km $\Delta\text{recBCD}::\text{P}tac\text{-gam-bet-exo cat}$* . Elimination of the λ -Red genes from LRT9 chromosome was attained by transducing the *argA::Tn10* allele from strain CF12489 (the *argA::Tn10* marker and the site where the *gam-bet-exo* genes were inserted are adjacent to each other on *E. coli* chromosome). Bacteria were selected for tetracycline resistance and tested for chloramphenicol sensitivity. The resulting strain thus carries the *eae::SPA-Km* fusion but not the λ -Red genes. LRT9 *eae::SPA $\Delta\text{relA}::\text{Cm}$* was obtained by P1 transduction from strain BS230.

Construction of LRT9 *bfpA::SPA* was essentially as described above for LRT9 *eae::SPA*, except that the hybrid primers used for recombineering (*bfpA*813(F)-SPA/*bfpA*893(R)-SPATR were specific for *bfpA*. To confirm that the SPA-tagging of intimin and BfpA was successful, LRT9 *bfpA::SPA-Km* and LRT9 *eae::SPA-Km* were assayed for BfpA and intimin by immunoblotting with a monoclonal anti-Flag antibody, as described below. Bands of the expected sizes, 110 kDa for intimin-SPA and 28 kDa for BfpA-SPA were observed and their levels were higher in DMEM at the exponential phase, and lower in cells grown in LB and in the stationary phase (not shown).

For the construction of plasmid pLG19, the *relA* ORF was amplified by PCR with primers relA6F/relA2633R and cloned in plasmid pTZ57R/T. The resulting plasmid was digested with BamHI/SacI and ligated in the same sites of plasmid pACT3. Plasmid pBS32 was obtained by ligating the GmR gene from pKNOCK-Gm digested with HincII into pLG19 DraI sites. The resulting plasmid pBS32 is Cm^{S} and Gm^{R} .

P1 Transduction

Transduction of chromosomal markers were performed with phage P1 *vir* essentially as described [27]. LRT9 can be transduced with P1 though less efficiently than *E. coli* K-12 strains [33].

ppGpp assay

For amino acid starvation, exponentially-growing cells were resuspended in T-salts minimal medium [28] supplemented with 0.2% glucose, 0.25 mM $^{32}\text{P-KH}_2\text{PO}_4$ and grown for 1 h. Amino acid starvation was started by adding 1 mg/ml serine hydroxamate (SH) to the cultures. Samples were collected at several time points following SH addition and immediately mixed with a 0.5 volume of cold formic acid. ppGpp under non-starving conditions was assayed by resuspending an overnight LB culture in DMEM containing only 0.25 mM KH_2PO_4 and 100 μCi ^{32}P . After 2 hours 0, 1, 10 or 100 μM IPTG were added. ppGpp was extracted and detected as described above. Following overnight incubation at -20°C , the extracts were centrifuged for 5 minutes at 10,000 rpm to precipitate cell debris, and 5 μl were applied to PEI-cellulose TLC-plates. The labeled nucleotides were resolved

Table 1. Bacterial strains, plasmids and DNA oligos used in this study.

Strains	Genotype	Source
BS230	BW25113 $\Delta relA::Cm$	This study
BW25113	<i>E. coli</i> K-12 carrying the recombineering plasmid pKD46	[30]
CF1652	MG1655 $\Delta relA::KmR$	M. Cashel
CF12489	<i>argA::Tn10 relA</i> ⁺	M. Cashel
CP01	MG1655 <i>lacZ::Tn5</i>	lab collection
KM32	$\Delta recBCD::Ptac-gam-bet-exo cat$	[54]
LRT9	EPEC O111:abH2	[33]
BS1230	LRT9 <i>bfpA::SPA-Km</i>	This study
BS1264	GMF204 <i>bfpA::SPA-Km</i>	This study
BS1298	LRT9 <i>eae::SPA-Km</i>	This study
BS1299	GMF204 <i>eae::SPA-Km</i>	This study
GMF204	LRT9 $\Delta relA::KmR$	This study
GMF302	LRT9 <i>argA::Tn10 relA</i> ⁺	This study
LG01	LRT9 <i>lacZ::Tn5</i>	This study
Plasmids		
pACT3	Cloning vector	[55]
pBS32	pLG19 GmR	This study
pGM17	<i>per</i> promoter cloned upstream to the promoterless <i>lacZ</i>	[33]
pJL148	SPA tag source	[31]
pKNOCK-Gm	Plasmid carrying the gentamycin resistance (GmR) gene	[56]
pLG19	<i>relA</i> ⁺ cloned in plasmid pACT3 under <i>Ptac</i>	This study
pTZ57R/T	Cloning vector	Thermo
Oligos		
bfpA813(F)-SPA	AAATACTGATTCAACCAATAAAGTTACATATTTTATGAAGTCCATGGAAAAGAGAAG	
bfpA813(F)-SPATR	CCCATAATAACGCCCAAAAACAGGGCGTATTATGTAGATTCATATGAATATCCTCCTTAG	
eae-lrt9(F)-SPA	TGGAGTAAACAATAAGAATGCTTTTTCTGTTTGTGAAAATCCATGGAAAAGAGAAGA	
eae-lrt9(R)-SPA	AATGAATTTTTATTTCCGGGATTTGAGATGTAATTAATTCATATGAATATCCTCCTTAG	
relA1-fw-P1-fw	ATGGTTGCGGTAAGAAGTGCACATATCAATAAGGCTGGTGAAGTGTAGGCTGGAGCTGCTTC	
relA1-rev-P1-rev	CTAACTCCCGTGCAACCGACGCGCTCGATAACATCCGGCACCATATGAATATCCTCCTTA	
relA6F	ATCCACCAGGTCAATCTTCAC	
relA2633R	AGGATATACCATTGCGCGAC	

doi:10.1371/journal.pone.0091703.t001

by one-dimensional TLC using 1.5 M KH_2PO_4 as a solvent. The amounts of ppGpp on the chromatograms were estimated by scanning the radioactivity of the spots in a Cyclone Plus storage phosphor system (Perkin Elmer) and calculating the level of ppGpp relative to that of GTP + ppGpp. The densitometric analysis was performed with the help of the software OptiQuant. Alternatively, the TLC plates were exposed to X-ray films and the densitometric analysis of the spots was performed with the software ImageJ [34].

$\Delta relA$ mutants are sensitive to the histidine analogue Amino-1,2,4-triazole (AT). Plates were prepared as described [29]. LRT9 derivatives were streaked on AT plates and incubated at 37°C for 24–48 h.

Adherence assay

A suspension containing 10^5 HEp-2 cells in 1 ml DMEM supplemented with 10% FCS was added to each well of a 24-well tissue plate and grown for 48 h at 37°C with 5% CO₂. The medium was removed from the cell monolayer and replaced with 1 ml of fresh DMEM supplemented with 2% FCS and 1% mannose. At this point, 5×10^7 bacteria previously grown for 18 h

in LB medium at 37°C were added to each well. After 3 h of incubation, the cell monolayer was washed six times with phosphate-buffered saline (PBS) to remove the non-adherent bacteria. The monolayer containing the adhered bacteria was treated with 1 ml 0.1% Triton X-100 in PBS for 5 minutes, bacteria were further diluted in PBS, plated onto LB-agar and incubated at 37°C for 24 h. On the next day, colonies were count and the number of colony forming units per ml (CFU/ml) was calculated.

β -galactosidase assay

Bacteria transformed with plasmid pGM17 were grown overnight in LB medium and diluted 100 times in DMEM. Samples were harvested every hour and assayed for growth (OD600) and β -galactosidase. When plasmid pLG19 was present, increasing concentrations of isopropyl β -D-1-thiogalactopyranoside (IPTG) were added to the cultures as specified in the text. The β -galactosidase assay was performed as described [27]. Briefly, 800 μ l of buffer Z (16.1 g/l Na_2HPO_4 , 5.5 g/l NaH_2PO_4 , 0.75 g/l KCl, 0.25 g/l $MgSO_4 \cdot 7H_2O$ and 2.7 ml/l β -mercaptoethanol)

were added to 200 μ l of permeabilized cells and the reaction was started with the addition of 200 μ l of a 4 mg/ml ortho-nitrophenyl-galactoside (ONPG) solution. Samples were incubated at 32°C until a yellow color developed and 500 μ l of a 1 M Na₂CO₃ solution was added to terminate the reaction. The reaction product was read at 420 nm and Miller units were calculated.

Western-blot analysis

Cells grown overnight in LB were diluted in 10 ml DMEM containing 2% FCS to a final OD₆₀₀ of 0.025 and incubated at 37°C without agitation up to the mid-late exponential phase. Proteins were extracted from 10⁹ bacteria by resuspending in 100 μ l of 5 \times diluted Lane Marker Sample Buffer (Thermo Scientific) and boiling for 5 minutes. 10–20 μ l of each sample were resolved by SDS-PAGE and transferred to a nitrocellulose membrane. Immunoblotting was performed with an anti-FLAG M2 primary antibody (Sigma) followed by a peroxidase conjugate of anti-mouse immunoglobulin G as secondary antibody. Protein bands were detected using the SuperSignal West Pico Chemiluminescent system (Pierce), as recommended by the manufacturer.

Sequencing of LRT9 *eae* 3'-end

For the sequencing of the highly polymorphic 3'-end of *eae* a DNA fragment of 369 bp was amplified by PCR with primers *eae*3036F/*eae*3536R using E2348/69 or LRT9 colonies as templates. The PCR products were purified directly with Wizard DNA Preps DNA purification system (Promega). DNA was sequenced by the Sanger method in an automatic sequencer type ABI Prism 3100 Genetic Analyzer (Applied Biosystems/Hitachi, Warrington, UK).

Results

Effect of Δ *relA* on adherence

To study the effect of *relA* on EPEC adherence, the Δ *relA*::Km allele was transferred to the EPEC strain LRT9. The LRT9 Δ *relA*::Km mutant did not accumulate ppGpp under amino acid starvation (Fig. S1) and was sensitive to amino-triazole (not shown), confirming the *relA* deficiency. To test the effect of Δ *relA* on EPEC adherence to epithelial cells, a standard *in vitro* adherence assay was conducted [33,35]. Bacteria grown overnight in LB were transferred to 24-well plates containing HEp-2 cells monolayers in DME medium and incubated for 3 hours to allow their adherence to the HEp-2 cells. The level of bacterial adherence was determined by counting the number of viable bacterial cells adhered to the HEp-2 cells monolayer at the end of the incubation period (Figure 1). Adherence of the Δ *relA* mutant was 75% lower than that of the *relA*⁺ parent (LRT9). Normal levels of adherence were restored when the *relA*⁺ allele was transduced back into the Δ *relA*::Km mutant (strain *relA*⁺ *argA*::Tn10). The inhibition in adherence caused by the *relA* deletion cannot be attributed to differences in growth rate, as the wild-type and the Δ *relA* mutant displayed very similar growth curves under the conditions employed in this assay (see below).

Effect of Δ *relA* on the expression of the *perABC* operon

In typical EPEC strains, both primary and intimate adherence are regulated by the products of the *perABC* operon. Transcription of the *bfp* and *perABC* operons [6,36] is dependent on PerA, while PerC is required for the synthesis of the master regulator Ler, which in turn activates the transcription of the LEE genes, including *eae* that encodes intimin [12,37]. To test whether the Δ *relA* mutation affects the expression of the *perABC* promoter

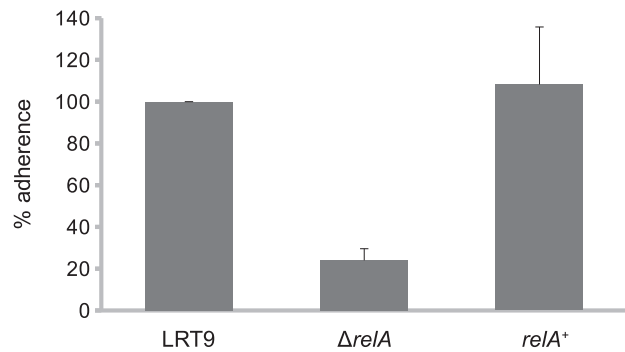


Figure 1. Effect of *relA* on the adherence of EPEC to epithelial cells. 5×10^7 bacteria grown overnight in LB broth were transferred to HEp-2 cells monolayers in DMEM supplemented with 2% FBS and incubated for 3 h. The cell wells were washed and the bacteria were released, diluted and plated on LB agar for CFU counting. To the wild-type strain was attributed the value of 100%. Each column represents the mean standard error of the mean (S.E.M.) of three independent experiments.

doi:10.1371/journal.pone.0091703.g001

plasmid pGM17 which harbors a *Pper-lacZ* fusion was transformed into LRT9 and into LRT9 Δ *relA*::Km. The same growth conditions used in a standard adherence assay (Fig. 1) were employed here. Bacteria grown overnight in LB, were diluted and further grown in DMEM without agitation for 8 h. Samples were harvested every hour and assayed for β -galactosidase activity (Fig. 2A). Expression of the *per* promoter (*Pper-lacZ*) in LRT9 increased three-fold as cells grew through the exponential phase reaching a peak at 4 h. The activity of *Pper-lacZ* in the Δ *relA* mutant was in average twice as lower as in the wild-type strain. For some unknown reason the *relA*⁺ *argA*::Tn10 transductant failed to fully restore the *Pper* original activity (not shown). To overcome this difficulty, the Δ *relA* mutant was transformed with plasmid pLG19 (*relA*⁺ cloned in a low-copy vector under *Ptac*). pLG19 not only complemented the mutation, but even slightly increased the expression of *Pper* when compared to the wild-type strain. This surplus activity is likely to be caused by the leakiness of *Ptac*, which is active even in the absence of IPTG increasing thus the cellular level of ppGpp (see below). Figure 2B shows the growth curves of the bacterial strains. The wild-type strain and the Δ *relA* mutant displayed similar growth curves throughout the experiment. At the exponential phase (1–6 h) the growth rate was $\mu = 0.51 \text{ h}^{-1}$ for LRT9, $\mu = 0.46 \text{ h}^{-1}$ for the Δ *relA* mutant and $\mu = 0.34 \text{ h}^{-1}$ for the pLG19 transformant. The slower growth of the latter is in part due to its slightly higher ppGpp level in the absence of IPTG (0.34 units of ppGpp in the transformant against 0.28 units of ppGpp in the non-transformed strain).

To further examine the effect of ppGpp on the expression of the *perABC* operon, plasmid pLG19 was transformed into LRT9 *lacZ*::Tn5 and into LRT9 Δ *relA*::Km *lacZ*::Tn5 carrying pGM17 (*Pper-lacZ*). β -galactosidase activity was followed as above, except that IPTG (0.001, 0.01, 0.1 or 1.0 mM) was added to the cultures (Figure 3). Similarly to what was observed in Figure 2A the mere presence of pLG19 increased *Pper-lacZ* expression when compared to the non-transformed wild-type strain. Addition of 1 μ M IPTG further boosted β -galactosidase activity, suggesting that a small increase in ppGpp positively affects *perABC* transcription. Higher concentrations of IPTG did not further stimulate *Pper-lacZ* activity because high levels of ppGpp cause growth arrest and general protein inhibition, limiting the beneficial effect of ppGpp on gene expression [23,38,39]. Indeed, the insert in Figure 3 shows that

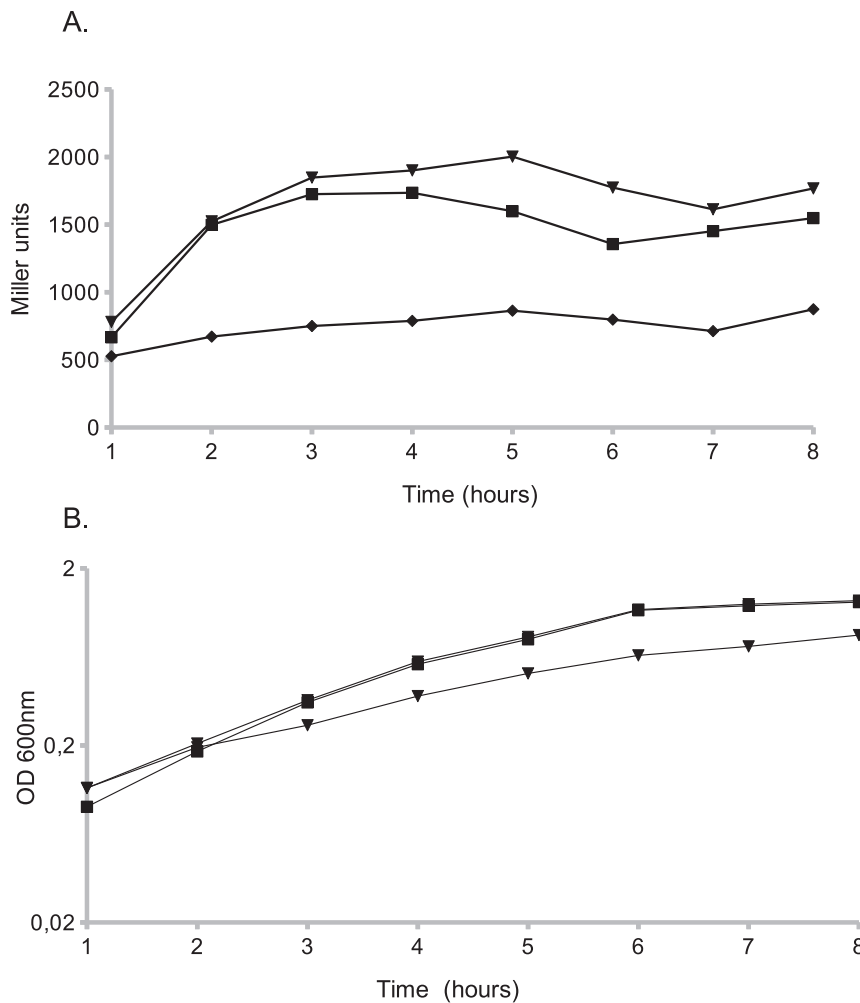


Figure 2. Effect of *relA* on *Pper* expression. Transformants carrying plasmid pGM17 (*Pper-lacZ*) were grown overnight in LB, resuspended in DMEM and grown for 8 hours without agitation. Samples were taken every hour and assayed for (A) β -galactosidase activity and (B) growth (OD_{600}). (■) wild-type (LRT9); (◆) $\Delta relA$ (GMF204); (▼) pLG19 ($relA^+$) in GMF204. Each point represents the average of two independent experiments. doi:10.1371/journal.pone.0091703.g002

addition of 0.1 and 1.0 mM IPTG caused a strong inhibition in growth rate. Figure S2 shows that LRT9 carrying pLG19 responds to increasing concentrations of IPTG in the medium by accumulating more ppGpp.

Effect of *relA* on EPEC adhesins

The type IV pilus BFP and the outer membrane protein intimin are the main adhesins of EPEC, both are positively regulated by the products of the *perABC* operon. Once ppGpp was shown to stimulate adherence and the transcription of the *perABC* operon, the next logical step was to test whether the synthesis of the EPEC adhesins was also affected by $\Delta relA$. A Sequential Peptide Affinity (SPA) tag [31] was inserted into the C-terminus of BfpA and intimin to detect the proteins by immunoblotting. The SPA DNA sequence was introduced by λ -*red* recombineering into the last codon of *bfpA* and *eae* ORFs (see Methods). Because the SPA-tag is linked to a kanamycin resistance gene, a new $\Delta relA::Cm^R$ mutant was constructed in a K-12 background and transduced into LRT9. The new LRT9 $\Delta relA::Cm^R$ mutant was tested for growth on amino-triazole and for ppGpp accumulation under amino acid limitation (data not shown). Likewise, the Cm^R marker in pLG19 ($relA^+$) was replaced by Gm^R to allow complementation *in trans* of

the $\Delta relA::Cm^R$ mutation. To assess the protein levels of BfpA and intimin, bacteria were grown under the same conditions used in the adherence assay, i.e., overnight growth in LB broth, followed by growth in DMEM without agitation. Proteins were extracted at the mid-late exponential phase, resolved by SDS-PAGE and immunoblotted with the anti-Flag monoclonal antibodies. Figure 4 shows that the bands corresponding to BfpA-SPA and intimin-SPA were considerably less intense in the $\Delta relA$ mutant than in the wild-type strain. Introduction of pBS32 ($relA^+$) not only complemented the mutation, but further increased the level of BfpA and intimin (compare the band intensities of the pBS32 transformant to that of the non-specific band). This indicates once more that a small increase in ppGpp has a positive effect on the synthesis of proteins associated with EPEC adherence. It can be concluded that *relA*/ppGpp play a positive role in the synthesis of the BFP fimbria and intimin and consequently, on EPEC adherence.

Discussion

The alarmone ppGpp controls many and diverse aspects of bacterial metabolism. Chiefly, ppGpp plays a pivotal role in maintaining the equilibrium between the intracellular amino acid

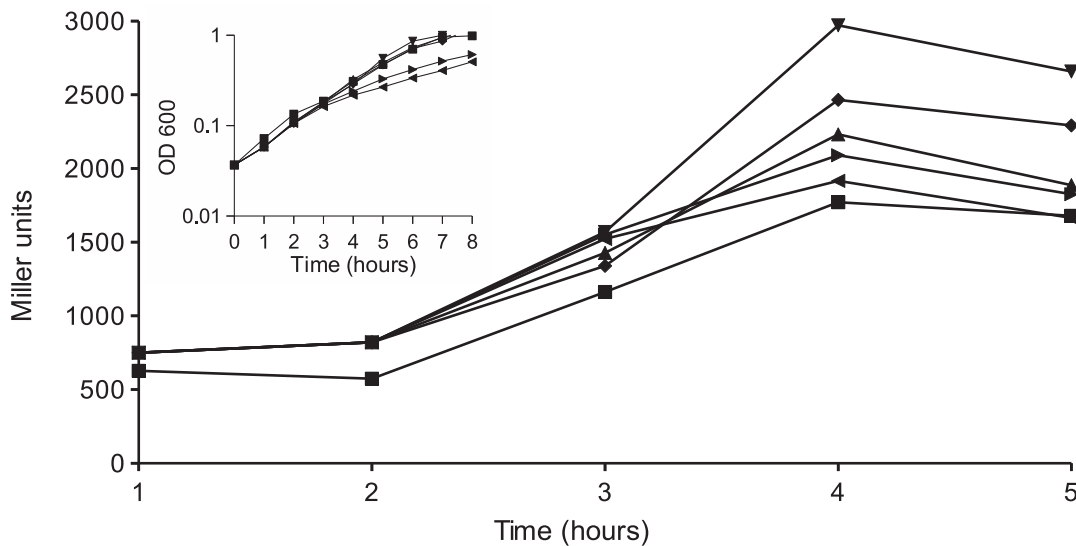


Figure 3. Effect of ppGpp overproduction on Pper. LG01 (LRT9 *lacZ::Tn5*) carrying pGM17 (*Pper-lacZ*) and LG01 carrying pGM17 and pLG19 (*Ptac-relA*⁺) were grown and sampled as in the legend to Figure 2. After 2 h of growth the bacteria were divided into five sub-cultures and (◆) 0, (▼) 0.001, (▲) 0.01, (▶) 0.1 or (◀) 1.0 mM IPTG were added; (■) represents the non-transformed wild-type strain. The insert shows the growth curves from 0 to 8 h. Each point represents the average of two independent experiments.
doi:10.1371/journal.pone.0091703.g003

pool and the rate of protein synthesis. This function is mediated by the ppGpp-synthetase RelA, which captures the signal from uncharged tRNAs at the ribosome A site and initiates the synthesis of ppGpp [16].

The present study provides evidence that the adherence of EPEC to epithelial cells and the expression of genes associated with adherence are positively regulated by ppGpp. This conclusion is supported by the following findings: deletion of *relA* significantly inhibited the adherence of EPEC to epithelial cells *in vitro*, reduced the protein levels of the adhesins Bfp and intimin and inhibited the expression of the *perABC* operon. Furthermore, mild overproduction of ppGpp had a beneficial effect on the expression of *bfp*, *eae* and *perABC*.

In typical EPEC, transcription of the LEE and of the *bfp* operon depends, respectively, on PerA [6,36] and PerC [12]. Sequence analysis of the *perABC* promoter in the LRT9 strain revealed an AT-rich discriminator sequence (-10 AAAATCAATAG +1), compatible with promoters activated by ppGpp [40,41]. In AT-

rich promoters ppGpp destabilizes the RNA polymerase (RNAP)-promoter complex allowing RNAP to escape and to carry out the elongation of the RNA chain [41]. It is thus been suggested that ppGpp stimulates EPEC adherence by enhancing the transcription of *perABC*, which in turn upregulates Bfp and intimin.

Another gene involved in ppGpp metabolism, *spoT*, encodes a bifunctional enzyme with a weak synthetase and a strong hydrolase activity [19]. *spoT* cannot be deleted from *E. coli* chromosome unless in a *relA* background because ppGpp over-accumulation would result in bacterial growth arrest. We managed to construct a ppGpp⁰ mutant ($\Delta relA \Delta spoT$) in LRT9. The inhibitory effect of the double mutant on adherence and on Pper-*lacZ* activity was, if anything, stronger than that of the *relA* single deletion (data not shown). However, the growth rate of the double mutant in DMEM was severely impaired, probably owe to the multiple auxotrophies caused by the complete absence of ppGpp in this strain [19]. It was therefore unclear to what extent the effect of the double deletion was caused by the lack of ppGpp or due to a deleterious effect on growth.

The assay of EPEC adherence to epithelial cells is usually performed by first growing the bacteria overnight in the nutrient-rich LB medium, followed by their transfer to HEp-2 cells monolayers seeded in DMEM [35]. The passage from an amino acid rich medium (LB) to a relatively poor medium such as DMEM mimics the pathway of EPEC through the digestive system, alternating portions of high nutrient content in the jejunum and the lower part of the small intestine, where nutrients are less abundant. Under these conditions RelA is likely to be activated, and a temporary increase in ppGpp ensues. Accordingly, EPEC adherence genes are poorly expressed in rich media [4,42] when ppGpp concentration is low and are activated upon transferring to DMEM.

The bacterial growth rates observed in the present study (0.4–0.5 h⁻¹) are similar to the one reported for *E. coli* in the host intestine [43], where *in vivo* adherence occurs. ppGpp is intimately associated with growth control and as shown here, it helps modulating the synthesis of EPEC adhesins. However, ppGpp level should not be as high as to cause complete growth arrest. We

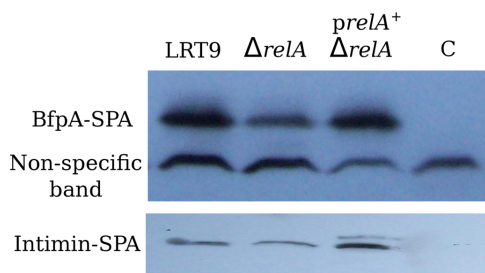


Figure 4. Effect of *relA* on the synthesis of BfpA and intimin. Bacteria grown overnight in LB broth were resuspended in DMEM and grown up to the mid-exponential phase. At this point the bacteria were harvested and immunoblotted with anti-SPA (FLAG 3X) antibodies. LRT9, wild-type strain; $\Delta relA$, LRT9 $\Delta relA::Cm$; *preLA*⁺ $\Delta relA$, pLG19 transformed in $\Delta relA::Cm$; C, control strain - LRT9 not carrying a *bfpA::SPA* or *eae::SPA* fusion. The picture is representative of two independent experiments.
doi:10.1371/journal.pone.0091703.g004

were able to reproduce the optimal concentration of ppGpp that results in the highest level of expression of the adherence-related genes by cloning *relA* in a low-copy plasmid under the control of *P_{tac}* and inducing the promoter with low concentrations of IPTG (Figs 3B and S2).

It has previously been shown that ppGpp enhances the expression of the LEE in enterohemorrhagic *E. coli* (EHEC) [44]. However, there are some important differences between EPEC and EHEC regarding the expression of the adhesin genes. First, even though both pathotypes share the LEE pathogenicity island, the EAF plasmid is not present in EHEC strains and the LEE genes are thus not regulated by PerC. Second, transcription of the LEE genes in EHEC begins at the mid-exponential phase and peaks at the late exponential/early stationary phase [45]. In contrast, in EPEC the LEE as well as the *perABC* and *bfp* operons are maximally activated at the mid-exponential-phase (Fig. 3 and [4]). Third, expression of the LEE genes as well as the adherence capacity of EHEC are higher in LB medium supplemented with bicarbonate [44], while EPEC neither adhere nor properly expresses the genes associated with adherence when grown in LB supplemented or not with bicarbonate ([4,11,42,46] and data not shown). Even though the environmental conditions required for the synthesis of EPEC and EHEC adherence factors are not identical, ppGpp positively affects the adherence of both lineages. This suggests that regulation by ppGpp is conserved regardless the specific mechanisms of control of adherence employed by the different diarrheogenic bacteria.

ppGpp is associated with bacterial virulence in several species. In most cases ppGpp plays a positive role and is required to fully induce the virulence genes. For instance in all *Proteobacteria* hitherto analyzed, such as *E. coli* (EHEC and UPEC), *Salmonella enterica*, *Yersinia pestis*, *Pseudomonas aeruginosa* *Francisella tularensis* and *Bordetella pertussis* a positive role for ppGpp was found [24,44,47–53]. This reinforces the notion that upregulation of bacterial virulence by ppGpp is an ancient evolutionary phenomenon.

Conclusions

The present study shows that the ppGpp synthetase RelA is required for the maximal adherence of EPEC to epithelial cells

References

- Moreno ACR, Filho AF, Gomes Tdo A, Ramos STS, Montemor LPG, et al. (2010) Etiology of childhood diarrhea in the northeast of Brazil: significant emergent diarrheal pathogens. *Diagn Microbiol Infect Dis* 66: 50–57.
- Dutta S, Guin S, Ghosh S, Pazhani GP, Rajendran K, et al. (2013) Trends in the prevalence of diarrheagenic *Escherichia coli* among hospitalized diarrheal patients in Kolkata, India. *PLoS One* 8: e56068.
- Scaletsky IC, Silva ML, Trabulsi LR (1984) Distinctive patterns of adherence of enteropathogenic *Escherichia coli* to HeLa cells. *Infect Immun* 45: 534–536.
- Puente JL, Bieber D, Ramer SW, Murray W, Schoolnik GK (1996) The bundle-forming pili of enteropathogenic *Escherichia coli*: transcriptional regulation by environmental signals. *Mol Microbiol* 20: 87–100.
- Donnenberg MS, Girón JA, Nataro JP, Kaper JB (1992) A plasmid-encoded type IV fimbrial gene of enteropathogenic *Escherichia coli* associated with localized adherence. *Mol Microbiol* 6: 3427–3437.
- Tobe T, Sasakawa C (2001) Role of bundle-forming pilus of enteropathogenic *Escherichia coli* in host cell adherence and in microcolony development. *Cell Microbiol* 3: 579–585.
- Cleary J, Lai LC, Shaw RK, Straatman-Iwanowska A, Donnenberg MS, et al. (2004) Enteropathogenic *Escherichia coli* (EPEC) adhesion to intestinal epithelial cells: role of bundle-forming pili (BFP), EspA filaments and intimin. *Microbiology* 150: 527–38.
- McDaniel TK, Jarvis KG, Donnenberg MS, Kaper JB (1995) A genetic locus of enterocyte effacement conserved among diverse enterobacterial pathogens. *Proc Natl Acad Sci* 92: 1664–1668.
- Donnenberg MS, Tzipori S, McKee ML, O'Brien AD, Alroy J, et al. (1993) The role of the *eae* gene of enterohemorrhagic *Escherichia coli* in intimate attachment in vitro and in a porcine model. *J Clin Invest* 92: 1418–1424.
- Chen HD, Frankel G (2005) Enteropathogenic *Escherichia coli*: unravelling pathogenesis. *FEMS Microbiol Rev* 29: 83–98.
- Martinez-Laguna Y, Calva E, Puente JL (1999) Autoactivation and environmental regulation of bfpT expression, the gene coding for the transcriptional activator of bfpA in enteropathogenic *Escherichia coli*. *Mol Microbiol* 33: 153–66.
- Gómez-Duarte OG, Kaper JB (1995) A plasmid-encoded regulatory region activates chromosomal *eaeA* expression in enteropathogenic *Escherichia coli*. *Infect Immun* 63: 1767–76.
- Cashel M (1969) The control of ribonucleic acid synthesis in *Escherichia coli*. iv. Relevance of unusual phosphorylated compounds from amino acid-starved stringent strains. *J Biol Chem* 244: 3133–3141.
- Edlin G, Donini P (1971) Synthesis of guanosine 5'-diphosphate, 2'-(or 3'-) diphosphate and related nucleotides in a variety of physiological conditions. *J Biol Chem* 246: 4371–4373.
- Spira B, Silberstein N, Yagil E (1995) Guanosine 3',5'-bispyrophosphate (ppGpp) synthesis in cells of *Escherichia coli* starved for Pi. *J Bacteriol* 177: 4053–8.
- Cashel M, Gentry D, Hernandez V, Vinella D (1996) *Escherichia coli* and *Salmonella*: Cellular and Molecular Biology, American Society for Microbiology, chapter The stringent response. pp. 1458–1496.
- Gentry DR, Hernandez VJ, Nguyen LH, Jensen DB, Cashel M (1993) Synthesis of the stationary-phase sigma factor sigma S is positively regulated by ppGpp. *J Bacteriol* 175: 7982–9.
- Goldman E, Jakubowski H (1990) Uncharged tRNA, protein synthesis, and the bacterial stringent response. *Mol Microbiol* 4: 2035–2040.
- Xiao H, Kalman M, Ikehara K, Zemel S, Glaser G, et al. (1991) Residual guanosine 3',5'-bispyrophosphate synthetic activity of *relA* null mutants can be eliminated by *spoT* null mutations. *J Biol Chem* 266: 5980–90.

and for the synthesis of the adhesins BFP and intimin. ppGpp positively affects the transcription of the *perABC* operon that encodes the main adherence regulators PerA and PerC. PerA positively controls the transcription of the *bfp* operon, while PerC activates the transcription of LEE1, whose product Ler, activates LEE5 (*cesT-tir-eae*) that encodes intimin. Therefore, by enhancing *perABC* ppGpp will ultimately upregulate adherence. EPEC can thus be added to the list of pathogenic bacteria whose virulence is enhanced by ppGpp.

Supporting Information

Figure S1 ppGpp accumulation in amino acid starved bacteria. Bacteria were treated with serine hydroxamate for 30 minutes. Samples were withdrawn at 15 and 30 minutes and ppGpp was resolved by TLC.

(EPS)

Figure S2 Accumulation of ppGpp caused by overproduction of *relA*. Exponentially growing bacteria (LRT9 carrying plasmid pLG19) in DMEM in the presence of 100 μ Ci 32P were treated with IPTG for 3 h. Samples were harvested each hour and assayed for ppGpp.

(EPS)

File S1 Supporting information.

(PDF)

Acknowledgments

The authors are very grateful to Mike Cashel for providing strains and very helpful comments.

Author Contributions

Conceived and designed the experiments: BS. Performed the experiments: BS GMF LGA. Analyzed the data: BS GMF LGA. Contributed reagents/materials/analysis tools: BS GMF LGA. Wrote the paper: BS.

20. Murray KD, Bremer H (1996) Control of *spoT*-dependent ppGpp synthesis and degradation in *Escherichia coli*. *J Mol Biol* 259: 41–57.
21. Spira B, Hu X, Ferenci T (2008) Strain variation in ppGpp concentration and RpoS levels in laboratory strains of *Escherichia coli* K-12. *Microbiology* 154: 2887–2895.
22. Ferenci T, Galbiati HF, Betteridge T, Phan K, Spira B (2011) The constancy of global regulation across a species: the concentrations of ppGpp and RpoS are strain-specific in *Escherichia coli*. *BMC microbiology* 11: 62.
23. Schreiber G, Metzger S, Aizenman E, Roza S, Cashel M, et al. (1991) Overexpression of the *relA* gene in *Escherichia coli*. *J Biol Chem* 266: 3760–7.
24. Dalebroux ZD, Svensson SL, Gaynor EC, Swanson MS (2010) ppGpp conjures bacterial virulence. *Microbiol Mol Biol Rev* 74: 171–199.
25. Hernandez VJ, Cashel M (1995) Changes in conserved region 3 of *Escherichia coli* sigma 70 mediate ppGpp-dependent functions in vivo. *J Mol Biol* 252: 536–49.
26. Lange R, Fischer D, Hengge-Aronis R (1995) Identification of transcriptional start sites and the role of ppGpp in the expression of *rpoS*, the structural gene for the sigma S subunit of RNA polymerase in *Escherichia coli*. *J Bacteriol* 177: 4676–4680.
27. Miller JH (1992) A Short Course In Bacterial Genetics: A Laboratory Manual And Handbook For *Escherichia coli* And Related Bacteria. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 876 pp.
28. Echols H, Garen A, Garen S, Torriani A (1961) Genetic control of repression of alkaline phosphatase in *E. coli*. *J Mol Biol* 3: 425–38.
29. Rudd KE, Bochner BR, Cashel M, Roth JR (1985) Mutations in the *spoT* gene of *Salmonella typhimurium*: effects on his operon expression. *J Bacteriol* 163: 534–42.
30. Datsenko KA, Wanner BL (2000) One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc Natl Acad Sci U S A* 97: 6640–5.
31. Zeghouf M, Li J, Butland G, Borkowska A, Canadien V, et al. (2004) Sequential peptide affinity (SPA) system for the identification of mammalian and bacterial protein complexes. *J Proteome Res* 3: 463–468.
32. Zhang WL, Köhler B, Oswald E, Beutin L, Karch H, et al. (2002) Genetic diversity of intimin genes of attaching and effacing *Escherichia coli* strains. *J Clin Microbiol* 40: 4486–4492.
33. Ferreira GM, Spira B (2008) The *pst* operon of enteropathogenic *Escherichia coli* enhances bacterial adherence to epithelial cells. *Microbiology* 154: 2025–2036.
34. Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image to ImageJ: 25 years of image analysis. *Nature Methods* 9: 671–675.
35. Cravioto A, Gross R, Scotland S, Rowe B (1979) Adhesive factor found in strains of *Escherichia coli* belonging to the traditional infantile enteropathogenic serotypes. *Curr Microbiol* 3: 95–99.
36. Bustamante VH, Calva E, Puente JL (1998) Analysis of cis-acting elements required for *bfpA* expression in enteropathogenic *Escherichia coli*. *J Bacteriol* 180: 3013–3016.
37. Bustamante VH, Santana FJ, Calva E, Puente JL (2001) Transcriptional regulation of type III secretion genes in enteropathogenic *Escherichia coli*: *Ler* antagonizes H-NS-dependent repression. *Mol Microbiol* 39: 664–678.
38. Svitil AL, Cashel M, Zyskind JW (1993) Guanosine tetraphosphate inhibits protein synthesis in vivo: a possible protective mechanism for starvation stress in *Escherichia coli*. *J Biol Chem* 268: 2307–11.
39. Spira B, Yagil E (1998) The relation between ppGpp and the PHO regulon in *Escherichia coli*. *Mol Gen Genet* 257: 469–477.
40. Potrykus K, Cashel M (2008) (p)ppGpp: still magical? *Annu Rev Microbiol* 62: 35–51.
41. Gummeson B, Lovmar M, Nyström T (2013) A proximal promoter element required for positive transcriptional control by ppGpp and DksA during the stringent response. *J Biol Chem*.
42. Kenny B, Abe A, Stein M, Finlay BB (1997) Enteropathogenic *Escherichia coli* protein secretion is induced in response to conditions similar to those in the gastrointestinal tract. *Infect Immun* 65: 2606–2612.
43. de Jong P, Vissers MMM, van der Meer R, Bovee-Oudenhoven IMJ (2007) In silico model as a tool for interpretation of intestinal infection studies. *Appl Environ Microbiol* 73: 508–515.
44. Nakanishi N, Abe H, Ogura Y, Hayashi T, Tashiro K, et al. (2006) ppGpp with DksA controls gene expression in the locus of enterocyte effacement (LEE) pathogenicity island of enterohaemorrhagic *Escherichia coli* through activation of two virulence regulatory genes. *Mol Microbiol* 61: 194–205.
45. Frankel G, Phillips AD, Rosenshine I, Dougan G, Kaper JB, et al. (1998) Enteropathogenic and enterohaemorrhagic *Escherichia coli*: more subversive elements. *Mol Microbiol* 30: 911–921.
46. Girón JA, Ho AS, Schoolnik GK (1991) An inducible bundle-forming pilus of enteropathogenic *Escherichia coli*. *Science* 254: 710–713.
47. Aberg A, Shingler V, Balsalobre C (2006) (p)ppGpp regulates type I fimbriation of *Escherichia coli* by modulating the expression of the site-specific recombinase FimB. *Mol Microbiol* 60: 1520–1533.
48. Pizarro-Cerdá J, Tedin K (2004) The bacterial signal molecule, ppGpp, regulates *Salmonella* virulence gene expression. *Mol Microbiol* 52: 1827–1844.
49. Sun W, Roland KL, Branger CG, Kuang X, Curtiss R III (2009) The role of *relA* and *spoT* in *Yersinia pestis* KIM5+ pathogenicity. *PLOS One* 4: e6720.
50. Erickson DL, Lines JL, Pesci EC, Venturi V, Storey DG (2004) *Pseudomonas aeruginosa relA* contributes to virulence in *Drosophila melanogaster*. *Infect Immun* 72: 5638–5645.
51. Faron M, Fletcher JR, Rasmussen JA, Long ME, Allen LAH, et al. (2013) The Francisella tularensis *migR*, *trmE*, and *cphA* genes contribute to *F. tularensis* pathogenicity island gene regulation and intracellular growth by modulation of the stress alarmone ppGpp. *Infect Immun* 81: 2800–2811.
52. Sugisaki K, Hanawa T, Yonezawa H, Osaki T, Fukutomi T, et al. (2013) Role of (p)ppGpp in biofilm formation and expression of filamentous structures in *Bordetella pertussis*. *Microbiology* 159: 1379–1389.
53. Kalia D, Meray G, Nakayama S, Zheng Y, Zhou J, et al. (2013) Nucleotide, c-di-GMP, c-di-AMP, cGMP, cAMP, (p)ppGpp signaling in bacteria and implications in pathogenesis. *Chem Soc Rev* 42: 305–341.
54. Murphy KC, Campellone KG, Potete AR (2000) PCR-mediated gene replacement in *Escherichia coli*. *Gene* 246: 321–330.
55. Dykxhoorn DM, St Pierre R, Linn T (1996) A set of compatible tac promoter expression vectors. *Gene* 177: 133–6.
56. Alexeyev MF (1999) The pKNOCK series of broad-host-range mobilizable suicide vectors for gene knockout and targeted DNA insertion into the chromosome of gram-negative bacteria. *Biotechniques* 26: 824–6, 828.