

Systematic and Phylogenetic Analysis of the Ole e 1 Pollen Protein Family Members in Plants

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1. Introduction

Pollen allergens are specific substances able to cause IgE-mediated hypersensitivity (allergy) after contact with the immune system [D'Amato et al. 1998]. To date, about 50 plant species have been registered in the official allergen list of the International Union of Immunological Societies (IUIS) Allergen Nomenclature Subcommittee <http://www.allergen.org> as capable of inducing pollen allergy in atopic individuals [Mothes et al. 2004]. These plants are usually grouped as (1) trees (members of the orders: *Fagales*, *Pinales*, *Rosales*, *Arecales*, *Scrophulariales*, *Junglandales*, *Salicales*, and *Myrtales*), (2) grasses (members of the families: *Bambusoideae*, *Arundinoideae*, *Chloridoideae*, *Panicoideae*, and *Poideae*), and (3) weeds (components of families *Asteraceae*, *Chenopodiaceae* and *Urticaceae*) [Hauser et al. 2010].

Allergens are proteins with a broad range of molecular weights (~5 to 50 kDa), which exhibit different features of solubility and stability. More than 10 groups of pollen allergens have been reported. Among all groups of pollen allergens, Pollen Ole e I (Ole) domain-containing proteins are the major allergens, included like-members of the "pollen proteins of the Ole e 1 family" (Accession number: PF01190) within the Pfam protein families database [Finn et al. 2010].

Ole e 1 was the first allergen purified from *Olea europaea* L. [Lauzurica et al. 1998] and named as such according to the IUIS nomenclature [King et al. 1994]. This protein is considered the major olive pollen allergen on the basis of its high prevalence among atopic patients and the high proportion it represents within the total pollen protein content, in comparison with other olive pollen allergens. These include at present another 10 allergens already identified and classified like Ole e 2 to Ole e 11 [Rodríguez et al. 2002, Barral et al. 2004, Salamanca et al. 2010]. Ole e 1 consists of a single polypeptide chain of 145 amino acid residues with a MW of 18–22 kDa, displaying acidic pI and different forms of N-glycosylation [Villalba et al. 1990, Batanero et al. 1994]. Heterologous proteins with a relevant homology have been described in other members of the *Oleaceae* family, such as fraxinus, lilac, jasmine and privet. The polypeptides encoded by the *LAT52* gene from tomato and the *Zmc13* gene from maize pollens also exhibit a high similarity to Ole e 1 [Twell et al. 1989, Hanson et al. 1989]. These plant pollen proteins are structurally related but their biological function is not yet known; though they have been suggested to be

involved in important events of pollen physiology, such as hydration, germination and/or pollen tube growth, and other reproductive functions [Alché et al. 1999, 2004, Tang et al. 2000, Stratford et al. 2001].

Structurally, the Ole domain contains six conserved cysteines which may be involved in disulfide bonds, since no free sulfhydryl groups have been detected in the native protein [Villalba et al. 1993]. Olive Ole e 1 exhibits a high degree of microheterogeneity, mainly concentrated in the third of the molecule closer to the N- terminus. The Ole e I (Ole) domain defining the pollen proteins Ole e I family signature or consensus pattern sequences PS00925 [Sigrist et al. 2010], is characterized by the amino acid sequence [EQT]-G-x-V-Y-C-D-[TNP]-C-R, where “x” could be any residue.

There is a high diversity of proteins sharing the Ole domain among plant species. To date, eleven Ole domain-containing genes have been isolated and characterized from olive pollens [Rodríguez et al. 2002]. Ole-containing proteins include proline-rich proteins, proteins encoding extensin-like domains, phosphoglycerate mutase, tyrosine-rich hydroxyproline-rich glycoprotein, and hydroxyproline-rich glycoprotein. These Ole-containing proteins can exhibit: (1) the pollen Ole signature exclusively, e.g. the ALL1_OLEEU P19963 protein from *Olea europaea* L., (2) both the pollen Ole signature and the replication factor A protein 3 motive pattern (PF08661), e.g. the O49527 pollen-specific protein-like from *Arabidopsis thaliana* (842 residues), (3) both the pollen Ole domain and the phosphoglycerate mutase (PGAM) motif, e.g. the Q9SGZ6 protein from *Arabidopsis thaliana*., and finally (4) both the pollen Ole signature and the reverse transcriptase 2 (RVT2) motif, e.g. the A5AJL0 protein from *Vitis vinifera*.

Several efforts have been made to develop an understandable and reliable systematic classification of the diverse and increasing number of different allergen protein structures. As mentioned above, the classification system widely established for proteins that cause IgE-mediated atopic allergies in humans (allergens) was defined by Chapman et al. (2007). This system uses the first three letters of the genus; a space; the first letter of the species name; a space and an Arabic number. Despite this classification system, protein databases are full of allergen proteins lacking this systematic and comprehensive nomenclature. In other cases, many of the proteins described here have not been described as allergens, or their naming makes no reference to the Ole e 1 family that facilitates their identification. Otherwise, naming in databases is frequently given randomly, on the basis of chromosome location, addressing structural features and functional characterizations or simply using the name of the entire family. In this study, we used a combination of functional genomics and computational biology to name and classify the entire Ole e 1 family, as well as to characterize structurally and functionally the proteins of this superfamily. Our data indicate that the Ole e 1 protein family consists of at least 109 divergent families, which will likely expand as more genomic studies are undertaken, and fully sequenced plant genomes become available.

2. Material and methods

2.1 Database search for Ole e 1 family genes

Sequences of Ole e 1 and Ole e 1-like genes were retrieved from the US National Center for Biotechnology Information (NCBI, <http://www.ncbi.nlm.nih.gov/>), the Uniprot database (<http://www.uniprot.org/>), and the non-redundant expressed sequence tag (EST)

databases using BLASTX, BLASTN and BLAST (low complexity filter, Blosum62 substitution matrix) [Altschul et al. 1997]. Searches were conducted using previously characterized *Olea europaea* L. *Ole e 1* (GenBank Accession number P19963), *Solanum lycopersicum* LAT52 (GenBank Accession number P13447), *Zea mays* *Zmc13* (GenBank Accession number B6T1A9), *Arabidopsis thaliana* pollen-specific protein-like (GenBank Accession number O49527), *Arabidopsis thaliana* PGAM containing domain protein (GenBank Accession number Q9SGZ6), and *Vitis vinifera* RVT2 containing domain protein (GenBank Accession number A5AJL0). Full-length amino acid sequences for Ole e 1 proteins were compiled and aligned using ClustalW [Thompson et al. 1994]. Genetic distances between pairs of amino acid sequences were calculated with Bioedit V7.0.5.3 [Hall 1999]. Consensus protein sequences were derived from these original alignment, and further analyzed for the presence of putative functional motifs using the PROSITE database [Sigrist et al. 2010], of biologically meaningful motif descriptors derived from multiple alignments and the ScanProsite program [de Castro et al. 2006], from the Expert Protein Analysis System (ExPASy) proteomics server of the Swiss Institute of Bioinformatics [Gasteiger et al. 2003]. Finally, the consensus protein sequences were submitted to BLASTP analysis to identify homologous proteins from other plant species.

2.2 Revised/unified nomenclature

In order to provide a revised and unified nomenclature for Ole e 1-like gene superfamily, we developed a sequence-based similarity approach to classify all the retrieved sequences using a previously developed gene nomenclature model [Kotchoni et al. 2010]. For this new nomenclature, Ole e 1 protein sequences that are more than 40% identical to previously identified Ole e 1 sequences compose a family, and sequences more than 60% identical within a family, compose a gene subfamily. Protein sequences that are less than 40% identical would describe a new Ole e 1 gene family. Taking olive protein Ole e 1_57A9 (previous name Ole e 1, major olive pollen allergen) as an example for the revised nomenclature (Table 1), Ole e 1 indicates the root; the digits (57) indicates a family and the first letter (A) a subfamily, while the final number (9) identifies an individual gene within a subfamily. The revised nomenclature is therefore composed of an assigned gene symbol (Ole e 1) (abbreviated gene name) for the whole gene superfamily. The gene symbol must be (i) unique and representative of the gene superfamily; (ii) contain only Latin letters and/or Arabic numerals, (iii) not contain punctuation, and (iv) without any reference to species. These newly developed criteria have been applied to database curators to generate the unified Ole e 1 gene families/classes regardless of the source of the cloned gene(s).

2.3 Sequence alignments and phylogenetic analyses

The retrieved Ole e 1 protein families were used to generate a phylogenetic tree using ClustalW [Thompson et al. 1994]. The alignment was created using the Gonnet protein weight matrix, multiple alignment gap opening/extension penalties of 10/0.5 and pairwise gap opening/extension penalties of 10/0.1. These alignments were adjusted using Bioedit V7.0.5.3 [Hall 1999]. Portions of sequences that could not be reliably aligned were eliminated. Phylogenetic tree was generated by the neighbourjoining method (NJ), and the branches were tested with 1,000 bootstrap replicates. The tree was visualized using Treedyn program [Chevenet et al. 2006].

2.4 Ole e 1 superfamily: Protein modeling and structural characterization

In order to study the structural and conformational variability between the Ole e 1 protein families, selected members of the Ole e 1 superfamily were modelled using SWISS-MODEL server, via the ExPASy web server [Gasteiger et al. 2003]. The initial modelled Ole e 1 structures were subjected to energy minimization with GROMOS96 force field energy [van Gunsteren et al. 1996] implemented in DeepView/Swiss-PDBViewer v3.7 [Guex and Peitsch 1997] to improve the van der Waals contacts and to correct the stereochemistry of the improved models. The quality of the models was assessed by checking the protein stereochemistry with PROCHECK [Laskowski et al. 1993] and the protein energy with ANOLEA [Melo et al. 1997, 1998]. Ramachandran plot statistics for the models were calculated to show the number of protein residues in the favoured regions.

3. Results

3.1 The Ole e 1 protein families: Revised and unified nomenclature

In order to provide a revised/international consensus and unified nomenclature for the Ole e 1 gene superfamily, we first retrieved all the Ole e 1 and Ole e 1-like gene sequences using PS00925 as the major molecular consensus defining the entire superfamily of Ole e 1 proteins. We next verified all annotated plant Ole e 1 open reading frames (ORFs) using Ole e 1 sequence domains. A complementary and comparative study was developed by using Uniprot database to validate the molecular function and previous denomination of each Ole e 1 protein. Our searches resulted in the identification of 571 sequences encoding Ole e 1 and Ole e 1 like proteins from a wide variety of plant species, with the diagnostic motif PS00925 (Table 1). According to the established criteria (see Material and Methods), these sequences integrated 109 Ole e 1 gene families which have been attributed to different functional categories including extensins and extensin-like proteins, proline-rich proteins, hydroxyproline-rich glycoproteins, tyrosine-rich/hydroxyproline-rich glycoproteins, hydrolases, phosphoglycerate mutases, arabinogalactan proteins, etc. (Table 1).

Among the sequences retrieved, Ole e 1₄₈ is the most extensive family with 63 gene members encoding for different pollen-specific protein C13 homologues, followed by Ole e 1₅₇ family with 42 gene homologues encoding Ole e 1 (the olive major pollen allergen), Ole e 1₁₆ with 26 gene members encoding proline-rich proteins, and Ole e 1₅₂ with 22 members encoding LAT52 homologues (Table 1). The number of Ole e 1 genes greatly varied from one plant species to another. The genus *Oryza* included the highest number of Ole e 1 genes (143), followed by *Arabidopsis* with 95 genes (Table 1). At present, more than half of the catalogued Ole e 1 families encoded a single Ole e 1/Ole e 1-like gene, which was in most cases “uncharacterized” (Table 1).

The total number of genes in the Ole e 1 superfamily is expected to increase steadily with time, mainly due to the genomic sequencing of additional species like *Olea europaea* L. (http://www.gen-es.org/11_proyectos/PROYECTOS.CFM?pg=0106&n=1). Regardless of the plethora of Ole e 1 genes yet to be identified/characterized, their classification and relationship to the entire extended Ole e 1 gene superfamily will be easy owing to this nomenclature building block that catalogues newly identified/characterized Ole e 1 gene products only on the basis of sequence similarity to previously characterized Ole e 1 gene products.

Ole e 1 Family	Revised annotation	Previous annotation	GeneBank Accession number	Source
1	Ole e 1_1A1	A14g17215	Q8RAZ6	ARATH
1	Ole e 1_1A2	-	Q8L8VR	ARATH
1	Ole e 1_1A3	ARALYDRAFT_493155	DFMC15	ARALY
1	Ole e 1_1A4	40.00006	Q2A9B5	BRAOL
1	Ole e 1_1A5	31.00008	Q2A9F3	BRAOL
1	Ole e 1_1B1	ARALYDRAFT_403053	D7LEF7	ARALY
1	Ole e 1_1B2	A12g40113	Q58FY6	ARATH
1	Ole e 1_1B3	-	Q8LEB2	ARATH
1	Ole e 1_1B4	A12g47825	Q29PT1	ARATH
1	Ole e 1_1B5	ARALYDRAFT_330672	D7MP26	ARALY
2	Ole e 1_2A1	POPTRDRAFT_818926	B9HCD0	POPTR
2	Ole e 1_2A2	POPTRDRAFT_776772	B9H1E6	POPTR
2	Ole e 1_2B1	VIT_00605138001	D755K6	VITVI
2	Ole e 1_2C1	-	C0T3E9	SOYBN
2	Ole e 1_2D1	RCOM_0860870	B8SA60	RICCO
3	Ole e 1_3A1	OsJ_33016	B9BG44	ORYSJ
3	Ole e 1_3A2	Os10g0206500	Q109X3	ORYSJ
3	Ole e 1_3B1	OSJNBa0014J14.3	Q7G7E7	ORYSJ
3	Ole e 1_3B2	OSJ1004_F02.9	Q8RV11	ORYSJ
3	Ole e 1_3C1	OSJNBa0014J14.29	Q8S6U0	ORYSJ
3	Ole e 1_3D1	Os10g0205600	Q109X1	ORYSJ
3	Ole e 1_3E1	OsJ_33026	B8B0S3	ORYSJ
3	Ole e 1_3J1	SORBIIDRAFT_01g013620	C5WR76	SORBI
3	Ole e 1_3F1	-	B4FE56	MAIZE
4	Ole e 1_4A1	SELMODRAFT_444621	D8SBK5	SEMLL
4	Ole e 1_4A2	SELMODRAFT_443385	D8SOX9	SEMLL
5	Ole e 1_5A1	ARALYDRAFT_481629	D7LGX1	ARALY
5	Ole e 1_5A2	A12g27385	Q6NLE8	ARATH
5	Ole e 1_5B1	-	C6SVU8	SOYBN
5	Ole e 1_5B2	-	C87B94	SOYBN
5	Ole e 1_5C1	-	C6SZD3	SOYBN
5	Ole e 1_5D1	POPTRDRAFT_821599	A9P157	POPTR
5	Ole e 1_5D2	RCOM_1281870	B8SCWA	RICCO
5	Ole e 1_5D3	VITISV_031957	A5BY12	VITVI
5	Ole e 1_5E1	A12g24230	Q9FMQ8	ARATH
5	Ole e 1_5E2	-	Q8LH14	ARATH
5	Ole e 1_5E3	ARALYDRAFT_351286	D7MBX5	ARALY
6	Ole e 1_6A1	-	B6TL01	MAIZE
6	Ole e 1_6A2	-	B4FQB6	MAIZE

Ole e 1 Family	Revised annotation	Previous annotation	GeneBank Accession number	Source	
6	Ole e 1_6B1	-	B6T4H9	MAIZE	
6	Ole e 1_6C1	-	Sh81g021840	CSXZ28	SORBI
6	Ole e 1_6D1	B1136H02.23	Q6EPW8	ORYSJ	
7	Ole e 1_7A1	SELMODRAFT_405039	D9QY88	SEMLL	
7	Ole e 1_7A2	SELMODRAFT_414879	D8RTV5	SEMLL	
8	Ole e 1_8A1	SELMODRAFT_448129	D8TAZ3	SEMLL	
8	Ole e 1_8B1	SELMODRAFT_409805	D8RC11	SEMLL	
8	Ole e 1_8C1	SELMODRAFT_448128	D8TAZ1	SEMLL	
9	Ole e 1_9A1	A12g21140	Q8SKP9	ARATH	
9	Ole e 1_9A2	Proline-rich protein 2	Q9M7P0	ARATH	
9	Ole e 1_9A3	ARALYDRAFT_300523	D7LLO3	ARALY	
9	Ole e 1_9B1	Extensin-like protein	Q9M6T6	ARATH	
9	Ole e 1_9B2	Proline-rich protein 1	Q9M7N9	ARATH	
9	Ole e 1_9B3	A12g33770/09A14_50	Q2T3U5	ARATH	
9	Ole e 1_9B4	ARALYDRAFT_430641	D7MFH2	ARALY	
10	Ole e 1_10A1	Proline-rich protein	Q9M6T7	NICGL	
10	Ole e 1_10B1	VIT_00024081001	D7USA0	VITVI	
10	Ole e 1_10B2	POPTRDRAFT_700888	B9HRA8	POPTR	
10	Ole e 1_10C2	POPTRDRAFT_156515	B9H154	POPTR	
10	Ole e 1_10D2	RCOM_0660490	B8BTC5	RICCO	
11	Ole e 1_11A1	Proline-rich protein	Q82066	SOLTU	
12	Ole e 1_12A1	VITISV_029041	A5BQP2	VITVI	
13	Ole e 1_13A1	VITISV_029838	A5BQP1	VITVI	
13	Ole e 1_13A2	VITISV_029837	A5BQP0	VITVI	
13	Ole e 1_13B1	VIT_00024076001	D7US97	VITVI	
14	Ole e 1_14A1	proline-rich protein	Q93WF4	ORYSA	
14	Ole e 1_14A2	Os10g0145100	C7J7T1	ORYSJ	
14	Ole e 1_14A3	proline-rich protein	Q93WV9	ORYSA	
14	Ole e 1_14A4	Os10g0145000	Q1XGT3	ORYSJ	
14	Ole e 1_14A5	proline-rich protein	Q8M1E8	ORYSA	
14	Ole e 1_14A6	Os10g0145200	Q1XGT1	ORYSJ	
14	Ole e 1_14A7	OsJ_30733	A3C2J9	ORYSJ	
14	Ole e 1_14A8	OsJ_12924	A2XKE9	ORYSJ	
14	Ole e 1_14A9	OsJ_12923	A2XKE7	ORYSJ	
14	Ole e 1_14A10	OsJ_12921	B8AP23	ORYSJ	
14	Ole e 1_14A11	OSJNBa0031A07.6	Q8M1H7	ORYSJ	
14	Ole e 1_14A12	OsJ_30737	A3C2K3	ORYSJ	
14	Ole e 1_14A13	Os10g0145400	Q1XGT0	ORYSJ	
14	Ole e 1_14A14	OsJ_30734	A3C2K0	ORYSJ	

Table 1. The Ole e 1 protein superfamily: new and unified nomenclature. ARATH: *Arabidopsis thaliana*; ARALY: *Arabidopsis lyrata*; BETPN: *Betula pendula*; BRAOL: *Brassica oleracea*; BRARP: *Brassica rapa*; CAPAN: *Capsicum annuum*; CARAS: *Cardaminopsis arenosa*; CHE1: *Chenopodium album*; CROSA: *Crocus sativus*; DAUCA: *Daucus carota*; EUPPU: *Euphorbia pulcherrima*; FRAEX: *Fraxinus excelsior*; GOSBA: *Gossypium barbadense*; GOSHE: *Gossypium herbaceum*; GOSHI: *Gossypium hirsutum*; GOSKI: *Gossypoides kirkii*; HYAOR: *Hyacinthus orientalis*; LigVu: *Ligustrum vulgare*; LILLO: *Lilium longiflorum*; LOLPE: *Lolium perenne*; MAIZE: *Zea mays*; MEDTR: *Medicago truncatula*; NICAL: *Nicotiana alata*; NICGL: *Nicotiana glauca*; NicLa: *Vitis pseudoreticulata*; OleEu: *Olea europaea*; ORYSI: *Oryza sativa*; PETCR: *Petroselinum crispum*; PETHY: *Petunia hybrida*; PHAVU: *Phaseolus vulgaris*; PHEPR: *Phleum pratense*; PHYPA: *Physcomitrella patens*; PICSI: *Picea sitchensis*; PLALA: *Platanus lanceolata*; POPTR: *Populus trichocarpa*; RICCO: *Ricinus communis*; SALKA: *Salsola kali*; SAMNI: *Sambucus nigra*; SELML: *Selaginella moellendorffii*; SOLLI: *Solanum lycopersicum*; SOLTU: *Solanum tuberosum*; SORBI: *Sorgum bicolor*; SOYBN: *Glycine max*; TOBAC: *Nicotiana tabacum*; TRISU: *Trifolium subterraneum*; VITVI: *Vitis vinifera*; 9ROSI: *Cleome spinosa*; (-): uncharacterized.

46	Ole e 1_48H8	-	B4FKQ2	MAIZE
46	Ole e 1_48H8	Pollen-specific protein C13	B6T728	MAIZE
46	Ole e 1_48I1	Putative pollen specific prot.C13	Q8RU50	ORYSJ
46	Ole e 1_48I2	Os10g0317000 protein	Q9Y39	ORYSJ
46	Ole e 1_48I3	-	A226J6	ORYSJ
46	Ole e 1_48I4	Pollen-specific protein C13	B6S340	MAIZE
46	Ole e 1_48I5	Pollen-specific protein C13	B6T594	MAIZE
46	Ole e 1_48I6	Sb0012s014630	C6JRR2	SORBI
46	Ole e 1_48I7	Pollen-specific protein	Q677C4	HYAOR
46	Ole e 1_48J1	Major pollen allergen Lol p 11	Q7M1X5	LOLAOR
46	Ole e 1_48J2	Pollen allergen Phi p 11	Q8H6L7	PHLPR
46	Ole e 1_48J3	Sb03g001020	C5XK86	SORBI
46	Ole e 1_48J4	Pollen allergen Phi p 11	B6T728	MAIZE
46	Ole e 1_48J5	-	A2YE17	ORYSJ
46	Ole e 1_48J6	Os06g055860 protein	Q5Z7I0	ORYSJ
46	Ole e 1_48J1	Sb03g007260	C5YUJ2	SORBI
46	Ole e 1_48K1	-	B4FC11	MAIZE
46	Ole e 1_48K3	Pollen allergen Phi p 11	B6T595	MAIZE
46	Ole e 1_48L1	-	B8BEUR	ORYSJ
46	Ole e 1_48L2	-	A3C1R9	ORYSJ
46	Ole e 1_48L3	Putative Pollen specific protein C13	Q850Z4	ORYSJ
46	Ole e 1_48L4	Os09g0572800 protein	Q8ZF60	ORYSJ
46	Ole e 1_48L5	Os07g0509500 protein	Q8ZLH6	ORYSJ
46	Ole e 1_48L6	-	A3BLQ7	ORYSJ
46	Ole e 1_48L7	-	A2YH83	ORYSJ
46	Ole e 1_48L8	Sb02g012930	C5X6P6	SORBI
46	Ole e 1_48A1	-	CS1355	SOYBN
50	Ole e 1_50A1	Pollen ole e 1 allergen	D7K9E2	ARALY
50	Ole e 1_50A2	Pollen ole e 1 allergen	D7K9E8	ARALY
50	Ole e 1_50A3	P-glycerate mutase 1 like prot.	Q8LD45	ARATH
50	Ole e 1_50A4	-	Q8H7B9	ARATH
50	Ole e 1_50A5	Pollen specific protein	Q42043	ARATH
51	Ole e 1_51A1	F28K14.76 protein	Q89G26	ARATH
52	Ole e 1_52A1	At1g29140	Q8HWDE	ARATH
52	Ole e 1_52A2	F28N24.16 protein	Q8LPA4	ARATH
52	Ole e 1_52B1	At5g45030	Q880L9	ARATH
52	Ole e 1_52B2	Ole e 1-like protein	Q8L9P9	ARATH
52	Ole e 1_52B3	-	D7M5G6	ARALY
52	Ole e 1_52B4	At1g18595	Q8NMJ2	ARATH
52	Ole e 1_52B5	Pollen ole e 1 allergen	D7M5C3	ARALY
52	Ole e 1_52B6	Ole e 1-like protein	Q9F4J8	ARATH

52	Ole e 1_52C1	Ole e 1-like protein	Q48E13	RICCO
52	Ole e 1_52D1	Pollen allergen Che a 1	B599A8	BETPN
52	Ole e 1_52E1	PN40024	D7JL11	VITI
52	Ole e 1_52F1	-	C8TL27	SOYBN
52	Ole e 1_52F2	-	B7FGN2	MEDTR
52	Ole e 1_52G1	Pollen allergen Che a 1	Q2GLR0	CHE1
52	Ole e 1_52G2	Pollen allergen Cro s 1	Q29W25	CROSA
52	Ole e 1_52H1	Sal k 4	E2D0Z8	SALKA
52	Ole e 1_52I1	-	B8N635	POPTR
52	Ole e 1_52J2	-	B8P8Z0	POPTR
52	Ole e 1_52J3	-	B8I1V1	POPTR
52	Ole e 1_52J1	Anther-specific prot. LAT52	B9SBK9	RICCO
52	Ole e 1_52K1	AS1	D7R0W3	GOSHI
52	Ole e 1_52L1	Anther-specific prot. LAT52	P13447	SOILL
53	Ole e 1_53A1	-	D7K9D8	ARALY
54	Ole e 1_54A1	Pollen-specific protein -like	Q495Z7	ARATH
55	Ole e 1_55A1	Rutafidin Ole e 1-like protein	A3FA46	NICLA
56	Ole e 1_56A1	Major pollen allergen Phi 1	P82242	PLALA
57	Ole e 1_57A1	Allergen Fra e 1,0101	Q7XAV4	FRAEX
57	Ole e 1_57A2	Fra e 1,0102 major allergen	Q5EXJ6	FRAEX
57	Ole e 1_57A3	Major pollen allergen Lig v 1	Q82015	LIGVU
57	Ole e 1_57A4	Ole e 1 olive pollen allergen	X76397	OleEu
57	Ole e 1_57A5	Ole e 1 olive pollen allergen	AF532765	OleEu
57	Ole e 1_57A6	Ole e 1 olive pollen allergen	AF532766	OleEu
57	Ole e 1_57A7	Ole e 1 olive pollen allergen	AF532767	OleEu
57	Ole e 1_57A8	Ole e 1 olive pollen allergen	X76396	OleEu
57	Ole e 1_57A5	Ole e 1 olive pollen allergen	P19963	OleEu
57	Ole e 1_57A10	Ole e 1 olive pollen allergen	Ole e 1 Educa	OleEu
57	Ole e 1_57A11	Ole e 1 olive pollen allergen	X76395	OleEu
57	Ole e 1_57A12	Ole e 1 olive pollen allergen	Y12426	OleEu
57	Ole e 1_57A13	Ole e 1 olive pollen allergen	Y12427	OleEu
57	Ole e 1_57A14	Ole e 1 olive pollen allergen	AF500908	OleEu
57	Ole e 1_57A15	Ole e 1 olive pollen allergen	AF515277	OleEu
57	Ole e 1_57A16	Ole e 1 olive pollen allergen	AF515278	OleEu
57	Ole e 1_57A17	Ole e 1 olive pollen allergen	AF515280	OleEu
57	Ole e 1_57A18	Ole e 1 olive pollen allergen	AF515279	OleEu
57	Ole e 1_57A19	Ole e 1 olive pollen allergen	AF515281	OleEu
57	Ole e 1_57A20	Ole e 1 olive pollen allergen	AF532755	OleEu
57	Ole e 1_57A21	Ole e 1 olive pollen allergen	AF532756	OleEu
57	Ole e 1_57A22	Ole e 1 olive pollen allergen	AF532757	OleEu
57	Ole e 1_57A23	Ole e 1 olive pollen allergen	AF532768	OleEu

57	Ole e 1_57A24	Ole e 1 olive pollen allergen	AF532753	OleEu
57	Ole e 1_57A25	Ole e 1 olive pollen allergen	AF532754	OleEu
57	Ole e 1_57A26	Ole e 1 olive pollen allergen	AY137487	OleEu
57	Ole e 1_57A26	Ole e 1 olive pollen allergen	AY137488	OleEu
57	Ole e 1_57A29	Ole e 1 olive pollen allergen	AY137469	OleEu
57	Ole e 1_57A30	Ole e 1 olive pollen allergen	S75766	OleEu
57	Ole e 1_57A31	Ole e 1 olive pollen allergen	Y13436	OleEu
57	Ole e 1_57A32	Ole e 1 olive pollen allergen	AF532758	OleEu
57	Ole e 1_57A33	Ole e 1 olive pollen allergen	AF532761	OleEu
57	Ole e 1_57A34	Ole e 1 olive pollen allergen	AF532762	OleEu
57	Ole e 1_57A35	Ole e 1 olive pollen allergen	AF532759	OleEu
57	Ole e 1_57A36	Ole e 1 olive pollen allergen	AF532764	OleEu
57	Ole e 1_57A37	Ole e 1 olive pollen allergen	AF532763	OleEu
57	Ole e 1_57A38	Ole e 1 olive pollen allergen	AY159888	OleEu
57	Ole e 1_57A39	Ole e 1 olive pollen allergen	AY159881	OleEu
57	Ole e 1_57A40	Allergen Fra e 1	Q6U740	FraEX
57	Ole e 1_57A41	Ole e 1 olive pollen allergen	X76541	OleEu
57	Ole e 1_57A42	Ole e 1 olive pollen allergen	X76540	OleEu
57	Ole e 1_57A43	Ole e 1 olive pollen allergen	X76539	OleEu
58	Ole e 1_58A1	-	B7FNF5	MEDTR
58	Ole e 1_58A2	-	B7FNF3	MEDTR
58	Ole e 1_58B1	-	C8YEB3	SOYBN
59	Ole e 1_59A1	Extensin-like protein	A9WFR9	PICSI
59	Ole e 1_59A2	-	A9HPL2	PICSI
59	Ole e 1_59A3	Extensin-like protein	E8Z662	PICSI
59	Ole e 1_59A4	Extensin-like protein	E8ZE90	PICSI
59	Ole e 1_59A5	Extensin-like protein	E8ZE78	PICSI
60	Ole e 1_60A1	AT1g2710017199_16	Q9AEJ3	ARATH
60	Ole e 1_60A2	-	Q8LJ22	ARATH
60	Ole e 1_60A3	Pollen ole e 1 allergen	D7MUX1	ARALY
60	Ole e 1_60A4	-	C6S0Q8	SOYBN
60	Ole e 1_60A5	-	C8T474	SOYBN
60	Ole e 1_60A6	-	A9P8A0	POPTR
60	Ole e 1_60A7	-	A9PFL1	POPTR
60	Ole e 1_60A8	-	B9SAF6	RICCO
60	Ole e 1_60A9	PN40024	D7T895	VITI
60	Ole e 1_60A10	-	A2X417	ORYSJ
60	Ole e 1_60A12	Os02g0317800 protein	Q6Z411	ORYSJ
60	Ole e 1_60A13	Sb07g009930	C5YJW7	SORBI
60	Ole e 1_60B1	-	B8T344	MAIZE
60	Ole e 1_60B1	-	BLR1F7	PICSI
61	Ole e 1_61A1	-	Q8RWG5	ARATH

61	Ole e 1_61A2	At2g16630	Q8SLF4	ARATH
61	Ole e 1_61A3	-	D7L7M3	ARALY
61	Ole e 1_61B1	-	B9H158	POPTR
61	Ole e 1_61B2	-	A9PG40	POPTR
61	Ole e 1_61B3	-	B9SQJ5	RICCO
61	Ole e 1_61B4	-	D7U593	VITI
62	Ole e 1_62A1	PN40024	A3W276	ORYSJ
62	Ole e 1_62A2	-	A3A255	ORYSJ
62	Ole e 1_62B1	-	B6T727	MAIZE
62	Ole e 1_62B2	-	B4F2U6	MAIZE
62	Ole e 1_62B3	-	B6TR02	MAIZE
63	Ole e 1_63A1	-	B6UFQ0	MAIZE
64	Ole e 1_64A1	-	A9RQ15	PHYPA
64	Ole e 1_64B1	-	A9SHJ0	PHYPA
65	Ole e 1_65A1	-	D8TAV3	SELM
65	Ole e 1_65A2	-	D8TDP3	SELM
66	Ole e 1_66A1	-	D7M720	ARALY
66	Ole e 1_66A2	proline-rich glycoprotein	O64586	ARATH
66	Ole e 1_66A3	-	D7LH37	ARALY
67	Ole e 1_67A1	-	D7LGE2	ARALY
67	Ole e 1_67A2	At2g33790	P93013	ARATH
67	Ole e 1_67B1	At2g28290	Q9FZA2	ARATH
67	Ole e 1_67B2	proline-rich protein	Q9WPA7	ARATH
67	Ole e 1_67B3	-	D7RCU8	ARALY
67	Ole e 1_67C1	HyPRP1	Q8PHW3	GOSHI
67	Ole e 1_67D1	Arabinogalactan protein	C8Y0U7	GOSHI
68	Ole e 1_68A1	-	C8TLD2	SOYBN
68	Ole e 1_68A2	proline-rich protein	Q41122	PHAVU
68	Ole e 1_68B1	-	B7F159	MEDTR
68	Ole e 1_68C1	-	A9PAW5	POPTR
68	Ole e 1_68C2	-	A9PA42	POPTR
68	Ole e 1_68D1	-	B9N307	POPTR
68	Ole e 1_68D2	-	B9H7T3	POPTR
68	Ole e 1_68D3	-	A9P8A5	POPTR
68	Ole e 1_68E1	Structural constituent of cell wall	B9RBC9	RICCO
68	Ole e 1_68F1	PN40024	D7GB84	VITI
68	Ole e 1_69G1	Arabinogalactan protein	Q8FSW6	DAUCA
69	Ole e 1_69A1	hybrid proline-rich protein PRP1	Q8XES6	TRISU
70	Ole e 1_70A1	Proline-rich protein	Q878R4	NICAL
70	Ole e 1_70A2	-	C8LLS3	PETHY
70	Ole e 1_70A3	Proline-rich protein 1	Q6QNA3	CAPAN

Table 1. (continued). The Ole e 1 protein superfamily: new and unified nomenclature.

71	Ole e 1_71A1	Pistil extensin-like protein	Q40385	NICAL
71	Ole e 1_71A2	Pistil-specific extensin-like prot.	Q03211	PEXLF
71	Ole e 1_71B1	Pistil extensin-like protein	Q40549	TOBAC
71	Ole e 1_71B1	Pistil extensin-like protein	Q40552	TOBAC
72	Ole e 1_72A1	120 kDa styly glycoprotein	Q49986	NICAL
72	Ole e 1_72A2	120 kDa pistil extensin-like prot.	Q49278	NiClA
72	Ole e 1_72A3	120 kDa pistil extensin-like prot.	Q49279	NiClA
72	Ole e 1_72A4	120 kDa pistil extensin-like prot.	Q49277	NiClA
72	Ole e 1_72A5	120 kDa pistil extensin-like prot.	Q49322	NiClA
72	Ole e 1_72A6	120 kDa pistil extensin-like prot.	Q49333	NICPL
72	Ole e 1_72A7	120 kDa pistil extensin-like prot.	Q49334	TOBAC
73	Ole e 1_73A1	120 kDa pistil extensin-like prot.	Q49330	NiClA
74	Ole e 1_73A1	Pollen ole e 1 allergen	D7M426	ARALY
74	Ole e 1_73A2	Atg92279	OB1417	ARATH
75	Ole e 1_75A1	-	C57977	SOYBN
75	Ole e 1_75A2	Drought resistance protein	E0A235	SOYBN
75	Ole e 1_75A3	-	C5T425	SOYBN
75	Ole e 1_75B1	-	B9P93	POPTR
75	Ole e 1_75B2	-	B9P92	POPTR
75	Ole e 1_75C1	-	B8M440	POPTR
75	Ole e 1_75C2	-	B9P97	POPTR
75	Ole e 1_75D1	-	B9GSD7	POPTR
76	Ole e 1_76A1	-	B9SAV3	RICCO
76	Ole e 1_76B1	Structural constituent cell wall	B9SAV4	RICCO
77	Ole e 1_77A1	-	B9GSD1	POPTR
77	Ole e 1_77A2	Structural constituent cell wall	B9SAV3	RICCO
77	Ole e 1_77B1	PN40024	D7U2C5	VITV1
77	Ole e 1_77B2	-	ASB127	VITV1
77	Ole e 1_77C1	PN40024	D7U2C3	VITV1
77	Ole e 1_77C2	-	ASB125	VITV1
77	Ole e 1_77D1	Pollen ole e 1 allergen	D7LGP2	ARALY
77	Ole e 1_77D2	Atg947540	Q32257	ARATH
78	Ole e 1_78A1	-	ASB126	VITV1
78	Ole e 1_78A2	PN40024	D7U2C4	VITV1
78	Ole e 1_78A1	-	D7KQ21	ARALY
78	Ole e 1_78A2	-	D7KQ24	ARALY
78	Ole e 1_78A3	Proline-rich protein 1	Q9F235	ARATH
78	Ole e 1_78A4	Proline-rich protein 1	Q9M7P1	ARATH
79	Ole e 1_79A5	Proline-rich protein	Q9LZJ7	ARATH
79	Ole e 1_79A6	Proline-rich protein 3	Q9M7N3	ARATH
79	Ole e 1_79A7	-	D7L786	ARALY
80	Ole e 1_80A1	-	D7LGP0	ARALY

80	Ole e 1_80A2	-	-	Q22258	ARATH
81	Ole e 1_81A1	-	-	D8TC68	SELM1
81	Ole e 1_81A2	-	-	D8TF48	SELM1
81	Ole e 1_81B1	-	-	D8TC67	SELM1
82	Ole e 1_82A1	-	-	D8TC60	SELM1
83	Ole e 1_83A1	Pollen ole e 1 allergen	-	D7LX78	ARALY
83	Ole e 1_83A2	Atg505500	-	Q8FFG5	ARATH
83	Ole e 1_83A3	-	-	B9HHU1	POPTR
83	Ole e 1_83B2	-	-	B9HSK7	POPTR
83	Ole e 1_83B3	-	-	B9SQR0	RICCO
83	Ole e 1_83B4	PN40024	-	D7T4LT	VITV1
83	Ole e 1_83B5	-	-	A5C9V2	VITV1
84	Ole e 1_84A1	-	-	A2WL03	ORYSJ
84	Ole e 1_84A2	-	-	Q8L1N2	ORYSJ
84	Ole e 1_84A3	-	-	A2WL01	ORYSJ
84	Ole e 1_84A4	-	-	B9ET17	ORYSJ
84	Ole e 1_84A5	B1189A09.32	-	Q5VR32	ORYSJ
84	Ole e 1_84A6	-	-	A2WL00	ORYSJ
84	Ole e 1_84A7	-	-	A2WL05	ORYSJ
84	Ole e 1_84A8	-	-	A2WL09	ORYSJ
84	Ole e 1_84A9	-	-	Q8LJM3	ORYSJ
84	Ole e 1_84A10	-	-	A2WL07	ORYSJ
84	Ole e 1_84A11	-	-	Q8LJM3	ORYSJ
84	Ole e 1_84A12	-	-	A2WL04	ORYSJ
84	Ole e 1_84A13	-	-	Q8LJN3	ORYSJ
84	Ole e 1_84A14	-	-	A2ZPK8	ORYSJ
84	Ole e 1_84A15	B1189A03.34	-	Q5VR30	ORYSJ
84	Ole e 1_84B1	-	-	A2WL02	ORYSJ
84	Ole e 1_84C1	-	-	A2WL12	ORYSJ
84	Ole e 1_84C2	-	-	Q8LJM5	ORYSJ
84	Ole e 1_84C3	-	-	B9ADE3	ORYSJ
84	Ole e 1_84C4	-	-	A2ZPL3	ORYSJ
84	Ole e 1_84C5	B1189A03.42	-	Q5VR13	ORYSJ
84	Ole e 1_84C6	-	-	A2WL11	ORYSJ
84	Ole e 1_84C7	-	-	Q8LJM6	ORYSJ
84	Ole e 1_84D1	B1189A09.38	-	Q5VR19	ORYSJ
84	Ole e 1_84E1	-	-	B9ET08	ORYSJ
84	Ole e 1_84F1	-	-	A2ZPL6	ORYSJ
84	Ole e 1_84F2	B1189A09.43	-	Q5VR17	ORYSJ
84	Ole e 1_84G1	-	-	A2WL13	ORYSJ
84	Ole e 1_84G2	Sb03g050960	-	C5XP43	SORBI
85	Ole e 1_85A1	-	-	D8SH48	SELM1

85	Ole e 1_85B1	-	-	D8T583	SELM1
86	Ole e 1_86A1	-	-	D8SHH9	SELM1
87	Ole e 1_87A1	-	-	D8T584	SELM1
88	Ole e 1_88A1	-	-	D8R4E3	SELM1
88	Ole e 1_88A2	-	-	D8RKB7	SELM1
89	Ole e 1_89A1	Pollen ole e 1 allergen	-	D7L8E4	ARALY
89	Ole e 1_89A2	-	-	Q3EBA2	ARATH
89	Ole e 1_89B1	-	-	B9M7K3	POPTR
89	Ole e 1_89B2	-	-	B9S0B7	RICCO
89	Ole e 1_89C1	-	-	C5T3U0	SOYBN
90	Ole e 1_90A1	PN40024	-	D7TW93	VITV1
90	Ole e 1_90A2	PN40024	-	D7TIX1	VITV1
91	Ole e 1_91A1	-	-	A2Z3X7	ORYSJ
91	Ole e 1_91A2	Oe1g0546100	-	Q9A2V1	ORYSJ
91	Ole e 1_91B1	Sb01g030980	-	C5WTH1	SORBI
92	Ole e 1_92A1	-	-	D7LH66	ARALY
92	Ole e 1_92A2	At2g41400	-	Q6D6F8	ARATH
92	Ole e 1_92A3	RAF122-03-M12	-	Q6Z171	ARATH
92	Ole e 1_92B1	At2g41400	-	Q4ZVC5	ARATH
92	Ole e 1_92B1	At2g41390	-	Q9ZVC4	ARATH
93	Ole e 1_93A1	At5g05020	-	Q9FF72	ARATH
94	Ole e 1_94A1	-	-	D7KW90	ARALY
95	Ole e 1_95A1	At3g16660	-	Q9QYV6	ARATH
95	Ole e 1_95A2	MGL5	-	Q5LUR8	ARATH
95	Ole e 1_95A3	-	-	D7L657	ARALY
95	Ole e 1_95B1	-	-	D7L658	ARALY
95	Ole e 1_95B2	AT3g16670-MGL6.12	-	Q4LUR6	ARATH
95	Ole e 1_95C1	-	-	B9HXT5	POPTR
95	Ole e 1_95C2	PH40024	-	D7SYD8	VITV1
95	Ole e 1_95C3	Phytoplanin	-	B8RT72	RICCO
95	Ole e 1_95D1	-	-	C6TK66	SOYBN
95	Ole e 1_95E1	Phytoplanin	-	Q5E5S9	PHYLL
95	Ole e 1_95F1	Phytoplanin	-	Q1PCF2	TOBAC
96	Ole e 1_96A1	-	-	B9APE1	ORYSJ
96	Ole e 1_96A2	-	-	B9F8E1	ORYSJ
96	Ole e 1_96B1	-	-	A2YV44	ORYSJ
96	Ole e 1_96B2	Os07g0874400 protein	-	Q6ZDW6	ORYSJ
96	Ole e 1_96B3	Sb02g042730	-	C5X19	SORBI
96	Ole e 1_96B4	-	-	B4FF5J	MAIZE
96	Ole e 1_96B5	Sb01g035810	-	C5XQ00	SORBI
97	Ole e 1_97A1	-	-	A2YV45	ORYSJ
97	Ole e 1_97A2	Os07g0674500	-	Q6ZDW7	ORYSJ

97	Ole e 1_97A3	Sb02g042740	-	C5X5J0	SORBI
98	Ole e 1_98A1	-	-	A2XGJ7	ORYSJ
98	Ole e 1_98A2	-	-	O1BLN4	ORYSJ
98	Ole e 1_98B1	-	-	A2XGJ8	ORYSJ
98	Ole e 1_98B2	Os03g0342100	-	Q1BLN2	ORYSJ
98	Ole e 1_98C1	Sb01g035830	-	C5X0Q2	SORBI
99	Ole e 1_99A1	-	-	B9UHT3	MAIZE
100	Ole e 1_100A1	-	-	A2JWU2	ORYSJ
100	Ole e 1_100A2	Os01g0725900	-	Q8S15E	ORYSJ
100	Ole e 1_100A3	-	-	A2ZXF1	ORYSJ
100	Ole e 1_100A4	Sb03g033350	-	C5XIF5	SORBI
100	Ole e 1_100B1	Sb03g033360	-	C5XIF6	SORBI
100	Ole e 1_100C1	QJ1131_E09.17	-	Q75K33	ORYSJ
100	Ole e 1_100A2	-	-	B9FH39	ORYSJ
100	Ole e 1_100C3	-	-	A2Y6T6	ORYSJ
100	Ole e 1_100C4	Os05g0531400 protein	-	Q8DGH6	ORYSJ
100	Ole e 1_100D1	Sb03g026510	-	C5YUF6	SORBI
100	Ole e 1_100D2	Arabinogalactan protein	-	B8SLV3	MAIZE
101	Ole e 1_101A1	-	-	A2Y6T4	ORYSJ
101	Ole e 1_101A2	Os05g0531200 protein	-	Q75K55	ORYSJ
101	Ole e 1_101B1	Pistil-specific extensin-like protein	-	B6UHM8	MAIZE
101	Ole e 1_101B2	-	-	B4FGH8	MAIZE
101	Ole e 1_101B3	-	-	C5Z1H9	SORBI
102	Ole e 1_102A1	-	-	A2WUJ3	ORYSJ
102	Ole e 1_102A2	Os01g0726100 protein	-	Q8S154	ORYSJ
102	Ole e 1_102B1	Sb03g033370	-	C5XIF7	SORBI
102	Ole e 1_102B2	Pistil-specific extensin-like prot.	-	B6UHE3	MAIZE
103	Ole e 1_103A1	-	-	D8ZL93	SELM1
104	Ole e 1_104A5	-	-	B4FMQ8	MAIZE
105	Ole e 1_105A1	-	-	D8RC12	SELM1
106	Ole e 1_106A1	-	-	D8T9S5	SELM1
107	Ole e 1_107A1	-	-	A9SC13	PHYPA
109	Ole e 1_108A1	-	-	D9Q767	SELM1
109	Ole e 1_109A1	-	-	D8SSV6	SELM1

Table 1. (continued). The Ole e 1 protein superfamily: new and unified nomenclature.

3.2 Phylogenetic analysis of the extended Ole e 1 protein families

A member of each retrieved full-length Ole e 1 sequences family was aligned to determine phylogenetic relationships within the Ole e 1 extended family. A phylogenetic tree of the Ole e 1 extended sequences is depicted in Figure 1.

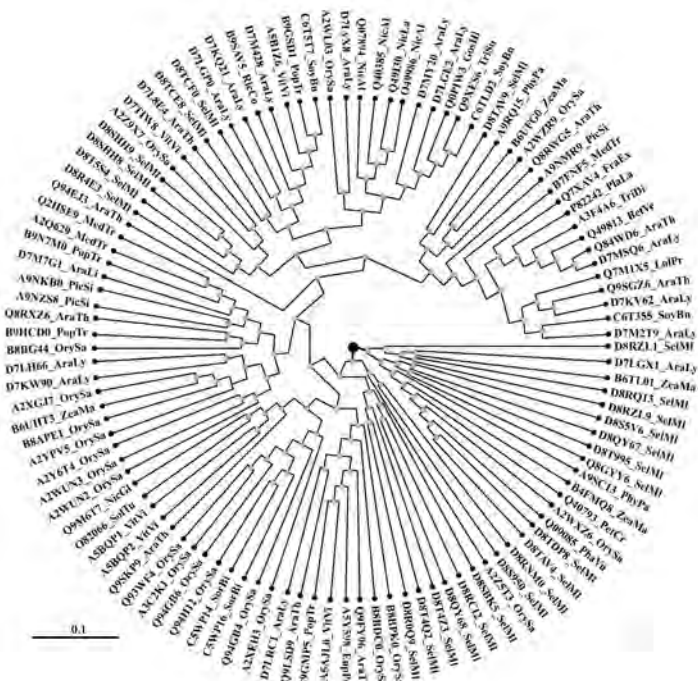


Fig. 1. Phylogenetic analysis of plant Ole e 1 proteins. Neighbour-Joining (NJ) method was used to perform a phylogenetic analysis of Ole e 1 proteins from 109 families. One representative sequence of each family was used, based in its higher consensus ability. Plant species analyzed included *Arabidopsis*, poplar, rice, spikemoss, tobacco, maize, potato, grape, *Sorghum*, kidney bean, barrel medic, *Pinus*, poinsettia, perennial ryegrass, soybean, white birch, ash, *Platanus*, *Physcomitrella*, cotton, subterranean clover, Persian tobacco and castor bean.

The phylogenetic tree shows that the 109 Ole e 1 extended families, although highly divergent, are split into two clades. The smaller clade was integrated by a few species like *Selaginella moellendorffii*, *Arabidopsis* and maize among others. The second clade included the majority of the Ole e 1 family proteins, clustering together almost all the biological functions (Figure 1). Numerous branches aroused from this clade.

3.3 Ole e 1 protein superfamilies: Structural and conformational variability

The crystallographic structural coordinates of relatively few proteins of the Ole e 1 family have been deposited in the Protein Database (PDB) up to date. To our knowledge, detailed comparative studies of the structural and conformational features of members of the Ole e 1

extended protein families have not been performed in higher plants. Using computational modelling analysis, we have determined and modelled the molecular-structural features of selected members of the Ole e 1 extended families. A first overview of the generated models (Figure 2) indicated a relatively high level of similitude.

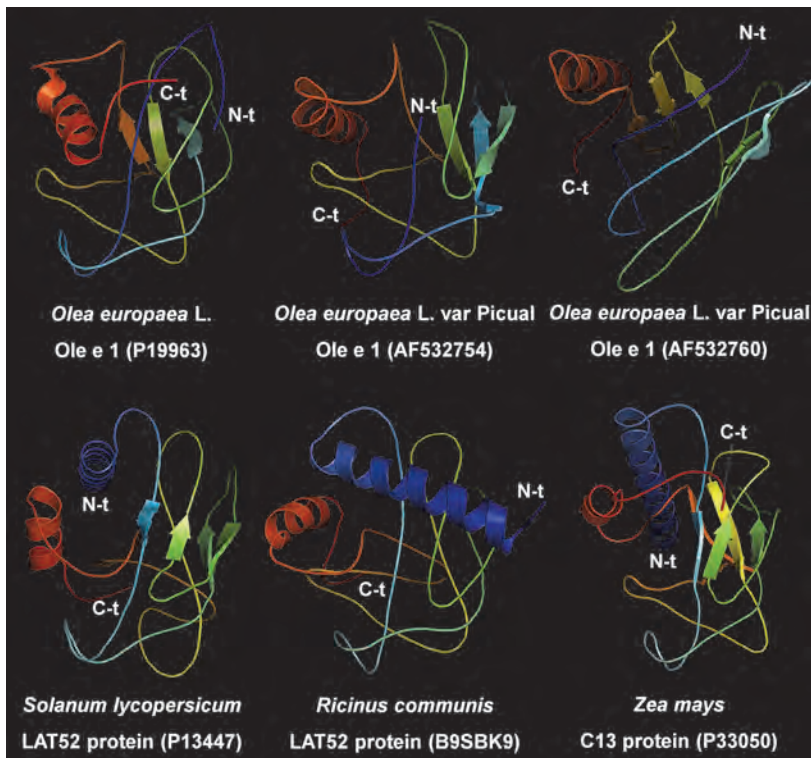


Fig. 2. Three-dimensional structure analysis of selected members of Ole e 1 family proteins. The model proteins are depicted as cartoon diagrams. The secondary elements of the crystallographic structures are rainbow coloured, with N-terminus in blue, and C-terminus in red.

However, a more detailed analysis allowed identifying certain differences in the generated models, particularly consisting in 2D structural features. These differences can be distinguished even between very close proteins like P19963, AF532754 and AF532760 (Ole e 1_57A9, Ole e 1_57A25 and Ole e 1_57A23 with the new nomenclature), corresponding to the olive pollen major allergen cloned from different varietal sources or even to different clones of the same cultivar (Figure 2). The differences become higher when models of the same protein obtained from different plant species are compared. This is the case of P13447 and B9SBK9 (Ole e 1_52L1 and Ole e 1_52J1), which correspond to the LAT52 gene product in tomato and *Ricinus*, respectively (Figure 2). Divergences are even more obvious between the models indicated above and that of a P33050 (Ole e 1_48H6), a different member of the Ole e 1 superfamily corresponding to a pollen protein from maize (C13 protein) (Figure 2).

4. Discussion

Research as regard to the proteins of the Ole e 1 family has been carried out steadily since its definition. At present, many genes from the allergen Ole e 1 family of proteins have been characterized, and data are available concerning the sequence, structure, expression and biological function (e.g. extensin-like proteins constituting part of the cell wall). However, and as depicted in this chapter, the precise identification of more than half members of this family remains uncompleted. Up to now, Ole e 1 and Ole e 1-like genes are deposited into the databases, many of them with repetitive or arbitrary naming system by authors. This nomenclature includes a variety of generic names, such as Ole e 1 major olive pollen allergen, putative Ole e 1-like protein, anther-specific Ole e 1-like protein, and others depending of the protein location in the chromosome, e.g. At3g26960, Os09g0508200, or simply giving a random name e.g. P1 clone: MOJ10. For those members of the Ole e 1 family which have been recognized like allergens, a more sustainable and precise nomenclature has been built, by following the recommendations of the International Union of Immunological Societies (IUIS) (<http://www.allergen.org/>). However, these allergenic proteins only represent a part of the members of the Ole e 1 family, and this nomenclature still does not display the relationships among these proteins. In several cases, it is still common for researchers to use different names for the same allergen. Allergen biochemistry is now entering a new time of structural biology and proteomics that will require sophisticated tools for data processing and bioinformatics, and might require further definition of the nomenclature. Increasingly, the wealth of structural information is enabling the biologic function of allergens to be established and the assignment of allergen function to diverse protein families. Therefore, the arbitrary nomenclature currently in use is not sustainable for adequate comparative mega-functional genomics studies, especially as the number of Ole e 1 genes has increased steadily and will continue with this upward trend with the completion of the sequencing projects corresponding to more plant genomes.

The implementation of modifications in the nomenclature as proposed here may assist further developments of allergy understanding and new clinical approaches. As an example, nomenclature and structural biology have been proposed to play a crucial role in defining allergens for research studies and for the development of new clinical products [Chapman et al. 2007]. Sequence comparisons and assignments to protein families provide a molecular basis for clinical cross-reactions between food, pollen, and latex allergens that give rise to oral allergy syndromes [Wagner et al. 2002, Scheiner et al. 2004, van Ree 2004]. For food and pollen allergens, intrinsic protein structure probably plays an important role in determining allergenicity by conferring, for example, heat stability or resistance to digestion in the digestive tract, e.g. storage proteins from seed/nuts or legumes [Orruño and Morgan 2011]. Interestingly, analysis of databases, e.g. pFAM shows that there are currently more than 120 molecular architectures that are responsible for eliciting IgE responses. It will be important to link nomenclature with classification of allergens into protein families and subfamilies to provide complete definition of allergens and their structure-functional relationships as part of a comprehensive bioinformatics database. The practical consequences of this approach are seen most clearly with genetically modified foods, in which sequence comparisons can be used for safety assessment of genetically modified organisms [Goodman and Tetteh 2011].

The success of our new and unified nomenclature lies in its simplicity, with genetic basis and structural-functional characterizations of the proteins, regardless of the species origin,

with the possibility to further nomenclature expansion, to include as-yet-unidentified protein allergens from different sources or species: mites, insects, pollens, molds and foods. It might be also possible to include in the system engineered protein molecules, such as hypoallergens, or others being described as non-protein allergens. Allergens entered into the nomenclature could be used to develop allergen-specific diagnostics and to formulate recombinant allergen vaccines that will benefit patients [Chapman et al. 2000, Ferreira et al. 2004, Jutel et al. 2005, Sastre 2010].

The proposed system may also assist to clarify the importance of allergen polymorphism. Allergens often display numerous variants. These are proteins with typically greater than 90% sequence identity, but with enough differences in their amino acid sequences to make worth individual structural and or functional characterization and identification. This polymorphism has been deeply analyzed in mites, as their allergens present an extensive number of isoforms: 23 for Der p 1 and 13 for Der p 2 [Smith et al. 2001, Smith et al. 2001]. Furthermore, these polymorphisms might affect T-cell responses or alter antibody-binding sites. These differences can be structurally characterized to distinguish isoforms in a well-defined nomenclature system, by mean of structural-functional differentiation, helping to design allergen formulations for immunotherapy [Jutel et al. 2005, Piboonpocanun et al. 2006]. In the case of pollen allergens, Ole e 1 from olive pollen is a clear example of extreme polymorphism, both in its peptide and in its carbohydrate moieties, as demonstrated by peptide mapping and N-glycopeptide analysis [Castro et al. 2010]. Olive cultivar origin is a major cause of polymorphism for Ole e 1 pollen allergen [Hamman-Khalifa et al. 2008, Castro et al. 2010]. The olive tree has an extremely wide germplasm, with over 1200 varieties cultivated over the world [Bartolini et al. 1994]. Therefore, the number of Ole e 1 isoforms yet to be characterized in olive pollen is expected to be enormous. A similar situation is also likely to occur in many other plant species.

Overall, our developed unified nomenclature system is helpful in a quick functional prediction of any newly cloned Ole e 1 gene(s), because from the nomenclature point of view, the newly sequenced gene(s) will always be characterized/named with sequence similarity with previously characterized Ole e 1 genes/proteins, as well as a protein structure-functional characterization and comparison. The changes that have been introduced reflect into which extended family or subfamily a certain Ole e 1 protein belongs. Accordingly, the new nomenclature will have no significant impact on already published data with old/arbitrary naming system. However, we urge scientists working on Ole e 1's to adopt this new and easy nomenclature system. In this regard, we have made an effort to preserve the user friendly linkage between the old and the new designations, which we hope will help researchers to adapt the new names. As the revised nomenclature should facilitate communication and understanding within the community interested in Ole e 1 allergen proteins, we advocate that this new naming system be used in all future studies.

The classification model used here has been developed under the basis of a previously designed gene nomenclature model for male fertility restorer (RF) proteins in higher plants [Kotchoni et al. 2010]. The increasing numbers of RF genes described in the literature represented an ongoing challenge in their clear identification and logical classification which was solved using the proposed nomenclature. Undoubtedly, similar approaches could be applied to numerous protein families involving relevant levels of nomenclature heterogeneity, many of them registered in specialized databases like pFam. In the case of allergens, other numerous protein families like profilins (Ole e 2 in the case of olive pollen)

prolamins, cupins, Bet v 1-related proteins etc., which are currently included in the AllFam database [Radauer et al. 2008] (<http://www.meduniwien.ac.at/allergens/allfam/>) could benefit of the use of similar approaches.

5. Conclusion

We propose for first time a unified naming system for Ole e 1-like genes and pseudogenes across all plant species, which accommodates the numerous sequences already deposited in several databases, offering the needed flexibility to incorporate additional Ole e 1-like proteins as they become available. Additionally, we provide an analysis of the phylogenetic relationships displayed by the members of the Ole e 1-like family and use computational protein modelling to determine structural features of selected members of this family. These data are of particular relevance for the understanding of their biological activity and allergenic cross-reactivity.

6. Acknowledgment

Support of the Spanish Ministry of Science and Innovation (ERDF-cofinanced project BFU2008-00629) and Andalusian Regional Government (ERDF-cofinanced Proyectos de Excelencia CVI5767 and AGR6274) is gratefully acknowledged.

7. References

- Alché, J.D.; Castro, A.J.; Olmedilla, A.; Fernández, M.C.; Rodríguez, R.; Villalba, M. and Rodríguez-García, M.I. (1999). The major olive pollen allergen (Ole e I) shows both gametophytic and sporophytic expression during anther development, and its synthesis and storage takes place in the RER. *Journal of Cell Science*, Vol.112, pp.2501-2509
- Alché, J.D.; M'rani-Alaoui, M.; Castro, A.J. and Rodríguez-García, M.I. (2004). Ole e 1, the major allergen from olive (*Olea europaea* L.) pollen, increases its expression and is released to the culture medium during in vitro germination. *Plant Cell Physiology*, Vol.45, pp.1149-1157
- Altschul, S.F.; Gish, W.; Miller, W.; Myers, E.W. and Lipman, D.J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*, Vol.215, No.3, pp.403-410
- Altschul, S.F.; Madden, T.L.; Schäffer, A.A.; Zhang, J.; Zhang, Z.; Miller, W. and Lipman, D.J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, Vol.25, No.17, pp.3389-402
- Barral, P.; Batanero, E.; Palomares, O.; Quiralte, J.; Villalba, M. and Rodríguez, R. (2004). A major allergen from pollen defines a novel family of plant proteins and shows intra- and interspecies cross-reactivity. *Journal of Immunology*, Vol.172, pp.3644-3651
- Bartolini, G.; Prevost, G. and Messeri, C. (1994). Olive tree germplasm: descriptor lists of cultivated varieties in the world. *Acta Horticulturae*, Vol.365, pp.116-118
- Batanero, E.; Villalba, M. and Rodríguez, R. (1994). Glycosylation site of the major allergen from olive tree. Allergenic implications of the carbohydrate moiety. *Molecular Immunology*, Vol.31, pp.31-37

- Castro, A.J.; Bednarczyk, A.; Schaeffer-Reiss, C.; Rodríguez-García, M.I.; Van Dorselaer, A.; Alché, J.D. (2010). Screening of Ole e 1 polymorphism among olive cultivars by peptide mapping and N-glycopeptide analysis. *Proteomics*, Vol. 10, No 5, pp.953-962
- Chapman, M.D.; Pomés, A.; Breiteneder, H. and Ferreira, F. (2007). Nomenclature and structural biology of allergens. *Journal of Allergy and Clinical Immunology*, Vol.119, No.2, pp.414-420
- Chapman, M.D.; Smith, A.M.; Vailes, L.D.; Arruda, K.; Dhanaraj, V. and Pomes, A. (2000). Recombinant allergens for diagnosis and therapy of allergic diseases. *The Journal of Allergy and Clinical Immunology*, Vol.106, pp.409-418
- Chevenet, F.; Brun, C.; Banuls, A.L.; Jacq, B. and Christen, R. (2006). TreeDyn: towards dynamic graphics and annotations for analyses of trees. *BMC Bioinformatics*, Vol.7, pp.439
- D'Amato, G.; Spiekma, F.T.; Liccardi, G.; Jager, S.; Russo, M.; Kontou-Fili, K.; Nikkels, H.; Wuthrich, B. and Bonini, S. (1998). Pollen-related allergy in Europe. *Allergy*, Vol.53, pp.67-78
- de Castro, E.; Sigrist, C.J.A.; Gattiker, A.; Bulliard, V.; Langendijk-Genevaux, P.S.; Gasteiger, E.; Bairoch, A. and Hulo, H. (2006) ScanProsite: detection of PROSITE signature matches and ProRule-associated functional and structural residues in proteins. *Nucleic Acids Research*, Vol.34, pp.362-365
- Ferreira, F.; Wallner, M. and Thalhamer, J. (2004). Customized antigens for desensitizing allergic patients. *Advances in Immunology*, Vol.84, pp.79-129
- Finn, R.D.; Mistry, J.; Tate, J.; Coggill, P.; Heger, A.; Pollington, J.E.; Gavin, O.L. Gunasekaran, P.; Ceric, G. Forslund, K.; Holm, L.; Sonnhammer, E.L.; Eddy, S.R. and Bateman, A. (2010). The Pfam protein families database. *Nucleic Acids Research*, Database Issue 38, pp.D211-222
- Gasteiger, E.; Gattiker, A.; Hoogland, C.; Ivanyi, I.; Appel R.D. and Bairoch A. (2003) ExPASy: the proteomics server for in-depth protein knowledge and analysis. *Nucleic Acids Research*, Vol.31, pp.3784-3788
- Goodman, R.E. and Tetteh, A.O. (2011). Suggested Improvements for the Allergenicity Assessment of Genetically Modified Plants Used in Foods. *Current Allergy and Asthma Reports*, doi: 10.1007/s11882-011-0195-6
- Guex, N. and Peitsch, M.C. (1997). SWISS-MODEL and the Swiss-PdbViewer: an environment for comparative protein modeling. *Electrophoresis*, Vol.18, No.15, pp.2714-2723
- Hall, T.A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, Vol.41, pp.95-98
- Hamman-Khalifa, A.M.; Castro A.J.; Jimenez-Lopez, J.C.; Rodríguez-García, M.I. and Alché, J.D. (2008). Olive cultivar origin is a major cause of polymorphism for Ole e 1 pollen allergen. *BMC Plant Biology*, Vol.8, 10
- Hanson, D.D.; Hamilton, D.S.; Travis, J.L.; Bashe, D.M. and Mascarenhas, J.P. (1998). Characterization of a pollen-specific cDNA clone from *Zea mays* and its expression. *Plant Cell*, Vol.1, pp.173-179
- Hauser, M.; Roulias, A.; Ferreira, F. & Egger, M. (2010). Panallergens and their impact on the allergic patient. *Allergy, Asthma & Clinical Immunology*, Vol.6, pp.1-

- Jutel, M.; Jaeger, L.; Suck, R.; Meyer, H.; Fiebig, H. and Cromwell, O. (2005). Allergenspecific immunotherapy with recombinant grass pollen allergens. *The Journal of Allergy and Clinical Immunology*, Vol.116, pp.608-613
- Laskowski, R.A.; MacArthur, M.W.; Moss, D.S. and Thornton, J.M. (1993). PROCHECK: A program to check the stereo-chemical quality of protein structures. *Journal of Applied Crystallography*, Vol.26, pp.283-291
- Lauzurica, P.; Gurbindo, C.; Maruri, N.; Galocha, B.; Diaz, R.; Gonzalez, J.; García, R. and Lahoz, C. (1988). Olive (*Olea europea*) pollen allergens—I. Immunochemical characterization by immunoblotting, CRIE and immunodetection by a monoclonal antibody. *Molecular Immunology*, Vol.25, pp.329-335
- King, T.P.; Hoffman, D.; Lowenstein, H.; Marsh, D.G.; Platts-Mills, T.A. and Thomas, W. (1994). Allergen nomenclature. WHO/IUIS Allergen Nomenclature Subcommittee. *International Archives of Allergy and Immunology*, Vol. 105, pp. 224-233
- Kotchoni, S.O.; Jimenez-Lopez, J.C.; Gachomo, W.E. and Seufferheld, M.J. (2010). A new and unified nomenclature for male fertility restorer (RF) proteins in higher plants. *PLoS ONE*, Vol.5, No.12, pp.e15906
- Melo, F. and Feytmans, E. (1997). Novel knowledge-based mean force potential at atomic level. *Journal of Molecular Biology*, Vol.267, No.1, pp.207-222
- Melo, F. and Feytmans, E. (1998). Assessing protein structures with a non-local atomic interaction energy. *Journal of Molecular Biology*, Vol.277, No.5, pp.1141-1152
- Mothes, N.; Horak, F. & Valenta, R. (2004). Transition from a botanical to a molecular classification in tree pollen allergy: implications for diagnosis and therapy. *International Archives of Allergy and Immunology*, Vol.135, pp.357-373
- Orruño, E. and Morgan, M.R.A. (2011). Resistance of purified seed storage proteins from sesame (*Sesamum indicum* L.) to proteolytic digestive enzymes. *Food Chemistry*, in press
- Piboonpocanun S, Malinual N, Jirapongsananuruk J, Vichyanond P, Thomas WR. (2006). Genetic polymorphisms of major house dust mite allergens. *Clinical & Experimental Allergy*, Vol.36, pp.510-516
- Radauer, C.; Bublin, M.; Wagner, S.; Mari, A. and Breiteneder, H. (2008). Allergens are distributed into few protein families and possess a restricted number of biochemical functions. *Journal of Allergy and Clinical Immunology*, Vol.121, pp.847-852
- Rodríguez, R.; Villalba, M.; Batanero, E.; González, E.M.; Monsalve, R.I.; Huecas, S.; Tejera, M.L. and Ledesma, A. (2002). Allergenic diversity of the olive pollen. *Allergy*, Vol.57, pp.6-16
- Rodríguez, R.; Villalba, M.; Monsalve, R.I.; Batanero, E.; González, E.M.; Monsalve, R.I.; Huecas, S.; Tejera, M.L. and Ledesma, A. (2002). Allergenic diversity of the olive pollen. *Allergy*, Vol.57, pp.6-15
- Salamanca, G.; Rodríguez, R. Quiralte, J.; Moreno, C.; Pascual, C.Y.; Barber, D. and Villalba, M. (2010). Pectin methylesterases of pollen tissue, a major allergen in olive tree. *FEBS Journal*, Vol.277, No.13, pp.2729-2739
- Sastre, J. (2010). Molecular diagnosis in allergy. *Clinical & Experimental Allergy*, Vol.40, No.10, pp.1442-1460
- Scheiner, O.; Aberer, W.; Ebner, C.; Ferreira, F.; Hoffmann-Sommergruber, K.; Hsieh, L.S.; Kraft, D.; Sowka, S.; Vanek-Krebitz, M. and Breiteneder, H. (1997). Cross-reacting

- allergens in tree pollen and pollen-related food allergy: implications for diagnosis of specific IgE. *International Archives of Allergy and Immunology*, Vol.113, pp.105-108
- Shultz, J.L.; Kurunam, D.; Shopinski, K.; Iqbal, M.J.; Kazi, S.; Zobrist, K.; Bashir, R.; Yaegashi, S.; Lavu, N.; Afzal, A.J.; Yesudas, C.R.; Kassem, M.A.; Wu, C.; Zhang, H.B.; Town, C.D.; Meksem, K. and Lightfoot, D.A. (2006). The Soybean Genome Database (SoyGD): a browser for display of duplicated, polyploid, regions and sequence tagged sites on the integrated physical and genetic maps of *Glycine max*. *Nucleic Acids Research*, Vol.34(suppl 1), pp.D758-D765
- Sigrist, C.J.A.; Cerutti, L.; de Castro, E.; Langendijk-Genevaux, P.S.; Bulliard, V.; Bairoch, A. and Hulo, N. (2010). PROSITE, a protein domain database for functional characterization and annotation. *Nucleic Acids Research*, Vol.38 (Database issue), pp.161-166
- Smith, A.S.; Benjamin, D.C.; Hozic, N.; Derewenda, U.; Smith, W.A.; Thomas, W.R.; Gafvelin, G.; van Hage-Hamsten, M. and Chapman, M.D. (2001). The molecular basis of antigenic cross-reactivity between the group 2 mite allergens. *The Journal of Allergy and Clinical Immunology*, Vol.107, pp.977-984
- Smith, W.A.; Hales, B.J.; Jarnicki, A.G. and Thomas W.R. (2001). Allergens of wild house dust mites: environmental Der p 1 and Der p 2 sequence polymorphisms. *The Journal of Allergy and Clinical Immunology*, Vol.107, pp.985-992
- Stratford, S.; Barne, W.; Hohorst, D.L.; Sagert, J.G.; Cotter, R.; Golubiewski, A.; Showalter, A.M.; McCormick, S. and Bedinger, P. (2001). A leucine-rich repeat region is conserved in pollen extensin-like (Pex) proteins in monocots and dicots. *Plant Molecular Biology*, Vol.46, pp.43-56
- Tang, B.; Banerjee, B.; Greenberger, P.A.; Fink, J.N.; Kelly, K.J. and Kurup, V.P. (2000). Antibody binding of deletion mutants of Asp f 2, the major *Aspergillus fumigatus* allergen. *Biochemical and Biophysical Research Communications*, Vol.270, pp.1128-1135
- Thompson, J.D.; Higgins, D.G. and Gibson, T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, Vol.22, pp.4673-4680
- Twell, D.; Wing, R.; Yamaguchi, J. and McCormick, S. (1989). Isolation and expression of an anther-specific gene from tomato. *Molecular and General Genetics*, Vol.217, pp.240-245
- van Gunsteren, W.F.; Billeter, S.R.; Eising, A.A.; Hünenberger, P.H.; Krüger, P.; Mark, A.E.; Scott, W.R.P. and Tironi, I.G. (1996). Biomolecular Simulations: The GROMOS96 Manual and User Guide. Zürich, VdF Hochschulverlag ETHZ
- van Ree R. (2004). Clinical importance of cross-reactivity in food allergy. *Current Opinion in Allergy & Clinical Immunology*. Vol.4, pp.235-240
- Villalba, M.; Batanero, E.; Lopez-Otin, C.; Sanchez, L.M.; Monsalve, R.I.; Gonzalez de la Pena, M.A.; Lahoz, C. and Rodriguez, R. (1993). The amino acid sequence of Ole e I, the major allergen from olive tree (*Olea europaea*) pollen. *European Journal of Biochemistry*, Vol.216, pp.863-869
- Villalba, M.; López-Otín, C.; Martín-Orozco, E.; Monsalve, R.I.; Palomino, P.; Lahoz, C. and Rodríguez, R. (1990). Isolation of three allergenic fractions of the major allergen from *Olea europaea* pollen and N-terminal amino acid sequence. *Biochemical and Biophysical Research Communications*, Vol.172, pp.523-528
- Wagner, S. and Breiteneder, H. (2002). The latex-fruit syndrome. *Biochemical Society Transactions*, Vol.6, pp.935-940