Case Report

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Chlorpromazine Combined with Cidofovir for Treatment of a Patient Suffering from Progressive Multifocal Leukoencephalopathy

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Key Words
JC viral load · Progressive multifocal leukoencephalopathy · Cidofovir · Chlorpromazine

Abstract
We report on a stem cell-transplanted patient with B cell chronic lymphatic leukemia who presented with a subacute onset of focal neurological deficits, gait abnormalities, emotional lability and dementia. Progressive multifocal leukoencephalopathy was diagnosed by magnetic resonance imaging (MRI) of the brain and detection of JC virus genome in the cerebrospinal fluid. Cidofovir and the 5HT2A receptor antagonist chlorpromazine were subsequently administered. A follow-up MRI of the brain 2 weeks after initiation of the antiviral therapy displayed progress of the demyelination, and the patient died 3 months after onset of the neurological symptoms. This report highlights the need for the development of novel and potent strategies for treatment of progressive multifocal leukoencephalopathy.

C.P. and K.H. contributed equally to this work.

Case Report

A 58-year-old human immunodeficiency virus (HIV)-negative man was diagnosed with B cell chronic lymphatic leukemia (CLL) of Binet stage C in 1995. In 2002, disease progression necessitated the administration of 6 cycles of fludarabine (FAMP; table 1). The patient’s health status deteriorated in October 2003, and chemotherapy was altered to the CHOP 21 regimen (doxorubicin, cyclophosphamide, vincristine and prednisone; table 1). Treatment was discontinued after 3 cycles due to unresponsiveness. After 2 relapses of B cell CLL, complete remission was achieved by treatment with alemtuzumab (table 1). The patient was subsequently referred for HLA-DR mismatched allogeneic stem cell transplantation (SCT) in late January 2004. Reduced-intensity conditioning consisted of FAMP, busulfan and alemtuzumab (table 1). Graft-versus-host disease (GvHD) prophylaxis was initially carried out with cyclosporin A, but was switched to tacrolimus due to side effects (table 1). Twelve days after SCT the patient developed acute skin GvHD, which responded to prednisolone therapy. On day 40 after SCT, cytomegalovirus reactivation occurred (pp65 antigenemia: 2 positive cells/100,000 leukocytes) and was successfully treated by valganciclovir medication for 5 weeks.
Four months after transplant, a relapse of skin GvHD required repeated steroid treatment with prednisolone. Two weeks later the patient complained of drowsiness, vertigo, slight tremor and erratic mood swings which worsened during the next days. Clinical examination revealed bilateral homonymous hemianopsia of his right field of vision. A brain magnetic resonance imaging (MRI) showed an area of demyelination in the left occipital lobe of the brain (fig. 1b, left panel). No typical signs of infarction or tumor were detected.

Biochemical analysis of the cerebrospinal fluid (CSF) revealed signs of nonspecific inflammation, with mild lymphocytic pleocytosis (18 megaparticles/l, normal <5 megaparticles/l) and a slight increase in glucose (8.07 mmol/l, normal 2.8–4.4 mmol/l), total protein (639 mg/l, normal 130–400 mg/l) and lactate (3.18 mmol/l, normal 1.2–2.1 mmol/l). In addition, neither bacterial nor fungal DNA was detectable in CSF by PCR, nor were neurotropic viruses (herpes simplex 1/2, varicella-zoster virus, cytomegalovirus, Epstein-Barr virus, human herpes virus

<table>
<thead>
<tr>
<th>Drug</th>
<th>Posology1</th>
<th>Year/time period relative to SCT2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fludarabine</td>
<td>25 mg/m²; 6 cycles</td>
<td>2002</td>
</tr>
<tr>
<td>CHOP (doxorubicin, cyclophosphamide, vincristine and prednisone)</td>
<td>doxorubicin 25 mg/m², cyclophosphamide 300 mg/m², vincristine 1 mg/m² and prednisone 40 mg/m²; 3 cycles in total</td>
<td>2003</td>
</tr>
<tr>
<td>Alemtuzumab (Campath-1H)</td>
<td>30 mg; 20 applications in total</td>
<td>2004</td>
</tr>
<tr>
<td>Fludarabine</td>
<td>30 mg/m²</td>
<td>2004/d–9 to d–5</td>
</tr>
<tr>
<td>Busulfan</td>
<td>4 × 1 mg/kg</td>
<td>2004/d–4 to d–3</td>
</tr>
<tr>
<td>Alemtuzumab (Campath-1H)</td>
<td>5 mg</td>
<td>2004/d–9</td>
</tr>
<tr>
<td></td>
<td>10 mg</td>
<td>2004/d–8</td>
</tr>
<tr>
<td></td>
<td>20 mg</td>
<td>2004/d–7 to d–5</td>
</tr>
<tr>
<td>Prednisolone</td>
<td>50 mg</td>
<td>2004/d–9 to d–5</td>
</tr>
<tr>
<td></td>
<td>80 mg tapered to 30 mg over 2 months</td>
<td>2004/d–9 to d–5</td>
</tr>
<tr>
<td></td>
<td>80 mg</td>
<td>2004/d–8</td>
</tr>
<tr>
<td></td>
<td>80 mg tapered to 20 mg</td>
<td>2004/d–7 to d–5</td>
</tr>
<tr>
<td>Cyclosporin A</td>
<td>2 × 150 mg</td>
<td>2004/d–1 to d+1</td>
</tr>
<tr>
<td></td>
<td>2 × 175 mg</td>
<td>2004/d–2 to d+3</td>
</tr>
<tr>
<td></td>
<td>2 × 200 mg</td>
<td>2004/d–4 to d+5</td>
</tr>
<tr>
<td></td>
<td>1 × 200/225 mg</td>
<td>2004/d–6 to d+11</td>
</tr>
<tr>
<td></td>
<td>2 × 200 mg</td>
<td>2004/d–12</td>
</tr>
<tr>
<td></td>
<td>2 × 150 mg</td>
<td>2004/d–13 to d+19</td>
</tr>
<tr>
<td>Tacrolimus</td>
<td>2 × 1 mg</td>
<td>2004/d+19</td>
</tr>
<tr>
<td>Cidofovir</td>
<td>5 mg/kg, once per week</td>
<td>2004/d+173 and d+180</td>
</tr>
<tr>
<td>Probenicid</td>
<td>2 g, 3 h before CDV application; 1 g, 2 and 8 h thereafter</td>
<td>2004/d+173 and d+180</td>
</tr>
<tr>
<td>Chlorpromazine</td>
<td>3 × 200 mg</td>
<td>2004/d+173</td>
</tr>
<tr>
<td></td>
<td>3 × 400 mg</td>
<td>2004/d+174 to d+186</td>
</tr>
<tr>
<td></td>
<td>3 × 200 mg</td>
<td>2004/d+187 to d+198</td>
</tr>
<tr>
<td></td>
<td>3 × 400 mg</td>
<td>2004/d+199 to d+201</td>
</tr>
<tr>
<td></td>
<td>3 × 200 mg</td>
<td>2004/d+202</td>
</tr>
</tbody>
</table>

1 The posology corresponds to the dose per day, except for cidofovir which is administered weekly.
2 The days before (d–) and after (d+) stem cell transplantation (SCT) are shown.
6/8, adeno- and enterovirus) except for JC virus (JCV). JC viral load was determined in both CSF and peripheral blood by real-time PCR using 2 different protocols [1, 2]. Mean values for CSF and peripheral blood were $7.3 \times 10^5$ and $7.2 \times 10^3$ copies/ml, respectively, indicating systemic infection (fig. 1a). Antiviral treatment including cidofovir (CDV) with concomitant probenecid, chlorpromazine and immunoglobulins was initiated (table 1). A follow-up brain MRI 2 weeks after therapy initiation evidenced multiple areas of demyelination (fig. 1b, right panel). The JC viral load in CSF and peripheral blood showed no significant decrease despite antiviral treatment (fig. 1a). The disease progressed to blindness, deafness and disorientation as well as fecal and urinary incontinence. The patient died from progressive multifocal leukoencephalopathy (PML) 8 months following transplant and 3 months after onset of the first neurological symptoms.

**Discussion**

We report on an HIV-negative patient diagnosed with PML following allogeneic SCT and immunosuppressive treatment (table 1), in whom JC viral load was monitored in plasma and CSF under anti-JCV-specific therapy. In
the course of 2-fold CDV application, a 40–60% decrease in JC viral load in plasma samples was observed shortly after drug administration (fig. 1a, left panel), whereas JC viral load in CSF samples in fact doubled (mean values: $7.3 \times 10^5$ vs. $1.5 \times 10^6$ genome equivalents/ml; fig. 1a, right panel). According to our observations, combined anti-JCV-specific therapy with CDV and chlorpromazine was ineffective in significantly lowering JCV plasma and CSF loads and thus providing a clinical benefit for our patient. In this context, possible mechanisms for the non-response of the anti-JCV-specific therapy are discussed.

PML is a subacute demyelinating disease of the central nervous system (CNS) with a grim prognosis [3] caused by the neurotropic virus JC, a double-stranded DNA virus of the Polyomaviridae family. In immunocompromised patients, reactivation of JCV leads to various symptoms ranging from acute febrile illness to PML [4]. So far, various treatment regimens have been discussed. They mainly consist of 2 therapeutic approaches: inhibition of viral replication in the host cell, and a most novel approach based on inhibition of virus entry into host cells.

The first therapeutic approach is based upon inhibition of viral replication in host cells by using drugs with antiviral activity against Polyomaviridae, such as CDV and cytosine arabinoside [5, 6]. CDV is a deoxycytidine monophosphate analogue, with broad in vitro and in vivo activity against DNA viruses of various families (Adenoviridae, Herpesviridae and Polyomaviridae) [5, 6]. In vitro studies showed that CDV inhibits replication of mouse polyomavirus and Simian virus 40 [5], and displays efficacy against JCV multiplication in persistently infected human fetal brain cell lines [7]. A scarce number of anecdotal reports describe possible benefits of CDV treatment in PML, specifically in HIV-negative CLL patients suffering from PML [8, 9].

However, it remains unclear whether the improvements of symptoms in those patients is the net result of the antiviral effects of CDV, or of the immune reconstitution that follows successful cytostatic CLL therapy. Of note, intravenous administration of CDV does not yield detectable CSF concentrations (<0.1 mg/ml, assay detection threshold, data from Gilead Sciences Inc.), indicating that CDV does not diffuse through the blood-brain barrier. Qalbumin (concentration of albumin in CSF/concentration of albumin in plasma) was normal in our patient, excluding a substantial CNS barrier dysfunction. However, small leaks in the blood-brain barrier allowing CDV to enter the CNS cannot be completely ruled out in PML despite normal Qalbumin. Whether sufficient CSF drug levels for inhibition of viral replication are reached is an open question. Due to limited specimens, intrathecal levels of CDV and its metabolites could not be measured in our case. In view of the rising JC viral load in CSF and disease progression despite CDV therapy, intrathecal drug concentrations may not have reached sufficiently high levels to display antiviral effects, thus contributing to treatment failure.

The second and novel therapeutic approach relies on the use of neurotropic drugs to inhibit JCV entry into the host cell and subsequent viral spread. The rationale for this approach is based on the observation that the serotonergic receptor 5HT2A acts as the cellular receptor for JCV on human glial cells [10]. 5HT2A receptor antagonists like chlorpromazine and clozapine effectively inhibit JCV infection of glial cells and the spread of JCV through tissues, by blocking the clathrin-dependent endocytosis that enables cellular JCV uptake [11]. However, the clinical efficacy of this drug class in treating PML has not been evaluated yet. In our case, progress of the demyelinating lesions was evidenced by brain MRI 2 weeks after initiation of the treatment, indicating that JCV spread within the white matter of the brain could not be impeded by chlorpromazine (fig. 1b, right panel). The continuous development of new brain foci as shown in our patient by MRI (fig. 1b, right panel) is concordant with a lytic infection including viral spread from cell to cell and high quantities of tissue-associated virus. Chlorpromazine interference with 5HT2A receptor-mediated clathrin-dependent endocytosis targets an early step in the viral life cycle. Drug application early in the course of the disease may thus be crucial to achieve a clinical benefit, as glial cells already harboring/having internalized the virus are left unaffected by chlorpromazine [11]. Furthermore, overall viral mass in tissue may constitute a critical factor as both virus particles and chlorpromazine compete for the same cell receptor. Newer atypical antipsychotics with less adverse effects, like risperidone or olanzapine, have proven to be significantly more potent 5HT2A receptor antagonists in vitro than chlorpromazine or clozapine [12], which may render them more ideal candidates for the treatment and/or prophylaxis of PML in immunodeficient patients.

Cell-mediated immunity is essential in the control of many viral infections in humans and has also been suggested to play a crucial role in the containment of JCV [13–15]. Therefore, it is likely that immunosuppressant drugs not only trigger JCV reactivation in immunocompromised patients but sustainably influence the course and prognosis of PML. Infectious complications associated with calcineurin inhibitors, for example cyclosporin...
A and tacrolimus, include other viral infections [16] of which BK virus reactivation represents a significant cause of allograft failure in renal transplant recipients [17]. Furthermore, cyclosporin A has been associated with the occurrence of PML in patients suffering from rheumatic diseases [18, 19]. The development of PML has also been observed in patients receiving corticosteroids for rheumatic disease [20–24]. A correlation between JC viruria and corticosteroid treatment has been demonstrated in one study [25]. Purine nucleoside analogues, primarily FAMP, inducing a rapid T lymphocyte decline render recipients particularly prone to various opportunistic infections [26] and have also been linked to the development of PML [9, 27–29]. Alemtuzumab, used as consolidation therapy in CLL patients, in preparation for SCT, or to avert acute and chronic GvHD, provokes increased vulnerability to infections due to profound and sustained lymphocyte depletion [30] as demonstrated by several clinical trials [31–33]. Polyomavirus — that is BK virus — infections have been described in renal transplant recipients [34], whereas JCV reactivation after administration of alemtuzumab has not been reported so far in the literature. Considering the immune-modulating therapy regimens given to our patient (table 1), it is conceivable that the underlying immunosuppression may account for the nonresponse of the anti-JCV-specific therapy by countervailing the antiviral effects of CDV and chlorpromazine.

In conclusion, the use of 5HT2A receptor antagonists like chlorpromazine in addition to CDV may represent, if administered early in the course of the disease or given prophylactically, a possible strategy for treatment/prevention of PML. Further studies providing both experimental and clinical data are needed to evaluate the benefit of this strategy for treatment and/or prophylaxis of PML.

Acknowledgements

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