Title:

Plant facilitation occurs between species differing in their associated arbuscular mycorrhizal fungi

Running headline: Phylogenetic signal in plant-AMF

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Summary

_ Complementary beneficial effects of different arbuscular mycorrhizal fungi(AMF) can result in a more efficient exploitation of the soil nutrients available, thus influencing plant communities. Here, we hypothesize that plant–AMF specificity is mediated by phylogenetic constraints defining possible interactions, and that plant–AMF interaction patterns can influence plant–plant facilitation specificity.

_ We reanalyzed previous data describing plant-plant and plant-AMF interaction at the community level to specifically test for a phylogenetic signal on plant and AMF interactions and for a relationship between plant-plant facilitation specificity and plant species differences in their AMF associates.

_ Closely related AMF operational taxonomical units (OTUs) tend to interact with the same plant species, but there is not a significant signal in the interaction through the plant phylogeny. This indicates that the similarity in theAMFassociates of two plant species is independent of their phylogenetic relatedness. Interestingly, plant–AMF interactions match plant facilitation specificity, with pairs of plant species recruiting more frequently under each other tending to have different AMF associates.

_ An increment of AMF diversity in the rhizosphere, as a result of plant–AMF and plant–plant selectivity, is suggested as a potential driver of plant–plant facilitation. This study highlights the role of plant–AMF interactions in shaping plant community assemblages.

Introduction

Plant-plant facilitation is an ecological process occurring in communities worldwide (Valiente-Banuet et al., 2006) required for the establishment of most woody plant species in semiarid environments (Valiente-Banuet & Verdú, 2008). Plant-plant facilitation is considered as a positive interaction in which the presence of one plant enhances the growth, survival or reproduction of a neighbor plant. Facilitation does not need to be a mutualistic interaction where both participants gain (+,+), but may occur only as a commensalism (+,0) in which just the facilitated species gets a benefit. However, mere species coexistence without any positive interaction (0,0) is not considered plant facilitation (Callaway, 2007). Plant facilitation has been experimentally shown to be species-specific, so that benefactor (nurse) species tend to promote the establishment of specific beneficiary (facilitated) species more strongly than others (Valiente-Banuet & Verdú, 2007, 2008; Castillo et al., 2010). Hereafter we will refer to it as plant-plant facilitation specificity. Plant-plant facilitation is patent when adult nurses are facilitating seedlings of the facilitated species. However, even though this positive interaction may turn into competition over time, a high percentage of specific positive plant-plant interactions remains when facilitated seedlings become adults (Valiente-Banuet & Verdú, 2008). The maintenance of plant-plant facilitation over time implies that the benefits of the association are not only related to germination and seedling establishment, but also extended to growth and long term survival. Understanding the mechanisms that promote and maintain plant-plant facilitation will contribute to a better understanding of assembly mechanisms in plant communities.

Mechanisms traditionally considered to underlie plant-plant facilitation involve avoidance of abiotic stresses such as an improved shade or moisture availability (Callaway, 2007). In this case plant-plant facilitation strength will be expected to be independent of ecological interactions (Fig. 1a for the specific case of plant-AMF interactions). However, it is unlikely to explain a stronger facilitation between specific plant species considering only abiotic stress avoidance. Plant-plant facilitation specificity is more likely explained by plant species-specific traits, including traits involved in ecological interactions. Plant-plant facilitation tends to occur among distantly related plant species (Valiente-Banuet & Verdú *et al.*, 2008). Simultaneously, phylogenetic relatedness could be underlying species similarities in traits influencing ecological interactions with both mutualistic (i.e. AMF) and antagonistic (i.e. pathogens) partners. A phylogenetic conservatism of ecological interactions is a recurrent phenomenon across the entire tree of life (Gómez et al., 2010). The tendency of related species to resemble each other may result in sharing interacting partner species (phylogenetic signal). Recent studies have started to hypothesize that the role of mycorrhizal fungi interconnecting plants are crucial to understand plant-plant facilitation processes (Van der Heijden & Horton, 2009; Van der Putten, 2009). One potential mechanism by which mycorrhizal fungi can promote plant-plant facilitation is that they can interconnect plant individuals from different species, genera and even families in natural communities (Newman, 1988). Plant interconnections provide pathways for the transference of nutrients such as Nitrogen (He et al., 2004, 2005), Phosphorus (Smith et al., 2001), Arsenic, Cesium and Rubidium (Meding & Zasoski, 2008). The mutualism between plant and arbuscular mycorrhizal fungi (AMF) can have considerable importance for seedling establishment, enhancing access to a nutrient absorption without requiring completely developed tissue structures (Kytoviita et al., 2003; Van der Heijden, 2004). Later on, the development of plant-plant facilitation could be maintained through the plant interconnections provided by AMF. Plant-plant facilitation may be stronger between plant species with similar AMF if there is an overall benefit due to an overlap of the AMF associated to each plant species (hereafter plant fungal-niche) driven by an increment in the abundance of the shared AMF (Fig. 1b). On the contrary, plant-plant facilitation specificity may be mediated by the degree of complementarity in the interacting plants fungal-niche. It has been shown in experimental approaches that plant co-existence and productivity increases with increasing numbers of AMF species, due to the added beneficial effect of each single AMF species (Van der Heijden et al., 1998; Hartnett & Wilson, 1999; Wagg et al., 2011). Increasing AMF diversity has been suggested to result in a more efficient exploitation of soil nutrients and in a better use of the resources available in the system (Van der Heijden et al., 1998). In this sense, plant-plant facilitation may be stronger between plant species which harbor different AMF, leading to an overall benefit due to an increase of AMF diversity in the shared rhizosphere (Fig. 1c). Host specificity is likely to be an important driver shaping AMF communities (Haussman & Hawkes, 2010), and likewise, AMF communities can influence plant species performance (Maherali & Klironomos, 2007) through strong bottom-up controls on plant community

composition (Grime *et al.*, 1987; Van der Heijden *et al.*, 1998a,b; Hartnett & Wilson, 1999). The role of microorganisms promoting plant species coexistence was first suggested considering avoidance of belowground antagonistic interactions (Janzen, 1970; Connell, 1971; Packer & Clay, 2000; Reinhart *et al.*, 2003; Van der Putten, 2009), but the influence of belowground mutualistic interactions promoting plant-plant facilitation has been less considered (but see Dickie *et al.*, 2002, 2005).

Plant-AMF interaction networks have been recently described for a semiarid community (Montesinos-Navarro et al., 2012) showing that there is a non-random interaction pattern between plant and AMF communities as revealed by significant network nestedness and modularity. Nestednessimplies that ecological-specialists (i.e. with the lower number of links) on one party (e.g. plants) tend to interact with a subset of the ecological-generalist (i.e. with the higher number of links) species on the other party (e.g. AMF). Specifically, in this plant-AMF network there are few AMF ecological-generalists which interact with almost every plant species in the community (Montesinos-Navarro et al., 2012). Modularity, in addition, reflects weakly interlinked subsets of species (modules) which internally consist of strongly connected species (Olesen et al., 2007). Plant and AMF species are grouped within a module on the basis of shared interactions, which means a similar interaction niche (set of species with which they interact) and in some cases this interaction pattern is independent of the spatial association of the species (Montesinos-Navarro et al., 2012). Under this scenario, this compartmentalization of the network can emerge, among other processes, from a phylogenetic conservatism of ecological interactions among taxa (Lewinsohn et al., 2006; Olesen et al., 2007), suggesting that specific plant-AMF association may be limited by phylogenetic constraints. Analyzing the phylogenetic conservatism in module membership is analogous to exploring phylogenetic conservatism in host selectivity (Gómez et al., 2010).

In this paper we test whether plant-AMF interactions are phylogenetically structured. In other words, if 1) there is a non-random phylogenetic pattern in plant-AMF interaction; 2) the modules of the network, previously shown to be independent of species spatial association (Montesinos-Navarro *et al.*, 2012) can emerge from such phylogenetic pattern (i.e. modules are composed by closely related plant and/or closely related AMF species); and 3) there is a relationship between facilitation specificity and plant species fungal-niche. We expect that as suggested for other ecological networks, there will be a significant phylogenetic signal in plant-AMF network resulting in a

phylogenetic conservatism of module membership. In addition, we expect that plantplant facilitation strength between specific species depends on their plant fungal-niche, potentially influencing the overall plants nutrients uptake in the shared rhizosphere.

Materials and methods

DATA BASE

This study is based on available data published by Montesinos-Navarro et al. (2012), collected in the semiarid Valley of Zapotitlán, in the state of Puebla, Mexico (18° 20N, 97° 28 W). It is a xeric shrubland dominated by the columnar cactus Neobuxbaumia tetetzo, Agave spp. and different species belonging to the families Fabaceae and Asteraceae. Non-lignified root segments from 130 individuals of 37 plant species, representing the relative abundance of each species, were collected to characterize AMF community (see further details in Montesinos-Navarro et al., 2012). Glomeromycota 18S (SSU) Internal Transcribed Spacer (SSU-full ITS) was amplified through a nested PCR (methods thoroughly described in Montesinos-Navarro et al. 2012). No amplification was obtained for the families Gigasporaceae and Acaulosporaceae. Less than 30% amplification success was obtained for Glomus group B primer-pair, whereas a 78.21% success was achieved for the primer-pair of Glomus group A, suggesting a predominance of Glomus A in the AMF communities in the study area. Glomus is the most common AMF in many field sites encompassing 70% of the AMF species identified al., 2002; Öpik et (range 60%-85%) (Helgason *et* al. 2009; Vandenkoornhuyse et al. 2002; Zhaoyong et al. 2006; Alguacil et al. 2009; Sonjak et al., 2009; Wilde et al., 2009; Öpik et al., 2010). Although subsequent sequencing of PCR products was continued only with the predominant monophyletic group of Glomus A due to financial constraints, the general pattern of interactions described with this subset of AMF has been previously shown to be largely generalized to other systems with higher AMF diversity (Montesinos-Navarro et al., 2012). The importance of using molecular techniques to prevent an underestimation of AMF richness has been increasingly highlighted. However, there is an unavoidable difficulty to precisely define AMF species using molecular techniques, due to the large lack of precise knowledge about intra-specific genetic variation for multiple species. Some studies have traditionally used a standard cut off of 3% of genetic dissimilarity to consider AMF

species, but the use of this or any other specific cut-off can be controversial. We defined Operational Taxonomic Units (OTUs) for AMF according to their DNA sequence dissimilarity at a wide range of reasonable cut-off values based on the range of intra and inter-specific genetic variation recently described in this genomic region for *Glomus* A (Stockinger *et al.* 2010, Schoch *et al.* 2012). Rarefaction curves performed at 1% to 10% DNA sequence dissimilarity cut-offs, reached the stabilization between 5% and 8% (Montesinos-Navarro *et al.*, 2012). All the analyses were performed at seven cut-offs ranging from 4 to 10% of DNA sequence dissimilarity. For the analyses regarding module species membership we used the modules defined by Montesinos-Navarro *et al.* (2012) which have been shown to be independent of spatial species association.

AMF AND PLANT PHYLOGENIES

For AMF, SSU-full ITS sequencing described above (Genbank accession numbers in Table S1), was used to build the AMF phylogenetic tree considering an SSU-fill ITS sequence of *Paraglomus* downloaded from Genbank (accession number FN555285) as the outgroup to root the phylogenetic tree. Phylogenetic analyses of the nuclear ribosomal SSU-full ITS sequences were carried out in the CIPRES (Cyberinfrastructure for Phylogenetic Research) web portal (Miller *et al.*, 2010) using the probabilistic Maximum Likelihood (ML) method, as implemented in RAxML blackbox with the default settings (Stamatakis, 2006; Stamatakis *et al.*, 2008). One hundred bootstrap trees were inferred to provide support values for the best-scoring ML tree (Fig. S1). We repeated the analyses to obtain another best-scoring tree and assess the robustness of our results to different phylogenetic trees. As the two best-scoring phylogenetic trees were very similar, with just a few tips showing different placements, results did not change substantially and thus, only those using the first tree will be shown. The tips of this tree which differ in less than a given cut-off (4 to 10% genetic dissimilarity) were collapsed in order to obtain the AMF phylogeny for each cut-off.

For plants, the phylogenetic distance matrix was obtained from the community phylogeny generated with Phylocom 4.2 (Webb *et al.*, 2008). This program produces a community phylogeny by matching the family names of our study species with those contained in a backbone phylogeny, which is themegatree of the Angiosperm Phylogeny Group III (Stevens, 2005). We then resolved the phylogenetic relationships

at the species level of the Cactaceae and Agavaceae families based on published phylogenies of Good-Avila (2006) and Hernández-Hernández et al. (2011). Our final tree includes all the plant species in which the amplification of AMF DNA was positive (35 out of the 37 species sampled) and was fully resolved with the exception of two polytomies (Fig. S2). The tree was calibrated with age estimates from Wikstrom et al. (2001) plus six ages obtained from the chronograms published by Good-Avila (2006)and Arakaki et al. (2011) for the nodes of Cactaceae (35 Mya), Mammillaria (6.3 Mya), and the splits between Mammillaria and Coryphantha (19.7 Mya), Agave karwinski and Agave macroacantha (6.7 Mya). Calibration was made with the phylocom bladj algorithm that evenly distributes the undated nodes between dated nodes or between dated nodes and terminals (Webb et al., 2008).

ANALYSES

PHYLOGENETIC SIGNAL OF THE INTERACTION

The phylogenetic signal of plant-AMF interactions was estimated with the estimated generalized least squares (EGLS) procedure of Ives and Godfray (2006). This procedure is similar to that developed for a single trait (Blomberg *et al.*, 2003) but considering the matrix of interactions as the target trait. The method calculates the strength of the phylogenetic signal in the plant-AMF interactions acting through both the AMF (d_{AMF}) and the plant (d_{plant}) species phylogenies. We used the association rate of plant species *k* on AMF OTU *i* (A_{ik}) as a measure of the strength of association between plant and AMF species following eq. 4 in Ives and Godfray (2006):

Aik=-log(1-Fik/Hi)

where *Hi* is the number of DNA sequences of the AMF OTU *i* and *F_{ik}* is the number of sequences of OTU *i* found in the plant species *k*. Note that there is a typo in the original article, indicating H_i/F_{ik} instead of the correct expression F_{ik}/H_i (A.G. Ives, pers. comm.).

 A_{ik} depends, as stated by Ives & Godfray (2006), on both the selectivity and abundance of species, two crucial variables explainingthe number of interactions occurring in facilitation networks (Verdú & Valiente-Banuet, 2008). In addition to quantitative matrices, we re-calculated the strength of association (A_{ik}) by transforming the interaction matrix into a binary matrix where 1/0 denotes the presence or absence of interaction, respectively. Although this approach may have other limitations, it avoids problems derived from assigning abundances on the basis of the number of identical copies of DNA in a given root sample, potentially coming from the same AMF individual. We perform the analyses with both quantitative and binary matrices to evaluate the robustness of the results.

The procedure estimates an EGLS model to fit A_{ik} in terms of the observed association strength and the separate effects of the plant and AMF species phylogenies. The model is based on the Ornstein-Uhlenbeck model of evolution, which incorporates stabilizing selection and drift, and detects the presence of phylogenetic signal through the parameter d. This parameter determines the strength of phylogenetic signal, with d=0 indicating the lack of phylogenetic correlation and d=1 corresponding to the Brownian motion assumption (i.e pure drift model). The goodness of fit of the different models was estimated by comparing the mean squared error calculated for 1) the full model (MSE), 2) a "star" phylogeny (MSE_{star}), and 3) a Brownian evolution model (MSE_b). The model minimizing the mean squared error was considered the best fit. We estimated d values for both plant (d_{plant}) and AMF (d_{AMF}) sets of species. Statistical significance was estimated by calculating bootstrap 95% confidence intervals as described in Ives and Godfray (2006). Analyses to assess the phylogenetic signal of the interaction were performed in Matlab (The MathWorks, Inc.) version 7.10.0.499.

PHYLOGENETIC SIGNAL OF MODULE MEMBERSHIP

Phylogenetic signal in module membership was determined following the method proposed by Maddison & Slatkin (1991). This test estimates whether the minimum number of evolutionary steps in a character on a phylogenetic tree is lower than

expected by chance. It was determined whether the steps occurred less than expected by chance using a null model in which data were reshuffled 1000 times across the tips of the phylogeny. The character was the module to which the species was ascribed by the annealing algorithm (Guimerà & Amaral, 2005a, b). An annealing algorithm uses a probabilistic function to find the optimum solutionbased on the "locality" of the preceding solution considering the improvement gained in each move. Module was considered as an unordered, multi-state factor. We mapped the evolution of module membership onto our phylogenetic trees. Analyses to assess phylogenetic signal in module membership were performed in R version 2.13.2 using the function "phylo.signal.disc" developed by Enrico Rezende and the species belonging to each module were extracted from Montesinos-Navarro*et al.* 2012.

RELATIONSHIP BETWEEN FACILITATION AND FUNGAL- NICHE OF PLANTS

Plant-plant facilitation has been corroborated for some species in this system using experimental approaches, showing that seedling establishment is enhanced in the understory of distantly related plant species (Castillo et al., 2010). In addition, studies considering the whole plant community have provided results supporting this facilitation pattern (Valiente-Banuet & Verdú, 2007, 2008, Verdú et al., 2010, Verdú & Valiente-Banuet, 2011). Plant-plant facilitation matrices from Valiente-Banuet & Verdú (2008) and Verdú et al. (2010) were used to characterize the strength of the facilitation interaction among each pair of plant species in the community. The strength of plantplant facilitation species was estimated for each pair of plant species as follows. Contingency analyses were used to compare the number of individuals on each facilitated-species recorded under each nurse species and in open spaces with the expected number of individuals derived from the proportions of area of plant-cover vs. open space considering the total cover of perennial plants and open space in four 1000m² transects (Verdú *et al.*,2010). This matrix was built considering only the plant-plant facilitation interactions (i.e seedling plant species recruiting under nurse plant species) that remain with time, resulting in plant-plant facilitation interactions among adult plant species (see Valiente-Banuet & Verdú, 2008 for a deeper description of the matrix). The plant-plant facilitation matrix was reduced to contain only the plant species on which information of their associated AMF was available (Montesinos-Navarro et al., 2012). Dissimilarity in plant species interaction niche was calculated based on the composition of AMF in each plant species roots (i.e. a plant fungal-niche). The number of AMF OTUs with which a given plant species interact (plant species degree) is influenced by the plant species relative abundance. However, there is a correlation between plant species degree and the mean number of AMF OTUs per individual plant (i.e. AMF load), indicating that plant abundance alone is not enough to explain the number of AMF interactions per species. In fact, both plant abundance and AMF load equally contribute to explain plant species degree (Montesinos-Navarro *et al.*, 2012). Pairwise values of plant species dissimilarity in their fungal niche were calculatedusing a Euclidean distance index. The statistical significance of the correlation among plant-plant facilitation strength and plant fungal-niche dissimilarity matrices was tested against a null model "frequency" in the PICANTE package implemented in R (Kembel *et al.* 2010). The correlation between log-transformed plant-plant facilitation and plant-fungal-niche matrices was tested for each cut-off using from 4% to 10% of genetic dissimilarity to define AMF OTUs.

CONTRIBUTION OF PLANT RELATIVE ABUNDANCE TO FACILITATION PATTERNS

Plant species relative abundance can be considered as a species specific intrinsic characteristic. As many other traits, the relative abundance can influence the species interaction pattern with other species or it could also be the result of its interaction pattern. In any case, including species relative abundance, reflected in the sampling design, is essential in order to approach species interaction patterns at the community level. An abundant plant species in the community will have a higher probability of interacting with a higher number of species. However, the combined effect of relative abundance with other ecological processes can be a better predictor of the interaction patterns, than the neutral process of abundance alone. Previous studies on plant facilitation in this system have shown that the frequency of interactions between a pair of plant species is by far better explained by the combined effect of relative abundance (Verdú & Valiente-Banuet, 2011). This indicates that there is a tendency of pairwise abundant plants to interact between them, but the final frequency of pairwise

interactions is shaped by an additional tendency to interact with distantly related species.

Similarly, we tested if the dissimilarity in plant-fungal-niche combined with plant abundance can better explain plant facilitation strength than solely plant species relative abundance. We evaluate the ability of abundance and dissimilarity in plant-fungal-niche to explain plant-plant facilitation strength by means of the likelihood approach developed by Vázquez *et al.* (2009).

The likelihood of the models including the following matrices to explain the observed matrix of facilitation interactions was estimated: a) null matrix; b) plant abundance matrix, c) plant-fungal-niche dissimilarity matrix; and d) abundance \times plant-fungal niche dissimilarity matrix.

The null matrix was defined as a matrix in which all pairwise interactions had the same probability. The probability matrix derived of plant abundances was constructed by multiplying the vectors of nurse and facilitated plant abundances recorded in 112 vegetation patches in the study area. Normalization was made in the resulting matrix so that their elements added up to one. The probability matrix derived of plant-fungal niches was constructed by normalizing the dissimilarity matrix in plant-fungal niche among plants so that their elements added up to one.

The models likelihood and AIC were recalculated considering every cut off from 4% to 10% to define AMF OTUs and using both quantitative and binary matrices to define the plant-AMF interaction pattern. The model with lower Akaike Information Criteria (AIC) was selected as the best model. As a rule of thumb, models whose AIC is less than 2 units larger than the best model have a substantial support, whereas those models resulting in AIC values >10 units larger have virtually no support (Burnham & Anderson, 2002). In addition, the likelihood of being a better model than the best model was estimated by means of AICs weights.

Results

Considering the number of modules across cut-offs, our plant-AMF interaction network has on average six modules. Focusing, for example, on the representative cut-off of 7%, from the AMF point of view, the two most ecological-generalist AMF OTUs were grouped in the same module with 18 ecological-specialist plants (open diamonds in Fig.

2), whereas from the plant perspective, the most ecological-generalist plants belonged to different modules (open and close circles and open squares modules in Fig. 2). In general terms, modules tend to be composed of ecological-generalists species of one party (either plants or AMF) with ecological-specialists of the other party.

A significant phylogenetic signal in the plant-AMF interactions is observed through the AMF phylogeny; the model considering the phylogenetic signal has a better fit than the models considering no phylogenetic covariances or Brownian motion for most of the cut-off values (4 to 8% considering both quantitative and binary matrices (Table 1). However, the phylogenetic signal of the interaction through plant phylogeny was close to zero and not significant for any cut-off and for both quantitative and binary matrices (Table 1). In other words, closely related AMF tend to interact with the same set of plant species, but the tendency of plant species to interact with the same set of AMF OTUs is independent of their phylogenetic relatedness.

When the membership of an AMF OTU to a particular module is mapped onto the AMF phylogeny (Fig. 2), a significant phylogenetic signal emerges for every cut-off from 4 to 9% (Table 2), indicating that closely related AMF tend to belong to the same module. In the case of plants (Fig. 2), the membership to a given module does not show a phylogenetic signal (Table 2), indicating that phylogenetically related plant species do not tend to belong to the same module. The convergence of results considering the phylogenetic signal in the plant-AMF interaction and phylogenetic conservatism of a module membership strengthens the conclusion that AMF phylogeny within Glomeraceae influence their pattern of interaction with plant species but this is not the case in plants.

The strength of plant-plant facilitation interactions was significantly positively correlated with dissimilarity in their fungal-niche. Pairs of plant species in which facilitation during their adult stage was recorded more frequently, tended to differ in their plant-fungal niche (r range for cut-offs 4% to 10% = 0.43-0-53 for quantitative interaction matrices; r range = 0.47-0-53 for binary interaction matrices; p<0.01 for every correlations) (Fig. 3). The null matrix was the worst predictor of the observed strength of plant-plant facilitation matrix (Table 3). Plant relative abundance alone was a better predictor of strength of plant-plant facilitation than dissimilarity in plant-fungal-niche abundance and dissimilarity in plant-fungal-niche probabilities. The combined effect of plant relative abundance and dissimilarity in plant-fungal-niche was

significantly better than abundance alone at every cut-off when quantitative plant-fungal matrices were considered and also when binary interaction matrices were used except for the cut-off of 10% (Table 3). The combined matrix represents the interaction probabilities expected if the species interact proportionally to both their relative abundance and dissimilarity in plant-fungal-niche. Although this combined matrix was the best predictor, it should be noted that much variation still remains unexplained as the differences in AIC's compared to the observed model suggests indicating that other factors beside these two are contributing to the strength of plant-plant facilitation.

Discussion

Our results show that closely related AMF OTUs tend to interact with the same set of plant species while the similitude in plant fungal-niches of two plant species is independent of their phylogenetic relatedness. We further show that although facilitation is more frequent among the most abundant plant species, this trend is significantly modulated by plant species fungal-niche. There is a tendency of plant-plant facilitation specificity to occur among plant species that differ in their fungal-niche, resulting in stronger facilitation between pairs of plant species with different AMF associated. We argue below that this might be a potential mechanism to increase AMF diversity in the shared rhizosphere which, by means of complementary beneficial effects of eachsingle AMF, can provide a more efficient exploitation of soil nutrients.

Previous studies have approached conservatism in plant-fungal interactions, considering mainly ectomycorrhizal associations within a particular plant phylogenetic clade, orchids (Shefferson *et al.*, 2007; Shefferson *et al.*, 2010; Jacquemyn *et al.*, 2011; Martos *et al.*, 2012). In this context, closely related plant species tend to interact with the same fungi, but closely related fungal species either do not share the same plant hosts (Jacquemyn *et al.*, 2011) or their phylogenetic signal is weaker than plant phylogenetic signal (Martos *et al.*, 2012). Although our results seem to challenge these previous results, it is important to remark that these studies are focused on a particular plant phylogenetic clade considering species that might not be co-occurring. Our study firstly approaches plant-fungal interactions at the community level, resulting in a wider range of plant phylogenetic diversity, due to the consideration of most of the coexisting plant species. In this framework, interestingly, closely related AMF tend to interact with the same plant species, and plant phylogenetic signal is not detected. This suggests that

although the pattern of interactions between plant and mycorrhizal fungi is evolutionarily conserved within a particular plant clade, this pattern does not scale when broader plant phylogenetic diversity is considered. This could be potentially due to convergent patterns of plant-fungal interaction across clades, but further studies with a community perspective, considering jointly a phylogenetic signal in the plant-AMF interactions will be required to confirm this hypothesis. Furthermore, this study in based on a group of fungal species within the genus Glomus. Although general plant-AMF interaction patterns described for this group of fungi can be generalized to a wider AMF phylogenetic diversity (Montesinos-Navarro et al. 2012), further studies considering the phylogenetic community interactions of other groups of AMF will shed light to the understanding of co-evolutionary patterns of plant and AMF. Nevertheless, the generality of host specificity in AMF remains speculative, with some plants showing repeatable AMF communities (e.g., Vandenkoornhuyse et al., 2002), and others showing variation with habitat and environmental conditions (e.g., Aldrich-Wolfe, 2007). In addition, the taxonomical delimitation of AMF species is controversial and AMF phylogenetic signal will be influenced by this limitation. Considering lower cutoffs to define OTUs can result in a phylogenetic signal either due to biological processes occurring at lower taxonomic levels or due to an artifact of considering intraspecific variation as different OTUs interacting with the same plant species. In a similar way, AMF phylogenetic signal can disappear at higher cut-offs that could potentially merge different genera or families within a given OTU, which might avoid the detection of plant-AMF specificity occurring at lower taxonomic levels. Our results report a decrease in AMF phylogenetic signal at the highest cut-off(10%) and a consistent significant phylogenetic signal in the rest of cut-offs. This suggests that the range of cutoffs considered might be covering a biological meaningful range of inter-specific variation.

It has been previously reported that phylogenetic conservatism in AMF traits can promote competition among closely related AMF species leading to a low contribution to plant biomass. Accordingly, plant performance decreases with the phylogenetic relatedness of the AMF species in their roots (Maherali & Klironomos, 2007). In our system, the two most generalist AMF -those interacting with almost all plant speciesare closely related, which according to Maherali & Klironomos (2007) could revert in fewer benefits to the plant. Species-specific patterns of plant-plant facilitation could compensate for this effect increasing AMF diversity in the shared rhizosphere. One possible mechanism may be by promoting associations with plant species which differ in their fungal-niche, potentially affecting ecosystem properties such as productivity (Cadotte *et al.*, 2008). In this study we present correlative evidence to support that plant-AMF interactions might be one of the underlying mechanisms influencing plantplant facilitation. Functional complementarity of AMF promoting plant productivity and plant-plant interactions have been so far reported for AMF belonging to different families (Hart & Reader, 2002; Maherali & Klironomos, 2007; Powell *et al.*, 2009). Our results firstly suggest that AMF phylogenetic diversity within the genus *Glomus* can also result in complementary functionality.

There is a tendency, supported by experimental (Castillo et al., 2010) and comparative evidence (Valiente-Banuet & Verdú, 2007) of plant-plant facilitation to occur among distantly related plant species. Interestingly, our results show that the similarity in the AMF associated between a pair of plant species is independent of the phylogenetic distance between them. Accordingly, the observed match between plantplant facilitation strength and fungal-niche cannot be attributed to a passive process in which distantly related plant species tend to differentiate in their plant-fungal niche. Furthermore, the combination of relative abundance and dissimilarity in plant-fungal niche substantially improves the prediction of facilitation specificity compared to solely plant species relative abundance. Taking all this together, it suggests that, among the possible plant-plant facilitation interactions (i.e. most abundant and distantly related species), the strength of facilitation increases when the involved plant species have a higher fungal-niche dissimilarity. Consequently, facilitation occurs also between distantly related plant species with similar plant-fungal niche but these interactions present weak facilitation strength. Specific plant-plant facilitation may be the result of facilitating species which differ in their overall fungal-niche, increasing AMF diversity and adding the beneficial effects of each AMF species (Van der Heijden et al., 1998; Hartnett & Wilson, 1999; Wagg et al., 2011). At the same time, plant species differing in their plant-fungal niche might be weaker resource competitors, and natural selection might positively select for these plant-plant interactions. Previous studies support the idea that both AMF host selectivity and plant fungal-niches can influence the emergent pattern of species-specificity in plant-AMF interaction, potentially influencing plantplant facilitation. AMF community composition can be highly influenced by the initial establishment of certainplant species (Hausmann & Hawkes, 2010), and seedling success can be affected by the presence of established AMF networks (Kytoviita et

al., 2003, Van del Heijden, 2004). Alternatively to plant-AMF interactions, other mechanisms can be underlying the observed correlation between plant facilitation strength and plant fungal niche. For example, an increase in AMF diversity in the rhizosphere can decrease the presence of plant pathogens (Van der Putten, 2009) resulting in more complex indirect effects underlying plant community assemblages.

Although our results are concordant with previous information on speciesspecificity in plant facilitation, experimental studies will be required to ultimately test for the specific biotic belowground mechanisms underlying plant-plant facilitation. Novel research lines are derived from our results exploring the potential implications of AMF networks in structuring plant community assemblages. Plant and AMF can regulate the resource allocation to the partner depending on the benefit received (Kiers *et al.*, 2011). If specific plant-plant facilitation allows a more efficient nutrient uptake (Van der Heijden *et al.*, 1998; Hartnett & Wilson, 1999; Wagg *et al.*, 2011) through an increase in AMF diversity, plants may increase their contribution to the plant-AMF mutualism resulting in higher resource allocation from AMF to the plants sharing a specific rhizosphere. Exploring the role of plant-AMF interaction as a potential mechanism promoting plant-plant facilitation specificity will contribute to a better understanding of assembly rules in plant communities.

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Tables

Table 1. Phylogenetic signal of the plant-AMF interaction using quantitative and binary matrices. Mean squared error calculated for the full model (MSE_d), a "star" phylogeny (MSE_{star}), and a Brownian evolution model (MSE_b). Strength of phylogenetic signal (d) for both plant (d_{plant}) and AMF (d_{AMF}) and the lower and upper values estimates of the confidence interval at 95% (in brackets) are shown.

			Quantitative			
Cut-off %	MSE _d	MSE _{Star}	MSE _{Brownian}	d _{AMF}	d _{plant}	
4	0.19 0.22 0.37	0.22	0.27	0.48[0.21-	0[0-0.05]	
4		0.74]	0[0-0.05]			
5	0.17	0.18	0.35	0.19[0.002-	0[0-0.01]	
				0.38]	0[0-0.01]	
6	0.16 0.17 0.33	0.17	0.22	0.44[0.13-	0[0-0.14]	
0		0.55	0.82]	0[0-0.14]		
7	0.12	0.14	0.28	0.49[0.12-	0[0-0.14]	
7	0.12	0.14	0.28	0.92]	0[0-0.14]	
8	0.12	0.15	0.31	0.57[0.1-1]	0[0-0.1]	
9	0.144 0.138 0.51	0.51	0.58[0.05-	0[0-0.16]		
)	0.144 0.158 0.51		0.31	1.22]	0[0-0.10]	
10	0.12	0.12	0.42	0.006[0-0.44]	0[0-0.34]	
			Binary			
4	0.18	0.21	0.35	0.46[0.20-	0[0-0.05]	
т	0.10	0.21	0.55	0.74]	0[0 0.05]	
5	0.16	0.17	0.34	0.18[0.002-	0[0-0.06]	
5	0.10	0.17	0.34	0.36]	0[0 0.00]	
6	0.15	0.16	0.31	0.42[0.09-	0[0-0.09]	
0	0.15	0.10	0.31	0.78]	0[0-0.07]	
7	0.09	0.10	0.20	0.49[0.06-	0[0-0.04]	
				0.94]	0[0-0.04]	
8	0.11	0.14 0.29		0.55[0.08-	010 0 201	
0	0.11	0.14	0.27	1.04]	0[0-0.20]	
9	0.12	0.12	0.33	0.17[0-0.61]	0[0-0.20]	

Table 2. Phylogenetic signal of the module membership for cut-offs of 4 to 10% genetic dissimilarity in DNA sequences. For both AMF and plants, it is presented: the number of modules detected at each cut-off, the number of observed evolutionary transitions in the module membership, the expected number of transitions under a null model in which data were reshuffled 1000 times across the tips of the phylogeny, and the *P*-value based on the comparison of observed vs. expected values (***: <0.001;**: 0.01; ns: >0.05).

		AMF		Plant	
Cut-off %	N Module s	Obs transition s	Mean Null (p-value)	Obs transitions	Mean Null (p-value)
4	6	18	27***	19	19 ^{ns}
5	8	14	22***	21	22 ^{ns}
6	7	12	19***	22	22^{ns}
7	6	10	15***	17	16 ^{ns}
8	6	11	15***	17	16 ^{ns}
9	6	10	13**	18	17 ^{ns}
10	5	8	9 ^{ns}	16	15 ^{ns}

Table 3. The effect of plant abundance, dissimilarity in fungal niche and their interaction on plant facilitation using quantitative and binary fungal niche matrices. Matrices were log transformed and the likelihood and Akaike information criteria is presented for: a) an observed model, using the same matrix as a predictor; b) a null model in which all pairwise interactions were equally probable; c) interaction probability determined solely by relative species abundance or d) solely fungal niche, for each cut-off, and d) the interaction of abundance and fungal niche matrices, for each cut-off. The number of parameters of each model is presented in parenthesis. The AIC weights of each candidate model with respect to the best fit model is lower than 0.05 except for abundance at the cut-off 10% (AIC weight = 0.31).

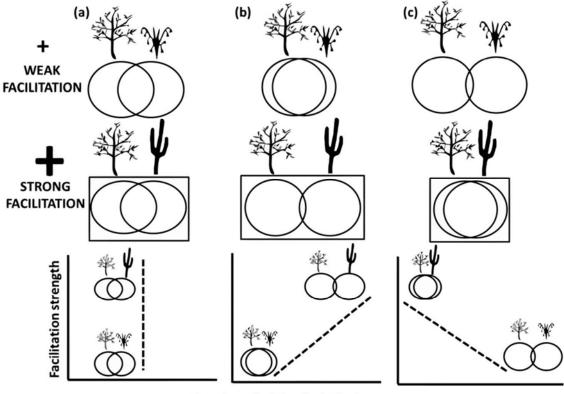
Likelihood AIC

a) Observed	l (1) 45.72	93.44				
b) Null (1)	169.20	340.41				
c) Abundan	ce (1) 139.69	281.37				
Quantitative						
	d) Plant-fungal	niche	e) Abundance x Plant -fungal			
	(1)		niche (2)			
Cut-off %			Cut-off			
	Likelihood	AIC	% Likelihood AIC			
4	150.86	303.73	4 126.51 257.02			
5	150.43	302.86	5 126.15 256.31			
6	149.86	301.73	6 125.73 255.47			
7	145.45	292.91	7 122.55 249.09			
8	145.47	292.94	8 122.56 249.11			
9	145.40	292.79	9 122.50 249.00			
10	140.77	283.54	10 120.61 245.22			
Binary						
			e) Abundance x Plant-fungal			
d) Plant-fungal niche (1)			niche (2)			
Cut-off %			Cut-off			
	Likelihood	AIC	% Likelihood AIC			
4	147.19	296.38	4 122.74 249.47			
5	147.74	297.49	5 123.26 250.51			
6	144.17	290.34	6 120.98 245.97			
7	148.43	298.87	7 128.53 261.07			
8	149.10	300.21	8 129.10 262.20			
9	146.06	294.12	9 127.39 258.77			
10	191.92	385.84	10 176.33 356.66			

Figures

Figure 1. Schematic representation of the potential relationships between plant-plant facilitation strength and other biotic interactions. Plant fungal-niche dissimilarity represents the differences in the set of AMF associated to any pair of plant species. The size of the positive sign indicates the strength of facilitation for a given pair of plant species. Circles represent plant fungal-niche, with high and low overlap indicating

similarity or difference in AMF communities respectively. The rectangle around circles represents the potential exploitation of soil nutrients. Different scenarios are represented (a) Members of plant pairs with different degree of facilitation strength have a similar plant fungal-niche, leading to an independence of plant-plant facilitation strength from this biotic interaction (plant-AMF association), (b) Higher facilitation strength between plant pairs with lower dissimilarity in plant fungal-niche (negative relationship), leading to a higher facilitation strength between plant-plant sharing a similar plant-fungal niche, and (c) Higher facilitation strength between plant pairs with higher dissimilarity in their plant fungal-niche (positive relationship), leading to a higher facilitation strength between plant pairs with higher dissimilarity in their access to resources (rectangle).



Plant fungal-niche dissimilarity

Figure 2. Plant-AMF interaction matrix combined with the phylogenetic topologies, using a cut-off of 7% for AMF. Black and white cells indicate presence and absence of the interaction. The six types of icons (open and close circles, diamonds and squares) on the tips represent the module to which each species belongs as described in Montesinos-Navarro *et al.* 2012.

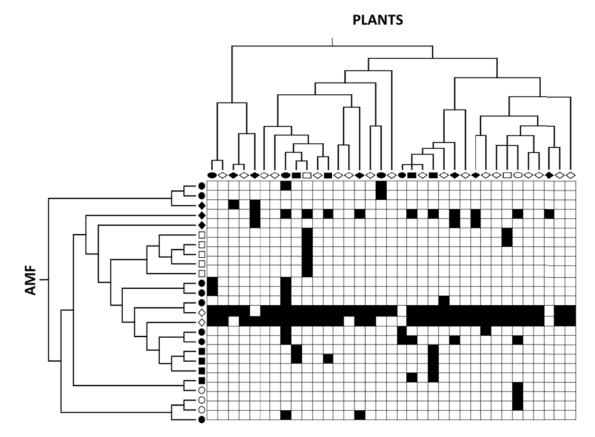
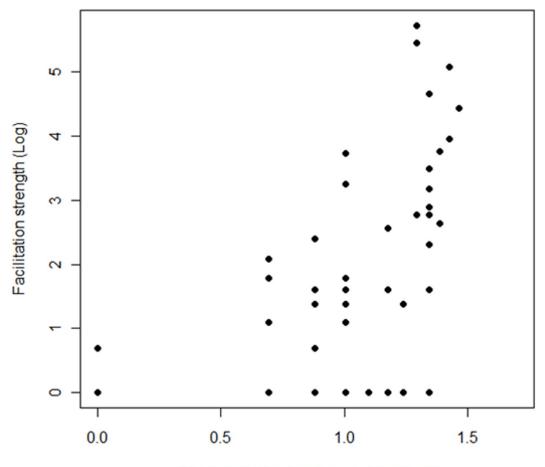


Figure 3. Relationship between strength of plant-plant among pairwise plant species and their dissimilarity regarding the AMF with which they interact. Data are presented for the representative cut-off of 7% of genetic dissimilarity to define AMF OTUS, after log transformation of plant-plant facilitation and dissimilarity plant-fungal-niche matrices. The strength of facilitation among each pair of plant species was estimated as the number of adults on each facilitated-species recorded under each nurse plant species (Verdú *et al.* 2010) and dissimilarity in plant fungal- niche was calculated based on the composition of AMF interacting with each plant species. The correlation coefficient between matrices was calculated against a null model randomizing plant-fungal-niche dissimilarities among plant species (r = 0.48; P < 0.001).



Dissimilarity in plant-fungal-niche (Log)

SUPPLEMENTARY MATERIAL

Table S1. Genebank accession numbers of all sequences used in the analyses.

Figure. S1. The best-scoring Maximum Likelihood arbuscular mycorrhizal fungi tree inferred from one hundred boostrap trees.

Figure. S2. Phylogeny of the plant species based on the megatree of the Angiosperm Phylogeny Group 3 (Stevens 2005) as the backbone phylogeny.