

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19

**Wine features related to safety and consumer health: an integrated  
perspective**

M. Ángeles Pozo-Bayón, María Monagas, Begoña Bartolomé,

M. Victoria Moreno-Arribas\*

Instituto de Fermentaciones Industriales (CSIC). Juan de la Cierva, 3. 28006 Madrid,

Spain

Tel. (34) 915622900

Fax: (34) 915644853

e-mail: [mvmoreno@ifi.csic.es](mailto:mvmoreno@ifi.csic.es)

*This work is dedicated to Prof. M.Carmen Polo, one of the main supporters of the  
research in Oenology at the Institute of Industrial Fermentations (CSIC), and whose  
guidance is greatly valuable in our daily work.*

1 *This review presents a global view of the current situation of the scientific knowledge*  
2 *about aspects of wine with possible repercussions (positive or negative) on consumer*  
3 *health and wine safety. The presence in wine of some potential harmful compounds such*  
4 *as phytosanitary products, trace metal compounds, sulfites and some toxics from*  
5 *microbial origin, such as ochratoxin A, ethyl carbamate and biogenic amines, is*  
6 *discussed. The different strategies and alternative methodologies that are being carried*  
7 *out to reduce or to avoid the presence of these substances in wines are also discussed.*  
8 *In recent years many work has been focused on establishing the scientific explanations*  
9 *for the positive biological effects of some wine compounds. In this review, we also*  
10 *examine the latest knowledge regarding wine and health, focusing in two types of*  
11 *compounds that have been related to the positive effects of moderate wine consumption*  
12 *such as phenolic compounds and bioactive peptides.*

13  
14

15 **Keywords:** Wine, Phytosanitary products, Trace metal compounds, Sulfur dioxide,  
16 Ochratoxin A, Ethyl carbamate, Biogenic amines, Phenolic compounds, Peptides

17

## 1 ***INTRODUCTION***

2

3           The intense changes in society's life style over the past few years have had  
4 significant effects on the basic concepts of our eating habits. Nowadays, we can go  
5 further than the traditional idea of what should be a 'suitable diet' in the sense that it  
6 should provide the sufficient nutrients to ensure individuals' survival, satisfy metabolic  
7 requirements and pleurably appease the sensation of hunger and wellbeing. Today, it  
8 is also necessary to comply with aspects of health and safety, emphasizing the potential  
9 of foods from a health perspective, to increase wellbeing and to reduce the risk of  
10 illnesses.

11           In an attempt to guarantee food safety for the consumer, several organizations  
12 such as the Food and Agriculture Organisation (FAO) of the United Nations and the  
13 World Health Organisation (WHO), among others, assess the health quality of foods in  
14 general, and also of wine. However, for aspects related to wine safety there is also  
15 another authority, the International Organisation of Vine and Wine (OIV), which,  
16 among other objectives, helps to protect consumer's health, improving and guiding  
17 research on health aspects and spreading out the results of this research to healthcare  
18 professionals. Within this Organisation, which encloses several Commissions and  
19 Subcomissions, the Safety and Health Commission, is included. This group of experts is  
20 mainly focused on food safety, wine consumption, nutrition and health.

21           In recent years, the topic 'wine and health' has aroused much interest but also  
22 considerable controversy. This is support by the large number of studies and scientific  
23 contributions that has been carried out within this area. In this review, we have  
24 summarised the main contributions of current knowledge about aspects of wine and its  
25 components that could affect safety and consumer's health. We do not specially focus

1 on alcohol, a wine ingredient with evident and well-known toxic effects, but instead on  
2 wine as a food, that can have beneficial effects on the health of healthy adult consumers  
3 when it is drunk in moderation. Nevertheless, wine may contain certain compounds that  
4 decrease the wholesomeness of wine such as chemical preservatives (sulphites),  
5 neurotoxins (ochratoxin A), potential carcinogens (ethyl carbamate) and allergens  
6 (biogenic amines), amongst others. Therefore, besides the need to examine the main  
7 causes of wine spoilage that may affect human health, this article also discusses the  
8 strategies followed to eliminate or minimise wine contamination. The latest studies  
9 about those compounds involved in positive health aspects of wine are also reviewed.

10

11 ***COMPOUNDS PRESENT IN WINE WITH POTENTIALLY NEGATIVE HEALTH***  
12 ***EFFECTS. METHODS FOR THEIR CONTROL AND PREVENTION***

13

14 Over the past few years, numerous research studies have focused on establishing  
15 the components of wine that can negatively affect the health of moderate wine drinkers,  
16 with the ultimate objective of searching for solutions to decrease or to avoid their  
17 presence in wines. The compounds that can damage wine's health quality include those  
18 derived from the grape, which can appear as contaminants (i.e. pesticides and other  
19 phytosanitary products, trace metal compounds, additives such as SO<sub>2</sub>, which is added  
20 during different steps of the wine production process), and those produced by different  
21 microorganisms during winemaking including, ochratoxin A, ethyl carbamate and  
22 biogenic amines.

23

24

25

## 1 ***Phytosanitary products***

2           Nowadays, a wide range of insecticides, fungicides and herbicides are used in  
3 grape production. In particular, grapes are especially vulnerable to moulds, which leads  
4 to the widespread use of antifungal agents. In Mediterranean countries, the principal  
5 parasites of vine are the grape moth (*Lobesia botrana*), downy mildew (*Plasmopora*  
6 *viticola*), powdery mildew (*Uncinula necator*) and gray mold (*Botrytis cinerea*). The  
7 latter, if conditions are appropriate, may attack bunches of grapes damaging the grape,  
8 by synthesizing the enzymes required to breakdown the cell walls of the skin and  
9 penetrate in the flesh. To control these parasites, insecticides and fungicides are  
10 commonly used. In practise, to tackle this and other alterations, it is much more  
11 effective to use preventive treatments in order to maintain the vine whole and improve  
12 the quality of the grape. Although the correct use of pesticides does not cause  
13 environmental problems or public health concerns, if an inappropriate and abusive  
14 pesticide treatment is carried out, undesirable residues may remain on the grapes after  
15 harvest. As a result, they may be transferred into the wine. A more detailed review in  
16 this area was published by Cabras and Angioni (2000).

17           With the aim to improve hygienic and sanitary characteristics of wines, the  
18 effect of some wine-making practices on the concentration and disappearance of  
19 pesticide residues during wine production has been evaluated (Cabras et al., 1995,  
20 Navarro et al., 1997, Soleas and Goldberg, 2000). Recently, some clarification and  
21 filtration processes have proved to be effective against four commonly used fungicides  
22 (cypronidil, fludioxonil, pyrimethanil, and quinoxifen) for the control of cryotogramic  
23 vineyard diseases (Fernández et al., 2005). The most common clarifying substances for  
24 wine are bentonite, charcoal, gelatin, polyvinylpyrrolidone, potassium caseinate, and  
25 colloidal silicon dioxide. While most of them show limited ability to decrease pesticide

1 residues, charcoal is especially recommended when the amount of residues is low,  
2 allowing the complete elimination of most pesticides. Its effectiveness generally  
3 decreases as the pesticide water solubility increases; therefore, pesticides highly soluble  
4 in water, such as dichlorvos and dimethoate, do not show appreciable changes (Cabras  
5 et al., 1995).

6         The activity of yeast and lactic acid bacteria involved in alcoholic and malolactic  
7 fermentation respectively, can be affected by the presence of pesticide residues in musts  
8 and wines (Cabras and Angioni, 2000). Nevertheless, while it has been shown that some  
9 wine yeasts are able to reduce the pesticide content, either by degradation or by  
10 adsorption (Cabras and Angioni, 2000), lactic acid bacteria do not possess this ability.

11         Although maximum residue limits for most pesticides in wine have not been set,  
12 several countries have established guidelines for the correct use of pesticides and for the  
13 maximum residual limits for the treatment of vines and grapes used in winemaking  
14 (Flamini and Panighel, 2006). Because the vinification process decreases the levels of  
15 pesticides, their content in wines is significantly lower than in grapes. Therefore,  
16 methods to detect pesticide residues must be very precise and sensitive. Pesticides have  
17 been analyzed in wines mostly by gas chromatography (GC) (Jiménez et al., 2001,  
18 Hyötyläinen et al., 2004) and high-performance liquid chromatography (HPLC) (Nozal  
19 et al., 2005; Braga et al., 2007) and more recently, by capillary electrophoresis (CE)  
20 (Molina-Mayo et al., 2006). Mass spectrometry represents an advantageous alternative  
21 for the simultaneous determination of different pesticides in wine. Recently, an  
22 interesting review about the applications of mass spectrometry for the determination of  
23 pesticides and other contaminants in grapes and wines has been reported (Flamini and  
24 Panighel, 2006).

1           One aspect that deserves special attention is that phytosanitary treatments could  
2 be reduced if the plants own defence mechanisms come into play. Phytoalexins are  
3 antimicrobial compounds produced by plants, such as grapevine, in response to biotic  
4 and abiotic stress factors and have received a lot of attention over the last decade  
5 (Jeandet et al., 1997; Jeandet et al., 2002). These compounds can be released from  
6 plants as a response to infections by pathogens and hinder their development. They are  
7 often synthesized within a few hours after stress exposure. They accumulate in the  
8 plant reaching a maximum content between 2 and 3 days after the induction. Therefore,  
9 stimulating the production of grapevine phytoalexins could be a strategy to limit the use  
10 of pesticides in vineyards. In the *Vitaceae* family, phytoalexins have been well  
11 characterized. They constitute a rather restricted group of molecules belonging to the  
12 stilbene family (for a review see Jeandet et al., 2002). Resveratrol (3,5,4'-  
13 trihydroxystilbene) is the major phytoalexin produced in grapevine as a general  
14 response to different agents: fungal attack, leaf UV-C irradiation, heavy metal exposure,  
15 and ozone treatments. Other biosynthetically related compounds that have been found in  
16 grapevine as a result of stress conditions are viniferins and pterostilbene (3,5-  
17 dimethoxy-4'-hydroxy stilbene) (Jeandet et al., 1997). A novel approach in the actual  
18 research on phytoalexins is the development of synthetic phytoalexins, similar to the  
19 natural ones, that can be used for plant disease control. Stilbene phytoalexines are  
20 formed on the phenylalanine/polymalonate pathway and the last step in their  
21 biosynthesis is catalyzed by the enzyme stilbene synthase (STS) (Jeandet et al., 2002).  
22 Therefore, another valuable scientific strategy that is being performed is focused on the  
23 transfer of STS gene from grapevine to numerous plants, with the objective of  
24 increasing their tolerance to pathogenic microorganisms. Following this strategy, it has  
25 been possible to enhance the resistance of grapevine rootstock 41B to the infection of

1 *Botrytis cinerea*. In addition to plants, the gene has been successfully transferred to  
2 yeast and mammalian cells (Zhang et al., 2006).

3

#### 4 ***Trace metal compounds***

5

6         Although trace elements are essential for humans, many of them (e.g. As, Sb, Bi,  
7 Pb, Sn, Se, Te) have unknown biological function and are regarded as toxic because  
8 they accumulate in the human organism. In fact, at slightly higher levels than when  
9 found in living beings, they can have toxic effects (Baluja-Santos and Gonzalez Portal,  
10 1992).

11         It is well known that some trace elements play a relevant role in winemaking, for  
12 example Zn is essential at low concentrations, for the correct development of alcoholic  
13 fermentation, while Cu, Fe, and Mn have organoleptic effects at increased levels,  
14 contributing to the haze and taste of wines (Riganakos and Velssitas, 2003). Excessive  
15 amounts of iron (10-20 mg/L, or more) can cause stabilization problems in wines, since  
16 it may be oxidized to a ferric form causing precipitation of pigmented materials ('Blue  
17 haze') or with orthophosphate ions ('White haze').

18         Nevertheless, as Ajtony et al., (2008) have stated, the concern about human  
19 exposure to trace metals in beverages and food products, including wines, has started to  
20 attract considerable attention, since the consumption of wines, especially in fairly large  
21 volumes, may significantly contribute to the daily dietary trace element intake by  
22 humans.

23         The primary natural mineral content of wines is determined by many factors such  
24 as type of soil, variety of grape, weather conditions, pollution and viticultural practices.  
25 In fact, the regionally varying trace metal content of wines can also be used for source

1 identification purposes (Pyrzynska, 2004). The secondary mineral contamination is  
2 derived from the use of pesticides and fertilizers or acquired during winemaking (in  
3 spite of the overall reduction in mineral content during this process) (Baluja-Santos and  
4 Gonzalez Portal, 1992). It has been shown for example, that while the origin of Cu in  
5 wines is associated with copper-based vineyard sprays, the content of As, Cd, and Pb  
6 reflects the differences in grape variety, environmental factors (e.g., soil, climate), and  
7 winemaking technology (Pyrzynska, 2004).

8 Sodium arsenite is employed in viticulture as fungicide against a grapevine  
9 necrotic disease known as *Eutypa dieback*, caused by the fungus *Eutypa lata*. Arsenic is  
10 a suspected carcinogen and little is known about its chronic sub-lethal effects. The  
11 concentration of As in wines mainly depends on factors such as soil composition, grape  
12 variety, climatic conditions, use of pesticides, winemaking technology and storage  
13 conditions (Flamini and Panighel, 2005). The OIV has established the maximum limit  
14 of As in wine at 200 pg/ $\mu$ l, but in general, the presence of only a few pg/ $\mu$ l of As in  
15 uncontaminated wines is accepted (Baluja-Santos and Gonzalez Portal, 1992; Pedersen  
16 et al., 1994; Bruno et al., 1994).

17 In the case of lead, its presence in wine could be the consequence of the airborne  
18 particulate matter of atmosphere on grapes, and/or by the grapevine intakes from ground  
19 water and soil. The metal may also be released from bronze tanks, taps, pumps and  
20 tubing containers use in winemaking (Flamini and Panighel, 2005). Lead adversely  
21 affects multiple enzyme systems of the body, as any ligand with sulfhydryl groups can  
22 be vulnerable. The best-known effect is that of the production of heme (Bornet and  
23 Teissedre, 2008) . Detection of lead in wine is usually focused on the determination of  
24 the total lead  $Pb^{2+}$ . In wine the structurally complex pectic polysaccharide  
25 rhamnogalacturonan II (RG-II), an anionic biomolecule released from the grape during

1 winemaking, can form coordination complexes with specific cations such as lead  
2 (Flamini and Panighel, 2005). This pectic polysaccharide could reduce intestinal  
3 absorption and tissue retention of Pb as has been reported in rats, with a potential  
4 beneficial effect by minimising toxicity (Tahiri et al., 2000; Tairi et al., 2002).

5         The toxic effects of cadmium are due to the inactivation of enzymes containing  
6 sulphhydryl groups and the uncoupling of oxidative phosphorylation in mitochondria.  
7 Cadmium may also compete with other metals such as zinc and selenium for inclusion  
8 into metallo-enzymes and it may compete with calcium for binding sites on regulatory  
9 proteins such as calmodulin (Bornet and Teissedre, 2008). The OIV establishes the  
10 maximum concentrations of lead and cadmium in wine at 150 µg/L and 10 µg/L,  
11 respectively.

12         Taking into consideration the importance of these elements regarding their  
13 potential toxicity and technological features, it is worth noticing the great amount of  
14 scientific work that has been focused in developing adequate analytical tools that can  
15 offer enough selectivity, sensitivity and robustness for the analysis of these compounds  
16 in wines. As consequence, there are many analytical techniques for quantification trace  
17 metal in wines, such as ion chromatography (IC), stripping potentiometry, inductively  
18 coupled plasma mass spectrometry (ICP-MS), inductively coupled plasma atomic  
19 emission spectrometry (ICP-AES), flame atomic absorption spectrometry and graphite  
20 furnace atomic absorption spectrometry (GFAAS). All of them have been extensively  
21 reviewed elsewhere (Pyrzynska, 2004; Flamini and Panighel, 2006; Ajtony et al., 2008).

22         The multiple sources involved in the content of metal trace elements in wines  
23 make difficult the establishment of appropriate strategies to prevent or to avoid their  
24 occurrence in wines. Obviously, an adequate soil for each grape variety and  
25 environmental conditions, the regulation in the use of fertilisers, fungicides and others

1 pesticides or an adequate winemaking equipment could greatly reduce the amount of  
2 these elements in wines. Recently, La Pera et al., (2008) in a study performed over two  
3 vintages (2003, 2004) have shown that treatments with the fungicides mancozeb,  
4 zoxamine and copper oxychloride significantly increased the levels of Mn, Zn, Cu, Pb  
5 and Cd of wines, grapes, marcs and grape stalks.

6 Some strategies that could be effective for the control of trace heavy metals  
7 imply the use of some polymers from natural sources. Chitin, chitosan and other  
8 derivatives are non-toxic biodegradable polymers that can remove metals and organic  
9 contaminants from food products (Bornet and Teissedre, 2005). Chitin exists widely in  
10 the cell walls of fungi, moulds and yeasts but also in the exoskeletons of some  
11 invertebrates such as molluscs, insects, etc. Chitosan is found only in a few species of  
12 fungi. Chitin and chitosan consist of 2-acetamido-2-deoxy- $\beta$ -D-glucose and 2-amido-2-  
13 deoxy-  $\beta$ -D-glucose as repeating units, respectively. Bornet and Teissedre (2008)  
14 showed that the addition of chitosan, chitin, chitin-glucan and chitin-glucan hydrolysate  
15 of fungal origin (until 2 g/L) to wine, can reduce the amount of Fe by 32-91%, Cd by  
16 11-57% , and Pb by 33-84 % , depending on the type of wine.

17 Furthermore, based on the parietal adsorption activity of yeasts (Caridi, 2007),  
18 the use of inactive yeast or yeast derivatives (walls, hulls, etc.) in wines, could also be  
19 considered in the future as a control mechanism for trace metal compounds to improve  
20 wine safety.

21

## 22 *Sulphur dioxide*

23

24 One compound that should be considered in relation to the health quality of  
25 wines, is sulfur dioxide (SO<sub>2</sub>). Because of the abundance of sugars, nitrogen

1 compounds, vitamins, mineral and salts, grape must is an ideal medium for microbial  
2 growth. SO<sub>2</sub> has been used since the times of Romans for disinfection and cleaning of  
3 wine cellars. Although its use as preservative in wines has been known for centuries, it  
4 has been only recently, when its use in prefermentation operations during winemaking  
5 has been spreading out. Most of the scientific knowledge about the use of sulphur  
6 dioxide in oenology has been acquired over the last decades. In wines, this compound  
7 has multiple and diverse properties. For example, it is an *antiseptic* agent against yeasts  
8 and bacteria. The *antioxidant* properties, are well established since it consumes oxygen  
9 and acts against non-enzymatic and enzymatic oxidation of wines. Moreover it behaves  
10 as a *solvent*, activating the extraction from the solid parts of the grape, (skins, seeds,  
11 stems). Finally, since it favours the static settling of wines, it can be considering as a  
12 *clarification agent*. These and other properties, make this compound widely used during  
13 the different steps of winemaking and storage.

14 During winemaking, SO<sub>2</sub> is mainly used on three occasions. Firstly, in the  
15 grapes or must during the prefermentation step, with the prime objective of preventing  
16 its oxidation; later, to inhibit microbial growth that can alter the wines, once the  
17 fermentation processes have finished and before the ageing or storage steps, and,  
18 finally, just before bottling, to stabilise the wines and prevent any alteration or accident  
19 in the bottles.

20 Today, the addition of SO<sub>2</sub>, is therefore, an essential treatment in winemaking  
21 technology. However, the use of this additive is strictly controlled, since high doses can  
22 cause organoleptic alterations in the final product (undesirable aromas from sulphurous  
23 gas or from the reduction products, hydrosulphate and mercaptanes) and, especially,  
24 owing to the risks for human health derived of its consumption.

1           Various studies have revealed that besides their multiple properties, sulphites  
2 may induce relevant adverse effects after their ingestion (Taylor et al., 1986). Moreover,  
3 it has been shown that considerable percentage of consumers, show intolerance or high  
4 sensitivity to sulphites. This risk is higher for asthmatics and children. It is estimated  
5 that almost 1% of the population and 5% of the asthmatics are concerned. Although  
6 sulphites do not cause a true allergic reaction, sulphite-sensitive people may experience  
7 some similar responses. The symptoms may develop quickly and can include flushed  
8 face; rash, red and itchy skin; headaches; eye, face, lip or throat swelling; difficulty for  
9 breathing, speaking or swallowing; anxiety, distress, faintness, weakness; cramps,  
10 diarrhoea, vomiting, among others.

11           The upper limit of sulfites permitted by the OIV ranges from 150 to 400 mg/l of  
12 total SO<sub>2</sub>, depending on the type of wine and its content in reducing matter. A first step  
13 to increase the food safety has been done by the EU through the approval of the  
14 legislation for regulation the use of sulphites as preservatives. Since then, directives  
15 2000/13/EC, 2003/89/EC and 2007/68/EC request the systematic labelling of allergens  
16 or similar (annexe III bis from directive 2000/13/EC) incorporated in food products. In  
17 the same way, since November 25<sup>th</sup> of 2005, is mandatory to declare the presence of  
18 sulphites in wines and other foodstuffs when the concentration exceeds 10 mg/L or 10  
19 mg/kg. In addition, directives 95/2/EC and 2006/52/EC precise the maximum sulphite  
20 concentration allowed in different foodstuffs (included wine) within the EU.

21           These legislative rules, together with consumer demands, have attracted the  
22 interest of scientific research on the study of other wine preservatives, harmless to  
23 health that can replace or reduce the amount of SO<sub>2</sub> in wines. One example is the  
24 production of 'ecological wines', which in recent years have gained considerable  
25 interest. They are produced by environmentally friendly methods, from integrated or

1 sustainable manufacture methods. During the production of these wines, only a series of  
2 strict oenological practises are permitted and no additive apart from SO<sub>2</sub> is possible.  
3 The final wine must have as low amount of sulphur dioxide as possible and must not,  
4 under any circumstance, exceed very strict limits (from 70 to 100 mg/l depending on the  
5 type of wine (EC Directive 2092/91).

6 Physical treatments, such as modified atmosphere packaging, control  
7 atmosphere storage, ozone, and other alternative non-conventional gas treatments have  
8 been applied to table grapes in order to prolong shelf life and storage, therefore reducing  
9 the SO<sub>2</sub> doses necessary in the harvest (Artés-Hernández et al., 2003; Artés-Hernández  
10 et al., 2006).

11 For wines, other methods for sulfites removal such as adsorption through the use  
12 of anion and cation exchangers (Brown, 1991) or the use of a membrane reactor in  
13 conjunction with non-diffusible oxidizing agents for sulfite oxidation (Seifter et al.,  
14 1994) have been tested. Although both methods can reduce the sulfite content of wines  
15 below 10 ppm, the lack of selectivity in their adsorption mechanisms can greatly  
16 decrease important flavor compounds negatively affecting wine quality. Another  
17 original alternative for reducing sulfite contents in wine is the use of wheatgrass  
18 (*Triticum aestivum*) chloroplasts for the oxidation of sulfites to innocuous sulfates. It  
19 has been shown that a crude chloroplasts preparation at a concentration as low as  
20 5mg/mL, is able to reduce sulfites in commercial white wines from 150 ppm to 7.5  
21 ppm, as well as up to 93 % of the initial sulfite content in red wines within 45 minutes  
22 (Lin and Georgiou, 2004). Although this simple bio-catalytic process seems to be highly  
23 effective, inexpensive and largely valuable for the winemaking industry, appropriate  
24 sensory analysis would be necessary to evaluate the quality of these wines.

1           The effects of electrochemical treatments and pulsed electric fields on the  
2 growth and metabolism of wine yeast have been studied (Ranalli et al., 2002) and  
3 applied for controlling grape must fermentation in winemaking (Lustrato et al., 2003).  
4 These treatments could be an alternative to the use of SO<sub>2</sub> since they are capable of  
5 destroying microorganisms and some enzymes while maintaining the characteristics of  
6 food products (Benicho et al., 2002). Nevertheless, more studies should be necessary to  
7 evaluate the potential use of these technologies in winemaking.

8           Regarding other chemical alternatives to the use of sulphites, the use of  
9 dimethyldicarbonate (DMDC) has been recently described. This compound, also known  
10 as E242, can inhibit the development of spoilage and fermentative yeasts, allowing to  
11 reduce the dose of SO<sub>2</sub> in some types of wines (Threlfall and Morris, 2002; Divol et al.,  
12 2005). It also acts on bacteria and mould development, inactivating some vital enzymes  
13 for these microorganisms, therefore causing their death. DMDC is also currently used  
14 for cold sterilization of different types of drinks.

15           Other alternatives focused on ‘natural antimicrobial agents’ have been developed  
16 to reduce the use of sulfite in wines. Among them, lysozyme (EC 3.2.1.17) has been  
17 recently introduced into the wine industry and offers important advantages for  
18 controlling malolactic fermentation in wine (Bartowsky, 2003; Pilatte, et al., 2000).  
19 Lysozyme (1,4-β-*N*-acetylmuramidase) has the ability to cleave the β-1,4-glycosidic  
20 bonds present in Gram-positive bacteria. It has no effect on yeasts, is not affected by  
21 alcohol and is active in the pH range of the winemaking process. As an organoleptically  
22 neutral alternative, lysozyme can be beneficial to further reduce SO<sub>2</sub> levels during  
23 winemaking. As wine pH increases, the antimicrobial activity of lysozyme increases as  
24 well, which makes it attractive for preventing spoilage in wines with high pH (Gao et  
25 al., 2002; Delfini et al., 2004). Despite lysozyme was firstly authorized as an additive in

1 winemaking due to the high costs of its application, its use has been largely reduced.  
2 Another aspect that deserve consideration, is the IgE-mediated immune reactions that  
3 lysozyme can cause in some individuals (Mine and Zhang, 2002). Therefore its presence  
4 in food products, including wine, can cause some health concern.

5         Currently, it is well known that other polypeptidic compounds, called  
6 bacteriocins, can also produce an inhibitory effect on the development of lactic acid  
7 bacteria. These products have received great attention in the manufacture of dairy  
8 products, since they are especially used as additives during cheese ripening (Martínez-  
9 Cuesta et al., 2003). Because of the lack of effects of bacteriocins on wine organoleptic  
10 properties and the absence of toxic effects, some studies to evaluate their use in the  
11 control of malolactic fermentation during winemaking have been conducted (Navarro et  
12 al., 2000; Bauer et al., 2003, 2005). Generally, bacteriocins can be introduced into the  
13 wine by direct addition of the peptide (normally in its purified form), by adding a  
14 supernatant of a bacterial culture that contains a crude extract of the peptide, or by  
15 inoculation of a starter culture of lactic acid bacteria. It has also been shown that after  
16 application of gene technology to transform laboratory strains of *S. cerevisiae* with the  
17 pediocin and leuocin structural genes, *pedA* and *lcaB* respectively, the transformed  
18 yeasts were able to produce biologically active bacteriocins, although in low levels  
19 (Schoeman et al., 1999). Despite the use of bacteriocins can be appropriate for  
20 inhibiting the growth of spoilage bacteria in wines, up to date, nisin is the only  
21 bacteriocin that can be obtained commercially. Although it has been shown to be  
22 effective (Radler et al., 1990a,b; Rojo-Bezares et al., 2007), its use has not been  
23 authorized in winemaking.

24         Another strategy that has been proposed to reduce the use of SO<sub>2</sub> in oenology, is  
25 by using antimicrobial metabolites such as hydrogen peroxide (Du Toit et al., 2000).

1 Glucose oxidase (GOX) from *Aspergillus niger*, which has GRAS status, is of  
2 considerable industrial importance. GOX metabolises glucose into gluconic acid and the  
3 antimicrobial compound that results from this conversion is hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>).  
4 H<sub>2</sub>O<sub>2</sub> has been shown to be active against both Gram-positive and Gram-negative  
5 bacteria. The *gox* gene was expressed in *S. cerevisiae* and the GOX-producing yeast  
6 transformant inhibited the growth of both acetic and lactic acid bacteria (Malherbe et  
7 al., 2003).

8         Currently there are increasing evidences of the potential application of some  
9 eukaryotic antimicrobial proteins and peptides as food preservatives (Rydlo et al.,  
10 2006). Among others, antimicrobial peptides derived from food proteins present clear  
11 advantages to be used in food preservation (Pellegrini, 2003). Milk is a particularly  
12 interesting source of antimicrobial peptides that can be released after digestion with  
13 protease. Among them, lactoferrin (LF) is a multifunctional iron glycoprotein that  
14 exhibits a diverse range of biological effects, including antimicrobial, antiviral,  
15 antioxidant, and immunomodulatory activities (Tomita et al., 2002). Recently, its  
16 antimicrobial properties against phytopathogen molds has also been described (Muñoz  
17 and Marcos, 2006). Several strategies based on the use of hydrolized LF have been  
18 also investigated to control microbial growth of *Saccharomyces cerevisiae* and other  
19 spoilage non *Saccharomyces* wine yeasts (*Cryptococcus albidus*, *Dekkera bruxellensis*,  
20 *Pichia membranifaciens*, *Zygosaccharomyces bailii* and *Zygosaccharomyces bisporu*)  
21 (Enrique et al., 2007). Further studies would be required to gain more knowledge about  
22 the mode of action of these antimicrobial agents in wines and their consequences for  
23 wine quality.

24         Recently, special attention has been paid to the potential of some natural  
25 constituents of grapes and wines, such as phenolic compounds, as an alternative to

1 sulphites (García-Ruiz, et al., 2008a). It has been suggested that phenolic compounds  
2 can behave as activators or inhibitors of bacterial growth depending on their chemical  
3 structure (substitutions in the phenolic ring) and concentration (Reguant et al., 2000;  
4 Vivas et al., 1997). However, data are scarce since research has mainly been focused on  
5 studying the effect of hydroxycinnamic and hydroxybenzoic acids and some flavonols  
6 (Stead, 1993; Rozes and Peres, 1998; Campos et al., 2003; Alberto et al., 2001). In a  
7 recent study, the evaluation of the dual antioxidant and antibacterial activity of 21  
8 phenolic compounds mainly present in *Vitis Vinifera L.* belonging to different groups  
9 (hydroxybenzoic acids and their derivatives, hydroxycinnamic acids, phenolic alcohols  
10 and other related compounds, flavonols, anthocyanins and stilbenes) was examined  
11 (García-Ruiz et al. 2008b). Through the use of structure-activity relationships, the  
12 antimicrobial and antioxidant properties of some of these wine phenolics was  
13 demonstrated, confirming the potential use of these compounds as an alternative to  
14 sulphites in winemaking. Nevertheless, more studies would be necessary to evaluate  
15 other phenolic structures present in wines and to elucidate the mechanism of action  
16 implicated on microbial inactivation.

17

### 18 ***Compounds resulting from microbial metabolism***

19

20 Winemaking cannot take place without the action of microorganisms, mainly  
21 yeasts, and also lactic acid bacteria. Yeasts are essential to obtain wine, and lactic acid  
22 bacteria to make an important contribution to its organoleptic quality. However, the  
23 winemaking process includes multiple stages at which spoilage microorganisms,  
24 mainly some strains and species of moulds, yeasts and lactic acid bacteria can develop  
25 and alter the quality and hygienic status of the wine, affecting the food safety for the

1 consumers. Some interesting examples include the production of ochratoxin A, ethyl  
2 carbamate and biogenic amines, that will be discuss as follows.

3

#### 4 ***Ochratoxin A***

5

6 Ochratoxin A (OTA) is one of the most common naturally occurring  
7 mycotoxins, which is receiving increasing attention due to its toxic effects and high  
8 incidence in a wide range of food products. The IUPAC developed formula of OTA is  
9 1-phenylalanina-*N*-[(5-chloro-3,4-dihydro-8-hidroxy-3-methyl-1-oxo-1H-2-  
10 benzopyran-7-yl)carbonyl]-isocoumarin (Ringot et al., 2006) (**Figure 1**). OTA is a  
11 mycotoxin considered to be a possible carcinogen (Class 2B) in humans (IARC, 1993)  
12 and it has been shown to be nephrotoxic, teratogenic, hepatotoxic and immunotoxic in  
13 animals, and possibly in humans (Zimmerli and Dick, 1996). It has also been associated  
14 with the Balkan Endemic Nephropathy (BEN) and with the development of urinary tract  
15 tumors. Unfortunately, the presence of this compound in blood from healthy humans  
16 confirms a continuous and widespread exposure (Creppy, 2002).

17 Since 1970 the occurrence of OTA in foods has been investigated and its  
18 presence has been confirmed in different food products such as cereals, coffee, some  
19 sauces such as soya sauce, cocoa, dried fruits, meat and beer (JECFA, 2001). In the  
20 European diet, wine and especially red wine, has been identified as the second major  
21 source of human exposure to OTA, following cereals. The occurrence of OTA in wine  
22 and grape has been reported in different countries, but higher levels of OTA were found  
23 in the southern regions of Europe than in northern regions (Zimmerli and Dick, 1996;  
24 Visconti et al., 1999; Otteneder and Majerus, 2000; López de Cerain et al., 2002;  
25 Bacaloni et al., 2005). Consequently, the European Commission has considered that is

1 necessary to impose regulatory limits establishing 2 µg/L as the maximum level of OTA  
2 in wine and grape products (EC N° 123/2005).

3         OTA is produced by the secondary metabolism of several *Aspergillus* and  
4 *Penicillium* species, mainly by *Aspergillus ochraceus* and *Penicillium verrucosum*  
5 (Chulze et al., 2006). Recently, black *Aspergillus* species (section *Nigri*), including  
6 *Aspergillus carbonarius*, *Aspergillus aculeatus* and *Aspergillus japonicus*, have been  
7 described as the main source for OTA contamination in grapes (Da Rocha Rosa et al.  
8 2002; Battiliani et al., 2003; Serra et al., 2003; Bellí et al., 2004; Leong et al., 2006a).  
9 Researchers agreed in considering *A. carbonarius* as the main causal agent of OTA  
10 presence in grapes in Europe and Israel (Battiliani et al., 2006; Belli et al., 2006).  
11 Molecular methods (for an overview see Abarca et al., 2004) have led to an important  
12 progress in the identification of the fungi species responsible for OTA contamination in  
13 grapes and wine.

14         Improvements in viticultural and winemaking practices are required to reduce  
15 OTA in wines. The great amount of work performed on this topic, has allowed to gain  
16 knowledge about the origin and mechanisms to explain the presence of OTA in wines  
17 and in the development of prevention and control strategies to avoid its occurrence.  
18 Recently with the aim to limit the presence of OTA in vine-derived products, the OIV  
19 has adopted a ‘Good Viticultural and Winemaking Practices Code’ based on  
20 recommendations and specific interventions for both harvest and cellars (OIV, 2005).  
21 The main measures considered by the OIV Code are summarized in **Table 1**.

22         Factors affecting the presence of ochratoxin A in wines have been widely  
23 reviewed (Blesa et al., 2006; Mateo et al., 2007). Different practices in grape cultivation  
24 such as the type of cultivars, and in winemaking, including the storage conditions of the  
25 harvested grapes and the length of the storage, can influence the accumulation of

1 mycotoxins (Zimmerli and Dick, 1996). Some grape varieties may exhibit a greater  
2 susceptibility than others to *Aspergillus* bunch rots. However, once damage is set, there  
3 are no inherent differences in resistance to mould growth between different grape  
4 varieties. The substantial processing that grapes undergo during wine production is one  
5 of the major factors that influence OTA levels in wine (Leong et al., 2006b).

6       Regarding preventive and corrective actions to reduce OTA content in wines, it  
7 should be taken into account that a good management of the vineyard guarantees a  
8 relevant decrease in OTA content at harvesting. A primary focus for continuing research  
9 in this subject is the development of vineyard management strategies to minimize the  
10 risk of OTA development in grapes and vineyard by reducing the incidence of black  
11 *Aspergillus* spp., in soils and grape bunches. One approach in this direction is the  
12 introduction of varieties that are more resistant to splitting and hence less susceptible to  
13 *Aspergillus* bunch rots. Further research should be conducted in order to examine the  
14 genetic relatedness of black *Aspergillus* isolates from various viticultural regions.  
15 Although additional OTA is not synthesised during winemaking, a careful management  
16 during wine production including corrective actions, would contribute to minimise OTA  
17 levels in wine. Data recently presented indicate that phenolic antioxidant compounds  
18 suppress OTA production in several important ochratoxigenic *Aspergillus* species  
19 (Palumbo et al., 2007). Whether searching for natural strategies for the reduction of  
20 OTA contaminations or the study of biosynthetic and regulatory genes involved in both  
21 OTA production and stress response in ochratoxigenic *Aspergilli*, can be useful tools for  
22 controlling OTA in wines.

23       During both red and white winemaking, the removal of precipitated yeast cells  
24 and other sediments after fermentation results in further reductions in OTA content.  
25 Binding and precipitation of OTA may be enhanced through the addition of fining

1 agents. The influence of some oenological practices on OTA concentration in wine has  
2 been evaluated (Gambutì et al., 2005). Microfiltration (0.45 µm) and the use of carbon  
3 are two useful treatments for wine decontamination, but the use of carbon results in the  
4 adsorption of several aroma compounds (Olivares-Marín et al., 2008). The feasibility of  
5 using selected adsorbents (bentonites, modified bentonites and chitosan) to reduce OTA  
6 levels in red wine has been recently investigated (Kurtbay et al., 2008). Quitine from  
7 yeast walls (2-acetamido-2-deoxy-B-D-glucose) has been shown to reduce OTA levels  
8 in wine (Bornet and Teissedre, 2008). In addition, yeast mannoproteins also play an  
9 important role for the removal of OTA because of their binding capacity (Bejaoui et al.,  
10 2004).

11         Subsequent operations during winemaking also reduce the OTA content of wine;  
12 This reduction has been shown to depend on several factors. In the case of red  
13 winemaking, the fermentation step reduces OTA content both *in vitro* and in true scale  
14 experiments; the efficacy of fermentations, both alcoholic and malolactic, in reducing  
15 OTA content is strictly dependent on the yeast and bacterial strains involved, and on the  
16 level of contamination (Mateo et al., 2007). In white winemaking, OTA content always  
17 decreases and the rate of decrease depends on the choices for yeasts during alcoholic  
18 fermentation and on the processing aids added for clarification.

19

## 20 ***Ethyl carbamate***

21

22         Ethyl carbamate, also known as urethane, has been used as antiseptic for many  
23 years in the alcoholic drink industry. At the beginning of the Seventies, a scientific  
24 article reported for the first time that this compound could have potentially carcinogenic  
25 properties in laboratory animals when present at high concentrations (Canas et al.,

1 1989). Since then, its use in food products is forbidden and considerable research  
2 regarding the mechanisms implicated in its formation in fermented and non-fermented  
3 foods, has gained interest. From the data obtained in experimental animals, it has been  
4 estimated that the daily dose for humans with no harmful effect is 0.3 ng/kg of body  
5 weight/day, which corresponds to 0.021 µg/day for a person of approximately 70 kg.

6 The formation of ethyl carbamate in wine is mainly attributed to the metabolism  
7 of the yeasts responsible for alcoholic fermentation. These microorganisms can  
8 sometimes produce a large amount of an intermediate metabolite, urea, that although in  
9 itself is not harmful to health, if the conditions are appropriate (high temperature,  
10 presence of ethanol and low pH), it can be transformed by a chemical reaction into ethyl  
11 carbamate during wine storage (Monteiro and Bisson, 1991). But also, it has been  
12 demonstrated that during malolactic fermentation, lactic acid bacteria can produce other  
13 precursors of ethyl carbamate such as citruline and carbamyl phosphate, which, in the  
14 aforementioned conditions can result in the production of this compound (Liu et al.,  
15 1995; Liu et al., 1996; Arena et al., 1999). In both cases, the initial substrate is arginine,  
16 one of the major amino acids present in grape must and wine (Lehtonen, 1996;  
17 Moreno-Arribas et al., 1998).

18 The formation of ethyl carbamate by lactic acid bacteria, has probably been the  
19 line of research most followed in recent years in this field, especially in an attempt to  
20 reduce its concentration in wines. Certain wine-associated lactic acid bacteria have the  
21 ability to utilise arginine; these include strains of *Oenococcus oeni* and  
22 heterofermentative lactic acid bacteria, such as *Lactobacillus brevis*, *L. buchneri*, *L.*  
23 *hilgardii*. Homofermentative lactic acid bacteria (e.g. *L. delbrueckii*, *L. plantarum*) and  
24 pediococci do not catabolise arginine. Now, both, the enzymatic route involved (i.e. the  
25 arginine deiminase pathway) and the genes that encode the corresponding enzymes of

1 oenological lactic acid bacteria species are known (Tonon et al., 2001). From the  
2 research performed, Mira de Orduña et al. (2001) concluded that the risk of citrulline  
3 formation by malolactic bacteria in wines with high residual arginine concentrations can  
4 be reduced by carrying out malolactic fermentation with pure oenococcal cultures and  
5 by precisely establishing complete malolactic conversion, which must be followed by  
6 inhibition of bacterial activity. Also, in this case, scientific results indicate the need for  
7 caution in the selection of starter cultures for MLF in wine, since citrulline formation  
8 from arginine degradation could result in ethyl carbamate production, even at normal  
9 temperatures, during prolonged storage. In addition, spontaneous malolactic  
10 fermentation by undefined strains should be avoided, as this may lead to formation of  
11 ethyl carbamate precursors.

12         Nowadays vine growers and winemakers have sufficient knowledge to prevent  
13 or reduce the formation of ethyl carbamate during the production process. The strategies  
14 for controlling the presence of this compound in wines are focused on the three factors  
15 involved in its production: vine growing and wine production practises that could  
16 increase the concentration of arginine in the grape; to control the winemaking process,  
17 especially the alcoholic and malolactic fermentations, and finally, the ageing and  
18 maturation of the wine. On the other hand, legislation authorises the use of an  
19 enological additive, the enzyme urease, in wines with high urea levels. The urease used  
20 is isolated from *Lactobacillus fermentum*. This enzyme has been effective on urea at  
21 doses of 50 mg/L in red wines and at doses of 25 mg/L in white wines (Bertrand, 1997).

22         A systematic study carried out to examine the ethyl carbamate levels of Spanish  
23 red wines from different grape varieties (Uthurry et al., 2004), showed that typical  
24 Spanish red wines had concentrations of ethyl carbamate below 30 ppb. During the  
25 alcoholic fermentation of the corresponding musts no ethyl carbamate was formed, even

1 in the presence of nitrogenous substrates, and low concentrations were recorded ( $\leq 1.3$   
2 ppb). Generally, regarding urea levels founded at the end of the alcoholic fermentation  
3 it was concluded that small quantities of ethyl carbamate would be formed under the  
4 normal storage conditions of wines (Uthurry et al., 2007) .

5

## 6 ***Biogenic amines***

7

8 Amines are widely occurring compounds in nature, containing one or more  
9 organic substituent bonded to a nitrogen atom. Biogenic amines are low molecular  
10 weight compounds, derived from aromatic or cationic amino acids and all of them have  
11 one or more positive charge and a hydrophobic skeleton. Depending on their chemical  
12 structure they can be classified in aliphatic (putrescine, cadaverine, spermine,  
13 spermidine), aromatic (tyramine, phenylethylamine) or heterocyclic (histamine,  
14 tryptamine) (**Figure 2**). They are endogenous of plants but can also be found in fresh  
15 fruit and vegetables. However, amines are mainly formed in foods by fermentative  
16 processes and during ageing and storage. Recently, knowledge concerning the presence  
17 of amines in wines has been reviewed (Ferreira and Pinho, 2006; Ancín-Azpilicueta et  
18 al., 2008).

19 Problems related to biogenic amine formation affect numerous fermented food  
20 products consumed more frequently than wine, such as cheese, beer, some fermented  
21 sausages and meat products among others (Fernández-García et al., 1999; Izquierdo-  
22 Pulido et al., 2000; Kaniou et al., 2001), which have higher levels of these compounds.  
23 However, in alcoholic drinks, especially in wine, biogenic amines have received more  
24 attention, because ethanol and acetaldehyde can increase their effects on health by  
25 directly or indirectly inhibiting the enzymes responsible for the detoxification of these

1 compounds in the intestinal tract (Maynard and Schenker, 1996). Although there are  
2 differences in individual susceptibility to intoxication by biogenic amines, several  
3 pharmacological reactions can take place after excess intake of these compounds. The  
4 most well-known reactions are those caused by histamine. More specifically, some  
5 histamine-induced symptoms include rash, edema, headaches, hypotension, vomiting,  
6 palpitations, diarrhoea and heart problems. Other amines, such as tyramine and  
7 phenylethylamine, can cause hypertension and other symptoms associated with  
8 vasoconstriction due to the release of noradrenaline (especially cerebral haemorrhages  
9 and migraine). Although putrescine and cadaverine are not themselves toxic, they can  
10 increase the toxicity of histamine, tyramine and phenylethylamine, since they interfere  
11 in detoxification reactions.

12         Some polyamines, such as putrescine, may already be present in grape berries  
13 (Halász et al., 1994; Bover-Cid et al., 2006; Del Prete et al., 2008) or be produced  
14 during fermentation processes, ageing or storage, when wine is exposed to the  
15 undesirable activity of decarboxylase-positive microorganisms. Contamination may also  
16 occur from poorly sanitary conditions of both grapes and processing equipment.  
17 Although yeasts may also contribute to the final levels of biogenic amines in wines  
18 (Torrea and Ancín, 2002; Caruso et al., 2002), most biogenic amine contamination of  
19 wine is believed to take place during malolactic fermentation from the presence of lactic  
20 acid bacteria strains that produce enzymes which decarboxylate the corresponding  
21 precursor amino acids (Marcobal et al., 2006a). For example, in the production of the  
22 biogenic amine histamine, the enzyme involved is histidine decarboxylase, which acts  
23 on the amino acid precursor, histidine.

24         It is well known that among the species and strains of wine's lactic acid bacteria,  
25 some of them are practically unable to produce biogenic amines whereas others are

1 characterized for their high capacity of production. It is often to detect aminobiogenic  
2 strains among heterofermentative lactobacilli (*Lactobacillus hilgardii* y *L. brevis*)  
3 (Moreno-Arribas et al., 2000), although among histamine producers strains, some  
4 *Pediococcus* (Landete et al., 2005; Bodmer and Gafner, 2000) and *O. oeni* strains have  
5 been also isolated (Coton et al., 1998). The latter have also been found to be a  
6 putrescine producer (Marcobal et al., 2004; Marcobal et al., 2006b). Although it has  
7 been demonstrated that oenococci could produce histamine and putrescine, this property  
8 seems to be variable and is only present in some strains, which explains why in other  
9 studies, strains of biogenic amine-producing *O. oeni* have not been found (Constantini  
10 et al., 2006; Moreno-Arribas and Polo, 2008).

11 Molecular methods for the detection of biogenic amine-producing bacteria have  
12 been recently reviewed by Landete et al., (2007). PCR techniques have been developed  
13 to detect bacterial amino acid decarboxylases in a rapid, sensitive and accurate way.  
14 They cannot determine quantitative (or qualitative) amounts of biogenic amines, but can  
15 be used to estimate the potential risk for amine formation. *Multiple* PCR systems that  
16 use different pairs of primers to simultaneously detect lactic acid bacteria producers of  
17 histamine, tyramine and putrescine, the major wine biogenic amines have been  
18 developed (Marcobal et al., 2005b; Constantini et al., 2006). This system detects the  
19 presence of genes that encode the enzymes responsible for synthesizing these amines in  
20 bacteria. Quantitative PCR (or real time PCR) allows knowing the number of potential  
21 amine producers bacteria in wine, therefore it is an interesting tool to evaluate the risk  
22 for biogenic amine production from a specific bacteria population (Lucas et al., 2008).

23 From the wide range of techniques described in the literature to determine the  
24 concentration of biogenic amines in wines, reverse phase high-performance liquid  
25 chromatography (RP-HPLC) is one of most used in laboratories, since it is possible to

1 obtain valuable information about the presence and concentration of amines in musts  
2 and wines (Lehtonen, 1996; Torrea and Ancín, 2002; Marcobal et al., 2005a; Gómez-  
3 Alonso et al., 2007). Moreover, other chromatographic techniques applicable to  
4 biogenic amine analysis, such as micellar liquid chromatography (MLC) (Gil-Augustí et  
5 al., 2007) and liquid-chromatography-electrospray ionisation ion trap mass  
6 spectrometry (Millán et al., 2007) have been developed. Enzymatic methods (ELISA or  
7 other enzymatic tools) for histamine analysis able to determine levels of this amine in a  
8 short time and do not require sophisticated equipment have lately appeared (Marcobal et  
9 al., 2005). In recent years there has also been a growing interest in the use of capillary  
10 electrophoresis (CE) and different CE methods with diverse detection systems (Santos  
11 et al., 2004; Simó et al., 2008).

12       Regarding enological practices and technological factors that may influence  
13 biogenic amine formation in wines, special attention should be paid to some oenological  
14 practices frequently used to enhance wine complexity (flavour and aroma), but that also  
15 can increase the concentration of the precursor amino acids, such as the ageing of wines  
16 with lees or longer maceration times (Martín-Álvarez et al., 2006). Yeasts can  
17 indirectly play an important role on the production of biogenic amines by lactic acid  
18 bacteria, since they affect the amino acid composition during the alcoholic fermentation  
19 (Soufleros et al., 1998) or during autolysis (Moreno-Arribas et al., 1998; Villamiel et  
20 al., 2008). These amino acids can act as precursor for the formation of biogenic amines  
21 during malolactic fermentation and during wine ageing (Martín-Álvarez et al., 2006;  
22 Marques et al., 2008). Yeast and bacteria lees can also be a source for decarboxylase  
23 enzymes that could be involved in amine production (Marcobal et al., 2004).

24       The use of bacteria starter with specific characteristics such a very reduced or  
25 null capacity to produce biogenic amines, is the most popular strategy to control amine

1 production during winemaking. *O. oeni* commercial starters are selected on the basis of  
2 their oenological properties and among them, the absence of amino acid decarboxylase  
3 enzymes is an important requirement. Research performed in *in vitro* studies (Moreno-  
4 Arribas et al., 2003) and in industrial conditions during winemaking (Marcobal et al.,  
5 2006a) have shown the viability of using bacteria starters without the gene involved in  
6 biogenic amine production to prevent the formation of amines in wines. Moreover, it  
7 has been shown that the co-inoculation of *O. oeni* starters simultaneously to the  
8 alcoholic fermentation as opposite to the conventional inoculation once the alcoholic  
9 fermentation has been completed, is more effective to avoid the production of biogenic  
10 amines (Smit, 2007).

11 The use of different clarification substances and oenological adjuvants, such as  
12 bentonite PVPP at the normal dose used in winemaking can affect the concentration of  
13 biogenic amines in wines, since these compounds have the ability for the adsorption of  
14 some of them (Alcaide-Hidalgo et al., 2008). Also the type container employed during  
15 malolactic fermentation (stainless steel or oak barrel) seems to affect biogenic amine  
16 content of wines (Alcaide-Hidalgo et al., 2007a). Some alternative techniques for wine  
17 ageing based on the use of wood chips or oenological tannins, can also influence the  
18 amine production depending on the product type and on the dose of these compounds  
19 applied to the wines (García-Ruiz et al., 2008b).

20 Biogenic amine formation is also determined by wine parameters, such as pH,  
21 ethanol and SO<sub>2</sub>, which have an important effect on the diversity of microorganisms,  
22 decarboxylase enzyme activity and decarboxylase gene expression (Lonvaud-Funel,  
23 1999; Leitao et al., 2000; Mazzoli et al., 2008).

24  
25

1 ***COMPOUNDS PRESENT IN WINE WITH POTENTIALLY BENEFICIAL***  
2 ***EFFECTS ON HEALTH***

3  
4 Since the postulation of the French paradox in 1992 by Renaud and de Lorgeril  
5 (1992), moderate consumption of red wine has been associated with reduce risk of  
6 developing cardiovascular heart disease (CHD). The findings that constitute the French  
7 paradox for CHD were confirmed in the MONICA project, a worldwide study for  
8 cardiovascular diseases organized by the World Health Organization, that showed that  
9 the mortality rate from CHD was much lower in France, than in other industrialised  
10 countries such as USA and UK, despite the presence of high level of risk factors, such  
11 as cholesterol, diabetes, hypertension, and high intake of saturated fat. These effects  
12 were partly attributed to the high intake of wine and in general to the so-called  
13 Mediterranean-type diet, which is particularly abundant in fruits, vegetables and olive  
14 oil. These findings were already in agreement with a previous epidemiological study  
15 reporting an inverse association between moderate wine consumption and CHD (St  
16 leger et al., 1979). Later studies have also confirmed the results of Renaud et al. (1992).  
17 Gronbaek et al. (2000) found that all-cause mortality was reduced in wine but not in  
18 non-wine drinkers in a study including 24,523 subjects over 10 years. In a meta-  
19 analysis on 209,523 subjects, di Castelnuovo et al. (2002) also reported a larger benefit  
20 for wine (~22% reduction) than for beer. Wine drinkers were found to have a lower  
21 mortality risk than beer o liquor drinkers in a follow-up study including 128,934  
22 subjects up to 20 years (Klatsky et al., 2003). The better effects of wine in comparison  
23 to other alcoholic beverages have been mainly attributed to the presence of antioxidant  
24 polyphenols that provide additional benefits. Other compounds such as low-molecular-  
25 weight peptides exhibiting antihypertensive activity could also contribute to these

1 beneficial effects. However, other prospective studies have indicated that the beneficial  
2 effects of alcohol intake were not dependent on the type of alcoholic beverage  
3 (Mukamal et al., 2003, Imhof et al., 2004). In a meta-analysis, moderate alcohol  
4 consumption (as much as 30 g alcohol/day) was related to lower risk (20-45%  
5 reduction) of CHD compared with abstention (Rimm et al., 1999). Discrepancies among  
6 studies have been attributed to the existence of many confounding variables related to  
7 life style and genetic factors. Health effects of moderate alcohol consumption ( $\leq 2$   
8 drinks/day) have been mainly related to an increase in HDL-cholesterol as well as to  
9 other protective mechanisms such as anti-platelet, anti-coagulatory, and anti-  
10 inflammatory effects (Rimm et al., 1999, Imhof et al., 2004). However, three or more  
11 drinks per day are associated with cardiomyopathy, hypertension, hypertriglyceridemia,  
12 and stroke. Although keeping in mind the importance of alcohol in the potentially  
13 beneficial effects of wine on human health, our dissertation will be focus on phenolic  
14 compounds and peptides as specific wine components with potentially beneficial  
15 effects.

16

### 17 *Phenolic compounds*

18

19 The term "phenolic" or "polyphenolic" describes the compounds that possess a  
20 benzenic ring substituted by one or several hydroxyl groups (-OH). Based on their  
21 carbon skeleton, polyphenols are classified in non-flavonoid and flavonoid compounds.  
22 Main non-flavonoid compounds present in wine are phenolic acids (hydroxybenzoic  
23 and hydroxycinnamic acids), as well as others phenolic derivatives such as stilbenes  
24 (**Figure 3**). Flavonoid compounds present in wine mainly include the flavonols, flavan-  
25 3-ols and anthocyanins, as well as flavanonols and flavones, in lower proportion

1 **(Figure 3)**. Due to their chemical structure, containing several hydroxyl groups,  
2 stilbenes and flavonoids have been demonstrated to be the most bioactive compounds in  
3 wine.

4 Hydroxylated stilbenes, as previously described in section #, are phytoalexins  
5 synthesized by the plant, especially in the skins, leaves and roots, in response to fungal  
6 infections and ultraviolet (UV) light (Jeandet et al., 1991; Korhammer et al., 1995).  
7 Stilbenes present in wine include *trans* and *cis* resveratrol (3,5,4'-trihydroxystilbene), as  
8 well as their glucose derivatives (*trans* and *cis* piceid). A series of mono-, di-, and  
9 tetrahydroxystilbenes have been also identified in wine (Lamikanra et al. 1996; Ribeiro  
10 de Lima et al. 1999; Baderschneider and Winterhalter, 2000). Considering that grapes  
11 and their derived products are the most important dietary sources of stilbenes (Mattivi et  
12 al., 1995), resveratrol metabolites have been recently proposed as a biomarkers of  
13 moderate wine consumption in humans (Zamora-Ros et al., 2006).

14 In relation to the flavonoid phenolic compounds found in wine, flavonols are  
15 yellow pigments mainly located in the vacuoles of the epidermal tissues. In wine they  
16 exist as the 3-*O*-glycosides of four main aglycones: myricetin, quercetin, kaempferol  
17 and isorhamnetin **(Figure 3)**. Flavan-3-ols or flavanols are found in wine in monomeric,  
18 oligomeric or polymeric forms in the solid parts of the grape; the latter two forms are  
19 also known as proanthocyanidins or condensed tannins. Flavanol monomeric units  
20 include (+)-catechin, (-)-epicatechin, (-)-epicatechin-3-*O*-gallate, (+)-gallocatechin and  
21 (-)-epigallocatechin **(Figure 3)**. Proanthocyanidins are constituted of flavanols units  
22 linked by C4-C6 or C4-C8 bonds. Proanthocyanidins with a mean degree of  
23 polymerization between 3 and 7 units have been reported in red wine (Fulcrand et al.,  
24 1999). Anthocyanins are pigment located in the grape skins of red grape varieties. Main  
25 anthocyanins identified in wine include the 3-*O*-monoglucosides and the 3-*O*-acylated

1 monoglucosides of five main anthocyanidins: delphinidin, cyanidin, petunidin, peonidin  
2 and malvidin (**Figure 3**). These compounds could be acylated in the C-6 position of the  
3 glucose molecule by esterification with acetic, *p*-coumaric and caffeic acids. Both,  
4 anthocyanins and flavanols are the most abundant flavonoids in red wines and they are  
5 largely responsible for the color, bitterness, astringency and chemical stability of red  
6 wines. A glass of red wine provides approximately 100 mg of polyphenols. In  
7 comparison to red wines, white wines are devoid of anthocyanins, and due to absence of  
8 skins and seed contact during maceration and fermentation in white wine technology,  
9 they possess a lower content of phenolic compounds, in particular of flavonoids.

10 *In vitro* and animal model studies have demonstrated that red wine polyphenols  
11 are protective agents against the development and progression of atherosclerosis  
12 (Dell'Agli et al., 2004). This protection is related to: *i) inhibition of LDL oxidation*  
13 (Frankel et al., 1993; Lourenço et al., 2008); *ii) vasorelaxation* (reduction of blood  
14 pressure; endothelium-dependent vasorelaxation by increased generation of NO,  
15 endothelium-derived hyperpolarizing factor and prostacyclin, or by reduction of the  
16 vasoconstrictor endothelin-1) (Fitzpatrick et al., 2000; Diebolt et al., 2001; Ndiaye et  
17 al., 2003, Schramm et al., 1997; Corder et al., 2001); *iii) inhibition of vascular smooth*  
18 *muscle cell migration and proliferation* [by possible inhibiting platelet-derived growth  
19 factor (PDGF)  $\beta$ -receptor and vascular endothelial growth factor (VEGF)] (Rosenkranz  
20 et al., 2002; Ferrera et al., 1999); and *iv) inhibition of platelet aggregation* (by  
21 stimulation of the synthesis of prostacyclin; suppression of platelet activation response  
22 to epinephrine; increase of platelet c-AMP levels) (Gryglewski et al., 1987; Rein et al.,  
23 2000; Russo et al., 2001). Administration of red wine, dealcoholized wine or grape juice  
24 has also been shown to attenuate the development of atherosclerotic lesions (da Luz.,  
25 1999; Vinson et al., 2001) and to prevent thrombosis (Demrow et al., 1995) in

1 experimental animals. Red wine polyphenols also exhibit antiviral (Takechi et al.,  
2 1985), antibacterial (Cueva et al., 2008), anticancer (Franke et al., 2002; Weyant et al.,  
3 2001) and antiulcer (Saito et al., 1998) activities. Finally, the implication of red wine  
4 polyphenols in the modulation of gene expression has also been reported (Sen and  
5 Bagchi, 2001).

6  
7 The health effects observed in *in vitro* and animal model studies have been  
8 partially confirmed in human subjects. A summary of short-term human intervention  
9 studies related to wine polyphenols is presented in **Table 2**. These studies have focused  
10 on confirming the hypothesis that: *i*) red wine provides more beneficial effects than  
11 other alcoholic beverage (red wine vs. spirits or beer); *ii*) red wine properties are beyond  
12 alcohol (red wine vs dealcoholized wine or grape juice); *iii*) red wine leads to greater  
13 beneficial effects than white wine, and *iv*) polyphenols are responsible for red wine  
14 properties. Most studies have been conducted on healthy men only, or on healthy men  
15 and women, with the exception of one study that was performed on healthy women  
16 (Sacanella et al., 2007). Intervention periods ranged between 14 and 30 days, although  
17 some acute intake (1 day) studies have also been performed. Alcohol doses were on the  
18 order of 20-40 g ethanol/day. Biomarkers significantly affected in these trials are  
19 related to red wine: *i*) *antioxidant effect* (inhibition of the LDL oxidation; reduction of  
20 plasma lipid peroxides, thiobarbituric acid-reactive substances, conjugated dienes, and  
21 urine isoprotanes levels; increase in plasma antioxidant capacity, plasma polyphenol  
22 content and LDL polyphenols); *ii*) *effects on lipid metabolism* (increment in plasma  
23 HDL cholesterol; reduction of LDL/HDL ratio and LDLox antibody); *iii*) *anti-*  
24 *thrombotic effects* (reduction of trombine-induced and collagen-induced platelet  
25 aggregation; reduction of plasmatic levels of tromboxane B2, fribinogen, factor VII, t-

1 PA, PAI); *iv) effects on vascular function* (reduction in flow-mediated dilatation of the  
2 brachial artery), and *vi) anti-inflammatory effects* (reduction of the expression of  
3 adhesion molecules on monocyte surface, including LFA-1, Mac-1, VLA-4 and MCP-1;  
4 reduction of serum levels of hs-CRP, VCAM-1, ICAM-1 and E-selectin; increment of  
5 TGF $\beta$ 1). The anti-inflammatory effects of white sparkling wine (cava) in comparison to  
6 gin have also been determined and were on favour of cava (Vázquez-Agell et al., 2007).  
7 However, it is important to highlight that the other studies failed to find positive  
8 outcomes for some parameters after red wine consumption. According to Djoussé et al.  
9 (1999), no significant changes were found in flow-mediated dilation of the brachial  
10 artery when red wine was drunk together with a fatty meal. Watzl et al. (2002, 2004) did  
11 not find changes in anti-inflammatory biomarkers after acute and chronic consumption  
12 of red wine. Van deer Gaag et al. (2000a,b) found an increase in plasma homocysteine  
13 levels after red wine consumption and no significant changes in the activity of  
14 antioxidant enzymes or plasmatic levels of some antioxidant vitamins.

15 The *in vivo* physiological importance derived from wine consumption is  
16 dependent on the bioavailability (absorption, distribution, metabolism and excretion) of  
17 wine polyphenols. The metabolic pathway of polyphenols is well established (Scalbert  
18 and Williamson, 2000). The bioavailability of wine polyphenols is influenced by their  
19 chemical structure. Monomers and dimeric procyanidins in a lower extent, are directly  
20 absorbed in the small intestine, whereas oligomers with a mean degree of polymerization  
21 (mDP) $>3$  and polymeric flavanols (proanthocyanidins) are not absorbed in their native  
22 forms or degraded under the acidic conditions of the stomach *in vivo* (Rios et al., 2002).  
23 These compounds reach the colon where they are metabolized by the intestinal  
24 microbiota into various phenolic acids, including phenylpropionic, phenylacetic and  
25 benzoic acid derivatives that can be further absorbed by enterohepatic recirculation via

1 the bile duct (Manach et al., 2004). In the other hand, glycosilated polyphenols such as  
2 resveratrol glucosides and anthocyanins, must be first hydrolyzed by intestinal  $\beta$ -  
3 glucosidases before they can be absorbed (Manach et al., 2004). However, in the case of  
4 wine anthocyanins, it has been demonstrated that they can be absorbed in their  
5 glycosilated form after oral ingestion (Matsumoto et al., 2001; Cao et al., 2001; McGhie  
6 et al., 2003).

7 After passing across the small intestine brush border, polyphenols are  
8 extensively conjugated first in the small intestine and then in the liver into methyl,  
9 glucuronide and sulphate derivatives by the enzymes catechol-*O*-methyltransferase,  
10 UDP-glucuronosyl transferase, and phenol sulfotransferases, respectively (Scalbert and  
11 Williamson, 2000). In general, glucuronide metabolites are the most abundant  
12 metabolites in plasma and only trace amounts of native forms are found. However, in  
13 the case of anthocyanins, although the existence of methyl, glucuronide and sulfate  
14 derivatives has been demonstrated (Wu et al., 2002; Felgines et al. 2003), intact  
15 glycosides were the only metabolites reported in previous studies, probably due to their  
16 degradation during sample conservation (Manach et al., 2004). This fact together with  
17 other issues related to the use of inappropriate analytical methods may have resulted in  
18 an underestimation of anthocyanin bioavailability in early studies.

19 Pharmacokinetic studies indicate that after the ingestion of 35 mg of catechin  
20 supplied in the form of red wine (120 mL), the maximum concentration ( $C_{\max}$ ) of  
21 catechin in plasma was on the order of 0.077-0.091  $\mu\text{mol/L}$ , and was achieved at a mean  
22 time ( $T_{\max}$ ) between 1.44-1.50 h. Urinary excretion ranged between 3-10% of the intake  
23 and the elimination half-life was approximately 3 h (Donovan et al., 1999, 2002; Bell et  
24 al., 2000). Concerning proanthocyanins, up to date there is no literature data of the  
25 absorption of these compounds after wine consumption. A plasma concentration of

1 0.0011  $\mu\text{mol/L}$  of procyanidin B1 has been reported after the ingestion of grape seed  
2 extract (18 mg of procyanidin B1) in humans (Sano et al., 2003). In relation to the  
3 microbial metabolism of proanthocyanins into phenolic acids, Gonthier et al. (2003)  
4 have shown that the DP has an inverse association with the extent of microbial  
5 degradation in rats fed purified monomeric, dimeric, trimeric or polymeric flavanols. In  
6 comparison with flavanols, anthocyanins are more rapidly absorbed. After the ingestion  
7 of 68 mg of malvidin-3-glucosides in the form of red wine (500 mL),  $T_{\text{max}}$  in plasma  
8 and urine was attained at 0.83h and <3h, respectively (Bub et al., 2001); data which  
9 seem to be consistent with absorption in the stomach. However the  $C_{\text{max}}$  was much  
10 lower (1.4 nmol/L) than for flavanols, as well as the urinary excretion (0.016%) (Bub et  
11 al., 2001). In another study, higher urinary excretion were found (1.5-5.1 %) after the  
12 consumption of 300 mL red wine (218 mg of total anthocyanins) (Lapidot et al., 1998).  
13 The bioavailability of resveratrol has not been determined after the consumption of red  
14 wine in humans. Oral administration of resveratrol (0.026 mg total resveratrol) to rats in  
15 the form of wine (4 mL), gave resveratrol levels of 0.1  $\mu\text{mol/L}$ , which is similar to that  
16 of catechin (Bertelli et al., 1996). A larger number of studies should be carried out in  
17 order to acquire more information about the metabolism of red wine polyphenols after  
18 acute or chronic consumption of wine and its correlation with the observed *in vivo*  
19 effects.

20

## 21 ***Peptides***

22 Peptides are important wine compounds involved in different properties, such as  
23 tensioactive (González-Llano et al., 2004), sensorial (Desportes et al., 2001) or functional  
24 (Pozo-Bayón et al., 2007) properties. In addition, they can act as yeasts and bacteria  
25 nutrients (Feuillat et al., 1977; Alexandre et al., 2001; Manca de Nadra et al., 2005;

1 Remize et al., 2006). However, wine peptides are the least known nitrogen compounds.  
2 The main reason for the dearth of studies on this fraction is due to their complexity and  
3 lack of specificity of the techniques used for their analysis.

4 Due to the wide range of structures (amino acid composition and sequence) in  
5 which these compounds may appear, peptides form a heterogeneous group of  
6 compounds. Generally speaking, the term oligopeptides, or low-molecular-weight  
7 peptides, refers to peptides with 10 or fewer residues of amino acids, while the term  
8 polypeptides is used for peptides with more amino acids. Although the transition point  
9 from polypeptide to protein is not well defined, proteins are normally considered to  
10 have at least 100 residues of amino acids ( $M_r > 10,000$ ).

11 In recent years, several studies have focused on their origin and identification  
12 (Acedo et al. 1994; Moreno-Arribas et al. 1998) and in some cases, the sequence of  
13 amino acids have also been elucidated (Yahai et al. 2003; Person et al. 2004; Desportes  
14 et al. 2001). Some of these studies have shown that the amino acids aspartic acid and/or  
15 asparagine and glutamic acid and/or glutamine are involved in the majority of wine  
16 peptides, independently of the type of wine or the analytical methodology employed in  
17 the analysis.

18 The wine peptide fraction is continuously changing both qualitatively and  
19 quantitatively. Some wine peptides come from the must, but most of them appear in the  
20 different stages of wine production. During alcoholic fermentation, especially if it takes  
21 place in nitrogen-deficient media, there is a reduction in their concentration. In the final  
22 stages of fermentation, peptides are released into the medium, reaching the maximum  
23 concentration after cell death (Usseglio-Tomasset and Bossia, 1990). In fact, it has been  
24 shown that, at least part of the peptide fraction of wines is derived from the yeasts  
25 involved in the alcoholic fermentation. These peptides can be hydrolysed by the action

1 of extracellular enzymes, being transformed into low molecular weight peptides  
2 (Moreno-Arribas et al., 1996; Martínez-Rodríguez et al., 2000, 2001). The existence of  
3 endo and exocellular proteases has been evidenced under winemaking conditions  
4 (Feuillat et al., 1980; Alexandre et al., 2001). This process is even more important in  
5 winemaking technologies based on ageing the wines with yeasts for long periods of  
6 time, such as biologically aged wines (Dos Santos et al., 2000; Villamiel et al., 2008)  
7 and sparkling wines manufactured by the *traditional* method (Moreno-Arribas et al.,  
8 1996, 1998 a,b).

9       Also, some wine lactic acid bacteria strains have shown proteolytic activity  
10 under winemaking conditions (Feuillat et al., 1980; Manca de Nadra et al., 1997; Leitão  
11 et al., 2000). This activity has been found even in the presence of ethanol and SO<sub>2</sub>  
12 (Manca de Nadra et al., 2005) and has been reported to be greater in red than in white  
13 wines (Manca de Nadra et al., 1999). The proteolytic activity is a very important  
14 characteristic for some bacteria strains allowing them to growth in nitrogen-deficient  
15 media, and to carry out the malolactic fermentation. According to Remize et al. (2005)  
16 and Remize et al., (2006), peptides from 0.5 to 10 kDa seem to be more favourable for  
17 the growth of wine lactic acid bacteria than other nitrogen sources (<0.5 kDa). Alcaide-  
18 Hidalgo et al., (2008) have observed a reduction in peptides during the malolactic  
19 fermentation of red wines followed by an increase during the ageing in barrels, that was  
20 more pronounce when the aging was in presence of lees. The occurrence of proteolytic  
21 activity in all the stages of wine manufacture may be responsible for the wide range of  
22 peptides that can be present in a wine at any time, and the discrepancies in most of the  
23 studies related to their identification.

24       Low molecular weight peptides can be implicated in important biological  
25 activities such antimicrobial and antioxidant. The antihypertensive activity is the

1 bioactive property of peptides that has been most studied . The increase in hypertension  
2 rates of the population in recent years, which is considered as one of the commonest  
3 chronic diseases in developed countries, has favoured these types of studies. Most of  
4 the antihypertensive peptides derived from foods act by inhibiting angiotensin-  
5 converting enzyme (ACE). In the organism, ACE is present in a large number of organs  
6 and tissues, such as lung, intestine, kidneys, testicles, heart, skeletal muscle, pancreas,  
7 spleen or placenta. It is thought that the ACE located in the lung belongs to the renin-  
8 angiotensin system, one of the main mechanisms that regulates blood pressure in the  
9 body. The mechanism of action of the enzyme operates at two levels, on one hand, it  
10 catalyses the transformation of angiotensin I to angiotensin II, which has a strong  
11 vasoconstrictor action in the sympathetic nervous system and on the blood vessels. One  
12 the other hand, ACE inactivates bradykinin, a strong vasodilator and natriuretic. Both  
13 mechanisms cause an overall increase in blood pressure. The use of ACE inhibitors as  
14 active ingredients is the strategy of current antihypertensive drugs (captopril), to  
15 control blood pressure and other related diseases.

16 It has been shown that some peptides from fermented foods products can act by  
17 inhibiting the activity of the ACE enzyme, and, therefore, causing a lowering of blood  
18 pressure after intake (Hartmann and Meisela, 2007). However, afterwards these  
19 peptides must be resistant to the action of gastric enzymes and must be absorbed to have  
20 an effect. Therefore, after isolating and identifying peptides with *in vitro* biological  
21 activity, *in vivo* studies are required to establish their true efficacy.

22 In wines, the first study focused on the occurrence of antihypertensive peptides  
23 was carried out by Takayanagi and Yokotsuka (1999), who determined the inhibitory  
24 activity of ACE (IACE) in two red wines and four white wines and in grapes of the red

1 Muscat Bailey A variety and during fermentation of the must. They showed a greater  
2 ACE inhibitor activity in red wines than white wines and a decrease in IACE during  
3 fermentation, although not providing any clear explanation for that.

4 In long-term trials carried out on normotensive and spontaneously hypertensive  
5 rats, Perrot et al., (2003) showed that the extract from the low molecular weight fraction  
6 of a Champagne wine had antihypertensive activity in hypertensive rats but no effects in  
7 normotensive rats. However, owing to the complexity of this fraction, the authors  
8 indicated that this finding could not be only attributed to a single compound present in  
9 this fraction. Pozo-Bayón et al., (2005), showed that amino acids aspartic acid and/or  
10 asparagine and glutamic acid and/or glutamine and valine, formed part of 5 from the 6  
11 fractions from white and red wines with IACE activity, while threonine and alanine  
12 formed part of 4 of them.

13 Due to the abundance of phenols and peptides in wines, it is difficult to attribute  
14 the IACE activity to one of these groups since both of them include individual  
15 compounds with this activity (Ling, 2003; Zhang et al., 2003). Although Pozo-Bayón et  
16 al., (2005) demonstrated that red wine fractions with high peptide and relative low  
17 phenolic contents highly contributed to IACE activity, it has also been found that red  
18 wines have greater ACE inhibitory activity than white wines (Takayanagi and  
19 Yokotsuka, 1999), suggesting the contribution of phenolic compounds to this activity.  
20 Lin (2003) has shown that phenols can be conjugated with proteins and produce non-  
21 competitive inhibition of the ACE. It is, therefore, necessary to carry out more studies to  
22 identify the compounds responsible for this activity in wines. Recently, ACE inhibitory  
23 activity has also been found in peptides released from *Saccharomyces cerevisiae* in a  
24 model wine (Alcaide-Hidalgo et al., 2007b). In this assay, there were not phenolic  
25 compounds, thus the IACE activity was exclusively attributed to yeast peptides.

1 Moreover, the same rich peptides fractions also showed antioxidant activity, suggesting  
2 that peptides released by *S. cerevisiae* during autolysis under wine conditions could  
3 present multifunctional activities.

4         Recently, prolylendopeptidase inhibitory peptides (PEP) have also been found in  
5 wines (Yanai et al., 2003). PEP may have a role in the degradation of biologically-  
6 active peptides containing proline, such as oxytocin, vasopressin, P substance,  
7 bradykinin, neurotensins and angiotensins. Two inhibitory peptides have been isolated  
8 and characterized, Pep A (Val-Glu-Ile-Pro-Glu) and Pep B (Tyr, Pro, Ile, Pro, Phe).  
9 Both of them showed PEP inhibition thereby suppressing the degradation of  
10 neuropeptides, vasopressin, substance P and the fragments 8-13 of neurotensin, which  
11 are involved in memory and neural communication.

12

### 13 ***CONCLUSIONS AND FUTURE OUTLOOK***

14

15         The aim of the present review has been to gather, from an integrated perspective,  
16 the scientific information concerning wine health and safety, a matter of emergent  
17 interest for researchers and consumers.

18         There is a wide variety of compounds that can affect wine safety. However to  
19 solve these problems there are different strategies currently available. For example, it  
20 should be pointed out that a large part of today's advance in control and prevention  
21 strategies for wine contaminants has been possible thanks to the funding from several  
22 research frameworks (special EU research projects and other bilateral grants). Specific  
23 funds to increase the scientific knowledge related to the presence of OTA, biogenic  
24 amines and sulphur dioxide in wines, among others, have concluded in relevant

1 scientific outputs. As a consequence, the winemaking industry has specific knowledge  
2 currently available to control and/or prevent the formation of harmful compounds.

3         Great progress has also been done in recent years, trying to elucidate the  
4 compounds associated to the health benefits of moderate wine consumption and their  
5 action mode. Most of these advances have been focused on the study of wine phenolic  
6 compounds, confirming their key role in some healthy aspects derived from wine  
7 consumption.

8         Laboratory, but mainly clinical research, will continue exploring the efficacy of  
9 these important wine constituents in the human body, their bioavailability and action  
10 mechanisms. In the future, some promising applications of wine phenolics, such as their  
11 use in prevention or conventional therapies to fight cancer and infections will gain  
12 interest. Clinical studies of how the human body interact with wine constituents and  
13 how such interactions can be modulated, will continue. Moreover, there is growing  
14 evidence of the biological properties of other wine constituents, such as peptides.  
15 Nevertheless, due to their complexity and low abundance in wines in comparison with  
16 other fermented foods, the scientific information regarding peptides as bioactive agents  
17 in wines is still scarce. Therefore, it is foreseeable that in the next years some progress  
18 related to the identity and health functions of specific wine peptides would increase.

19         As a summary, it is possible to confirm for consumers, that utmost efforts are  
20 being made in research, technology and administrative control to ensure that wine is a  
21 safe, pleasant and healthy drink when consumed responsibly.

22  
23  
24  
25

1 **ACKNOWLEDGEMENTS**

2 The authors are grateful to AGL2006-04514, PET2007\_0134 and CSD2007-00063  
3 Consolider Ingenio 2010 FUN-C-FOOD Projects (Ministerio de Educación y Ciencia)  
4 and S-505/AGR-0153 ALIBIRD Project (Comunidad Autónoma de Madrid) for the  
5 financial support for this work. M.A. P.-B. and M.M. greatly acknowledge CSIC for  
6 their respective research contracts.

7  
8

9 **REFERENCES**

10

11 Abarca M.L.; Accensi F.; Cano J. and Cabañes F. (2004). Taxonomy and significance  
12 of black aspergilli. *Antonie Van Leeuwenhoek*. **86**:33-49.

13 Acedo, M.I.; Pueyo, E. and Polo, M.C. (1994). Preliminary studies on peptides in wine by  
14 HPLC. *Am. J. Enol. Vitic.*, **45**: 167-172.

15 Ajtonya, Z.; Szoboszlai, N. ; Suskó, E.K.; Mezeic, P. ; György K. and Bencs L.  
16 Direct sample introduction of wines in graphite furnace atomic absorption  
17 spectrometry for the simultaneous determination of arsenic, cadmium, copper and  
18 lead content. *Talanta*, **76**: 627-634.

19 Alberto, M.R.; Farias, M.E. and Manca de Nadra, M.C. (2001). Effect of gallic acid  
20 and catechin on *Lactobacillus hilgardii* 5w growth and metabolism of organic  
21 compounds. *J. Agric. Food Chem.*, **49**:4359-4363

22 Alcaide-Hidalgo J., Moreno-Arribas, M.V., Polo, M.C. and Pueyo, E. (2008). Partial  
23 characterization of peptides from red wines. Changes during malolactic fermentation  
24 and aging with lees. *Food Chem.*, **107**:622-630.

25 Alcaide-Hidalgo J.; Moreno-Arribas, M.V; Martín-Álvarez, P.J. and Polo, M.C. (2007a).  
26 Influence of malolactic fermentation, postfermentative treatments and ageing with  
27 lees on nitrogen compounds of red wines. *Food Chem.*, **103**: 572-581.

1 Alcaide-Hidalgo, J.M.; Pueyo, E.; Polo, M.C. and Martínez-Rodríguez, A.J. (2007b).  
2 Bioactive peptides released from *Saccharomyces cerevisiae* under accelerated  
3 autolysis in a model wine system. *J. Food Sci.*, **72** : 276-279

4 Alexandre, H.; Heintz, D.; Chassagne, D.; Guilloux-Benatier, M.; Charpentier, C. and  
5 Feuillat, M. (2001). Protease A activity and nitrogen fractions released during  
6 alcoholic fermentation and autolysis in enological conditions. *J. Ind. Microbiol.*  
7 *Biotechnol.*, **26**: 235-240.

8 Ancín-Azpilicueta, C.; González-Marco, A. and Jiménez-Moreno, N. (2008). Current  
9 knowledge about the presence of biogenic amines in wine. *Cr. Rev. Food Sci. Nutr.*  
10 **48**:257-27

11 Arena, M.E.; Saguir, F.M. and Manca de Nadra, M.C. (1999) Arginine, citrulline and  
12 ornithine metabolism by lactic acid bacteria from wine. *Int. J. Food Microbiol.* **52**:  
13 155-16

14 Artés-Hernández, F.; Artes, F. and Tomás-Barberán, F.A. (2003). Quality and  
15 enhancement of bioactive phenolics in Cv. Napoleon table grapes exposed to  
16 different postharvest gaseous treatments. *J. Agric. Food Chem.* **51**: 290-5295

17 Artés-Hernández, F.; Tomás-Barberán, F.A.; Artes, F.; (2006). Modified atmosphere  
18 packaging preserves quality of SO<sub>2</sub>-free 'Superior seedless' table grapes.  
19 *Postharvest Biol. Tec.* **39**: 146-154

20 Avellone, G.; Di Garbo, V.; Campisi, D.; De Simone, R. ; Raneli, G. ; Scaglione, R. and  
21 Licata, G. (2006). Effects of moderate Sicilian red wine consumption on  
22 inflammatory biomarkers of atherosclerosis. *Eur. J. Clin. Nutr.*, **60**: 41-47.

23 Bacaloni, A.; Cavaliere, C.; Faberi, A.; Pastoriani, E.; Samperi, R.; Laganá, A. (2005).  
24 Automated on-line solid phase extraction-liquid chromatography-electrospray

1 tandem mass spectrometry method for the determination of ochratoxin A in wine  
2 and beer. *J. Agric Food Chem.*, **53**:5518-5525.

3 Baderschneider, B. and Winterhalter, P. (2000). Isolation and characterization of novel  
4 stilbene derivatives from Riesling wine. *J. Agric. Food Agric.*, **48**: 2681-2686.

5 Badía, E.; Sacanella, E.; Fernández-Solá, J.; Nicolás, J.M.; Antúnez, E.; Rotilio, D.; de  
6 Gaetano, G.; Urbano-Márquez, A. and Estruch R. (2004). Decreased tumor necrosis  
7 factor-induced adhesion of human monocytes to endothelial cells after moderate  
8 alcohol consumption. *Am. J. Clin. Nutr.* **80**: 225-230.

9 Baluja-Santos, C. and Gonzalez Portal, A. (1992). Application of hydride generation to  
10 atomic-absorption spectrometric analysis of wines and beverages: a review. *Talanta*,  
11 **39**: 329-339.

12 Bartowsky, E. (2003). Lysozyme and winemaking. *Aus. J. Grape Wine Res.*, **473a**:101-  
13 104.

14 Battiliani, P.; Giorni, P. And Pietri, A. (2003). Epidemiology of toxin producing fungi  
15 and ochratoxin A occurrence in grape. *Eur. J. Plant Pathol.* **109**: 715-722

16 Battliani, P.; Magan, N.; Logrieco, A. (2006). European research on ochratoxin A in  
17 grapes and wine. *Int. J. Food Microbiol.*, **119**: 79-83.

18 Bauer, R., Hannes, A.N. and Dicks, L.M.T. (2003). Pediocin PD-1 as a method to  
19 control growth of *Oenococcus oeni* in wine. *Am. J. Enol. Vitic.*, **54**: 86-91

20 Bauer, R.; Chikindas, M.L. and Dicks, L.M.T. (2005). Purification, partial amino acid  
21 sequence and mode of action of pediocin PD-1, a bactericin produced by  
22 *Pediococcus damnosus* NCFB 1832. *Int. J. Food Microbiol.*, **101**:17-27

23 Bejaoui, H. ; Mathieu, F.; Taillandier, P. ; Lebrihi, A. (2004). Ochratoxin A removal in  
24 synthetic and natural grape juices by selected oenological *Saccharomyces* stains. *J.*  
25 *Appl. Microbiol.*, **97**: 1038-1044.

- 1 Bell, J.R.; Donovan, J.L.; Wong, R.; Waterhouse, A.L.; German, J.B.; Walzem, R.L.  
2 and Kasim-Karakas, S.E. (2000). (+)-Catechin in human plasma after ingestion of a  
3 single serving of reconstituted red wine. *Am. J. Clin. Nutr.*, **71**: 103-108.
- 4 Bellí, N., Marín, S., Coronas, I., Sanchis, V., & Ramos, A.J. (2007). Skin damage, high  
5 temperature and relative humidity as detrimental factors for *Aspergillus carbonarius*  
6 infection and ochratoxin A production in grapes. *Food Control*, **18**: 1343-1349.
- 7 Bellí, N.; Marin, S. ; Duaigues, A. ; Ramos, A.J.; Sanchis, V. (2004). Ochratoxin A in  
8 wines, musts and grape juices from Spain. *J. Sci. Food Agric.*, **84**:591-594.
- 9 Benicho, S.; Barbosa-Cánovas, G.V. and Martín, O. (2002). Milk processing by high  
10 intensity pulsed electric fields. *Trends Food Sci. Tech.*, **13**:195-204.
- 11 Bertelli, A.A.; Giovannini, L.; Stradi, R.; Urien, S. ; Tillement, J.P. and Bertelli, A.  
12 (1996). Kinetics of trans- and cis-resveratrol (3,4',5-trihydroxystilbene) after red  
13 wine oral administration in rats. *Int. J. Clin. Pharmacol. Res.*, **16**: 77-81.
- 14 Bertrand, A. (1997). Le carbamate d'éthyle dans les vins, observations sur son origine,  
15 possibilité d'en diminuer la teneur. Rapport OIV. Décembre 1997.
- 16 Blesa, J.; Soriano, J.M.; Moltó, J.C.; Mañes, J. (2006). Factors affecting the presence of  
17 ochratoxin A in wines. *Crit. Rev. Food Sci. Nutr.* **46**:473-478.
- 18 Bodmer, S. and Gafner, H. (2000). Histamin in wein. *Schweizerische Zeitschrift für*  
19 *Obst- und Weinbau* **21**:546-547.
- 20 Bornet A. and Teissedre P.L. (2008) Chitosan, chitin-glucan and chitin effects on  
21 minerals (iron, lead, cadmium) and organic (ochratoxin A) contaminants in wines.  
22 *Eur. Food Res. Technol.*, **226**: 681-689.
- 23 Bornet A. and Teissedre PL (2005). Applications and interest of chitin, chitosan and  
24 their derivatives in enology. *J. Int. Sci. Vigne Vin*, **39**: 199-207.

- 1 Bover-Cid, S., Izquierdo-Pulido, M., Mariné-Font, A. and Vidal-Carou, M.C. (2006).  
2 Biogenic, mono-, di- and polyamine contents in Spanish wines and influence of a  
3 limited irrigation. *Food Chem.*, **96**:43-47.
- 4 Braga, J.W.B.; Bottoli, C.B.G.; Jardim, I.C.S.F.; Goicoechea, H.C.; Oliveri, A.C. and  
5 Poppi, R.J. (2007). Determination of pesticides and metabolites in wine by high  
6 performance liquid chromatography and second-order calibration methods. *J.*  
7 *Chromatogr. A.* **1148**:200-210.
- 8 Brown S.T. (1991). Method of removing sulfites from standard wine. US patent,  
9 5,071,664.
- 10 Bruno S.N.F; Campos R.C and Curtius A.J. (1994). Determination of lead and arsenic  
11 in wines by electrothermal atomic-absorption spectrometry. *J. Anal. Atom. Spectrom.*,  
12 **9**: 341-344.
- 13 Bub, A.; Watzl, B.; Heeb, D.; Rechkemmer, G. and Briviba, K. (2001). Malvidin-3-  
14 glucoside bioavailability in humans after ingestion of red wine, dealcoholized red  
15 wine and red grape juice. *Eur. J. Nutr.*, **40**: 113-120.
- 16 Cabras, P. and Angioni, A. (2000). Pesticide residues in grapes, wine and their processing  
17 products. *J. Agric. Food Chem.*, **48**: 967-973.
- 18 Cabras, P. ; Garau, V.L.; Melis, M.; Pirisi, F.M. and Tuberoso, C.I.G. (1995). The effect  
19 of clarifying substances on organophosphorous insecticide residues in wine. *J. Wine*  
20 *Res.* **6**:201-205.
- 21 Campos, F.M.; Couto, J.A. and Hogg, T.A. (2003). Influence of phenolic acids on  
22 growth and inactivation of *Oenococcus oeni* and *Lactobacillus hilgardii*. *J. Appl.*  
23 *Microbiol.*, **94**:167-174.

1 Canas, B.J., Harvey, D.C.; Robinson, L.R.; Sullivan, M.P.; Joe F.L. Jr. and Diachenko,  
2 G.W., (1989). Ethyl carbamate levels in selected foods and beverages. *J. Ass. Offic.*  
3 *Anal. Chem.*, **72**: 873-876.

4 Caridi A. (2007). New perspectives in safety and quality enhancement of wine through  
5 selection of yeasts based on the parietal adsorption activity. *Int. J. Food Microbiol.*,  
6 **120**: 167-172.

7 Caruso, M., Fiore, C., Contrusi, M., Salzano, G., Paparella, A. and Romano, P. (2002).  
8 Formation of biogenic amines as criteria for the selection of wine yeast. *World J.*  
9 *Microbiol. Biotech.*, **18**:159-163.

10 Chulze, S.N.; Magnoli, C.E.; Dacero, A.M. (2006). Occurrence of ochroatoxin A and  
11 ochratoxigenic mycoflora in grapes and dried vine fruits in South America. *Int. J.*  
12 *Food Microbiol.*, **111**: S88-S92.

13 Constantini, A., Cersosimo, M., Del Prete, V. and García-Moruno, E. (2006).  
14 Production of biogenic amines by lactic acid bacteria: screening by PCR, thin-layer  
15 chromatography, and high-performance liquid chromatography of strains isolated  
16 from wine and must. *J. Food Prot.*, **69**: 391-396.

17 Coton, E., Rollan, G., Bertrand, A. and Lonvaud-Funel, A. (1998). Histamine-producing  
18 lactic acid bacteria in wines; early detection, frequency and distribution. *Am. J.*  
19 *Enol. Vitic.*, **49**:199-204.

20 Creppy, E.E. (2002). Update of survey, regulation and toxic effects of mycotoxins in  
21 Europe. *Toxicology Lett.*, **17**: 19-28.

22 Da Luz, P.L.; Serrano Júnior, C.V.; Chacra, A.P.; Monteiro, H.P.; Yoshida, V.M.;  
23 Furtado, M.; Ferreira, S.; Gutierrez, P. and Pileggi, F. (1999). The effect of red wine  
24 on experimental atherosclerosis: lipid-independent protection. *Exp. Mol. Pathol.*, **65**:  
25 150-159.

1 Da Rocha Rosa, C.A.; Palacios, V.; Combina, M.; Fraga, M.E.; De Oliveira Rekson,  
2 A.; Magnoli, C.E. and Dalcero, A.M. (2002). Potential ochratoxin A producers from  
3 wine grapes in Argentina and Brazil. *Food Addit. Contam.*, **19**: 408-414.

4 Dell'Agli, M.; Buscialà, A. and Bosisio, E. (2004). Vascular effects of wine  
5 polyphenols. *Cardiovasc Res.*, **63**: 593-602.

6 Del Prete, V.; Constantini, A.; Cecchini, F.; Morassut, M.; García-Moruno, E. (2008).  
7 Occurrence of biogenic amines in wine: The role of grapes. *Food Chem.*, **112**:474-  
8 481.

9 Delfini, C. (2004). Resistance screening assay of wine lactic acid bacteria on lysozyme:  
10 efficacy of lysozyme in unclarified grape musts. *J. Agric. Food Chem.*, **52**:1861-  
11 1866.

12 Demrow, H.S.; Slane, P.R. and Folts, J.D. (1995). Administration of wine and grape  
13 juice inhibits in vivo platelet activity and thrombosis in stenosed canine coronary  
14 arteries. *Circulation*, **91**: 1182-1188.

15 Desportes, C. ; Charpentier, M. ; Duteurtre, B., Maujean, A. and Duchiron, F. (2001).  
16 Isolation, identification, and organoleptic characterization of low-molecular-weight  
17 peptides from white wine. *Am. J. Enol. Vitic.*, **52**: 376-380.

18 Di Castelnuovo, A.; Rotondo, S.; Iacoviello, L.; Donati, M.B. and De Gaetano, G.  
19 (2002). Meta-analysis of wine and beer consumption in relation to vascular risk.  
20 *Circulation*, **105**: 2836-2844.

21 Diebolt, M.; Bucher, B. and Andriantsitohaina, R. (2001). Wine polyphenols decrease  
22 blood pressure, improve NO vasodilatation, and induce gene expression.  
23 *Hypertension*, **38**: 159-165.

- 1 Djoussé, L. ; Ellison, R.C. ; McLennan, C.E.; Cupples, L.A.; Lipinska, I.; Tofler, G.H.;
- 2 Gokce, N. and Vita, J.A. (1999). Acute effects of a high-fat meal with and without
- 3 red wine on endothelial function in healthy subjects. *Am. J. Cardiol.*, **84**: 660-664.
- 4 Divol, B. ; Strehaiano, P. and Lonvaud-Funel, A. (2005). Effectiveness of
- 5 dimethyldicarbonate to stop alcoholic fermentation in wine. *Food Microbiol.*,
- 6 **22** :169-178.
- 7 Donovan, J.L.; Bell, J.R.; Kasim-Karakas, S.; German, J.B.; Walzem, R.L.; Hansen,
- 8 R.J. and Waterhouse, A.L. (1999). Catechin is present as metabolites in human
- 9 plasma after consumption of red wine. *J. Nutr.*, **129**: 1662-1668.
- 10 Donovan, J.L.; Kasim-Karakas, S.; German, J.B. and Waterhouse, A.L. (2002). Urinary
- 11 excretion of catechin metabolites by human subjects after red wine consumption.
- 12 *Br. J. Nutr.*, **87**: 31-37.
- 13 Dos Santos A.M.; Feuillat M. and Charpentier C. (2000). Flor yeast metabolism in a
- 14 model system similar to cellar ageing of the french 'Vin jaune': Evolution of some
- 15 by-products, nitrogen compounds and polysaccharides. *Vitis*, **39**: 129-134.
- 16 Du Toit, M., du Toit, C., Krieling, S.J. and Pretorius, I.S. (2002). Bio-preservation of wine
- 17 with antimicrobial peptides. *Bull. OIV*, **855-856** :284-320.
- 18 Enrique, M.; Marcos, J.M.; Yuste, M.; Martínez, M.; Valles, S. and Manzanares, P.
- 19 (2007). Antimicrobial action of synthetic peptides towards wine spoilage yeasts.
- 20 *Int. J. Food Microbiol.*, **118**: 318-325.
- 21 Estruch, R.; Sacanella, E.; Badia, E.; Antúnez, E.; Nicolás, J.M.; Fernández-Solá, J.;
- 22 Rotilio, D.; de Gaetano, G.; Rubin, E. and Urbano-Márquez, A. (2004). Different
- 23 effects of red wine and gin consumption on inflammatory biomarkers of

1 atherosclerosis: a prospective randomized crossover trial. Effects of wine on  
2 inflammatory markers. *Atherosclerosis*. **175**: 117-123.

3 European Commission (2005). Commission Regulation No. 123/2005 of 26 of January  
4 of 2005 amending Regulation 466/2001 as regards ochratoxin A. Official Journal  
5 of the European Union L25, 3-5.

6 Felgines, C.; Talavéra, S.; Gonthier, M.P.; Texier, O.; Scalbert, A.; Lamaison, J.L. and  
7 Rémésy, C. (2003). Strawberry anthocyanins are recovered in urine as glucuro- and  
8 sulfoconjugates in humans. *J. Nutr.*, **133**: 1296-1301.

9 Fernández, M.J.; Oliva, J.; Barba, A. and Cámara, M.A. (2005). Fungicide dissipation  
10 curves in wine-making process with and without maceration step. *J. Agric. Food*  
11 *Chem.*, **53**:804-811.

12 Fernández-García, E., Tomillo, J. and Nuñez, M. (1999). Effect of added proteinases  
13 and level of starter culture on the formation of biogenic amines in raw milk  
14 Manchego cheese. *Int. J. Food Microbiol.*, **52**:189-196.

15 Ferrera, N. (1999). Molecular and biological properties of vascular endothelial growth  
16 factor. *J Mol Med.*, **77**: 527-543.

17 Ferreira, I.M. and Pinho, O. (2006). Biogenic amines in Portuguese traditional foods  
18 and wines. *J. Food Prot.* **69**:2293-2303.

19 Feuillat, M. ; Brillant, G. and Rochard, J. (1980). Mise en évidence d'une production de  
20 proteases exocellulaires par les levures au cours de la fermentation alcoolique du  
21 moût de raisin. *Conn. Vigne Vin*, **14**: 37-52.

22 Feuillat, M.; Bidan, P. and Rosier, V. (1977). Croissance des bactéries lactiques à partir  
23 des principaux constituants azotés du vin. *Ann. Technol. Agric.*, **28**: 435-447.

- 1 Fitzpatrick, D.F.; Fleming, R.C.; Bing, B.; Maggi, D.A. and O'Malley, R.M. (2000).  
2 Isolation and characterization of endothelium-dependent vasorelaxing compounds  
3 from grape seeds. *J. Agric. Food Chem.*, **48**: 6384-6390.
- 4 Flamini, R. and Panighel, A. (2006). Mass spectrometry in grape and wine chemistry.  
5 Part II: The consumer protection. *Mass Spectrom. Rev.*, **25**:741-744.
- 6 Franke, A.A.; Custer, L.J.; Cooney, R.V.; Tanaka, Y.; Xu, M.; Dashwood, R.H. (2002).  
7 Inhibition of colonic aberrant crypt formation by the dietary flavonoids (+)-catechin  
8 and hesperidin. *Adv. Exp. Med. Biol.*, **505**: 123-133.
- 9 Frankel, E.; Kanner, J.; German, J.B.; Parks, E.; Kinsella, J.E. (1993). Inhibition of  
10 oxidation of human low-density lipoprotein by phenolic substances in red wine.  
11 *Lancet*, **341**: 454-457.
- 12 Fulcrand, H.; Remy, S.; Souquet, J.M.; Cheynier, V. and Moutounet, M. (1999b). Study  
13 of wine tannin oligomers by on-line liquid chromatography electrospray ionization  
14 mass spectrometry. *J. Agric. Food Chem.*, **47**: 1023-1028.
- 15 Gambuti, A.; Strollo, D.; Genovese, A.; Ugliano, M.; Ritieni, A.; Moio, L. (2005).  
16 Influence of oenological practices on ochratoxin A concentration in wine. *Am. J.*  
17 *Enol. Vitic.*, **56**, 155-162.
- 18 Gao, Y.C.; Zhang, G.; Krentz, S.; Dariu, S.; Power, J. and Lagarde, G. (2002).  
19 Inhibition of spoilage lactic acid bacteria by lysozyme during wine alcoholic  
20 fermentation. *Aus. J. Grape Wine Res.*, **8**: 76-83;
- 21 García-Ruiz, A., Bartolomé, B., Martínez-Rodríguez, A., Pueyo, E., Martín-Álvarez,  
22 P.J. and Moreno-Arribas, M.V. (2008a). Potential of phenolic compounds for  
23 controlling lactic acid bacteria growth in wine. *Food Control*, **19**:835-841.

- 1 García-Ruiz, A., López-Expósito, I., Díaz, S., Bartolomé, B., Pozo-Bayón, M.A.,  
2 Martín-Álvarez, P.J. and Moreno-Arribas, M.V. (2008b). Evaluation of the dual  
3 antibacterial and antioxidant activities of wine polyphenols. *Proceeding of the*  
4 *'WAC2008' International conference*. David Chassagne, (Eds.) Oenoplurimedia,  
5 Francia, pp. 36-38.
- 6 Gil-Agustí, M., Carda-Brochm, S., Monferrer-Pons, L. and Esteve-Romero, J. (2007).  
7 Simultaneous determination of tyramine and tryptamine and their precursor amino  
8 acids by micellar liquid chromatography and pulsed amperometric detection in  
9 wines. *J. Chromatogr. A.*, **1156**:288-295.
- 10 Gómez-Alonso, S., Hermosín-Gutiérrez, U. and García-Romero, E. (2007).  
11 Simultaneous HPLC analysis of biogenic amines, amino acids and ammonium ion  
12 as animoenone derivatives in wine and beer simples. *J. Agric. Food Chem.*, **55**:608-  
13 613.
- 14 Gonthier, M.P.; Donovan, J.L.; Texier, O.; Felgines, C.; Remesy, C. and Scalbert, A.  
15 (2003). Metabolism of dietary procyanidins in rats. *Free Radic. Biol. Med.*, **35**: 837-  
16 844.
- 17 González-Llano, D.; Herraiz, T. and Polo, M.C. (2004). Peptides. In Leo M.L. Nollet  
18 (Ed), *Handbook of Food Analysis*, Vol. 1: Physical Characterization and Nutrient  
19 Analysis. Chapter 6 (pp 125-166). New York: Marcel Dekker.er.Grønbaek, M. (2000).  
20 Wine and mortality. Evidence for causal inference?. *Dan Med Bull.*, **47**: 271-282.
- 21 Gryglewski, R.J.; Korbut, R.; Robak, J. and Swies, J. (1987). On the mechanism of  
22 antithrombotic action of flavonoids. *Biochem. Pharmacol.*, **36**: 317-322.
- 23 Halász, A., Baráth, Á., Simon-Sarkadi, L. and Holzapfel, W. (1994). Biogenic amines  
24 and their production by microorganisms in food. *Trends Food Sci. Tech.*, **5**:42-49.

- 1 Hartmann R. and Meisela H. (2007) Food derived peptides with biological activity:  
2 from research to food applications. *Curr. Opin. Biotech.*, **18**: 163-169.
- 3 Hyötyläinen, T.; Lüthje, K.; Ratiainen-Rämä, M. and Riekkola, M.L. (2004).  
4 Determination of pesticides in red wines with on-line coupled microporous  
5 membrane liquid-liquid extraction-gas chromatography. *J. Chromatogr. A.*,  
6 **1056**:267-271.
- 7 IARC (International Agency for Research on Cancer) (1993). Some naturally occurring  
8 substances: food items and constituents, heterocyclic aromatic amines and  
9 mycotoxins (vol. 56). Lyon, France: World Health Organization.
- 10 Imhof, A.; Woodward, M.; Doering, A.; Helbecque, N.; Loewel, H.; Amouyel, P.;  
11 Lowe, G.D. and Koenig, W. (2004). Overall alcohol intake, beer, wine, and  
12 systemic markers of inflammation in western Europe: results from three MONICA  
13 samples (Augsburg, Glasgow, Lille). *Eur. Heart J.*, **25**: 2092-2100.
- 14 Izquierdo-Pulido, M., Mariné-Font, A. and Vidal-Carou, M.C. (2000). Effect of  
15 tyrosine on tyramine formation during beer fermentation. *Food Chem.*, **70**:329-332.
- 16 Jeandet, P.; Bessis, R. and Gautheron, B. (1991). The production of resveratrol (3,5,4'-  
17 trihydroxystilbene) by grape berries in different developmental stages. *Am J Enol*  
18 *Vitic.*, **42**: 41-46.
- 19 Jeandet P.; Breuil A.C.; Adrian M.; Weston L.A.; Debord S.; Meunier P.; Maume G.  
20 and Bessis R. (1997). HPLC analysis of grapevine phytoalexins coupling  
21 photodiode-array detection and fluorimetry. *Anal. Chem.* **69**:5172–5177
- 22 Jeandet, P.; Douillet.Breuil, A.C.; Bessis, R.; Debord, S.; Sbaghi, M. and Adrian, M.  
23 (2002). Phytoalexin from the Vitaceae: biosynthesis, phytoalexin gene expression in

1 transgenic plants, antifungal activity and metabolism *J. Agric Food Chem.*, **50**:  
2 2731-2341.

3 JECFA (Joint FAO/WHO Expert Committee of Food Additives) (2001). Ochratoxin A.  
4 In: *Safety Evaluation of Certain Mycotoxins in Food*. Prepared by the Fifty-sixth  
5 meeting of the JECFA. FAO Food and Nutrition Paper 74, Food and Agriculture  
6 Organization of the United Nations, Rome, Italy.

7 Jiménez, J.J.; Bernal, J.L.; del Nozal, M.J.; Toribio, L. and Bernal, J. (2001). Use of  
8 SPE-GC/EIMS for residue analysis in wine elaborated from musts spiked with  
9 formulations of chlorpyrifos-methyl, methiocarb, dicofol, and cyproconazol. *J.*  
10 *Sep. Sci.*, **30**:547-556

11 Kaniou, I., Samouris, G., Mouratidou, T., Eleftheriadou, A. and Zantopoulos, A. (2001).  
12 Determination of biogenic amines in fresh unpacked and vacuum-packed beef  
13 during storage at 4°C. *Food Chem.*, **74**:515-519

14 Klatsky, A.L.; Friedman, G.D.; Armstrong, M.A. and Kipp, H. (2003). Wine, liquor,  
15 beer, and mortality. *Am. J. Epidemiol.*, **158**: 585-595.

16 Korhammer, S.; Reniero, F. and Mattivi, F. (1995). An oligostilbene from *Vitis* roots.  
17 *Phytochem.*, **38**: 1501-1504.

18 Kurtbay, H.M.; Bekçi Z.; Merdivan M.; and Yurdakoç K. (2008). Reduction of  
19 ochratoxin a levels in red wine by bentonite, modified bentonites, and chitosan. *J.*  
20 *Agric. Food Chem.* **56**:2541-5.

21 Lamikanra, O.; Grimm, C.C.; Rodin, J.B. and Inyang, I.D. (1996). Hydroxylated  
22 stilbenes in selected american wines. *J. Agric. Food Chem.*, **44**: 1111-1115.

23 Landete, J.M., de las Rivas, B., Marcobal, A. and Muñoz, R. (2007). Molecular methods  
24 for the detection of biogenic amine-producing bacteria on foods. *Int. J. Food*  
25 *Microbiol.*, **117**: 258-269.

- 1 Landete, J.M; Ferrer, S. and Pardo, I. (2005). Which lactic acid bacteria are responsible  
2 of histamine production in wine?. *J. Appl. Microbiol.*, **99**:580-586.
- 3 La Pera, L.; Dugo, C.; Rando, R.; Di Bella, G.; Maisano, R. and Salvo (2008).  
4 Statistical analysis of the influence of fungicide treatments (mancozeb, zoxamide  
5 and copper oxychloride) on heavy metal concentrations in Sicilian red wine. *Food*  
6 *Addit. Contam.*, **25**: 302-313.
- 7 Lapidot, T.; Harel, S.; Granit, R.; Kanner, J. (1998). Bioavailability of red wine  
8 anthocyanins as detected in human urine. *J. Agric. Food Chem.* 46: 4297-4302.
- 9 Lavy, A.; Fuhrman, B.; Markel, A.; Dankner, G.; Ben-Amotz, A.; Presser, D. and  
10 Aviram, M. (1994). Effect of dietary supplementation of red or white wine on  
11 human blood chemistry, hematology and coagulation: favorable effect of red wine  
12 on plasma high-density lipoprotein. *Ann. Nutr. Metab.* **38**: 287-294.
- 13 Lehtonen, P. (1996). Determination of amines and amino acids in wine – a review. *Am.*  
14 *J. Enol. Vitic.*, **47**:127-133.
- 15 Leitáo, M.C.; Teixeira, H.C.; Barreto Crespo, M.T. and San Romáo, M.V. (2000).  
16 Biogenic amines occurrence in wine. Amino acid decarboxylase and proteolytic  
17 activities expression by *Oenococcus oeni*. *J. Agric. Food Chem.*, **48**: 2780-2784.
- 18 Leong, L.S., Hocking, A.D. and Scott, E.S. (2006b). The effect of juice clarification,  
19 static or rotary fermentation and fining on ochratoxin A in wine. *Aust. J. Grape*  
20 *Wine Res.*, **12**: 245-252.
- 21 Leong, L.S., Hocking, A.D., Pitt, J.I., Kazi, B.A., Emmett, R.W., and Scott, E.S.  
22 (2006a). Black *Aspergillus* Species in Australian Vineyards: From Soil to  
23 Ochratoxin A in Wine. In: Hocking, A.D., Pitt, J.I., Samson, R.A., Thrane, U.  
24 (eds). *Adv. Exp. Med. Biol.*, **571**:153-171.

- 1 Lin S.-C. and George G. (2004). A biocatalyst for the removal of sulfite from alcoholic  
2 beverages. *Biotechnol. Bioeng.*, **89**: 123-127.
- 3 Lin, J.Y. (2003). Antihypertensive effects of tannins isolated from traditional Chinese  
4 herbs as non-specific inhibitors of angiotensin converting enzyme. *Life Sci.*, **73**:  
5 1543-1555.
- 6 Liu, S.Q.; Pritchard, G.G.; Hardman, M.J. and Pilone, G.J. (1995) Occurrence of  
7 arginine deiminase pathway enzymes in arginine catabolism by wine lactic acid  
8 bacteria. *Appl. Environ. Microbiol.*, **61**: 310-316.
- 9 Liu, S.-Q.; Pritchard, G.G.; Hardman, M.J. and Pilone, G.J. (1996). Arginine catabolism  
10 in wine lactic acid bacteria: is it via the arginine deiminase pathway or the arginase-  
11 urease pathway?, *J. Appl. Bacteriol.*, **81**: 486-492.
- 12 Lonvaud-Funel, A. (1999). Biogenic amines in wines: role of lactic acid bacteria.  
13 *FEMS Microbiol. Lett.*, **199**:9-13.
- 14 López de Cerain, A.; González-Peñas, E.; Jiménez, A.M.; Bello, J. (2002). Contribution  
15 of the study of ochratoxin A in Spanish wines. *Food Addit. Contam.* **19**, 1058-1064.
- 16 Lourenço, C.F.; Gago, B.; Barbosa, R.M.; de Freitas, V and Laranjinha, J. (2008). LDL  
17 isolated from plasma-loaded red wine procyanidins resist lipid oxidation and  
18 tocopherol depletion. *J. Agric Food Chem.*, **56**: 3798-3804.
- 19 Lucas, P.M. ; Claisse, O. ; Lonvaud-Funel, A. (2008). High frequency of histamine-  
20 producing bacteria in the enological environment and instability of the histidine  
21 decarboxylase production phenotype. *Appl. Environ. Microbiol.*, **74**:81-817
- 22 Lustrato, G.; Alfano, G.; Belli, C.; Grazia, L.; Iorizzo, M.; Maiuro, L.; Massarella, F. ;  
23 Zanardini, E. and Ranalli, G.(2003). Controlling grape must fermentation in early  
24 winemaking phases: the role of electrochemical treatment. *J. Appl. Microbiol.*,  
25 **95**:1087-1095.

1 Malherbe, D.F.; Du Toit, M.; Cordero, R.R., Van Rensburg, P. and Pretorius, I.S.  
2 (2003). Expression of the *Aspergillus niger* glucose oxidase gene in *Saccharomyces*  
3 *cerevisiae* and its potential application in wine production. *Appl. Environ.*  
4 *Biotechnol.*, **62**:205-511.

5 Manach, C.; Scalbert, A.; Morand, C.; Rémésy, C. and Jiménez L. (2004). Polyphenols:  
6 food sources and bioavailability. *Am. J. Clin Nutr.*, **79**: 727-747.

7 Manca de Nadra, M.C., Farias, M., Moreno-Arribas, M.V., Pueyo, E., and Polo, M.C.  
8 (1997). Proteolytic activity of *Leuconostoc oenos*: Effect on proteins and polypeptides  
9 from white wine. *FEMS Microbiol. Lett.*, **150**: 135-139.

10 Manca de Nadra, M.C.; Farias, M.E.; Moreno-Arribas, V.; Pueyo, E. and Polo, M.C.  
11 (1999). A proteolytic effect of *Oenococcus oeni* on the nitrogenous macromolecular  
12 fraction of red wine. *FEMS Microbiol. Lett.*, **174**: 41-47.

13 Manca de Nadra, M.C.; Farias, M.E.; Pueyo, E. and Polo, M.C. (2005). Protease activity  
14 of *Oenococcus oeni* viable cells on red wine nitrogenous macromolecular fraction in  
15 presence of SO<sub>2</sub> and ethanol. *Food Control*, **16**: 851-854.

16 Marcobal, A., de las Rivas, B., Moreno-Arribas, M.V. and Muñoz, R. (2004).  
17 Identification of the ornithine decarboxylase gene in the putrescine-producer  
18 *Oenococcus oeni* BIFI-83. *FEMS Microbiol. Lett.*, **239**, 213-220.

19 Marcobal, A., de las Rivas, B., Moreno-Arribas, M.V. and Muñoz, R. (2005b). Multiplex-  
20 PCR method for the Simultaneous Detection of Acid Lactic Bacteria Producing  
21 Histamine, Tyramine and Putrescine, Three Major Biogenic Amines. *J. Food Prot.*,  
22 **68**: 874-878.

23 Marcobal, A., de las Rivas, B., Moreno-Arribas, M.V. and Muñoz, R. (2006b).  
24 Evidence for horizontal gene transfer as origin of putrescine-production in  
25 *Oenococcus oeni* RM83. *Appl. Environ. Microbiol.*, **72**: 7954-7958.

- 1 Marcobal, A., Martín-Álvarez, P.J., Polo, M.C., Muñoz, R., and Moreno-Arribas, M.V.  
2 (2006a). Formation of biogenic amines throughout the industrial manufacture of red  
3 wine. *J. Food Prot.*, **69**:391-396.
- 4 Marcobal, A., Polo, M.C., Martín-Álvarez, P.J. and Moreno-Arribas, M.V. (2005a).  
5 Biogenic amines content of red Spanish wines. Comparison of a Direct ELISA and an  
6 HPLC method for the determination of histamine in wines. *Food Res. Int.*, **38**, 387-  
7 394.
- 8 Marques, A.P., Leitao, M.C. and San Romao, M.V. (2008). Biogenic amines in wines:  
9 Influence of oenological factors. *Food Chem.*, **107**: 853-860.
- 10 Martín-Álvarez, P.J., Marcobal, A., Polo, C. and Moreno-Arribas, M.V. (2006).  
11 Technological factors influencing biogenic amine production during red wine  
12 manufacture. *Eur. Food Res. Technol.*, **222**, 420-424.
- 13 Martínez-Cuesta, M.C.; Requena-Rolania, M.T.; Pelaez-Martínez, M.C. (2003).  
14 Bacteriocin-forming *Lactococcus lactis* suitable for use as a starter for acceleration  
15 of ripening of cheese. PN: ES 2 170 723 BA
- 16 Martínez-Rodríguez, A.J. and Polo, M.C. (2000). Characterization of the nitrogen  
17 compounds released during yeast autolysis in a model wine system. *J. Agric. Food*  
18 *Chem.*, **48**: 1081-1085.
- 19 Martínez-Rodríguez, A.J.; Carrascosa, A.V. and Polo, M.C. (2001). Release of nitrogen  
20 compounds to the extracellular medium by three strains of *Saccharomyces cerevisiae*  
21 during induced autolysis in a model wine system. *Int. J. Food Microbiol.*, **68**: 155-  
22 160.
- 23 Mateo, R., Medina, A., Mateo, E.M., Mateo, F., and Jiménez, M. (2007). An overview  
24 of ochratoxin A in beer and wine. *Int. J. Food Microbiol.*, **119**:79-83.

1 Mattivi, F.; Reniero, F. and Korhammer, S. (1995). Isolation, characterization, and  
2 evolution in red wine vinification of resveratrol monomers. *J. Agric. Food Chem.*,  
3 **43**: 1820-1823.

4 Matsumoto, H.; Inaba, H.; Kishi, M.; Tominaga, S.; Hirayama, M. and Tsuda, T.  
5 (2001). Orally administered delphinidin 3-rutinoside and cyanidin 3-rutinoside are  
6 directly absorbed in rats and humans and appear in the blood as the intact forms. *J.*  
7 *Agric. Food Chem.*, **49**: 1546-1551.

8 Maynard, L.S. and Schenker, V.J. (1996). Monoamine-oxidase inhibition by ethanol in  
9 vitro. *Nature*, **196**:575-576.

10 Mazzoli, R.; Lamberti, C.; Coisson, J.D.; Purroti, M.; Arlorio, M.; Giuffrida, M.G.,  
11 Gounta, C. and Pessione, E. (2008). Influence of ethanol, malate and arginine on  
12 histamine production of *Lactobacillus hilgardii* isolated from an Italian red wine.  
13 *Amino acids* (in press, doi: 10.1007/s00726-008-0035-8).

14 McGhie, T.K.; Ainge, G.D.; Barnett, L.E.; Cooney, J.M. and Jensen, D.J. (2003).  
15 Anthocyanin glycosides from berry fruit are absorbed and excreted unmetabolized  
16 by both humans and rats. *J. Agric. Food Chem.*, **51**: 4539-4548.

17 Miyagi, Y.; Miwa, K. and Inoue, H. (1997). Inhibition of human low-density lipoprotein  
18 oxidation by flavonoids in red wine and grape juice. *Am. J. Cardiol.*, **80**: 1627-1631.

19 Mukamal, K.J.; Kronmal, R.A.; Mittleman, M.A.; O'Leary, D.H.; Polak, J.F.; Cushman,  
20 M. and Siscovick, D.S. (2003) Alcohol consumption and carotid atherosclerosis in  
21 older adults: the Cardiovascular Health Study. *Arterioscler Thromb Vasc Biol.*, **23**:  
22 2252-2259.

23 Millán, S., Sanpedro, M.C., Unceta, N., Goicolea, M.A. and Barrio, R.J. (2007). Simple  
24 and rapid determination of biogenic amines in wine by liquid chromatography-

1       electrospray ionization ion trap mass spectrometry. *Anal. Chim. Acta*, **584**:145-  
2       152.

3       Mine, Y. and Zhang, J.W. (2002). Comparative studies on antigenity and allergenicity of  
4       native and denatured egg white proteins. *J. Agric. Food Chem.*, **50** :2679-2683

5       Mira de Orduña, R.; Liu, S.-Q. ; Patchet, M.L. and Pilone, G.J. (2001). Growth and  
6       arginine metabolisms of the wine lactic acid bacteria *Lactobacillus buchneri* and  
7       *Oenococcus oeni* at different pH values and arginine concentrations. *Appl. Environ.*  
8       *Microbiol.*, **67** :1657-1662.

9       Molina-Mayo, C.; Hernández-Borges, J.; Borges-Miquel, T.M. and Rodríguez-Delgado,  
10       M.A. (2006). Determination of pesticides in wine using micellar electrokinetic  
11       chromatography with UV detection and simple stacking. *J. Chromatogr. A*. **25**: 348-  
12       355.

13       Monteiro, F. and Bisson, L.F. (1991). Amino acid utilization and urea formation during  
14       vinification fermentations. *Am. J. Enol. Vitic.*, **42**:199-208.

15       Moreno-Arribas, M. V., Polo, M.C., Jorganes, F. and Muñoz, R. (2003). Screening of  
16       biogenic amine production by lactic acid bacteria . 2003. Screening of biogenic  
17       amine production by lactic acid bacteria isolated from grape must and wine. *Int. J.*  
18       *Food Microbiol.*, **84**:117-123.

19       Moreno-Arribas, M.V. and Polo, C. (2008). Occurrence of lactic acid bacteria and  
20       biogenic amines in biologically aged wines. *Food Microbiol.*, **25**:875-881.

21       Moreno-Arribas, M.V.; Bartolomé, B., Pueyo, E. and Polo, M.C. (1998a). Isolation and  
22       characterization of individual peptides from wine. *J. Agric. Food Chem.*, **46**: 3422-  
23       3425.

- 1 Moreno-Arribas, V., Pueyo, E., Polo, M.C. and Martín-Álvarez, P.J. (1998b). Changes  
2 in the amino acid composition of the different nitrogenous fractions during the aging of  
3 wine with yeast. *J. Agric. Food Chem.*, **46**: 4042-4051.
- 4 Moreno-Arribas, V., Torlois, S., Joyeux, A., Bertrand, A. and Lonvaud-Funel, A.  
5 (2000). Isolation, properties and behaviour of tyramine-producing lactic acid  
6 bacteria from wine. *J. Appl. Microbiol.*, **88**:584-593.
- 7 Moreno-Arribas, V.; Pueyo, E. and Polo, M.C. (1996). Peptides in musts and wines.  
8 Changes during the manufacture of cava (sparkling wines). *J. Agric. Food Chem.*, **44**:  
9 3783-3788.
- 10 Muñoz, A. and Marcos, J.F. (2006). Activity and mode of action against fungal  
11 phytopathogens of bovine lactoferricin-derived peptides. *J. Appl. Microbiol.* **101**:  
12 1199-1207.
- 13 Navarro, L.; Zarazaga, M.; Sáenz, J.; Ruiz-Larrea, F. and Torres, C. (2002). Bacteriocin  
14 production by lactic acid bacteria isolated from Rioja red wines. *J. Appl.*  
15 *Microbiol.* **88**: 44-51.
- 16 Navarro, S.; García, B.; Navarro, G.; Oliva, J. and Barba, A. (1997). Effect of  
17 winemaking practices on the concentrations of fenarimol and penconazole in rose  
18 wines. *J. Food Prot.*, **60**:1120-1124.
- 19 Nigdikar, S.V.; Williams, N.R.; Griffin, B.A. and Howard, A.N. (1998). Consumption  
20 of red wine polyphenols reduces the susceptibility of low-density lipoproteins to  
21 oxidation in vivo. *Am. J. Clin. Nutr.*, **68**: 258-165.
- 22 Nozal, M.J.; Bernal, J.L.; Jiménez, J.J.; Martín, M.T. and Bernal, J. (2005).  
23 Determination of azolic fungicides in wine by solid-phase extraction and high-  
24 performance liquid chromatography-atmospheric pressure chemical ionization-mass  
25 spectrometry. *J. Chromatogr. A.*, **27**:90-96.

1 Ndiaye, M.; Chataigneau, T.; Andriantsitohaina, R.; Stoclet, J.C. and Schini-Kerth, V.B.  
2 (2003). Red wine polyphenols cause endothelium-dependent EDHF-mediated  
3 relaxations in porcine coronary arteries via a redox-sensitive mechanism. *Biochem.*  
4 *Biophys. Res. Commun.*, **310**: 371-377.

5 OIV (2005). Code of sound vitivinicultural practices in order to minimise levels of  
6 Ochratoxin A in vine-based products. Resolution Viti-Oeno 1/2005. Assessed at  
7 [http://news.reseau-concept.net/images/oiv\\_uk/Client/VITI-OENO\\_1-](http://news.reseau-concept.net/images/oiv_uk/Client/VITI-OENO_1-2005_EN.pdf)  
8 [2005\\_EN.pdf](http://news.reseau-concept.net/images/oiv_uk/Client/VITI-OENO_1-2005_EN.pdf).

9 Olivares-Marín, M.; Del Prete, V.; Garcia-Murno, E.; Fernández-González, C.; Macías-  
10 García, A. and Gómez-Serrano, V. (2008). The development of an activated  
11 carbon from cherry stones and its use in the removal of ochratoxin A from red  
12 wine. *Food Control*, doi: 10.1016/j.foodcont.2008.05.008.

13 Ottener, H., and Majerus, P. (2000). Occurrence of ochratoxin A (OTA) in wines:  
14 Influence of the type of wine and its geographical origin. *Food Addit. Contam.*,  
15 **17**: 793-798.

16 Ough, C.S.; Huang, Z.; An, D. and Stevens, D. (1991) Amino acid uptake by four  
17 commercial yeasts at two different temperatures of growth and fermentation: effects  
18 on urea excretion and reabsorption. *Am. J. Enol. Vitic.*, **42**: 26-40.

19 Pace-Asciak, C.R.; Rounova, O.; Hahn, S.E.; Diamandis, E.P. and Goldberg, D.M.  
20 (1996). Wines and grape juices as modulators of platelet aggregation in healthy  
21 human subjects. *Clin Chim Acta.*, **246**: 163-182.

22 Palumbo, J.D.; O’Keeffe, T.L. and Mahoney, N.E. (2007). Inhibition of ochratoxin A  
23 production and growth of *Aspergillus* species by phenolic antioxidant compounds.  
24 *Mycopathologia*, **164**: 241-248.

1 Papamichael, C.; Karatzis, E.; Karatzi, K.; Aznaouridis, K.; Papaioannou, T.;  
2 Protogerou, A.; Stamatelopoulos, K.; Zampelas, A.; Lekakis, J. and Mavrikakis, M.  
3 (2004). Red wine's antioxidants counteract acute endothelial dysfunction caused by  
4 cigarette smoking in healthy nonsmokers. *Am. Heart. J.*, **147**: 274.

5 Pedersen, G.A.; Mortensen, G.K. and Larsen E.H. (1994). Beverages as a source of  
6 toxic trace element intake. *Food Addit. Contam.*, **11**: 351-363.

7 Pellegrini, A. (2003). Antimicrobial peptides from food proteins. *Curr. Pharm. Design*,  
8 **9**:1225-1238.

9 Pellegrini, N.; Pareti, F.I.; Stabile, F.; Brusamolino, A. and Simonetti P. (1996). Effects  
10 of moderate consumption of red wine on platelet aggregation and haemostatic  
11 variables in healthy volunteers. *Eur. J. Clin. Nutr.*, **50**: 209-213.

12 Perrot, L.; Dukic, S.; Charpentier, M; Duteurtre, B.; Duchiron, F. and Kaltenbach, M.L.  
13 (2003). Antihypertensive effect of a low molecular weight fraction (1 kDa) of  
14 champagne wine in spontaneously hypertensive rats. In. A. Lonvaud-Funel, G.  
15 Revel, P. Darriet (Eds), *Oenologie 2003* (pp 688-691) Paris: TEC and DOC.

16 Person, M. de; Sevestre, A.; Chaimbault, P. ; Perrot, L. ; Duchiron, F. and Elfakir, C.  
17 (2004). Characterization of low-molecular weight peptides in champagne wine by  
18 liquid chromatography/tandem mass spectrometry. *Anal. Chim. Acta*, **520**: 149-158.

19 Pilatte, E.; Nygaard, M.; Gao, Y.C.; Krentz, S.; Power, J. and Lagarde, G. (2000).  
20 Studies on effects of lysozyme on different strains of *Oenococcus oeni*. Applications  
21 in control of malolactic fermentation. *Rev. Fr. Oenol.*, **185**: 26-29.

22 Pignatelli, P.; Ghiselli, A.; Buchetti, B.; Carnevale, R.; Natella, F.; Germanò, G.;  
23 Fimognari, F.; Di Santo, S.; Lenti, L. and Violi F. (2006). Polyphenols  
24 synergistically inhibit oxidative stress in subjects given red and white wine.  
25 *Atherosclerosis*, **188**: 77-83.

1 Pignatelli, P.; Lenti, L.; Pulcinelli, F.M.; Catasca, R.; Saccani, G.; Germanò, G.;  
2 Marcoccia, A.; Silvestri, M.A.; Ghiselli, A. and Violi, F. (2002). Red and white  
3 wine differently affect collagen-induced platelet aggregation. *Pathophysiol*  
4 *Haemost. Thromb.*, **32**: 356-358.

5 Pozo-Bayón, M.A., Alcaíde, J.M. ; Polo, M.C., and Pueyo, E. (2005). Angiotensin I-  
6 converting enzyme inhibitory compounds in white and red wines. *Food Chem.*, **100**:  
7 43-47.

8 Pyrzynska K. (2004). Analytical Methods for the Determination of Trace Metals in  
9 Wine *Crit Rev. Anal. Chem*, **34**: 69-83.

10 Radler, F. (1990). Possible use of nisin in winemaking. II. Experiments to control lactic  
11 acid bacteria in the production of wine. *Am. J. Enol. Vitic.* **41**:7-11

12 Ranalli, G.; Iorizo, M.; Lustrato, G.; Zanardii, E. and Grazia, L. (2002). Effects of low  
13 electric treatments on yeast microflora. *J. Appl. Microbiol.*, **93**: 877-883.

14 Rein, D.; Paglieroni, T.G.; Pearson, D.A.; Wun, T.; Schmitz, H.H.; Gosselin, R. and  
15 Keen, C.L. (2000). Cocoa and wine polyphenols modulate platelet activation and  
16 function. *J Nutr.*, **130**: 2120S-2126S.

17 Remize, F.; Augagneur, Y.; Guilloux-Benatier, M. and Guzzo, J. (2005). Effect on  
18 nitrogen limitation and nature of the feed upon *Oenococcus oeni* metabolism and  
19 extracellular protein production. *J. Appl. Microbiol.*, **98**: 652-661.

20 Remize, F.; Gaudin, A.; Kong, Y.; Guzzo, J.; Alexandre, H.; Krieger, S. and Guilloux-  
21 Benatier, M. (2006). *Oenococcus oeni* preference for peptides: Qualitative  
22 assimilation and quantitative analysis of nitrogen assimilation. *Arch. Microbiol.*,  
23 **185**: 459-469.

24 Renaud, S. and de Lorgeril, M. (1992). Wine, alcohol, platelets, and the French paradox  
25 for coronary heart disease. *Lancet*, **339**: 1523-1526.

- 1 Ribeiro de Lima, M.T.; Waffo-Téguo, P.; Teissedre, P.L.; Pujolas, A.; Vercauteren, J.;  
2 Cabanis, J.C. and Mérillon, J.M. (1999). Determination of stilbenes (trans-astringin,  
3 cis- and trans-piceid, and cis- and trans-resveratrol) in portuguese wines. *J. Agric.*  
4 *Food Chem.*, **47**: 2666-2670.
- 5 Riganakos K. A. and Veltsistas P. G. (2003). Comparative spectrophotometric  
6 determination of the total iron content in various white and red Greek wines. *Food*  
7 *Chem.*, **82**: 637-643.
- 8 Rimm, E.B.; Williams, P.; Fosher, K.; Criqui, M. and Stampfer, M.J. (1999). Moderate  
9 alcohol intake and lower risk of coronary heart disease: meta-analysis of effects on  
10 lipids and haemostatic factors. *BMJ.*, **319**: 1523-1528.
- 11 Ringot, D., Chango, A., Schneider, Y.J., and Larondelle, Y. (2006). Toxicokinetics and  
12 toxicodynamics of ochratoxin A, an update. *Chem. Biol.*, **159**: 18-46.
- 13 Rios, L.Y.; Bennett, R.N.; Lazarus, S.A.; Rémésy, C.; Scalbert, A.; Williamson, G.  
14 (2002). Cocoa procyanidins are stable during gastric transit in humans. *Am. J.*  
15 *Clin. Nutr.*, **76**: 1106-1110.
- 16 Rojo-Bezares, B.; Sáez, Y.; Zarazaga, M.; Torres, C. and Ruiz-Larrea, F. (2007).  
17 Antimicrobial activity of nisin against *Oenococcus oeni* and other wine bacteria.  
18 *Int. J. Food Microbiol.* **116**:32-36.
- 19 Rosenkranz, S.; Knirel, D.; Dietrich, H.; Flesch, M.; Erdmann, E. and Böhm, M. (2002).  
20 Inhibition of the PDGF receptor by red wine flavonoids provides a molecular  
21 explanation for the "French paradox". *FASEB J.*, **16**: 1958-1960.
- 22 Rozès, N. and Perez, C. (1998). Effects of phenolic compounds on the growth and the  
23 fatty acid composition of *Lactobacillus plantarum*. *Appl. Microbiol. Biotechnol.*,  
24 **49**:108-111.

- 1 Russo, P.; Tedesco, I.; Russo, M.; Russo, G.L.; Venezia, A. and Cicala, C. (2001).  
2 Effects of de-alcoholated red wine and its phenolic fractions on platelet  
3 aggregation. *Nutr. Metab. Cardiovasc Dis.*, **11**: 25-29.
- 4 Rydlo, T., Miltz, J. and Mor, A. (2006). Eukaryotic antimicrobial peptides: Promises  
5 and premises in food safety. *J. Food Sci.* **71**: R125-R135.
- 6 Sacanella, E.; Vázquez-Agell, M.; Mena, M.P.; Antúnez, E.; Fernández-Solá, J.;  
7 Nicolás, J.M.; Lamuela-Raventós, R.M.; Ros, E. and Estruch, R. (2007). Down-  
8 regulation of adhesion molecules and other inflammatory biomarkers after  
9 moderate wine consumption in healthy women: a randomized trial. *Am. J. Clin.*  
10 *Nutr.*, **86**: 1463-1469.
- 11 Saito, M.; Hosoyama, H.; Ariga, T.; Kataoka, S.; Yamaji, N. (1998). Antiulcer Activity  
12 of Grape Seed Extract and Procyanidins. *J. Agric. Food Chem.*, **46**: 1460-1464.
- 13 Sano, A.; Yamakoshi, J.; Tokutake, S.; Tobe, K.; Kubota, Y. and Kikuchi, M. (2003).  
14 Procyanidin B1 is detected in human serum after intake of proanthocyanidin-  
15 rich grape seed extract. *Biosc. Biotechnol. Biochem.*, **67**: 1140-1143.
- 16 Santos, B. Simonet, B.M., Rios, A. and Valcárcel, M. (2004). Direct automatic  
17 determination of biogenic amines in wine by flow injection-capillary  
18 electrophoresis-mass spectrometry. *Electrophoresis*, **25**:3427.
- 19 Scalbert, A. and Williamson, G. (2000). Dietary intake and bioavailability of  
20 polyphenols. *J Nutr.*, **130**: 2073S-2085S.
- 21 Schoeman H.; Vivier M.A.; Du Toit, M., Dicks, L.M. and Pretorius I.S. (1999). The  
22 development of bactericidal yeast strains by expressing the *Pediococcus acidilactici*  
23 pediocin gene (pedA) in *Saccharomyces cerevisiae*. *Yeast*, **15**(8):647-56.

1 Schramm, D.D.; Pearson, D.A. and German, J.B. (1997). Endothelial cell basal PGI(2)  
2 release is stimulated by wine in vitro: One mechanism that may mediate the  
3 vasoprotective effects of wine. *J. Nutr. Biochem*, **8**: 647-651.

4 Seifter E.; Padawer, J. and Lalezari, I. (1994). Method and system for removing  
5 impurities from aliments. US patent 5,358,732.

6 Sen, C.K. and Bagchi, D. (2001). Regulation of inducible adhesion molecule expression  
7 in human endothelial cells by grape seed proanthocyanidin extract. *Mol. Cell.*  
8 *Biochem.*, **216**: 1-7.

9 Serafini, M.; Maiani, G.; Ferro-Luzzi, A. (1998). Alcohol-free red wine enhances  
10 plasma antioxidant capacity in humans. *J. Nutr.*, **128**: 1003-1007.

11 Serra, R., Cabañes, J.F., Perrone, G., Castellá, G., Venâncio, A., Mulè, G., and  
12 Kozakiewicz, Z. (2006). *Aspergillus ibericus*: A new species of section *Nigri*  
13 isolated from grapes. *Mycologia*, **98**: 295-306.

14 Simó, C., Moreno-Arribas, M.V. and Cifuentes, A. (2008). Ion-trap vs. Time-of-flight  
15 mass spectrometry coupled to capillary electrophoresis to analyze biogenic amines  
16 in wine. *J. Chromatogr. A* , **1195**:150-156.

17 Smit, A.Y. (2007). Evaluating the influence of winemaking practices on biogenic amine  
18 production by wine microorganisms. Master Thesis. Stellenbosch University.  
19 Stellenbosch, South Africa.

20 Spaak, J.; Merlocco, A.C.; Soleas, G.J.; Tomlinson, G.; Morris, B.L.; Picton, P.;  
21 Notarius, C.F.; Chan, C.T. and Floras, J.S. (2008). Dose-related effects of red wine  
22 and alcohol on hemodynamics, sympathetic nerve activity, and arterial diameter. *Am*  
23 *J. Physiol. Heart Circ. Physiol.*, **294**: H605-H612.

- 1 Soleas, G.C. and Goldberg, D.M. (2000). Potential role of clarifying agents in the  
2 removal of pesticide residues during wine production and their effects upon wine  
3 quality. *J. Wine Res.* **11**:19-34.
- 4 Soufleros, E., Barrios, M.L. and Bertrand, A. (1998). Correlation between the content of  
5 biogenic amines and other wine compounds. *Am. J. Enol. Vitic.*, **49**:266-278.
- 6 Stead, D. (1993). The effect of hydroxycinnamic acids on the growth of wine-spoilage  
7 lactic acid bacteria. *J. Appl. Bacteriol.*, **75** :135-141.
- 8 Stein, J.H.; Keevil, J.G.; Wiebe, D.A.; Aeschlimann, S. and Folts, J.D. (1999). Purple  
9 grape juice improves endothelial function and reduces the susceptibility of LDL  
10 cholesterol to oxidation in patients with coronary artery disease. *Circulation*, **100**:  
11 1050-1055.
- 12 St Leger, A.S.; Cochrane, A.L. and Moore, F. (1979). Factors associated with cardiac  
13 mortality in developed countries with particular reference to the consumption of  
14 wine. *Lancet*, **1**: 1017-1020.
- 15 Tahiri M.; Tressol JC.; Doco T; Rayssiguier, Y. and Coudray C. (2002). Chronic oral  
16 administration of rhamnogalacturonan-II dimer, a pectic polysaccharide, failed to  
17 accelerate body lead detoxification after chronic lead exposure in rats. *Brit. J. Nutr.*,  
18 **87**: 47-54.
- 19 Tahiri, M.; Pellerin, P.; Tressol, J.C. ; Doco T.; Pepin, D. ; Raissiguier, Y. and Coudray  
20 C. (2000). The rhamnogalacturonan-II dimer decreases intestinal absorption and  
21 tissue accumulation of lead in rats. *J. Nutr*, **130**: 249-253.
- 22 Takayanagi, T. and Yokotsuka, K. (1999). Angiotensin I converting enzyme-inhibitory  
23 peptides from wine. *Am. J. Enol. Vitic.*, **50**: 65-68.
- 24 Takechi, M.; Tanaka, Y.; Nonaka, G.I. and Nishioka, I. (1985). Structure and  
25 antiherpetic activity among the Tannins. *Phytochem.*, **24**: 2245-2250.

- 1 Taylor, S L; Higley, N A and Bush, R K. (1986). Sulfites in foods: uses, analytical  
2 methods, residues, fate, exposure assessment, metabolism, toxicity, and  
3 hypersensitivity. *Adv. Food Res.* **30**:1-76.
- 4 Threlfall, R.T. and Morris, J.R. (2002). Using dimethyldicarbonate to minimize sulphur  
5 dioxide for prevention of fermentation from excessive yeast contamination in juice  
6 and semi-sweet wine. *J. Food Sci.* **67**:2758-2761.
- 7 Tomita, M., Wakabayashi, H., Yamauchi, K., Teraguchi, S. and Hayasawa, H. (2002).  
8 Bovine lactoferrin and lactoferricin derived from milk: production and applications.  
9 *Biochem. Cell Biol.* **80**:109-112.
- 10 Tonon, T.; Bourdineau, J.P. and Lonvaud-Funel A., (2001). The arcABC gene cluster  
11 encoding the arginine deiminase pathway of *Oenococcus oeni*, and arginine  
12 induction of a CRP-like gene. *Res. Microbiol.*, **152**: 653-661.
- 13 Torrea, D. and Ancín, C. (2002). Content of biogenic amines in a Chardonnay wine  
14 obtained through spontaneous and inoculated fermentations. *J. Agric. Food*  
15 *Chem.*, **50**:4895-4899.
- 16 Ursini, F.; Zamburlini, A.; Cazzolato, G.; Maiorino, M.; Bon, G.B. and Sevanian, A.  
17 (1998). Postprandial plasma lipid hydroperoxides: a possible link between diet and  
18 atherosclerosis. *Free Radic. Biol. Med.*, **25**: 250-252.
- 19 Usseglio-Tomasset, L. and Bosia, P.D. (1990). Amino acids and oligopeptides  
20 development from the must to the wine. *Bull. OIV (707-708)* 21-46.
- 21 Uthurry, C.A.; Suárez-Lepe, J.A. Lombardero, J.; García del Hierro, J.R. (2007). Ethyl  
22 carbamate production induced by selected yeasts and lactic acid bacteria in red  
23 wine. *Food Chem.* **94**:262-270

1 Uthurry, C.A.; Varela, F.; Colomo, B.; Suárez-Lepe, J.A. Lombardero, J.; García del  
2 Hierro, J.R. (2004). Ethyl carbamate concentrations in typical Spanish red wines.  
3 *Food Chem.* **88**: 329-388.

4 van der Gaag, M.S.; Ubbink, J.B.; Sillanaukee, P.; Nikkari, S. and Hendriks, H.F.  
5 (2000a). Effect of consumption of red wine, spirits, and beer on serum  
6 homocysteine. *Lancet*, **355**: 1522.

7 van der Gaag, M.S.; van den Berg, R.; van den Berg, H.; Schaafsma, G. and Hendriks,  
8 H.F. (2000b). Moderate consumption of beer, red wine and spirits has counteracting  
9 effects on plasma antioxidants in middle-aged men. *Eur. J. Clin. Nutr.*, **54**: 586-91.

10 Vázquez-Agell, M.; Sacanella, E.; Tobias, E.; Monagas, M.; Antúnez, E.; Zamora-Ros,  
11 R.; Andrés-Lacueva, C.; Lamuela-Raventós, R.M.; Fernández-Solá, J.; Nicolás, J.M.  
12 and Estruch, R. (2007). Inflammatory markers of atherosclerosis are decreased after  
13 moderate consumption of cava (sparkling wine) in men with low cardiovascular  
14 risk. *J. Nutr.*, **137**: 2279-2284.

15 Villamiel, M., Polo, M.C. and Moreno-Arribas, M.V. (2008). Nitrogen compounds and  
16 polysaccharides changes during the biological ageing of sherry wines. *LWT-Food*  
17 *Sci. Technol.*, **41**:1842-1846.

18 Vinson, J.A.; Teufel, K. and Wu, N. (2001). Red wine, dealcoholized red wine, and  
19 especially grape juice, inhibit atherosclerosis in a hamster model. *Atherosclerosis*,  
20 **156**: 67-72.

21 Visconti, A.; Pascale, M. and Centonze, G. (1999) Determination of ochratoxin A in  
22 wine by means of immunoaffinity column clean-up and high-performance liquid  
23 chromatography. *J. Chromatogr. A.*, **864**, 89-101.

- 1 Vivas, N., Lonvaud-Funel, A. and Glories, Y. (1997). Effect of phenolic acids and  
2 anthocyanins on growth, viability and malolactic activity of a lactic acid  
3 bacterium. *Food Microbiol.* **14**:291-300.
- 4 Watzl, B.; Bub, A.; Briviba, K. and Rechkemmer, G. (2002). Acute intake of moderate  
5 amounts of red wine or alcohol has no effect on the immune system of healthy  
6 men. *Eur. J. Nutr.*, **41**: 264-270.
- 7 Watzl, B.; Bub, A.; Pretzer, G.; Roser, S.; Barth, S.W. and Rechkemmer, G. (2004).  
8 Daily moderate amounts of red wine or alcohol have no effect on the immune  
9 system of healthy men. *Eur. J. Clin. Nutr.*, **58**: 40-45.
- 10 Weyant, M.J.; Carothers, A.M.; Dannenberg, A.J. and Bertagnolli, M.M. (2001). (+)-  
11 Catechin inhibits intestinal tumor formation and suppresses focal adhesion kinase  
12 activation in the min/+ mouse. *Cancer Res.*, **61**: 118-125.
- 13 Wu, X.; Cao, G. and Prior, R.L. (2002). Absorption and metabolism of anthocyanins in  
14 elderly women after consumption of elderberry or blueberry. *J. Nutr.*, **132**: 1865-  
15 1871.
- 16 Yanai, T; Suzuki, I. and Sato, M. (2003). Prolyl endopeptidase inhibitory peptides in  
17 wine. *Biosc. Biotech. Biochem.*, **67**: 380-382.
- 18 Zamora-Ros, R.; Urpí-Sardà, M.; Lamuela-Raventós, R.M.; Estruch, R.; Vázquez-  
19 Agell, M.; Serrano-Martínez, M.; Jaeger, W. and Andres-Lacueva, C. (2006).  
20 Diagnostic performance of urinary resveratrol metabolites as a biomarker of  
21 moderate wine consumption. *Clin. Chem.*, **52**: 1373-1380.
- 22 Zhang, Y.; Choi, H.J.; Han, H.S.; Park, J.H.; Kim, S.; Bae, J.H.; Kim, H.K. and Choi, C.  
23 (2003). Polyphenolic compounds from korean pear and their biological activities.  
24 *Food Sci. Biotech.*, **12**: 262-267.

- 1 Zhang, Y.S.Z.; Li, J.; Li, X.; Pan, R.E.; Cahoon, R.E.; Jaworski J.G.; Wang X.; Jez  
2 J.M., Chen F., and Yu O.(2006). Using unnatural protein fusions to engineer  
3 resveratrol biosynthesis in yeast and Mammalian cells. *J. Am. Chem. Soc.*, **128**:  
4 13030-13031.
- 5 Zimmerli, B., and Dick, R. (1996). Ochratoxin A in table wine and grape-juice:  
6 Occurrence and risk assessment. *Food Add. Contam.*, **13**: 665-668.
- 7

1 **Table 1.** Summary of the most important measures to prevent or reduce OTA  
2 accumulation in wines, according the *Code of Sound Vitivinicultural Practices* of the  
3 OIV (OIV, 2005, Resolution VITI-OENO 1/2005)

4

---

***Cultivation practices in the vineyards***

- To train producers with regards to risk of mould and mycotoxins, the identification of ochratoxigenic fungus and period of infection, and knowledge of preventive measures to be applied to vineyard and wineries

***Practices at harvest***

- To train the harvesting staff to reject rotten bunches, particularly those affected by dark brown black moulds
- To vinify the mechanical harvested grapes separately, when the sanitary quality of the crop is poor

***Treatment at the winery***

- To monitor, in large wineries and cooperatives, the sanitary quality of the grapes in order to favour with fair pricing the sanitary quality of grapes, and to process the grapes accordingly
- To avoid long periods of maceration and to use enological adsorbents, such as activated charcoal or yeast hulls, in red wine, and bentonite, in white wine, when the crop has a relevant percentage of rotten grapes
- To perform a rapid grape drying and avoid water condensation overnight for grapes used in the vinification of dessert wines
- To implement a complete HACCP plan, from the vine to the bottled wine or raisin, in the wine regions where the OTA occurrence is higher

---

5

1 **Table 2.** Human intervention studies related to wine consumption.

2

Reference	Subjects*	Intervention test products	Dose/day	Duration (days)	Positive outcomes	Negative outcomes
Lavy et al. (1994)	20 men	-Red wine -White wine	400 mL (40g ethanol/day)	15	-Red wine: Increment in plasma HDLc and apo A1	No changes in plasma total cholesterol, LDLc, VLDLc
Pace-Asciak et al. (1996)	24 men	-Red wine -White wine -Grape juice -Grape juice enriched with resveratrol	400 mL (40g ethanol/day)	30	-Red wine/white wine: reduction of trombine-induced platelet aggregation and plasmatic levels of tromboxane B2  -White wine: reduction of ADP-induced platelet aggregation -Enriched grape juice: Reduction of trombine-induced platelet aggregation	-Red wine: no reduction of ADP-induced platelet aggregation
Pellegrini et al. (1996)	11 men	-Red wine -Fruit juice + alcohol -Dealcoholized red wine	-320 mL (30g ethanol/day) -320 mL (30g ethanol/day) -320 mL	30	-Red wine / fruit juice+alcohol: Reduction in collagen-induced platelet aggregation and fibrinogen levels	No changes in ADP-induced platelet aggregation, t-PA antigen, vWF and plasminogen levels
Miyagi et al. (1997)	20 men	-Red wine -White wine -Beer -Grape juice	-300 mL -450 mL -750 mL -300 mL	1	-Red wine: Inhibition of the LDL oxidation after 1 and 2 h of ingestion	
Nigdikar et al. (1998)	9 men	-Red wine -White wine -Red wine polyphenols (capsule) -Red wine polyphenols in white wine -Alcoholic drink	-375 mL -375 mL -1g/day= 375 ml red wine -1g/day -40g ethanol /day	14	-Red wine, red wine polyphenols, red wine polyphenols+white wine: Reduction of plasma lipid peroxides, thiobarbituric acid-reactive substances, and conjugated dienes. Increase in plasma and LDL polyphenols -White wine: Increase in thiobarbituric acid-reactive substances	No changes in plasma triacylglycerol, LDL or HDL cholesterol
Serafini et al. (1998)	10 men	-Dealcoholized red wine - Dealcoholized white wine -Water	113 mL	1	-Dealcoholized red wine: Increment in plasma antioxidant capacity and polyphenol content after 50 min of ingestion	

Reference	Subjects*	Intervention test products	Dose/day	Duration (days)	Positive outcomes	Negative outcomes
Ursini et al. (1998)	9 men	-fatty meal -red wine+ fatty meal	300 mL	1	Red wine+fatty meal: Reduction of post-prandial plasma lipid hydroperoxide levels	
Djoussé et al. (1999)	13 men/women	-Red wine+fatty meal (0.8 g fat/kg body weight) -Isocaloric beverage	3mL/kg	1		No significant changes in FMD of the brachial artery after consumption of red wine +fatty meal
Stein et al. (1999)	15 men/women with coronary artery disease	Grape juice	7.7±1.2 mL/kg	15	Improvement of FMD and decrease of LDL susceptibility to oxidation	
Cuevas et al. (2000)	6 men	-Red wine + fatty meal -Fatty meal	240 mL	30	-Red wine: reduction in FMD of the brachial artery	
van deer Gaag et al. (2000a)	9 men	-Red wine -spirits -Beer -Water	40 g ethanol /day	21	-Beer: Decrease in plasma homocysteine levels	-Red wine/spirits: Increment in plasma homocysteine levels
van deer Gaag et al. (2000b)	11 men	-Red wine -spirits -Beer -Water	40 g ethanol /day	21		No effects on the enzyme activities of serum glutathion peroxidase, erythrocyte glutathion reductase and superoxide dismutase, and plasma concentrations of $\alpha$ - and $\gamma$ -tocopherol, lutein, zeaxanthin, $\beta$ -cryptoxanthin, lycopene and $\alpha$ -carotene
Pignatelli et al. (2002)	20 men/women	-Red wine -White wine	300 mL	15	-Red wine: higher reduction of collagen-induced platelet aggregation	
Watzl et al. (2002)	6 men	-Red wine -12% ethanol dilution	500 mL	1		No effects on: TNF- $\alpha$ , IL-2, IL-4, phagocytosis activity of neutrophils and lymphocyte proliferation
Watzl et al. (2004)	24 men	-Red wine -12% ethanol dilution -Dealcoholized red wine -Red grape juice	500 mL	15		No effects on: TNF- $\alpha$ , TNF- $\alpha$ mRNA, IL-2, IL-4, TGF- $\beta$ , phagocytosis activity of neutrophils, lymphocyte proliferation, lytic activity of natural killer cells, and percentage of apoptotic lymphocytes

Reference	Subjects*	Intervention test products	Dose/day	Duration (days)	Positive outcomes	Negative outcomes
Badía et al. (2004)	8 men	-Red wine -Gin	-320 mL -100 mL (30 g ethanol/day)	28	-Red wine: Reduction of the expression of VLA-4 in monocyte surface. -Both: Reduction of the TNF- $\alpha$ induced adhesion of monocyte to an endothelial cell line. More pronounce for red wine.	
Estruch et al. (2004)	40 men	-Red wine -Gin	-320 mL -100 mL (30 g ethanol/day)	28	-Red wine: reduction in the expression of LFA-1, Mac-1, VLA-4 and MCP-1 on monocyte surface. Reduction of serum levels of hs-CRP, VCAM-1 and ICAM-1. -Both: Reduction in C-reactive protein and IL1- $\alpha$ levels	
Papamichael et al. (2004)	16 men/women	-Cigarette (1) -Cigarette (1) +red wine -Cigarette (1) + dealcoholized red wine	250 mL	1	Cigarette smoking + red wine or dealcoholized red wine: No changes in flow mediated dilatation after 15, 30, and 60 minutes after the inhalation of smoke compared to baseline levels	
Avellone et al. (2006)	48 men/women	-Red wine	250 mL	30	Reduction of LDL/HDL ratio, fibrinogen, factor VII, C-reactive protein, LDLox antibody. Increment in HDLc, apoA1, TGF $\beta$ 1, t-PA, PAI and plasma antioxidant capacity	
Pignatelli et al. (2006)	20 men/women	-Red wine -Alcohol wine	300 mL	15	-Both: Increment in plasma antioxidant capacity and decrease in urine isoprotanes levels.	
Sacanella et al. (2007)	35 women	-Red wine -White wine	20g ethanol/day	30	-Red wine: Reduction of serum levels of VCAM-1 and E-selectin. -Both: Increment in HDLc. Reduction of serum concentrations of hs-CRP, ICAM-1, CD40L, and IL-6	

1  
2

Reference	Subjects*	Intervention test products	Dose/day	Duration (days)	Positive outcomes	Negative outcomes
Vázquez-Agell et al. (2007)	20 men	-Sparkling white wine (cava) -Gin	-300 mL -100 mL (30g ethanol/day)	28	-Cava: Reduction of the expression LFA-1, VLA-4, SLe <sup>x</sup> and CD40 on monocyte surface. Reduction of serum levels of hsCRP, ICAM-1, IL-6, MCP-1 and CD40L -Gin: Reduction of while only SLe <sup>x</sup> on monocyte surface. -Both: reduction of serum levels of VCAM-1, E-selectin and P-selectin.	
Imhof et al. (2008)	49 men/women	Red wine (12.5% alcohol; 275 mg/L polyphenols) -Ethanol (concentration 12.5% alcohol) -Beer (5.6% alcohol; 160 mg/L polyphenols) -De-alcoholized red wine (275 mg/L polyphenols) -De-alcoholized beer (171 mg/L polyphenols) -Water	30g ethanol/day for men and 20 g/d for women	21	-Ethanol/ de-alcoholized red: Reduction on MCP-1-induced monocyte migration and in FMLP-induced migration	No effects on MCP-1 receptor expression
Spaak et al. (2008)	30 men/women	-Red wine -Ethanol -Water	-120 ml -44 ml (one drink)	1	-Ethanol/wine: Increase in cardiac output and MSNA after 2 drinks of each. Increase in brachial artery diameter after both one and two drinks. -Red wine: Increment in heart rate and attenuation in FMD after 2 drinks.	No effects on blood pressure

3  
4  
5  
6  
7  
8  
9

\*When not stated, studies were conducted on healthy subjects

ADP: adosin diphosphate; ApoA1: apolipoprotein A1; FMD: flow mediated vasodilation; FMLP: N-formyl-methionyl-leucyl-phenylalanine; HDLc: high density lipoprotein cholesterol; hs-CRP: High-sensitivity C-reactive protein; ICAM-1: intercellular adhesion molecule-1; IL-2, IL1- $\alpha$ , IL-4, IL-6: interleukin 2, 1 $\alpha$ , 4, 6; LDLc: low density lipoprotein cholesterol; LDLox: Oxidized LDL; LFA-1: lymphocyte function-associated antigen-1; Mac-1: CD11b/CD18; MCP-1: monocyte chemoattractant protein; MSNA: muscle sympathetic nerve activity; PAI : plasminogen activator inhibitor; SLe<sup>x</sup>: Sialyl-Lewis<sup>x</sup>; TGF $\beta$ 1: transforming growth factor  $\beta$ 1; TNF- $\alpha$ : tumor necrosis factor  $\alpha$ ; t-PA: tissue-type plasminogen activator antigen; VCAM-1: vascular adhesion molecule-1; VLA-4: very late activation antigen-4; VLDLc: very low density lipoprotein cholesterol; vWF: von Willebrand factor

1 **FIGURE LEGENDS**

2

3 **Figure 1.** Chemical structure of ochratoxin A.

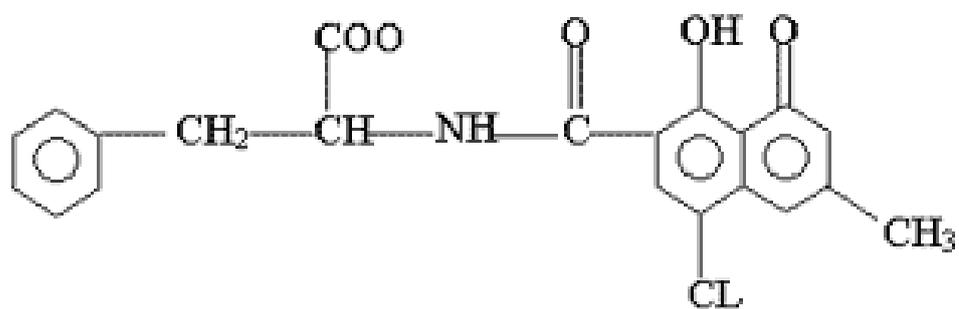
4 **Figure 2.** Chemical structure of main biogenic amines present in wine.

5 **Figure 3.** Chemical structure of main non-flavanoid and flavanoid phenolic compounds  
6 present in wine.

7

1 Figure 1

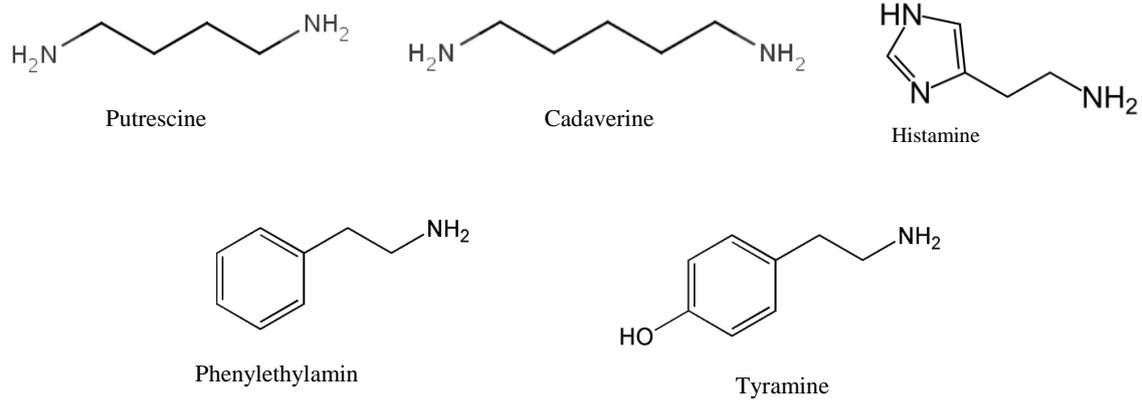
2



3

4

5



1

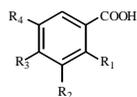
Figure 2

2

Figure 3

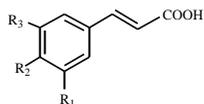
## NON-FLAVANOID PHENOLIC COMPOUNDS

### Hydroxybenzoic acids



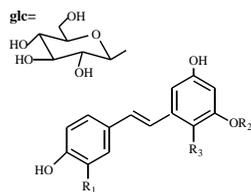
Benzoic acid	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>
<i>p</i> -Hydroxybenzoic	H	H	OH	H
Protocatechuic	H	OH	OH	H
Vanillic	H	OCH <sub>3</sub>	OH	H
Gallic	H	OH	OH	OH
Syringic	H	OCH <sub>3</sub>	OH	OCH <sub>3</sub>
Salicylic	OH	H	H	H
Gentisic	OH	H	H	OH

### Hydroxycinnamic acids



Hydroxycinnamic acid	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
<i>p</i> -Coumaric	H	OH	H
Caffeic	OH	OH	H
Ferulic	OCH <sub>3</sub>	OH	H
Sinapic	OCH <sub>3</sub>	OH	OCH <sub>3</sub>

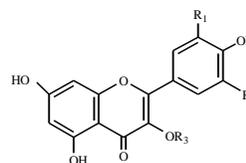
### Stilbenes



Stilbene	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
<i>Trans</i> -resveratrol	H	H	H
<i>Trans</i> -resveratrol-3- <i>O</i> -glucoside (piceid)	H	glc	H
<i>Trans</i> -resveratrol-2- <i>C</i> -glucoside	H	H	glc
<i>Trans</i> -astringin	OH	glc	H

## FLAVANOID PHENOLIC COMPOUNDS

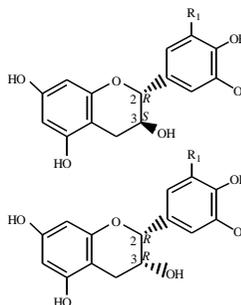
### Flavonols



Flavonol	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
Kaempferol	H	H	H
Kaempferol-3- <i>O</i> -glucoside	H	H	glc
Kaempferol-3- <i>O</i> -galactoside	H	H	gal
Kaempferol-3- <i>O</i> -glucuronide	H	H	gluc
Quercetin	OH	H	H
Quercetin-3- <i>O</i> -glucoside	OH	H	glc
Quercetin-3- <i>O</i> -glucuronide	OH	H	gluc
Myricetin	OH	OH	H
Myricetin-3- <i>O</i> -glucoside	OH	OH	glc
Myricetin-3- <i>O</i> -glucuronide	OH	OH	gluc
Isorhamnetin	OCH <sub>3</sub>	H	H
Isorhamnetin-3- <i>O</i> -glucoside	OCH <sub>3</sub>	H	glc

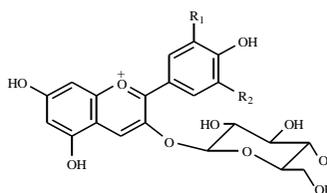
gal= galactose; glc= glucose; gluc= glucuronide acid

### Flavan-3-ols or flavanols



Flavan-3-ol	R <sub>1</sub>	C-2	C-3
(+)-Catechin	H	<i>R</i>	<i>S</i>
(+)-Gallocatechin	OH	<i>R</i>	<i>S</i>
(-)-Epicatechin	H	<i>R</i>	<i>R</i>
(-)-Epigallocatechin	OH	<i>R</i>	<i>R</i>

### Anthocyanins



Anthocyanidin	R <sub>1</sub>	R <sub>2</sub>
Cyanidin	OH	H
Delphinidin	OH	OH
Peonidin	OCH <sub>3</sub>	H
Petunidin	OCH <sub>3</sub>	OH
Malvidin	OCH <sub>3</sub>	OCH <sub>3</sub>

R<sub>3</sub>= -CO-CH<sub>3</sub> (-acetyl)    -CO-CH=CH- (-*p*-coumaroyl)    -CO-CH=CH- (-caffeoyl)