

# Minireview: Role of Glia in Neuroendocrine Function

LUIS M. GARCIA-SEGURA AND MARGARET M. MCCARTHY

*Instituto Cajal (L.M.G.-S.), Consejo Superior de Investigaciones Científicas, 28002 Madrid, Spain; and Department of Physiology and Program in Neuroscience (M.M.M.), University of Maryland, Baltimore, School of Medicine, Baltimore, Maryland 21201*

Long relegated to the backwaters of neuroendocrinology, it is becoming increasingly apparent that glial cells of the central and peripheral nervous system are key participants because they are capable of both sending and receiving hormonal signals. Hormones are also a critical component of neuronal/glia cross talk, leading to neuromodulatory and neurotrophic actions under physiological and pathological conditions. In the peripheral nervous system, hormonal actions on Schwann cells and hormonal metabolites produced by these glial cells promote myelin formation and the remyelination and regen-

eration of injured nerves. In the central nervous system, glial cells participate in the hormonal regulation of synaptic function, synaptic plasticity, myelin formation, cognition, sleep, and the response of nervous tissue to injury. In addition, central glial cells participate in the regulation of hormonal secretion by hypothalamic neurons. Therefore, glial cells are a key element to understanding hormonal actions in the nervous system and the regulation of neuroendocrine events. (*Endocrinology* 145: 1082–1086, 2004)

THE NAME OF glia derives from the German word for glue because Rudolf Virchow, in his original description of these cells in 1846, considered that they were the “glue” of the brain that gives structural support to neurons. Now, our view of glial cells is very different (1). We know that these cells are functional components of the neural tissue that express receptors for neurotransmitters and show excitability based on intracellular  $Ca^{2+}$  variations. Glial cells integrate signals emanating from neurons and other glial cells, including hormonal inputs. They also regulate extracellular concentrations of ions, metabolites, and neurotransmitters to coordinate the differentiation, metabolism, and excitability of neurons and modulate synaptic transmission (1). There is limited evidence that glial cells contain vesicles and release transmitters in much the same way as neurons (2). The preferential localization of glia around synapses further enhances their ability to modulate excitability in the brain and has led to the coining of the term “tripartite synapse” (3, 4).

Hormones regulate the development and function of the nervous system to influence physiology and behavior. Fifty years of research have focused on the effects of hormones on neurons or neuronal systems to modulate physiology and behavior, with scarcely a mention of glial cells of any variety. This is not a condemnation of the science of neuroendocrinology because the same was more or less true for all of neuroscience. We now know that hormonal actions in the brain are exerted equally on neurons and glial cells, and that the ability of glia to both respond to and produce hormones has profound functional implications in virtually every neuroendocrine system. Glial cells express many of the same

hormone receptors as found on neurons, such as receptors for melatonin, thyroid and steroid hormones, vasopressin, oxytocin, leptin, corticotropin-releasing factor, glucagon, insulin, and IGF-I. The closer we look, the more it becomes apparent that glial cells play a critical role as mediators of these hormonal messages to the nervous system. Glial cells regulate the activity of neurosecretory neurons, participate in the control of hormonal release, and can even serve as the source of hormone themselves. Here we review some of the recent advances in the study of the role of glial cells in mediating hormonal actions in the nervous system and in the regulation of hormonal secretion. In our view, the additional role of hormones in modulating the tripartite synapse lends a level of complex control that maximizes the ability of the organism to respond to dynamic changes in the internal as well as external environment.

## Who Is Who among Glial Cells?

Glial cells are classified in two main groups: microglia and macroglia. Microglia are macrophage-like cells that regulate the inflammatory response of the neural tissue to injury or infection. Macroglia are subdivided in four specialized cell types: ependymal cells, Schwann cells, oligodendroglia, and astroglia. Ependymal cells line the cerebral ventricular cavities and the central canal of the spinal cord and are in direct contact with the cerebrospinal fluid. Schwann cells and oligodendroglia are responsible for the formation of myelin, an essential component facilitating action potential propagation. Schwann cells form the myelin in the peripheral axons and oligodendroglia in central axons. In addition, both cell types regulate axonal excitability. Astroglia refers to a heterogeneous population that includes astrocytes, marginal glia, radial glia in the developing brain, Bergmann cells in the cerebellar cortex, Müller cells in the retina, pituicytes in the neurohypophysis, and tanocytes in the hypothalamus. All of these cells are grouped as astroglia because they have in common the expression of glial fibrillary acidic protein

Abbreviations: GABA,  $\gamma$ -Aminobutyric acid; GFAP, glial fibrillary acidic protein;  $PGE_2$ , prostaglandin  $E_2$ .

*Endocrinology* is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.

(GFAP). However, this is probably an artificial classification that groups together cells with different functions. Astroglial processes maintain contact with blood vessels, neurons, and other glial cell types. Astrocytic processes rarely extend beyond a 50- $\mu\text{m}$  radius, allowing them to exert largely local control. However, propagation of  $\text{Ca}^{2+}$  waves through networks of astrocytes, via the release of signaling molecules such as ATP, may affect the activity of distant neurons integrated in different neuronal circuits (1, 3, 4).

### Glia Cells Mediate Hormonal Signaling

Integration of hormonal signaling by glial cells occurs in at least two fundamental ways: 1) the hormone acts directly on the glia, which in turn signals to the neuron to modulate its function (5, 6). Signaling to the neuron may involve secretion of a growth factor, neurohormone, or transmitter-like substance (a variation of this involves the glia signaling to the neuron by physically interacting with it, meaning that the membranes become immediately juxtaposed or disassociated); and 2) the hormone acts first on the neuron, which then releases a substance to signal to the glia, which presumably then signals back to that or to other neurons (5, 6).

Untangling precisely what type of cross talk between neurons and glial cells is occurring is extremely challenging, and the distinctions we have made are largely operationally defined. It is plausible that in some cases where steroids appear to be acting directly on astrocytes, further examination may reveal a previously undiscovered preliminary step involving neurons. Nonetheless, examples for each can be found. For instance, the profound influences of thyroid hormone ( $\text{T}_3$ ) on brain development are in part due to its direct actions on astroglia and oligodendroglia (6, 7).  $\text{T}_3$  signaling is necessary to promote the complete differentiation of oligodendrocyte precursor cells and, therefore, the formation of central myelin (7, 8).  $\text{T}_3$  also regulates Schwann cell function and differentiation, thereby affecting peripheral myelin (9). Progesterone and progesterone derivatives such as dihydroprogesterone and tetrahydroprogesterone, also promote central and peripheral myelination acting on oligodendrocytes (10) and Schwann cells (11), respectively.

There is limited evidence that gonadal hormones also act directly on glial cells to regulate the building and remodeling of synaptic contacts (5). For instance, estradiol reorganizes astrocytic laminin into extracellular fibrillar arrays that facilitate neurite extension (12), although the precise mechanisms of the estrogen effect on laminin have not been established. The hormone also increases expression of TrkA receptors in astrocytes. As a consequence, the hormone may regulate axonal growth and synaptic plasticity because nerve growth factor, the ligand for TrkA, stimulates astrocytes to function as substrates for axon growth (13). In addition, gonadal hormones interact with growth factors secreted by glial cells to promote axonal growth of developing neurons in the central nervous system, and this likely involves the hormone acting directly on the neuron but requiring the involvement of the glial cell (14). Finally, direct hormonal actions on microglia and reactive astroglia may affect the endogenous inflammatory response of the nervous system and the outcome of neurodegenerative processes (6, 15–18).

Examples that involve steroids acting first on neurons to initiate a circular conversation between neurons and astrocytes that ultimately returns to neurons are found in the developing brain. Gonadal steroid-mediated sex differences in astrocyte morphology have been correlated with changes in dendritic spine synapse density in the neonatal brain (19, 20). In the case of the developing arcuate nucleus, it is clear that the primary effect of the steroid is at the neuron, to promote release of  $\gamma$ -aminobutyric acid (GABA), which then promotes the extension and branching of astrocytic processes (21). A second example is found in the nearby preoptic area where astrocytes can induce dendritic spine synapse formation by local release of glutamate. Of critical importance is that the glutamate release from the astrocytes is signaled by prostaglandin  $\text{E}_2$  ( $\text{PGE}_2$ ), which is synthesized in neurons. The control of  $\text{PGE}_2$  synthesis is via estradiol acting on the rate-limiting enzyme cyclooxygenase 2 (22). Thus, an endocrine signal initiated in the neuron is transmitted to the astrocyte, which then feeds back on the neurons to regulate synapse formation.

### Glia Cells Synthesize Active Hormonal Metabolites

Glial cells metabolize hormones as well as synthesize active metabolites that affect neuronal function. Combined metabolism and synthesis is the case for thyroid hormones, where astrocytes and tanocytes play a key role in the conversion of  $\text{T}_4$  to the active metabolite  $\text{T}_3$  (6). Glial cells can also convert steroids into neuroactive steroids. In particular, glial cells metabolize native steroid hormones into their 5 $\alpha$ - and 3 $\alpha$ -hydroxy-5 $\alpha$  reduced derivatives via the enzymatic complex formed by the 5 $\alpha$ -reductase and the 3 $\alpha$ -hydroxysteroid dehydrogenase. Thus, testosterone is converted into dihydrotestosterone and then into 5 $\alpha$ -androstane-3 $\alpha$  and 17 $\beta$ -diol, progesterone is converted into dihydroprogesterone and subsequently into tetrahydroprogesterone, corticosterone into dihydrocorticosterone, and deoxycorticosterone into dihydrodeoxycorticosterone and then into tetrahydrodeoxycorticosterone (23, 24). The presence of the enzyme steroid 21-hydroxylase, which is required for the conversion of progesterone to its downstream metabolites, was recently confirmed in cultured astrocytes (25).

The steroids produced by glial cells may serve a paracrine role regulating synaptic function as these steroids are known to modulate anxiety, cognition, sleep, ingestion, aggression, and reinforcement (23, 24). Some of them are positive modulators of *N*-methyl-D-aspartate receptors and enhance cognitive performance. Other steroids produced by glial cells, such as tetrahydroprogesterone and tetrahydrodeoxycorticosterone, are highly selective and extremely potent modulators of the GABA<sub>A</sub> receptor and elicit marked anxiolytic and stress reducing effects, as well as increase feeding and promote sleep (23, 24). The metabolism of steroid hormones by glial cells is important under pathophysiological conditions. For instance, progesterone and progesterone derivatives produced by Schwann cells promote axonal regeneration and remyelination after peripheral nerve lesions, whereas steroid metabolites produced by central glial cells are protective against neurodegenerative stimuli (23, 26).

### Glial Cells Modulate Hormonal Secretion by Hypothalamic Neurons

In addition to being a target for hormonal action, glial cells act as modulators of hypothalamic neurosecretory neurons. The best example has been provided by the laboratory of Sergio Ojeda, showing that neuron-glia signaling mediated by growth factors of the epidermal growth factor family and the expression of their associated erbB tyrosine kinase receptors in hypothalamic astrocytes are a requisite for the timely initiation of mammalian puberty in female animals (27, 28). Astrocytes control GnRH neurons by different mechanisms. These glial cells mediate estrogen-induced synaptic plasticity in GnRH cells and in hypothalamic areas that project to GnRH neurons (5, 6, 12). In addition, astrocytes and tanyocytes, in response to hormonal steroids, release factors that control the activity of GnRH neurons, such as PGE<sub>2</sub>, the TGF $\alpha$ ,  $\beta$ 1 (TGF $\beta$ 1),  $\beta$ 2 (TGF $\beta$ 2), the basic fibroblast growth factor, and the IGF-I (12, 29, 30). Astrocytes also regulate the local intraneuronal formation of steroids able to intervene as negative or positive signals in the feedback control of GnRH neurons (31). Changes during the estrous cycle in the extension of tanyocyte cell processes in the external zone of the median eminence, modulate the access of GnRH nerve terminals to the portal vasculature (32, 33). In addition tanyocytes release factors, such as IGF-I that are involved in the estrogen-induced synaptic remodeling of hypothalamic neurons involved in the control of GnRH cells (33) (Fig. 1).

Involvement of glial cells in the regulation of hypothalamic hormone release has also been demonstrated for the magnocellular neurosecretory neurons of the supraoptic and paraventricular nuclei (34, 35). Under conditions of intense neurohypophysial hormone secretion, such as lactation, par-

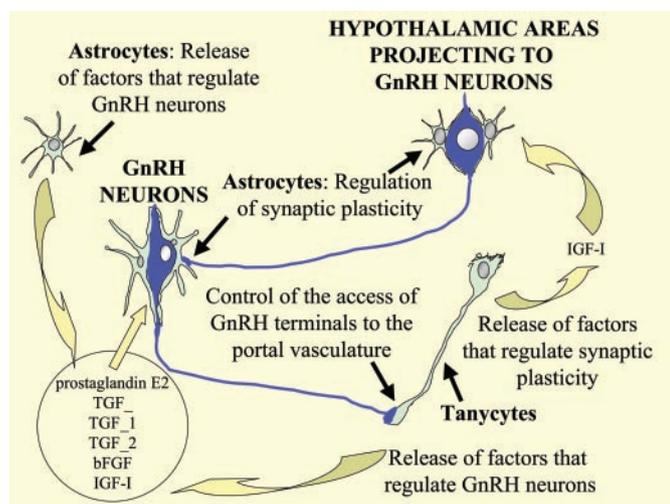


FIG. 1. Summary of the sites of action and the mechanisms involved in the regulation of GnRH secretion by astroglia. 1) Astrocytes secrete PGE<sub>2</sub> and growth factors [TGFs, basic fibroblast growth factor (bFGF), IGF-I] that act on GnRH neurons (6, 28–30, 33). 2) Astrocytes regulate synaptic connectivity on GnRH neurons and in hypothalamic areas projecting to GnRH neurons, such as the hypothalamic arcuate nucleus (5, 33) Tanyocytes control the access of GnRH terminals to portal vasculature in the median eminence and release factors that affect GnRH neurons or modulate synaptic plasticity of hypothalamic neurons involved in the control of GnRH cells (6, 28–30, 33).

turition, and chronic dehydration, astrocytic processes in these nuclei retract and the glial coverage of neuronal membranes decreases. This change in glial coverage is accompanied by a remodeling of synaptic contacts and probably has additional important consequences for neuronal excitability because it modifies extracellular ionic homeostasis and glutamate neurotransmission (34, 35). Similar changes in the extension of glial processes associated with hormone release have been observed in pituicytes, the glial cells of the neurohypophysis. Under conditions of low hormonal demand, pituicyte processes surround neurosecretory axon terminals at the neurovascular contact zone. In contrast, when hormonal demand is enhanced, pituicytes withdraw their processes, apparently playing a permissive role for hormonal

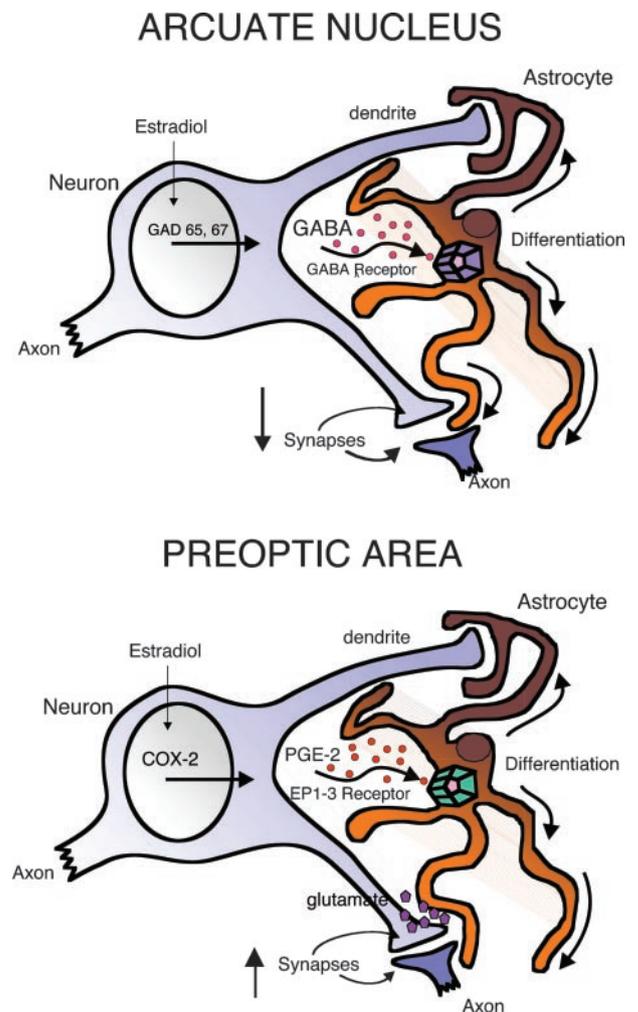


FIG. 2. Neuronal/glial cross talk is region specific. In the developing arcuate nucleus, estradiol-mediated increases in neuronal GABA synthesis and release leads to increased stellation of neighboring astrocytes. The increased complexity of astrocytes is inversely correlated with the density of dendritic spine synapses on arcuate neurons, but a mechanistic relationship between these two architectures has not yet been established (19–21). In the developing preoptic area, estradiol-mediated increases in PGE<sub>2</sub> synthesis and release results in increased stellation of astrocytes and a corresponding increase in the density of dendritic spines on neighboring neurons. The increase in dendritic spines induced by PGE<sub>2</sub> is dependent on activation of AMPA receptors by glutamate, which is presumably released from the astrocytes (22). COX-2, Cyclooxygenase 2; EP1–3, PGE<sub>2</sub> 1–3 receptors.

release (34, 35). Furthermore, astrocytes and pituicytes may release taurine in response to hypo-osmotic stimulation. Taurine, in turn, activates glycine receptors in neurosecretory neurons and inhibits vasopressin release (36). The interaction of pituicytes with vasopressinergic axons represents a good example of the interaction between glia and neurons to regulate a defined physiological function.

### Glial/Neuronal Cross Talk Is Regionally and Developmentally Specific

Neuroendocrine function is tightly regulated by discrete groups of neurons, and the astrocytes that interact with those populations can be expected to have a distinct phenotype as well. Regional variation in development has offered a unique window into site specificity by the simple nature of the fact that different areas of the brain develop at different rates. This is in part reflected by the astrocytes themselves. As mentioned, previously, expression of GFAP is a distinguishing characteristic for a large and varied group of astroglia. GFAP is also a hallmark of astroglia maturation and as such its expression pattern varies from region to region. Interestingly, both the preoptic area and arcuate nucleus, two brain regions importantly involved in neuroendocrine regulation and highly sexually differentiated, have robustly GFAP-expressing astrocytes as early as the day of birth. Hormonal modulation of synaptic development in both these brain regions involves cross talk between astrocytes and neurons (Fig. 2). In contrast, the mediobasal hypothalamus has immature astrocytes at birth and synaptogenesis is static between males and females at this developmental stage. Thus, differences in the maturation of astrocytes in particular brain regions may preclude or allow hormonal modulation of developmental processes (37).

We have mentioned the variety of receptor types expressed by astrocytes, and just as with neurons, it would be expected that these would vary regionally and developmentally as well. An additional critical mediator of hormonal modulation in astrocyte-neuron cross talk is the presence or absence of nuclear receptors. Hormonal nuclear receptors in glia show developmental regulation and have regionally specific distributions (6, 37–47). The relative contribution to neuroendocrine function of regionally specific glia phenotype and developmental regulation of hormone receptors in glia will no doubt be the continued focus of future studies and lead inevitably to questions of how such variance is established and maintained.

In summary, although there is still much to be learned on the role of glial cells in neuroendocrine regulation and hormonal signaling, we know that glial cells are able to respond to hormonal and neuronal signals and then to propagate the activation to other glial cells and to neurons for long distances. This provides glial cells with the capacity to amplify and expand hormonal effects to distant uncoupled neuronal circuits. It also provides glial cells with the capacity to coordinate over time, for phasic hormonal release, the activity of distant neurons involved in neuroendocrine regulation. Furthermore, glial cells may produce local active hormonal metabolites when and where they are needed under physiological and pathological conditions. Developmentally, glial

cells may contribute to the establishment and permanent maintenance of sexually dimorphic synaptic patterning that mediates adult differences in sexual behavior and neuroendocrine function. Therefore, glial cells represent a very relevant cellular element to be taken in consideration to understand the mechanisms of neuroendocrine regulation.

### Acknowledgments

Received October 15, 2003. Accepted December 2, 2003.

Address all correspondence and requests for reprints to: Margaret M. McCarthy, Department of Physiology and Program in Neuroscience, University of Maryland, Baltimore, School of Medicine, Baltimore, Maryland 21201

Research presented in this review was supported in part by grants from the National Institutes of Mental Health (MH52716) and the National Science Foundation (IBN-9511328) (to M.M.M.) and from the Ministerio de Ciencia y Tecnologia (SAF 2002-00652) and the Commission of the European Communities, specific RTD program "Quality of Life and Management of Living Resources" (QLK6-CT-2000-00179) (to L.M.G.-S.).

### References

1. Araque A, Carmignoto G, Haydon PG 2001 Dynamic signaling between astrocytes and neurons. *Annu Rev Physiol* 63:795–813
2. Krzan M, Stenovec M, Kreft M, Pangrsic T, Grlic S, Haydon PG, Zorec R 2003 Calcium-dependent exocytosis of atrial natriuretic peptide from astrocytes. *J Neurosci* 23:1580–1583
3. Araque A, Parpura V, Sanzgiri RP, Haydon PG 1999 Tripartite synapses: glia, the unacknowledged partner. *Trends Neurosci* 22:208–215
4. Volterra A, Magistretti PJ, Haydon PG 2002 The tripartite synapse. *Glia in synaptic transmission*. New York: Oxford University Press
5. Garcia-Segura LM, Chowen JA, Parducz A, Naftolin F 1994 Gonadal hormones as promoters of structural synaptic plasticity: cellular mechanisms. *Prog Neurobiol* 44:279–307
6. Garcia-Segura LM, Chowen JA, Naftolin F 1996 Endocrine glia: roles of glial cells in the brain actions of steroid and thyroid hormones and in the regulation of hormone secretion. *Front Neuroendocrinol* 17:180–211
7. Rodriguez-Pena A 1999 Oligodendrocyte development and thyroid hormone. *J Neurobiol* 40:497–512
8. Baas D, Legrand C, Samarut J, Flamant F 2002 Persistence of oligodendrocyte precursor cells and altered myelination in optic nerve associated to retina degeneration in mice devoid of all thyroid hormone receptors. *Proc Natl Acad Sci USA* 99:2907–2911
9. Mercier G, Turque N, Schumacher M 2001 Rapid effects of triiodothyronine on immediate-early gene expression in Schwann cells. *Glia* 35:81–89
10. Ghomari AM, Ibanez C, El-Etr M, Leclerc P, Eychemen B, O'Malley BW, Baulieu EE, Schumacher M 2003 Progesterone and its metabolites increase myelin basic protein expression in organotypic slice cultures of rat cerebellum. *J Neurochem* 86:848–859
11. Azcoitia I, Leonelli E, Magnaghi V, Veiga S, Garcia-Segura LM, Melcangi RC 2003 Progesterone and its derivatives dihydroprogesterone and tetrahydroprogesterone reduce myelin fiber morphological abnormalities and myelin fiber loss in the sciatic nerve of aged rats. *Neurobiol Aging* 24:853–860
12. Rozovsky I, Wei M, Stone DJ, Zanjani H, Anderson CP, Morgan TE, Finch CE 2002 Estradiol (E2) enhances neurite outgrowth by repressing glial fibrillary acidic protein expression and reorganizing laminin. *Endocrinology* 143:636–646
13. McCarthy JB, Barker-Gibb AL, Alves SE, Milner TA 2002 TrkA immunoreactive astrocytes in dendritic fields of the hippocampal formation across estrous. *Glia* 38:36–44
14. Dhandapani KM, Mahesh VB, Brann DW 2003 Astrocytes and brain function: implications for reproduction. *Exp Biol Med* (Maywood) 228:253–260
15. Stone DJ, Rozovsky I, Morgan TE, Anderson CP, Hajian H, Finch CE 1997 Astrocytes and microglia respond to estrogen with increased apoE mRNA in vivo and in vitro. *Exp Neurol* 143:313–318
16. Mor G, Nilsen J, Horvath T, Bechmann I, Brown S, Garcia-Segura LM, Naftolin F 1999 Estrogen and microglia: a regulatory system that affects the brain. *J Neurobiol* 40:484–496
17. Bruce-Keller AJ, Keeling JL, Keller JN, Huang FF, Camondola S, Mattson MP 2000 Antiinflammatory effects of estrogen on microglial activation. *Endocrinology* 141:3646–3656
18. Vegeto E, Bonincontro C, Pollio G, Sala A, Viappiani S, Nardi F, Brusadelli A, Viviani B, Ciana P, Maggi A 2001 Estrogen prevents the lipopolysaccharide-induced inflammatory response in microglia. *J Neurosci* 21:1809–1818

19. **Mong JA, McCarthy MM** 1999 Steroid-induced developmental plasticity in hypothalamic astrocytes: implications for synaptic patterning. *J Neurobiol* 40:602–619
20. **Mong JA, Roberts RC, Kelly JJ, McCarthy MM** 2001 Gonadal steroids reduce the density of axospinous synapses in the developing rat arcuate nucleus: an electron microscopy analysis. *J Comp Neurol* 432:259–267
21. **McCarthy MM, Amateau SK, Mong JA** 2002 Steroid modulation of astrocytes in the neonatal brain: implications for adult reproductive function. *Biol Reprod* 67:691–698
22. **Amateau SK, McCarthy MM** 2002 A novel mechanism of dendritic spine plasticity involving estradiol induction of prostaglandin-E2. *J Neurosci* 22:8586–8596
23. **Melcangi RC, Magnaghi V, Galbiati M, Martini L** 2001 Formation and effects of neuroactive steroids in the central and peripheral nervous system. *Int Rev Neurobiol* 46:145–176
24. **Baulieu EE, Robel P, Schumacher M** 2001 Neurosteroids: beginning of the story. *Int Rev Neurobiol* 46:1–32
25. **Lovelace M, Watson TG, Spehenson GL** 2003 Steroid 21-hydroxylase expression in cultured rat astrocytes. *Brain Res Bull* 61:609–615
26. **Schumacher M, Guennoun R, Mercier G, Desarnaud F, Lacor P, Benavides J, Ferzaz B, Robert F, Baulieu EE** 2001 Progesterone synthesis and myelin formation in peripheral nerves. *Brain Res Brain Res Rev* 37:343–359
27. **Ojeda SR, Ma YJ** 1999 Glial-neuronal interactions in the neuroendocrine control of mammalian puberty: facilitatory effects of gonadal steroids. *J Neurobiol* 40:528–540
28. **Ojeda SR, Prevot V, Heger S, Lomniczi A, Dziedzic B, Mungenast A** 2003 Glia-to-neuron signaling and the neuroendocrine control of female puberty. *Ann Med* 35:244–255
29. **Ojeda SR, Dissen GA, Junier MP** 1992 Neurotrophic factors and female sexual development. *Front Neuroendocrinol* 13:120–162
30. **Melcangi RC, Martini L, Galbiati M** 2002 Growth factors and steroid hormones: a complex interplay in the hypothalamic control of reproductive functions. *Prog Neurobiol* 67:421–449
31. **Micevych P, Sinchak K, Mills RH, Tao L, LaPolt P, Lu JK** 2003 The luteinizing hormone surge is preceded by an estrogen-induced increase of hypothalamic progesterone in ovariectomized and adrenalectomized rats. *Neuroendocrinology* 78:29–35
32. **Prevot V** 2002 Glial-neuronal-endothelial interactions are involved in the control of GnRH secretion. *J Neuroendocrinol* 14:247–255
33. **Garcia-Segura LM, Naftolin F, Hutchison JB, Azcoitia I, Chowen JA** 1999 Role of astroglia in estrogen regulation of synaptic plasticity and brain repair. *J Neurobiol* 40:574–584
34. **Theodosis DT** 2002 Oxytocin-secreting neurons: a physiological model of morphological neuronal and glial plasticity in the adult hypothalamus. *Front Neuroendocrinol* 23:101–135
35. **Miyata S, Hatton GI** 2002 Activity-related, dynamic neuron-glial interactions in the hypothalamo-neurohypophysial system. *Microsc Res Tech* 56:143–157
36. **Hussy N, Bres V, Rochette M, Duvoid A, Alonso G, Dayanithi G, Moos FC** 2001 Osmoregulation of vasopressin secretion via activation of neurohypophysial nerve terminals glycine receptors by glial taurine. *J Neurosci* 21:7110–7116
37. **Mong JA, Glaser E, McCarthy MM** 1999 Gonadal steroids promote glial differentiation and alter neuronal morphology in the developing hypothalamus in a regionally specific manner. *J Neurosci* 19:1464–1472
38. **Lima FR, Gervais A, Colin C, Izembart M, Neto VM, Mallat M** 2001 Regulation of microglial development: a novel role for thyroid hormone. *J Neurosci* 21:2028–2038
39. **Carlson DJ, Strait KA, Schwartz HL, Oppenheimer JH** 1996 Thyroid hormone receptor isoform content in cultured type 1 and type 2 astrocytes. *Endocrinology* 137:911–917
40. **Bury F, Carre JL, Vega S, Ghandour MS, Rodriguez-Pena A, Langley K, Sarlieve LL** 2002 Coexpression of thyroid hormone receptor isoforms in mouse oligodendrocytes. *J Neurosci Res* 67:106–113
41. **Billon N, Jolicoeur C, Tokumoto Y, Vennstrom B, Raff M** 2002 Normal timing of oligodendrocyte development depends on thyroid hormone receptor  $\alpha 1$  (TR $\alpha 1$ ). *EMBO J* 21:6452–6460
42. **Finley SK, Kritzer MF** 1999 Immunoreactivity for intracellular androgen receptors in identified subpopulations of neurons, astrocytes and oligodendrocytes in primate prefrontal cortex. *J Neurobiol* 40:446–457
43. **Magnaghi V, Cavarretta I, Galbiati M, Martini L, Melcangi RC** 2001 Neuroactive steroids and peripheral myelin proteins. *Brain Res Brain Res Rev* 37:360–371
44. **Blurton-Jones M, Tuszyński MH** 2001 Reactive astrocytes express estrogen receptors in the injured primate brain. *J Comp Neurol* 433:115–123
45. **Garcia-Ovejero D, Veiga S, Garcia-Segura LM, DonCarlos LL** 2002 Glial expression of estrogen and androgen receptors after rat brain injury. *J Comp Neurol* 450:256–271
46. **Kruijver FP, Balesar R, Espila AM, Unmehopa UA, Swaab DF** 2002 Estrogen receptor- $\alpha$  distribution in the human hypothalamus in relation to sex and endocrine status. *J Comp Neurol* 454:115–139
47. **Platania P, Laureanti F, Bellomo M, Giuffrida R, Giuffrida-Stella AM, Catania MV, Sortino MA** 2003 Differential expression of estrogen receptors  $\alpha$  and  $\beta$  in the spinal cord during postnatal development: localization in glial cells. *Neuroendocrinology* 77:334–340

*Endocrinology* is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.