

The use of demineralisation and torrefaction to improve the properties of biomass intended as a feedstock for fast pyrolysis

Tansy Wigley*, Alex C.K. Yip, Shusheng Pang

Department of Chemical and Process Engineering, University of Canterbury, New Zealand

Abstract

Pre-treatments of biomass were investigated to reduce its undesirable properties which may affect the quality of fast pyrolysis bio-oil. A pre-treatment sequence was developed in this study to incorporate both biomass demineralisation and torrefaction. Demineralisation was performed by dilute acid leaching, primarily to reduce the inorganic concentration in raw biomass, whereas torrefaction targeted a reduction of the carboxyl, moisture and oxygen content. The liquid produced during torrefaction was recycled back as the leaching reagent for demineralisation. This solution contained dilute organic acids; therefore, the viability of leaching with organic acids (acetic and formic acid) compared to commonly used mineral acids (sulphuric, nitric and hydrochloric acid) was validated. Synthetic leaching solutions reduced the inorganic content in raw biomass from 0.41 wt% to 0.14 wt% when leached with 1% formic acid and to 0.16 wt% when leached with 1% acetic acid, which was comparable to leaching with the mineral acids. Recycled torrefaction liquid that contained other acidic compounds in small quantities reduced the inorganic content to 0.14 wt%, suggesting it is effective to use the recycled torrefaction liquid as the leaching solution. From the experimental results, the optimal conditions for biomass torrefaction were 260 °C for 20 min to minimise the char formation during pyrolysis, based on the increase in the acid-insoluble fraction of the biomass. However, the torrefaction temperature may be increased to 280 °C if further reductions in acetyl and oxygen content are required. Higher temperatures are associated with severe biomass loss and the initiation of hydrogen loss. It should be noted that even at 280 °C, the oxygen reduction is minimal. If oxygen reduction is the principal target when pre-treating biomass, it is suggested that torrefaction alone is not a suitable method to obtain bio-oil with a low oxygen content due to the low pyrolysis yields obtainable. This study demonstrated that the combined use of demineralisation and torrefaction as biomass pre-treatments has the ability to decrease the inorganic, acetyl and moisture content of biomass, which reduces undesirable catalytic reactions during fast pyrolysis to improve the quality of bio-oil produced.

Keywords: demineralisation; acid leaching; torrefaction; pre-treatments; pyrolysis

1 Introduction

1 *Pinus radiata* residues are a major by-product of the wood processing industry in New Zealand. In
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3 2001, approximately 1.3 Mm³ of wood processing residues remained unused and additional forest
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5 residues were not extracted [1]. New Zealand already has many sources of renewable electricity
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7 (such as solar, hydro, wind, geothermal and wave power [2, 3]). It would be beneficial if a
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9 transportation fuel could be produced from the wood residues. Since conversion of softwoods to
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11 ethanol is not currently effective [4], focus should be placed on processes such as gasification and
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13 fast pyrolysis.

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15 Fast pyrolysis is a thermal process in which biomass polymers are fragmented into light compounds
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17 that are able to volatise, thereby producing a product referred to as bio-oil. High heating rates and
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19 short vapour residence times employed during fast pyrolysis are preferred to maximise the liquid
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21 yield, opposed to slow pyrolysis which enhances char production [5]. The focus for bio-oil
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23 production is typically as an alternative to petroleum transportation fuels; however, the high
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25 acidity, reactivity, solids content, water content, viscosity and distillation residue restrict the direct
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27 use of bio-oil in combustion engines. The high oxygen content of biomass is reflected in the bio-
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29 oils elementary composition, subsequently decreasing its heating value and stability during storage
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31 or upon heating. The water content can reach 30 wt%, further decreasing the heating value to
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33 approximately half that of petroleum fuels. Typical properties for bio-oils are summarised and
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35 compared to those of a heavy fuel oil in Table 1.

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37 Upgrading crude bio-oil through either catalytic cracking or hydroprocessing has been extensively
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39 studied [14-27], however, the upgrading process is complicated due to the complexity of the bio-
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41 oil, resulting in low yields and high processing costs. Pre-treating biomass prior to pyrolysis has the
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43 potential to improve the quality of crude pyrolysis bio-oil and thus, increase the efficiency of
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45 upgrading via catalytic cracking and hydroprocessing processes through reduced reactor clogging
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47 and catalyst deactivation. The approach of combining demineralisation and torrefaction as pre-
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49 treatments has the capability to reduce the oxygen, carboxyl, water and ash content of biomass,
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51 subsequently decreasing the negative attributes of bio-oil that limit the efficiency of conventional
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53 refining techniques. If small, de-centralised pyrolysis plants could directly produce bio-oil of
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55 sufficient quality, the bio-oil could then be transported to existing petroleum refineries for co-
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57 processing with petroleum crude. Figure 1 presents a simplified version of the proposed system.
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59 The primary objective of this paper was to investigate the effectiveness of pre-treatments for
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61 reducing the negative attributes of biomass for subsequent fast pyrolysis.
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1 In the pre-treatment stages, biomass is initially demineralised during acid leaching, followed by the
2 removal of excess moisture. Biomass is then torrefied, which also represents the drying stage. The
3 liquid produced during torrefaction mainly contains water and organic acids therefore, this liquid is
4 recycled back into the demineralisation unit as the acid leaching reagent. The use of dilute organic
5 acids (< 1%) compared to commonly used mineral acids has not been reported so far; only 10%
6 acetic acid solutions have been tested previously [28]; therefore the secondary objective of this
7 study was to compare the efficiency for demineralisation with organic acids to mineral acids.
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10 11 12 *1.1 Acid leaching*

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14 Demineralisation typically involves leaching biomass in water or a dilute acid to reduce the
15 inorganic content (ash). It is well documented that inorganics catalyse dehydration and cracking
16 reactions during pyrolysis, lowering the bio-oil quality [29-33]. Additionally, inorganics facilitate
17 coke formation during the catalytic upgrading of bio-oil. Alkali and alkaline earth metals (AAEMs)
18 are typically considered as detrimental compounds in ash. Encinar *et al* [34] demonstrated that
19 certain transition metals actually decreased the bio-oil yield more than common AAEMs. Non-
20 metals and halogens (with the exception of silica, which is generally considered inert [35]) can also
21 have a catalytic effect during pyrolysis; therefore, the inorganics targeted during demineralisation
22 were considered as the total inorganic fraction, assuming that silica decreases proportionally to the
23 other elements.
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27 Previous studies indicate that water washing cannot fully remove inorganics from biomass,
28 suggesting that they are present in at least two forms in biomass; either soluble salts or as cations
29 bound to reactive sites in biomass. The latter is most likely as functionalities of cellulose or of acid
30 groups associated with lignin, hemicellulose and extractives. Soluble salts can be removed through
31 simple water leaching, whereas cations require the presence of an acid for ion exchange to occur
32 [29]. The removal of soluble salts via water leaching has been reported to have negligible effect on
33 the bio-oil yield and composition. Cations are bound more tightly to biomass polymers and are
34 thought to interact more readily during pyrolysis by facilitating dehydration and cracking reactions.
35 The removal of acid-soluble cations typically decreases the amount of pyrolytic water, increases the
36 bio-oil yield and significantly alters its composition, primarily by increasing the levoglucosan yield
37 [29, 36].
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40 41 42 *1.2 Torrefaction*

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44 Torrefaction is a mild version of slow pyrolysis, characterised by slow heating rates, long residence
45 times and temperatures between 200 and 320 °C. The process is relatively new, with development
46 aimed at increasing the energy density of biomass for pelletising [37-39]. The predominant product
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is a darkened biomass, with minor by-products of a liquid and non-condensable gas [40].

Torrefaction can be classified as mild, moderate or severe depending on the temperature and residence time. During mild torrefaction ($< 220\text{ }^{\circ}\text{C}$), non-reactive drying and particle shrinkage occur initially, followed by reactive drying, bond breaking, and volatilisation of lipophilic extractives and other light compounds [39]. Moderate torrefaction ($220\text{ to }250\text{ }^{\circ}\text{C}$) is distinguished by the release of water, CO, CO₂, CH₄, light volatiles and organic acid [41]. Under severe torrefaction conditions ($> 250\text{ }^{\circ}\text{C}$) the majority of hemicellulose is decomposed, with cellulose and lignin decomposition initiating when residence times are prolonged [37]. The fibrous biomass structure is destroyed during torrefaction. This reduces the energy for grinding by up to 90% [41]. The solid product from torrefaction exhibits increased particle sphericity when ground, which improves the flow characteristics and increases the bulk density [38, 42-44].

Several authors have considered torrefaction as a pre-treatment for pyrolysis biomass [37, 41, 45-48]. It was reported that the quality of the bio-oil improved due to a reduced content of water, oxygen, organic acids and light components [45]. Meng *et al.* [41] observed that the bio-oil under consideration was rich in pyrolytic lignin and anhydrosugars and low in light oxygenates and aldehydes. Chang *et al.* [47] reported that the levoglucosan yield increased after torrefaction, and speculated that this was caused by the reduction of catalytic interactions between the major biomass components resulting from the slightly disturbed wood structure. Under severe torrefaction conditions, the total liquid yield (torrefaction plus pyrolysis) decreases due to the formation of stable carbon-carbon cross-links during torrefaction that can undergo further polycondensation reactions to form char during pyrolysis [48]. Cross-links are formed when hydroxyl groups are removed during dehydration reactions. Adjacent cross-links form stable compounds that appear as lignin in the fibre analysis; therefore, the degree of cross-linking can be determined by the increase in acid insoluble fibres during biomass hydrolysis [47, 49].

2 Experimental

2.1 Materials

Pinus radiata wood chips ($< 6\text{ mm}$) were obtained from a local sawmill near Christchurch, New Zealand. After the chips were received, they were dried in a controlled room with a relative humidity of 50% and temperature of $40\text{ }^{\circ}\text{C}$, after which the final biomass moisture content was 8.4% (dry basis). The dried biomass was then knife-milled to $< 2\text{ mm}$ and sieved to remove the fines with diameters of less than $295\text{ }\mu\text{m}$. Removal of the fines reduced errors during the pre-treatments, as these could become embedded in, or permeate through the filters. For investigations

of the effect of the particle size during leaching, particles up to 6 mm were used, as received.

Finally, the dried biomass was stored in air-tight containers until use.

2.2 Acid leaching

Prior to acid leaching, the biomass was further dried overnight in an oven at 105 °C to remove residual moisture and then cooled in a desiccator. Leaching experiments were carried out in capped 2 L conical flasks. A total of 700 mL of leaching solution was added to the flask, and 70 g of oven-dry biomass was added to this solution. The flasks containing biomass and leaching solution were heated using magnetic hot plates with a stirring speed of 250 rpm. The leaching reagents used are listed in Table 2, and the experiments were carried out at internal liquid temperatures of 30 °C and 90 °C, respectively, for 4 h unless otherwise stated. After the leaching was completed, samples were neutralised with deionised water under vacuum using a Buchner funnel with a poly-cotton filter. Neutralised biomass was dried in an oven overnight at 105 °C and then analysed for polymer composition and inorganic elements. Three duplicates were averaged for each reagent.

2.3 Torrefaction

A 300 mL Gallenkamp bomb calorimeter was modified for torrefaction experiments. Two ¼ inch Swagelok fittings were welded to the top of the bomb, and these fittings were used to attach carrier gas and vapour removal tubes. Pressure was monitored with a manual pressure gauge (in case of line blockage). The internal temperature was monitored using a type K thermocouple. A single pass counter-current glass condenser (215 mm long and 6 mm in internal diameter) was used to condense vapours exiting the reactor. Condensed liquor entered a 100 mL flask with a cotton wool filter at the top which captured persistent vapours and aerosols. The operational temperature limit for the bomb was 316 °C. The reactor was heated from below using a magnetic hot plate with a 40 mm thick concentric aluminium block on top to increase the heat transfer rate up the sides of the reactor. A flea containing four 15 mm in diameter and 5 mm thick samarium cobalt magnets was designed with the addition of a paddle to provide agitation of the biomass during torrefaction. Employment of the stirrer increased the amount of biomass treated per run to 37.5 g, and improved the consistency of the torrefied biomass. The moisture content of all samples prior to torrefaction was 25% to simulate torrefaction representing the drying stage (25% was used, as this value is near the fibre saturation point for *Pinus radiata*). Three duplicates were averaged for each run.

2.4 Combined acid leaching and torrefaction

In order to optimise the combined pre-treatment of biomass, all of the 1% leached samples listed in Table 2 were torrefied at 240 °C for 20 min. Samples were dried after leaching to obtain accurate yields but the moisture content was then increased to 25% prior to torrefaction. Next, 1% acetic

1 acid leached samples (at 30 °C) were torrefied at temperatures between 240 and 280 °C at 10 °C
2 increments. Figure 2 provides an overview of the pre-treating process and apparatus.

3 Separate experiments were conducted by recycling the torrefaction liquid as the demineralisation
4 solution. The results obtained from these experiments were compared to those obtained with 1%
5 acetic acid leached biomass. The liquid was collected from 5 torrefaction experiments at 240 °C,
6 each with 37.5 g of biomass (moisture content of 25%); these aliquots were combined to yield a
7 total of approximately 35 mL of leaching reagent with an acetic acid concentration of 1.65%. The
8 liquid was then diluted to a pH of 2.75 which is equivalent to a 1% acetic acid solution, yielding a
9 total volume of 50 mL, to which 5 g of dry biomass was added, and the solution was leached at 30
10 °C for 4 h.

11 2.5 Analysis

12 *ICP-OES:* 300 ± 10 mg of biomass was digested in 2.5 mL of nitric acid and 2.5 mL of 30%
13 hydrogen peroxide in a CEM MARS Xpress microwave digester. The temperature was ramped to
14 90 °C over 15 min, held for 5 min, then ramped again to 180 °C over 10 min and held for 15 min.
15 The digested samples were analysed using a Varian 720 ICP-OES at Lincoln University in
16 Christchurch.

17 *Elemental analysis:* The carbon, hydrogen, oxygen and nitrogen content of the solid samples were
18 determined through complete oxidation [50] with the elementary analyser at CRL Energy Ltd. in
19 Wellington, New Zealand. The carbon, hydrogen, and nitrogen content were determined using
20 standard ISO 29541:2010, and the oxygen content was calculated by the difference.

21 *Biomass ashing:* Biomass was heated in a muffle furnace at 625 ± 10 °C for at least 16 h. Due to
22 the inherent variability in biomass and possible bark and stone contamination, the analysis result
23 for each sample was an average value from 9 repetitions. The residence time was relatively long
24 (approximately 6 h is common [16, 51]) because a constant weight could not be assessed without at
25 least 2 h of cooling before removing the sample from the furnace.

26 *Biomass structural analysis via hydrolysis:* Biomass samples were extracted prior to hydrolysis
27 using an automatic Dionex ASE 350 extractor with ethanol as the solvent. Conditions for extraction
28 followed standard NREL/TP-510-42619. In brief, samples were heated in 33 mL extraction cells to
29 100 °C at 1,500 psi for 7 min, followed by flushing with 150 vol% of ethanol. Three cycles were
30 repeated for each sample. After this, samples were dried overnight to obtain the extractive yield.

31 The procedure for hydrolysis followed standard NREL/TP-510-42618, with slight alterations. In
32 brief, 0.30 ± 0.01 g of dry biomass was hydrolysed for 1 h in 3 mL of 72% sulphuric acid and
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1 stirred every 10 min. The samples were diluted to 4% sulphuric acid by adding 84 mL of deionised
2 water and then heated to 113 °C (1.5 bar) in Duran[®] GL45 Pressure Plus laboratory glass bottles for
3 90 min. After hydrolysis was completed, the samples were vacuum filtered using Buchner funnels,
4 and the liquor was collected for HPLC and UV-Visible spectrophotometer analysis. The remaining
5 hydrolysis solids (Klason lignin) were washed with deionised water until a neutral pH was reached
6 before being dried overnight at 105 °C. Finally, the samples were cooled in a desiccator and
7 weighed to obtain the Klason lignin yield. A portion of the inorganics in biomass dissolve during
8 hydrolysis [52]; therefore, the lignin was ashed following hydrolysis, and the lignin yield was
9 adjusted accordingly.
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16 *UV-Visible spectrophotometer:* The hydrolysis liquor was analysed for acid-soluble lignin by
17 absorbance at 240 nm with an absorptivity of 12 Lg⁻¹cm⁻¹. A wavelength of 280 nm is commonly
18 used [53-55], but degraded carbohydrate products such as furfural and hydroxymethylfurfural
19 strongly absorb at a wavelength of 280 nm, whereas a wavelength of 240 nm results in minimal
20 absorbance. Only total lignin yields are reported, representing the sum of Klason and acid-soluble
21 lignin.
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28 *HPLC:* The hydrolysis liquor was neutralised with calcium carbonate and filtered through 0.22 µm
29 nylon syringe filters prior to the HPLC analysis. A Supelcogel C-610H carbohydrate column was
30 used for the sugar and organic acid analysis with a refractive index detector and a mobile phase of
31 0.1% phosphoric acid. The column temperature was maintained at 30 °C. The sample injection size
32 was 17 µL with a residence time of 20 min. Standards were run for cellobiose, glucose, xylose,
33 galactose, arabinose, mannose, acetic acid, formic acid, levulinic acid and methanol. The column
34 was chosen for optimal carboxylic acid analysis opposed to optimal sugar analysis, as accurately
35 quantifying the acids content was deemed more important.
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42 43 **3 Results and Discussion**

44 45 *3.1 Biomass Leaching*

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48 *Leaching reagents:* The total yields and structural analysis of biomass samples following leaching
49 with different reagents are presented in Table 3. During leaching, biomass sugar polymers
50 depolymerised if conditions were sufficiently severe for hydrolysis to initiate. The mass yields at
51 90 °C were related to the pH of the leaching solution (in decreasing order: hydrochloric, nitric,
52 sulphuric, formic and acetic acid). Mineral acids all reduced the biomass yield significantly during
53 leachings at 90 °C. Leaching with hydrochloric acid was the most severe, with hemicellulose being
54 partially hydrolysed and the acetyl concentration significantly reduced. Mass yields from various
55 leaching experiments with organic acids and water at 90 °C were approximately the same,
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indicating that water soluble extractives were removed and minimal carbohydrate degradation occurred.

The inorganic fraction in biomass was reduced from 0.45 to 0.27 wt% with water washing alone, indicating that approximately 40% of the inorganics in biomass are present as soluble salts.

Reduction of the inorganics ionically bound to biomass was accomplished using dilute acid solutions. The remaining inorganics were inaccessible within the wood matrix and could not be removed even with severe leaching procedures for *Pinus radiata*. Further reduction in inorganic content has been reported by other researchers with different feedstocks [28, 56]. The reduction of inorganics was slightly better for mineral acids compared to the organic acids at 30 °C. Increasing the leaching temperature to 90 °C increased the removal for the organic acids but had only a minor effect for the mineral acids, partially due to the increased mass loss. This result indicates that the reduction in inorganics can be achieved using the dilute organic acids obtained from torrefaction.

The ICP-OES results given in Table 4 indicate that only S, P, Na (and Zn for formic acid) were reduced to the same degree by organic acids as by mineral acids at 30 °C. The results at 90 °C imply that the alkali metals (K and Na) were reduced approximately equally with all leaching acids but that the alkaline earth metals (Mg and Ca) were not. The result could be beneficial, as alkaline earth metals have been used as catalysts for de-oxygenation during pyrolysis [57, 58], by favouring depolymerisation reactions over dehydration reactions [59]. Sulphuric acid caused a large increase in S, indicating that either some of the acid was not washed out during the neutralising step or S became incorporated into the biomass and thus could not be removed through water washing alone. This suggests a potential benefit for using organic acids compared to some mineral acids during leaching.

A mass balance for the elements in biomass leached with 1% acetic acid at 30 °C is given in Table 5. The discrepancy between the calculated mass balance for the elements removed from the biomass and the experimentally measured values in the leachate indicate that additional ions were present in the leachate which were not generated from the biomass. These could originate from the acetic acid or deionised water, but both sources would be minimal based on the assay of the raw solutions. The most likely introduction of additional ions would be from either the vessel in which the leaching solution was collected or stored.

Effect of organic acid concentration: The concentration of acetic and formic acid was varied to determine the minimal acid loading required to reduce the inorganic content. Figure 3 indicates that the effect of low acid concentrations was more pronounced for acetic acid compared to formic acid, but increasing the concentration above 1% had minimal effect for either acid when the biomass was

1 leached at 30 °C for 4 h. This result implies that a fraction of the inorganics present in biomass
2 cannot be removed at 30 °C with formic or acetic acid under the given leaching conditions. This
3 may be correlated with to the pH obtained with the leaching solution as shown in Figure 4. It
4 appears that the inorganic reduction is related to the pH of the leaching solution, regardless of the
5 actual acid type. Figure 4 can be used to predict the inorganic content after leaching for any given
6 acid leaching reagent under the same leaching conditions.
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10 *Effect of leaching residence time:* The residence time for 1% acetic acid leached samples was
11 assessed between 1 and 8 h. The total inorganic fraction of the biomass following leaching is given
12 in Figure 5. The results indicate that at least 4 h is required to reduce the ion concentration
13 significantly, but increasing in the residence time beyond 4 h provides minimal benefit.
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18 *Effect of biomass particle size:* Because torrefaction significantly reduces grinding costs, it would
19 be beneficial for leaching and torrefaction to be carried out on larger chips; afterward, a final size
20 reduction can be implemented if necessary. Wood chips with a particle size of 6 mm or less were
21 leached to compare the efficiency of ion removal to the standard wood chips with a particle size of
22 2 mm or less. The samples were leached using 1% acetic acid at standard conditions (30 °C for 4 h,
23 stirred at 250 rpm). It was interesting to find that leaching was slightly more efficient with the
24 larger chips and reduced the inorganic content to 0.15 ± 0.02 wt%, indicating that the system was
25 not limited by internal mass transfer. This finding could be because the 6 mm chips were milled
26 predominantly in the longitudinal direction, whereas the 2 mm chips were knife-milled equally in
27 the longitudinal, radial and tangential directions. Knife milling against the grain could close
28 elongated tracheid cell ends, thereby reducing the mass transfer rate of leaching solution into the
29 biomass and the rate of ion transfer out. Figure 6 provides a representation of the different chip
30 sizes used. The 6 mm chips could reach 20 mm the longitudinal direction due to the chipping
31 technique used, whereas the 2 mm chips were approximately equal in all directions.
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45 3.2 Torrefaction

46 *Effect of residence time:* The residence time for torrefaction at 245 °C was varied from 15 to 120
47 min, with the results displayed in Figure 7. The acetyl content decreased as the residence time was
48 extended up to 120 min. The decrease was not significantly large to warrant an extension beyond
49 20 min due to the additional heating costs and decrease in biomass carbohydrate content.
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Effect of temperature: Torrefaction targets the reduction of highly oxygenated side braches
associated with hemicellulose and lignin, particularly carboxyl compounds. It is reported that a

1 harsher severity of torrefaction by increasing the temperature enhances the cleavage of acetyl
2 branches associated with galactoglucomannan in the hemicellulose fraction of softwood biomasses
3 [60, 61]. Figure 8 indicates a reduction in acetyl branches from 1.51 wt% to 0.43 wt% when
4 comparing raw biomass to biomass torrefied at 290 °C for 20 min. Mild pyrolysis occurs when
5 torrefaction conditions are severe, thereby decreasing the overall potential bio-oil yield during
6 pyrolysis. Furthermore, severe torrefaction initiates the formation of cross-linked carbohydrate
7 polymers that are stable under pyrolysis conditions, leading to char formation instead of further
8 polymeric breakdown to bio-oil. This effect was quantified by the ‘apparent’ increase in lignin after
9 a full acid hydrolysis of the biomass as cross-linked sugar polymers are acid insoluble. At 290 °C,
10 the total ‘apparent lignin’ content reached 59 wt%; the pyrolytic char yield could not be reliably
11 predicted from this result, but it may be close to the yield from true lignin, which can exceed 46%
12 [62].
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21 The structural analysis of the torrefied biomass also indicated that the acid-soluble fraction of lignin
22 increased for torrefied biomass, possibly due to the production of compounds during hydrolysis that
23 have the same UV-Visible absorbance as lignin by slightly altered cellulose and hemicellulose
24 polymers. Alternatively, the acid soluble lignin content could indicate the partial degradation of
25 lignin during torrefaction, which would increase the acid soluble fraction.
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31 An elemental analysis of torrefied biomass is presented in Table 6. Elemental analysis was another
32 method used to quantify the severity of torrefaction. It also provides an indication of the oxygen
33 reduction relative to the hydrogen reduction. Because pyrolysis is a hydrogen deficient process
34 [63], the removal of oxygen through the production of water is undesirable during pre-treating;
35 furthermore, such removal leads to severely cross-linked biomass. Most research suggests that there
36 is no chemical elimination of water from biomass below 220 °C [59], thus hydroxyl groups are not
37 removed from sugar polymers to produce a cross-linked structure. However, carboxyl cleavage
38 from hemicellulose and oxygen reduction were minimal in this low temperature regime. Higher
39 temperatures were required for significant acetyl reduction and oxygen reduction, as well as to
40 breakdown the fibrous structure of the biomass. Thus, the optimum torrefaction temperature was
41 proposed to be approximately 250 °C to prevent significant biomass loss during torrefaction
42 (<10%) and to minimise the char yield during pyrolysis. The torrefaction temperature could be
43 increased to a maximum of 280 °C (before hydrogen loss becomes significant) if oxygen reduction
44 and acetyl removal were considered more important than maintaining high bio-oil yields in the
45 subsequent pyrolysis. Significant oxygen reduction, however, is difficult to achieve during
46 torrefaction without severe biomass losses. For example, after torrefaction at 290 °C with a biomass
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yield of 65.3 wt% the oxygen content was only decreased to 35.7%. Therefore, it is suggested that torrefaction alone is not a suitable method to obtain a low oxygen bio-oil at reasonable yields.

3.3 Combined acid leaching and torrefaction

Biomass leached at 30 °C and 90 °C (leaching reagents and biomass analysis given in Table 3) was torrefied at 240 °C for 20 min to determine the effect of leaching temperature on the subsequent effect torrefaction. The yields after torrefaction are provided in Table 7. The liquid yields varied on a dry basis due to the retention of a fraction of the condensate in the condenser. For the 1% nitric acid solution at 90 °C, it appeared that there was no liquid produced but the collected solution still had a low pH and was the normal light yellow colour. Yield deviations from the torrefaction of raw wood appear minimal for leaching procedures at 30 °C, although slight deviations were observed when the leaching temperature was increased to 90 °C. There was a general trend of decreased severity of torrefaction with slightly higher biomass yields and a higher pH of the torrefaction liquid.

The acetyl content in the biomass after the combined pre-treatments is given in Table 8. The relative reduction in acetyl compounds during torrefaction was lower for samples leached at 90 °C compared to samples leached at 30 °C or biomass not previously leached. Torrefaction may appear less severe due to the loss of easily cleavable compounds during leaching, especially for the mineral acids. There was no reduction in the acetyl content for DI-water leached biomass at 90 °C; therefore, the decreased acetyl removal cannot be solely attributed to previous cleavage. Inorganics could play a catalytic role in acetyl cleavage during torrefaction. Acetyl side-branches of galactoglucomannan are easily cleaved by alkali species during leaching [61]; a similar mechanism may apply during torrefaction. However, the inorganic reduction for DI-water leached biomass at 90 °C was considerably less than the reduction by all of the acid leaching reagents at 30 °C. Therefore, elevated temperatures during demineralisation must alter the morphology of the biomass and make it less susceptible to acetyl degradation during torrefaction. Performing torrefaction following biomass leaching at 90 °C would provide minimal benefits; instead biomass leaching at 30 °C is recommended when both acid leaching and torrefaction are employed as biomass pre-treatments.

Sample leached with 1% acetic acid were torrefied between 240 and 280 °C at 10 °C increments for 20 min. The results are compared to the torrefaction of raw biomass in Table 9. The severity of torrefaction appears to be less affected by the leaching pre-treatment as the torrefaction temperature increases. The overall biomass loss from acid leaching and torrefaction combined was still less than the loss for torrefaction alone. The acetyl reduction improved, possibly due to weakening of biomass polymers, which facilitated cleavage during torrefaction. Sugar degradation was reduced

1 when samples had been previously leached, and therefore, the oxygen content was higher due to the
2 high oxygen content in cellulose and hemicellulose; approximately 49 wt% and 54 wt%
3 respectively [8]. When acid leaching precedes torrefaction, the temperature of torrefaction can be
4 increased to 260 °C while still maintaining high overall biomass yields, but with a further reduction
5 in acetyl content to 0.83 wt% and lower lignin and oxygen contents compared to biomass solely
6 torrefied at 250 °C.
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10 *Recycling torrefaction liquor:* Liquor from 5 torrefaction runs (240 °C for 20 min) was combined
11 and diluted to obtain a solution with a pH of 2.75. This solution was used to leach biomass at 30 °C
12 for 4 h. The inorganic content in the leached biomass was reduced to 0.14 ± 0.02 wt%, which was
13 lower than for biomass leached with 1% acetic acid. This low inorganic content could be due to the
14 presence of other acidic compounds in small quantities, which increased cation removal. The
15 torrefaction liquor was qualitatively analysed using gas chromatography-mass spectrometry (GC-
16 MS), which indicated the presence of hexanoic, heptanoic, octanoic and nonanoic acid in small
17 quantities. Torrefaction does not produce adequate liquid to leach an equivalent amount of biomass;
18 therefore, the leachate would need to be recycled from acid leaching. The leachate could be
19 regeneration using pyrolysis chars, because their hydrophilic surface is suitable for removing ions
20 from water [64]. A portion of the leachate is lost during the regeneration; this would be replenished
21 with the new torrefaction liquor. The recycling and regeneration of leachate would minimise
22 environmental issues concerning the disposal of the mobile phase after leaching due to possible
23 trace amounts of hazardous elements.
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37 Demineralisation procedures require subsequent rinsing of the biomass to ensure complete removal
38 of the leaching reagent. Rinsing is necessary because mineral acids reduce yields during pyrolysis
39 and increase pyrolytic water through dehydration, condensation and cross-linking reactions [65].
40 Rinsing is also necessary because acids in the feed material directly influence the pH of the bio-oil.
41 The influence organic acids during pyrolysis is ambiguous, but preliminary pyrolysis experiments
42 indicate that the bio-oil yield is significantly reduced in their presence. This could lead to
43 economical drawbacks at larger scales due to the amount of water required for rinsing and disposal
44 of the contaminated solution during the rinsing process. No rinsing step is required in the proposed
45 pre-treatment sequence, as the organic acids will volatilise during torrefaction.
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54 *Effect of pre-treatments on pyrolysis:* Detailed pyrolysis results will be reported in a subsequent
55 paper but in brief; the dynamics of fast pyrolysis were altered through the targeted reduction of
56 three natural biomass catalysts (moisture, inorganics and organic acids). The bio-oil yield increased
57 by 20% on a water free basis. Pyrolytic water was significantly lower through decreased
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1 dehydration reactions, and with no moisture in the feed, the total water content of the bio-oil was
2 under 5%. The acetic acid content in the bio-oil was reduced by 96% through the removal of acetyl
3 branches prior to pyrolysis, additionally carboxylic acids produced during pyrolysis decreased due
4 to minimal ring opening and cracking reactions of primary products catalysed by inorganics [66].
5 The average molecular weight and pyrolytic lignin content of the oil decreased due to reduced re-
6 polymerisation reactions during pyrolysis. The inorganic content of the bio-oil decreased and the
7 stability improved. However, the oxygen content of the bio-oil reflected that of the feedstock,
8 therefore was only marginally decreased for pyrolysis of pre-treated biomass. Further research will
9 investigate other techniques for reducing the oxygen content of the bio-oil without incurring
10 significant mass losses. Together, these improvements to the bio-oil properties may increase the
11 efficiency when refining bio-oil using current technologies such as common fluid cracking catalysts
12 or hydroprocessing.
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22 **4 Conclusions**

23 This paper investigated the individual effects of demineralisation and torrefaction on *Pinus radiata*
24 biomass, and the integrated effects of combining the two pre-treatments. This study proposed to
25 recycle the liquid fraction produced during torrefaction as the leaching reagent; therefore, weak
26 organic acids were compared to the commonly used mineral acids in terms of their capacity to
27 reduce the inorganic fraction during demineralisation. The reduction capacity was slightly lower at
28 30 °C but equivalent when samples were leached at 90 °C. When the torrefaction liquid was diluted
29 to a pH of 2.75 and recycled back for leaching, the reduction in the inorganic content was similar to
30 that of biomass leached with formic acid.
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40 Torrefaction targeted a reduction in the acetyl and oxygen contents and complete moisture removal
41 from the biomass, although it was realised that only slight oxygen reductions could be achieved
42 without large biomass losses from torrefaction alone. Prolonged residence times during torrefaction
43 had a minimal effect on the severity of torrefaction compared to the effect produced by increasing
44 the temperature. Severe torrefaction (temperatures above 250 °C) was associated with a large mass
45 loss and increased the apparent lignin content of the biomass; therefore when torrefaction was the
46 sole pre-treatment, the optimal temperature was 250 °C. The temperature of torrefaction could be
47 increased to a maximum of 280 °C if the quality of the pyrolysis oil required further acetyl, oxygen
48 and water reduction, but such improvements would occur at the expense of bio-oil yields. Above
49 280 °C, cross-linking of the biomass was excessive, and hydrogen was lost due to the formation of
50 water; therefore, such temperatures are not recommended.
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1 When 1% acetic acid leached biomass was torrefied between 240 and 280 °C, the severity of
2 torrefaction decreased in terms of biomass, oxygen and sugar loss; therefore, the optimal
3 torrefaction temperature was increased to 260 °C, which lead to a further reduction in the acetyl
4 content of the biomass.
5

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Table 1: Properties of bio-oil compared to a heavy fuel oil and the effects of these properties on the oil's quality

Property	Pyrolysis oil [6]	Heavy fuel oil [6]	Effect on the quality in terms of fuel use
HHV (MJkg ⁻¹)	14-20	40	Larger volumes required [7-9]
Water content (wt%)	15-30	0.1	Lowers the heating value, viscosity, density, and ignition rate [10, 11]
pH	2-3	-	Corrosive to pipes and vessels [10, 11]
Solids (wt%)	0.2-1	1	Enhances bio-oil aging, corrosion and equipment blockages [10]
Ash (wt%)	0-0.2	0.1	High temperature corrosion, hard deposits, and bio-oil aging [11, 12]
Viscosity (cP at 50 °C)	40-100	180	High pressure drop, increased equipment costs, and ruptures [10]
Density (kgL ⁻¹)	1.2	0.85	Higher density can cause pumping issues [7]
Oxygen content (wt%)	30-40	-	Bio-oils immiscible with petroleum based fuels [11]
Safety (flammable class)	3	3	Potentially harmful [13]

Table 2: Leaching reagents for demineralisation at 30 and 90 °C

Leaching reagents
DI-water
0.5, 1, 2.5, 5, 10% acetic acid ^a
0.5, 1, 2.5, 5, 10% formic acid ^a
1 % sulphuric acid
1% hydrochloric acid
1% nitric acid

^a0.5, 2.5, 5 and 10% only tested at 30 °C

Table 3: Composition of biomass after leaching with different leaching acids

	Leaching yield (%)		Inorganic content (%)		Acetyl content (%)		Lignin (%)		Cellulose (%)		Hemicellulose (%)	
Raw wood	-	-	0.41 ± 0.04		1.51 ± 0.03		28 ± 1		43 ± 1		26 ± 1	
<i>Leaching temperature (°C)</i>	<i>30</i>	<i>90</i>	<i>30</i>	<i>90</i>	<i>30</i>	<i>90</i>	<i>30</i>	<i>90</i>	<i>30</i>	<i>90</i>	<i>30</i>	<i>90</i>
DI-water	99.3	97.7	0.27	0.29	1.53	1.50	30	28	42	42	26	27
1% acetic acid	99.3	98.0	0.16	0.12	1.48	1.46	28	28	43	42	27	27
1% formic acid	99.0	97.9	0.14	0.11	1.53	1.41	28	31	42	42	26	26
1% sulphuric acid	99.3	92.0	0.11	0.10	1.36	0.80	29	32	42	43	27	22
1% hydrochloric acid	98.4	85.0	0.11	0.10	1.38	0.40	29	32	42	49	27	17
1% nitric acid	99.0	88.0	0.12	0.13	1.48	0.54	28	32	44	46	27	21

Table 4: ICP-OES results for biomass leached at 30 and 90 °C

Element^b (ppm)	Al	B	Ca	Fe	K	Mg	Mn	Na	P	S	Zn
Raw wood	70.9	3.1	756.0	81.5	524.9	204.0	51.4	60.3	146.6	63.0	6.8
Leachings at 30 °C											
DI-water	48.3	2.2	647.6	44.6	142.1	173.1	45.6	50.7	107.2	40.5	6.1
1% acetic acid	58.2	2.1	198.6	76.9	18.1	38.6	9.1	42.7	111.8	41.2	2.7
1% formic acid	54.6	1.9	133.8	63.8	19.2	18.5	3.4	44.8	114.5	43.7	1.4
1% sulphuric acid	42.1	1.2	52.8	44.3	9.5	12.3	1.4	42.4	112.0	82.8	2.4
1% nitric acid	48.9	0.8	47.5	51.3	10.5	11.9	1.3	42.4	111.3	45.6	1.2
1% hydrochloric acid	62.8	0.9	84.3	68.4	11.6	18.4	1.8	43.4	115.2	46.3	0.8
Leachings at 90 °C											
DI-water	46.7	1.3	685.2	48.5	99.8	172.6	45.2	51.7	100.9	41.9	9.8
1% acetic acid	39.8	0.5	190.9	32.2	15.5	30.1	8.4	43.8	100.7	41.1	4.6
1% formic acid	47.6	0.3	90.4	23.9	9.4	14.4	2.0	43.5	102.6	44.2	1.2
1% sulphuric acid	40.6	0.3	32.6	12.9	7.7	7.3	0.7	42.9	91.2	71.1	1.3
1% nitric acid	39.6	0.3	33.6	11.7	7.9	7.4	1.0	42.7	88.1	47.5	1.2
1% hydrochloric acid	48.0	0.3	32.2	12.8	9.6	7.7	0.9	43.2	86.9	46.4	1.1

^bElements below 2 ppm are not displayed, these include Ba, Cd, Cr, Cu, Li, Ni and V. Elements tested and not detected were As, Co and Pd.

Table 5: Mass balance of elements for 1% acetic acid leached biomass at 30 °C

Element (mg)	Al	B	Ca	Fe	K	Mg	Mn	Na	P	S	Zn
In 70 g of raw biomass	5.0	0.2	52.9	5.7	36.7	14.3	3.6	4.2	10.3	4.4	0.5
In 70 g of 1% acetic acid leached biomass	4.1	0.1	13.9	5.4	1.3	2.7	0.6	3.0	7.8	2.9	0.2
Calculated mass balance for ion removal	0.9	0.1	39.0	0.3	35.5	11.6	3.0	1.2	2.4	1.5	0.3
Measured amount in the leachate	1.7	2.0	44.9	0.6	46.6	12.6	3.0	9.6	3.3	1.9	0.5

Table 6: Elemental analysis of biomass after torrefaction at varying temperatures

Temperature (°C)	Carbon (%)	Hydrogen (%)	Nitrogen (%)	Oxygen (%)
Raw	50.4 ± 0.7	6.0 ± 0.2	0.10 ± 0.00	43.1 ± 0.8
220	51.1	5.9	0.09	42.9
230	51.1	6.0	0.11	42.8
240	51.5	5.9	0.11	42.5
250	52.4	5.9	0.11	41.6
260	53.4	5.9	0.11	40.6
270	54.8	5.9	0.11	39.3
280	55.9	5.8	0.11	38.2
290	58.7	5.6	0.09	35.7

Table 7: Yields from the torrefaction of demineralised biomass

Leaching reagent	Raw	DI-water		1% acetic acid		1% formic acid		1% nitric acid		1% sulphuric acid		1% hydrochloric acid	
<i>Leaching temperature (°C)</i>	-	30	90	30	90	30	90	30	90	30	90	30	90
Moisture content (wt%)	24.9	24.8	24.9	24.8	24.9	25.2	24.9	24.6	24.8	24.9	24.7	24.9	24.7
Yields (g)													
Liquid	7.3	7.2	7.2	7.4	7.5	7.4	7.8	7.7	6.5	7.9	7.5	7.5	7.3
Biomass	27.5	27.4	27.4	27.2	27.5	27.6	27.4	27.2	27.7	27.3	27.5	27.5	28.1
Gas (by difference)	1.5	1.3	1.2	1.0	0.8	1.1	0.6	0.9	1.3	0.9	0.7	0.9	0.8
Yields, dry Basis (wt%)													
Liquid	0.5	0.4	0.7	0.9	1.2	0.4	2.2	2.2	0.0	2.3	1.9	1.3	0.5
Biomass	94.5	95.3	95.7	95.3	96.1	95.7	95.6	94.6	97.3	94.5	96.0	95.6	96.9
Gas (by difference)	5.1	4.3	3.7	3.7	2.7	3.9	2.1	3.2	2.7	3.2	2.1	3.1	2.6
Analysis													
pH	2.5	2.5	2.6	2.5	2.7	2.6	2.6	2.6	2.6	2.5	2.7	2.5	2.9

Table 8: Total loss in yield and structural analysis of biomass after demineralisation and torrefaction

	Overall biomass yield (%)		Acetyl after leaching (%)		Acetyl after torrefaction (%)		Acetyl change ^c (%)		Lignin (%)		Cellulose (%)		Hemicellulose (%)	
	30	90	30	90	30	90	30	90	30	90	30	90	30	90
Torrefaction only	94.5 ± 1.5		1.51 ± 0.03		1.32 ± 0.05		14.4		34.7 ± 0.5		41.5 ± 0.9		21.2 ± 0.6	
<i>Leaching temperature (°C)</i>	30	90	30	90	30	90	30	90	30	90	30	90	30	90
DI-water	94.5	93.5	1.53	1.50	1.34	1.39	14.2	7.9	35.0	32.6	39.0	42.2	23.5	23.5
1 % acetic acid	94.6	94.3	1.48	1.46	1.29	1.40	14.7	4.3	34.5	37.0	40.9	37.1	23.5	23.5
1 % formic acid	94.8	92.9	1.53	1.41	1.31	1.24	16.8	13.7	36.3	38.0	39.7	38.1	22.9	22.9
1 % nitric acid	93.7	85.6	1.48	0.54	1.29	0.54	14.7	0.0	35.9	38.6	41.6	39.4	20.5	20.5
1 % sulphuric acid	93.8	88.3	1.36	0.80	1.23	0.73	10.6	9.6	36.7	36.8	42.1	41.0	19.7	19.7
1 % hydrochloric acid	94.1	82.4	1.38	0.40	1.18	0.42	16.9	-4.8	37.4	41.7	39.6	41.1	14.9	14.9

^cChange in acetyl content following torrefaction of leached biomass - relative to the acetyl content after leaching

Table 9: Comparison between torrefaction of raw biomass and 1% acetic acid leached biomass

Torrefied biomass (no leaching prior)							
Temperature (°C)	Biomass loss (wt%)	Acetyl (wt%)	Lignin (wt%)	Total sugars (wt%)	Carbon (wt%)	Hydrogen (wt%)	Oxygen (wt%)
240	5.5 ± 1.5	1.32 ± 0.05	34.7 ± 0.5	62.7 ± 1.1	51.5	5.9	42.5
250	8.6 ± 1.0	1.23 ± 0.03	38.7 ± 2.7	59.0 ± 1.5	52.4	5.9	41.6
260	11.4 ± 2.1	1.16 ± 0.18	40.6 ± 3.2	57.9 ± 1.1	53.4	5.9	40.6
270	15.6 ± 2.0	0.95 ± 0.11	44.0 ± 2.8	53.8 ± 2.2	54.8	5.9	39.3
280	19.8 ± 1.5	0.76 ± 0.08	49.2 ± 4.2	50.1 ± 2.9	55.9	5.8	38.2
1% acetic acid leached and torrefied biomass							
Temperature (°C)	Biomass loss^d (wt%)	Acetyl (wt%)	Lignin (wt%)	Total sugars (wt%)	Carbon (wt%)	Hydrogen (wt%)	Oxygen (wt%)
240	5.4 ± 1.1	1.29 ± 0.13	34.5 ± 3.9	64.0 ± 1.7	51.4	5.9	42.6
250	7.6 ± 0.7	1.12 ± 0.16	34.9 ± 1.8	64.2 ± 1.8	52.2	5.9	41.8
260	10.3 ± 2.1	0.83 ± 0.15	35.5 ± 1.1	62.5 ± 2.5	53.4	5.8	40.7
270	14.0 ± 0.6	0.74 ± 0.03	39.5 ± 5.8	59.1 ± 1.9	53.6	5.8	40.6
280	17.8 ± 1.5	0.71 ± 0.05	45.8 ± 4.4	51.8 ± 1.8	55.2	5.8	39.0

^dOverall mass loss from leaching and torrefaction

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Figure 7: Biomass fibre analysis when varying the residence time of torrefaction at 245 °C

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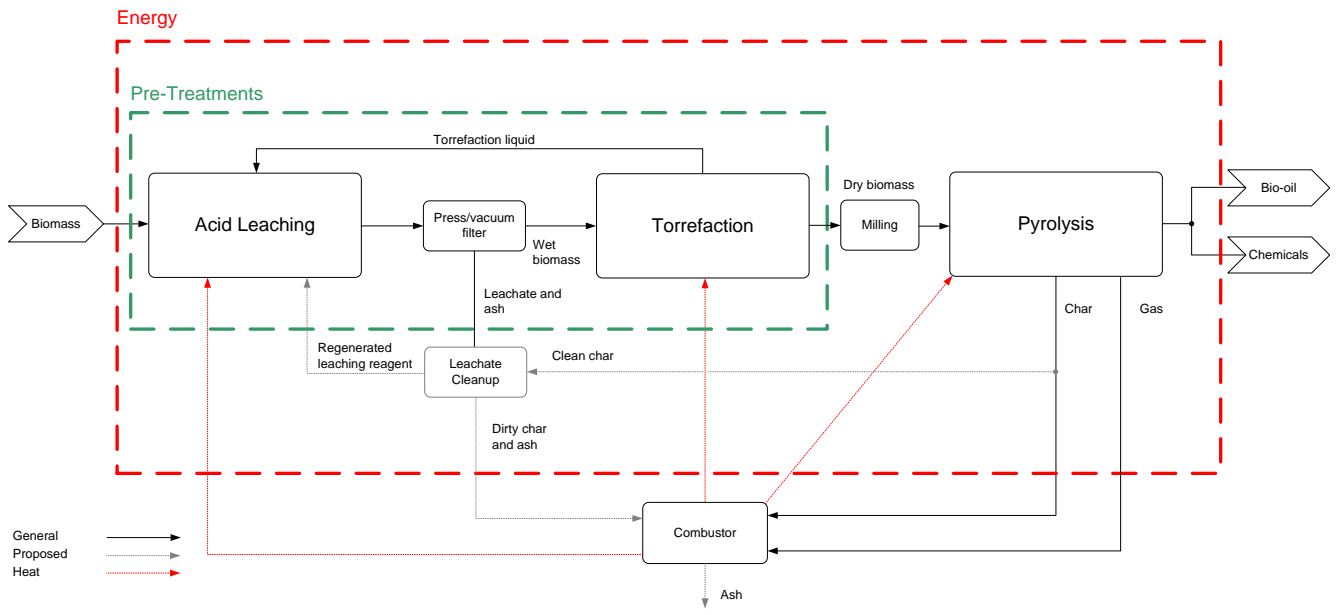


Figure 1: Pre-treatment sequence for the fast pyrolysis of biomass to bio-oil

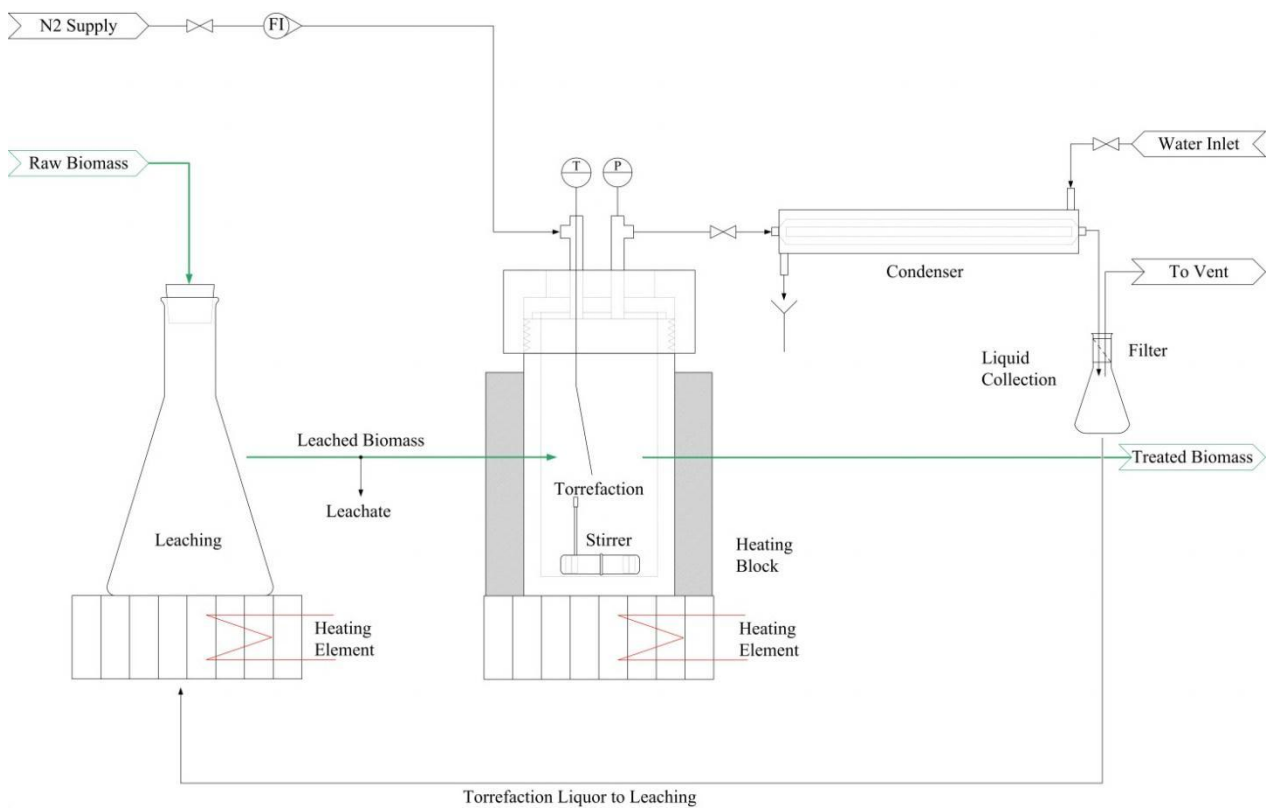


Figure 2: Apparatus for pre-treating biomass prior to pyrolysis

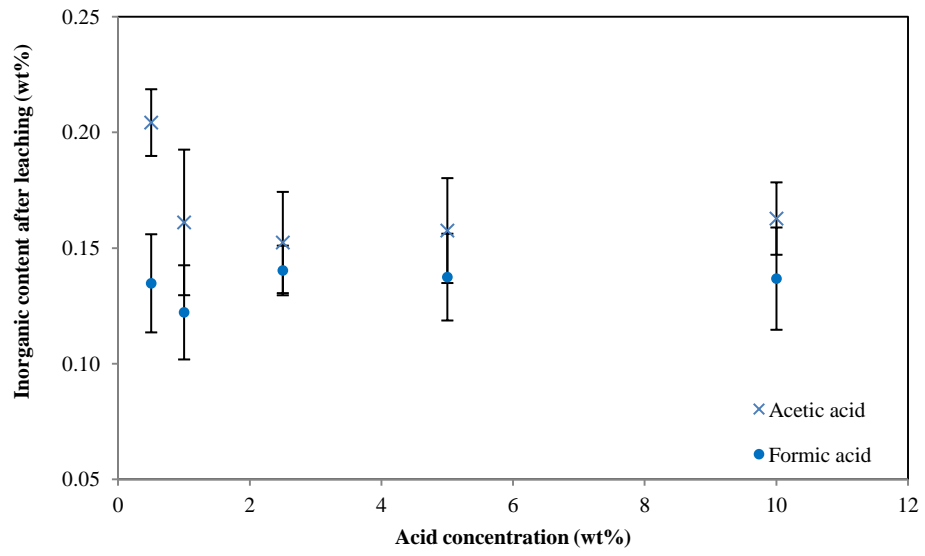


Figure 3: Inorganic reduction after leaching with varying acetic and formic acid concentrations

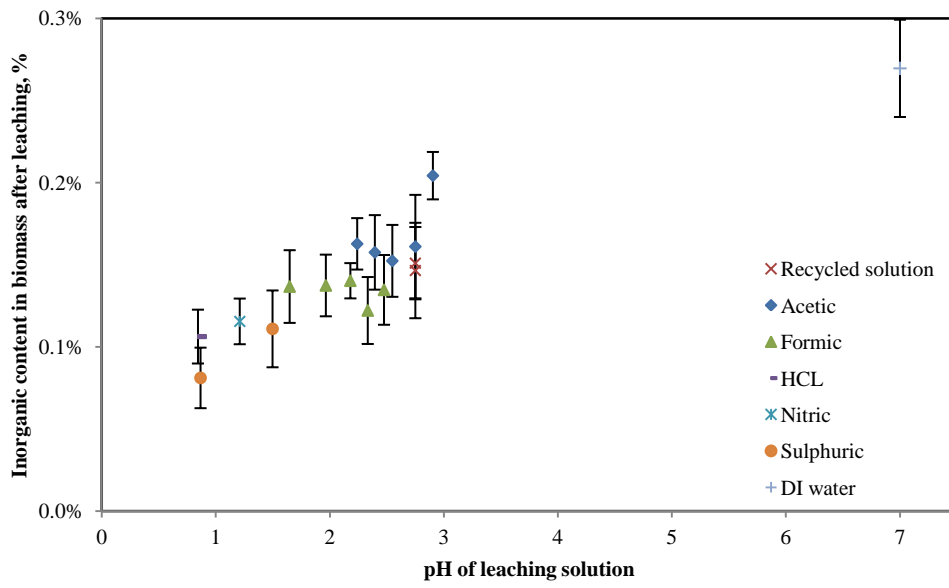


Figure 4: Inorganic reduction after leaching with varying acid types and concentrations

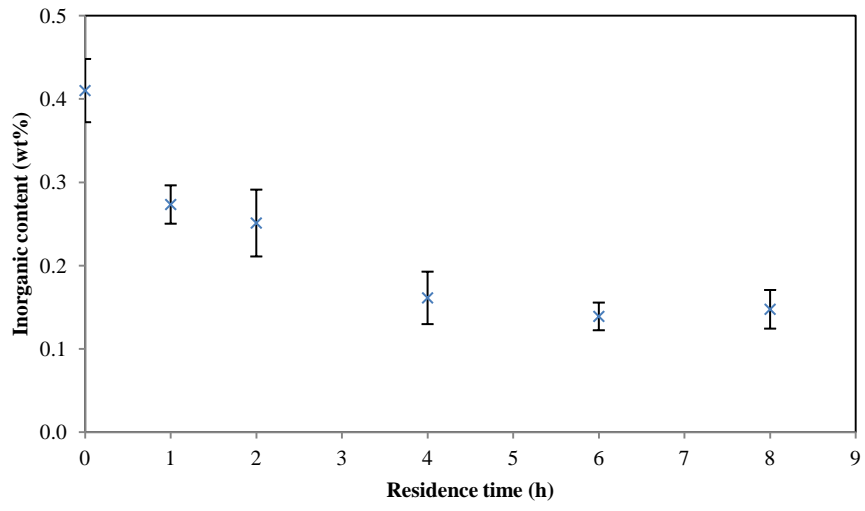


Figure 5: Inorganic fraction in biomass after leaching with 1% acetic acid for various residence times



Figure 6: Wood chips 6 mm and smaller (left) and biomass knife-milled to under 2 mm (right)

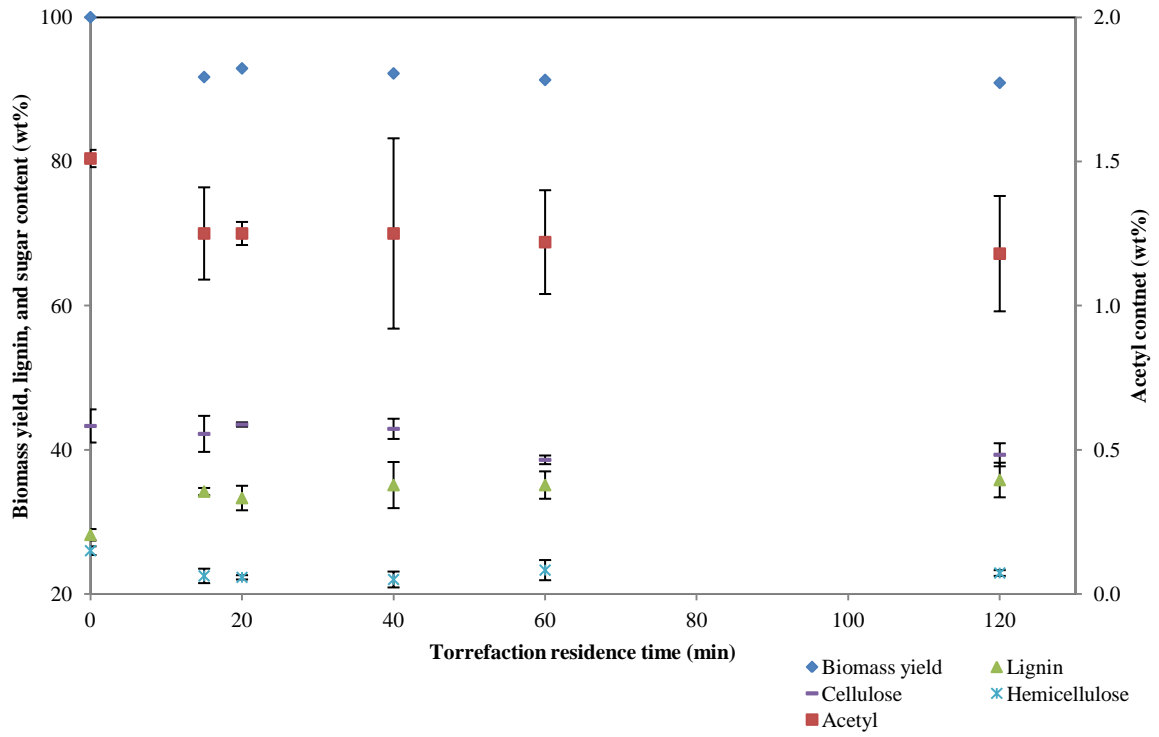


Figure 7: Biomass fibre analysis when varying the residence time of torrefaction at 245 °C

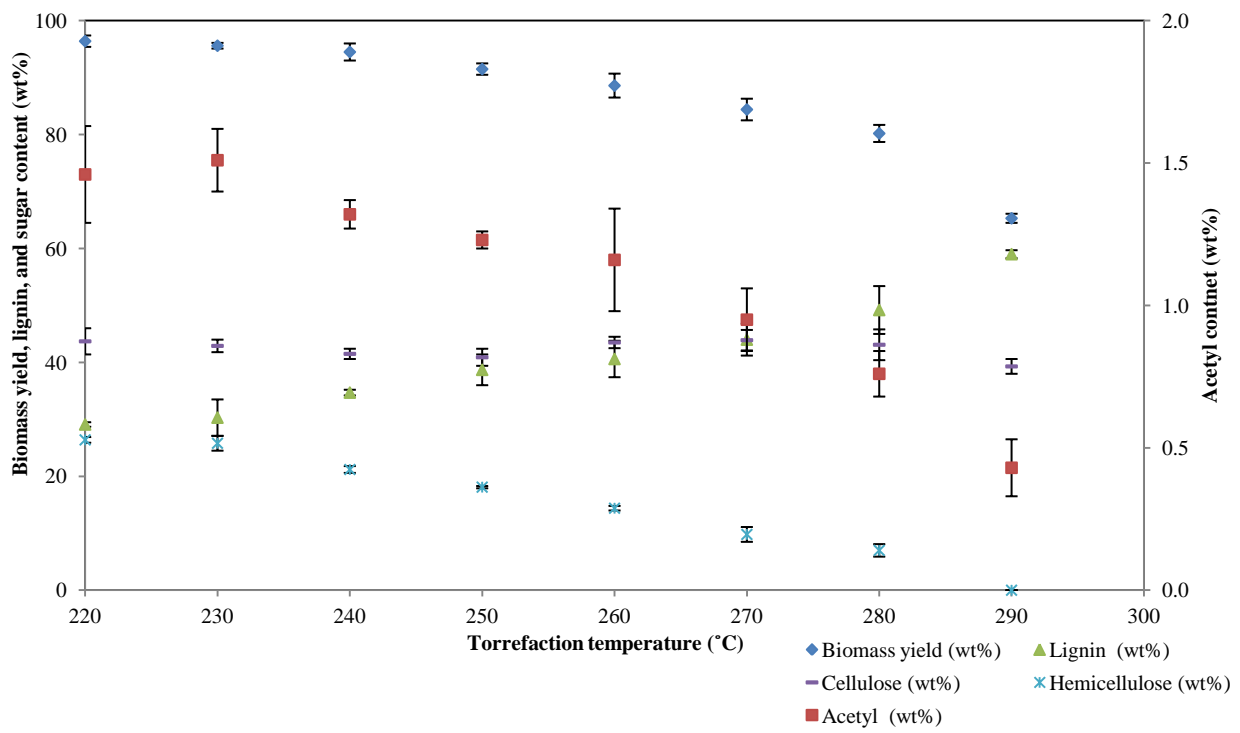


Figure 8: Effect of temperature during torrefaction for 20 min

Highlights

- Pre-treatments were used to improve the properties of biomass
- These consisted of acid leaching and/or torrefaction
- Torrefaction liquor was recycled as the acid leaching reagent
- Pre-treatments decreased the ash, oxygen, moisture and acetyl content of raw biomass