

## APPENDIX

### THE DIATOM FLORA OF THE STEENBRUGGE-CLAY (EEMIAN)

We received a continuous sampling of the Steenbrugge-Clay of Eemian age which was studied by J. Vandenberghe et al. (1973). A complete pollen diagram being available these authors concluded that this schorre-clay could well be correlated with the Eemian zone 4b of Zagwijn (1961). As only few diatom flora's are described from the Eemian it may be worthwhile to publish here the detailed list of the determinations and the quantitative data.

#### METHOD

For the quantitative purpose, the method given by Crommelin (1945) seemed the most suitable. From each sample a quantity of 5 g was boiled during twenty minutes in concentrated sulphuric acid to destroy the organic material. At the end potassium nitrate was added to become the whitish-gray colour. The contents of the porcelain discs in which the samples were treated, were added slowly to glasses filled with 1 l distilled water. After 24 hours of sedimentation the supernatant was carefully siphoned over, and the samples were rinsed again with 1 l distilled water, after which they were stored during 24 hours for sedimentation. This treatment was repeated 5 times to remove the gypsum. At the end the residue was replenished to 0,5 l.

Before the microscopical observation the samples (0.5 l) were shaken thoroughly to become a suspension as homogeneous as possible. From each sample three drops with a volume of 0.041 ml were taken immediately. With each drop a preparation was made in which the diatoms were counted. The results of these counts are given in table I. The number of diatoms in 1 mg of fresh material is given by the formula :

$$N = 0.8 X$$

where  $N$  = number of diatoms in 1 mg

$X$  = the total of the numbers of diatoms counted in the three preparations.

The table I gives at the same time the list of observed diatoms.

TABLE I  
Number of Observed Diatoms

Species	type (1)	D <sub>1</sub>	D <sub>2</sub>	D <sub>3</sub>	D <sub>4</sub>	D <sub>5</sub>	D <sub>6</sub>	D <sub>7</sub>	D <sub>8</sub>
<b>CENTRALES</b>									
Actinocyclus Ehrenbergii Ralfs	M	—	1	—	2	1	—	1	—
Actinoptychus undulatus (Bailey) Ralfs	M B	5	6	2	5	11	2	13	6
Auliscus sculptus (W. Smith) Ralfs	M	—	—	1	—	1	—	—	—
Biddulphia pulchella Gray	(M)	—	—	1	—	—	—	—	—
B. rhombus (Ehrenberg) W. Smith	M	—	4	—	2	1	—	17	2
Coscinodiscus excentricus Ehrenberg	M B	2	1	—	3	1	—	6	—
C. Kützingii A. Smith	(M)	—	—	—	1	—	—	—	—
C. radiatus Ehrenberg	(M)	—	—	—	—	—	—	1	—
Melosira sulcata (Ehrenberg) Kützing	M	44	44	29	40	33	22	68	25
Podosira Stelliger (Bailey) Mann	M	1	5	2	1	1	3	2	—
Triceratium alternans Bailey	M	—	—	—	—	—	1	—	—
T. favus Ehrenberg	M	2	12	4	4	2	1	6	—
<b>PENNALES</b>									
Achnantes brevipes Agardh	B M	—	1	—	1	2	—	1	3
Caloneis formosa (Gregory) Cleve	B	—	—	1	—	25	24	11	—
Campylodiscus clypeus Ehrenberg	B	—	—	—	—	—	—	1	—
C. echeneis Ehrenberg	B	—	—	—	—	1	—	—	—
Cocconeis scutellum Ehrenberg	(M)	2	—	—	—	1	—	3	—
Diploneis didyma Ehrenberg	M B	67	5	7	340	347	285	222	54
D. interrupta (Kützing) Cleve	B	—	—	—	—	—	—	1	1
D. ovalis (Hilse) Cleve	Z B	—	1	—	3	67	224	82	5
Navicula hungarica Grunow	Z B	—	—	—	—	—	—	—	3
N. latissima Gregory	M	—	—	—	—	—	—	—	4
Nitzschia acuminata (W. Smith) Grunow	M B	—	—	—	—	1	—	—	—
N. navicularis (De Brébisson) Grunow	B	88	11	24	211	181	133	288	33
N. punctata (W. Smith) Grunow	B M	6	—	4	23	41	3	38	15
N. sigmoidea (Ehrenberg) W. Smith	Z B	—	—	—	—	3	2	7	1
N. tryblionella Hantzsch	B Z	—	—	—	—	—	—	3	—
Rhaphoneis amphiceros Ehrenberg	M B	5	2	—	—	1	1	—	1
Rh. surirella (Ehrenberg) Grunow	M B	—	—	1	—	—	—	—	—
Rhopalodia musculus (Kützing) O. Müller	B	1	1	—	3	—	—	1	—
Scoliopleura tumida (De Brébisson) Rabenh.	B M	—	—	—	—	—	—	4	4
Stauroneis phoenicenteron Ehrenberg	Z B	1	—	—	1	10	45	30	2
Surirella fastuosa Ehrenberg	(M)	—	—	1	80	14	11	11	—
not classified		6	5	4	11	7	5	13	10
Total		230	99	81	731	752	762	830	169
number of frustules per mg		184	79	65	585	602	610	664	135

Explanation by table I :

- (1) The indication of the type of salinity-dependence is copied from A. Van der Werff and H. Huls (1957) with exception of the data between brackets. For the latter the information is based upon the other cited works.

The symbols mean :

M = marine  
MB = marine-brackish  
BM = brackish-marine

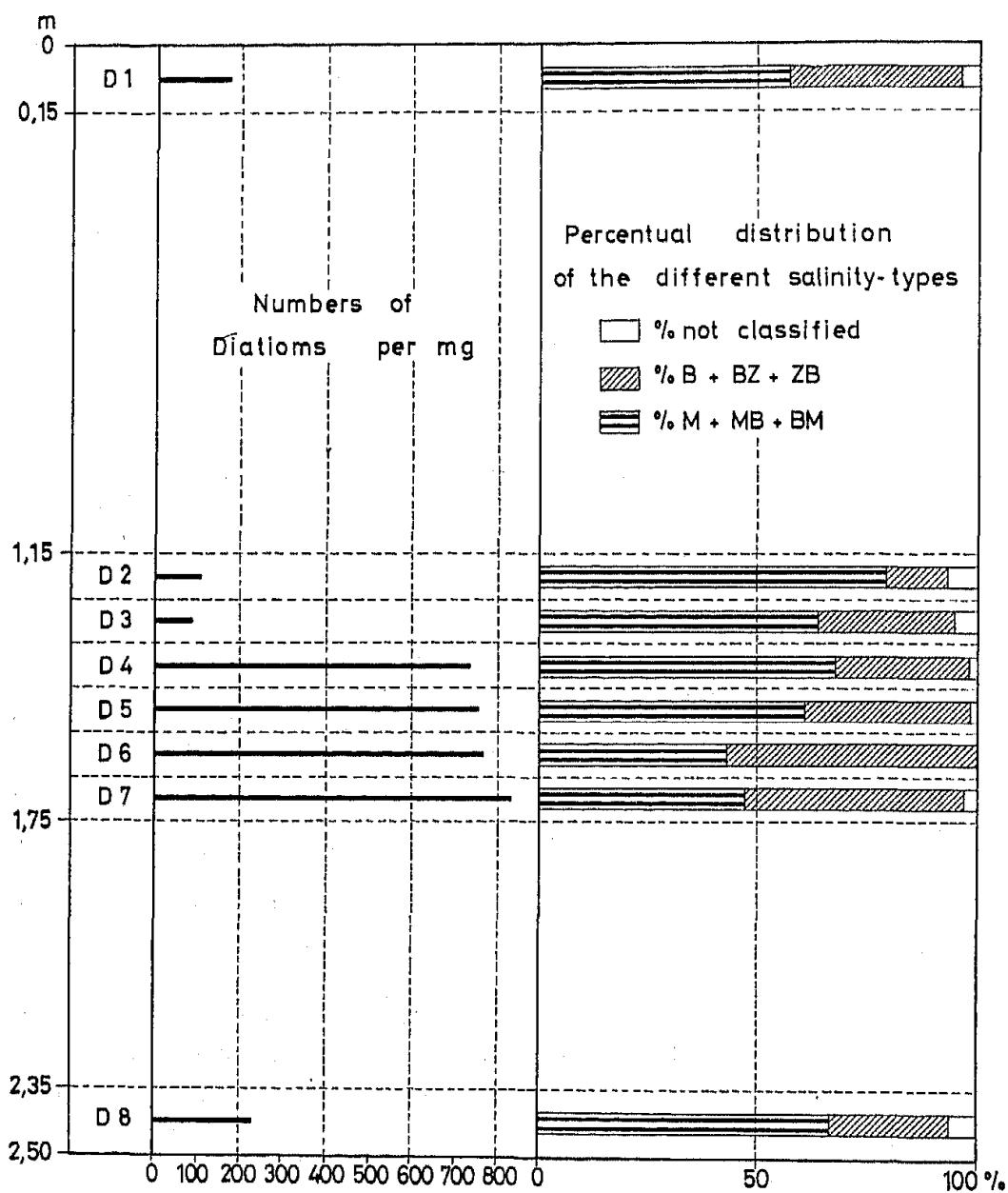
B = brackish  
BZ = brackish-fresh  
ZB = fresh-brackish.

## DISCUSSION

### 1. The total number of diatoms per mg

In comparison with some literature data the totals given in table I seem rather low. For this fact two explanations are acceptable :

- 1° The samples contain a relatively high quantity of organic material.
- 2° A great number of frustules are broken and it is impossible to make an estimation of the number of complete frustules represented by the fragments. Anywhere, most of the fragments could be identified as



belonging to the cited species. The number of fragments was moreover in proportion with the number of complete frustules so that at any rake a comparison can be made between the different results.

By comparison of the results (fig.), it is clear that the samples can be divided in two groups : the totals in D<sub>7</sub>, D<sub>6</sub>, D<sub>5</sub> and D<sub>4</sub> are strikingly higher than in D<sub>1</sub>, D<sub>2</sub>, D<sub>3</sub> and D<sub>8</sub>. The high values are mainly due to the marine-brackish species *Diploneis didyma* and the brackish species *Nitzschia navicularis*. The fresh-brackish species *Diploneis ovalis* reach a climax in D<sub>6</sub>.

If one takes into account that fresh water contains more phytoplankton than brackish water and brackish more than seawater (Louis and Clarysse, 1971, p. 312), the samples D<sub>7</sub>, D<sub>6</sub>, D<sub>5</sub> and D<sub>4</sub> must be considered as brackish sediments, and the samples D<sub>1</sub>, D<sub>2</sub>, D<sub>8</sub> and D<sub>3</sub> as marine (littoral) sediments. The most striking difference lies between samples D<sub>4</sub> and D<sub>3</sub>, where in a distance of only 10 cm the picture of the diatom population is completely altered.

## 2. Salinity dependence

For each sample the observed diatom frustules are added up per type. The values are converted to become the percent of each type of the total population. To make a separation between the fresh water and the sea water influence the contributions of the marine, marine-brackish and brackish-marine types are added up on the one side, the brackish, brackish-fresh and fresh-brackish on the other side.

TABLE II  
Percentual distribution of the different salinity-types

Type Sample	M %	MB %	BM %	Sub-total %	B %	BZ	ZB %	Sub-total %	Unclassified %
D <sub>1</sub>	21	34	3	58	39	—	—	39	4
D <sub>2</sub>	66	14	1	81	12	—	1	13	5
D <sub>3</sub>	47	12	5	64	31	—	—	31	5
D <sub>4</sub>	18	48	3	69	29	—	11	30	2
D <sub>5</sub>	7	48	6	61	27	—	11	38	1
D <sub>6</sub>	5	38	—	43	21	—	36	57	1
D <sub>7</sub>	13	29	5	47	36	—	14	50	2
D <sub>8</sub>	18	36	13	67	20	—	7	27	4

The results are given in table II (see also figure).

The fresh water influence reaches a climax in sample D<sub>6</sub> and the marine influence in D<sub>2</sub>. A more profound knowledge of the ecological characteristics of the different species should make further interpretations possible.

### 3. Comparison between the observed species and the actual diatom populations at the Belgian Coast

Because there is a very great periodical and geographic variability in the occurrence of diatom populations, even on a small scale; it is rather difficult to compare the observations, given in this paper, with the actual situation. A great part of the species correspond with those observed by Brockmann (1928) in Eemian sediments. However, most of these species are living nowadays in our littoral and brackish waters (Van Heurck, 1899; Allaerts, 1967; Van der Wielen, 1967; De Pauw, 1969; Louis en Clarysse, 1971; Petes, 1972; Ramboer, 1972).

In this respect, it is not obvious to speak of a „fossil vegetation”. Further conclusions may be based upon larger series of investigations.

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Dr. R. CLARYSSE  
Nationale Plantentuin van België

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