HARMONIZATION OF THE EVALUATION OF TUMOR INFILTRATING LYMPHOCYTES (TILs) IN BREAST CANCER: RECOMMENDATIONS BY AN INTERNATIONAL TILs-WORKING GROUP 2014.


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Abstract

Background: The morphological evaluation of tumor infiltrating lymphocytes (TILs) in breast cancer is gaining momentum as evidence strengthens for the clinical relevance of this immunological biomarker. Accumulating evidence suggests that the extent of lymphocytic infiltration in tumor tissue can be assessed as a major parameter by evaluation of (H&E) hematoxylin and eosin-stained tumor sections. TILs have been shown to provide prognostic and potentially predictive value, particularly in triple negative and HER2-overexpressing breast cancer.

Design: A standardized methodology for evaluating TILs is now needed as a prerequisite for integrating this parameter in standard histopathological practice, in a research setting as well as in clinical trials. This article reviews current data on the clinical validity and utility of TILs in breast cancer in an effort to foster better knowledge and insight in this rapidly evolving field, and to develop a standardized methodology for visual assessment on H&E-sections, acknowledging the future potential of molecular/multiplexed approaches.

Conclusion: The methodology provided is sufficiently detailed to offer a uniformly applied, pragmatic starting point and improve consistency and reproducibility in the measurement of TILs for future studies.

Key words

Tumor infiltrating lymphocytes (TILs), breast cancer (BC), prognosis, prediction, clinical validity.

Word count: 4401 words (Microsoft Word 2010).
Key message

Several translational studies based on phase III clinical trials have demonstrated that stromal tumor infiltrating lymphocytes (TILs) are a robust prognostic factor in TNBC. For HER2-positive breast cancer, TILs in baseline samples have been associated with higher responses to trastuzumab and chemotherapy treatment. There is therefore a need for harmonizing the histological approach to assess the percentage of stromal TILs in primary tumor specimens prior to therapy. The goal of this article is to facilitate the use of TILs as a biomarker in research and clinical trial settings, and ultimately in diagnostic practice. Herein, the current fundamental concepts for TILs-evaluation are laid out by pathologists to foster their widespread use and consistent interpretation at this early stage. The recommendations illustrated in this manuscript focus on: 1) what areas to examine in the tumor, 2) how to score the TILs, and 3) why TILs are clinically relevant. This information should provide a platform for pathologists to further engage in efforts leading to procedural standardization of this biomarker assay.
Context of research:

A panel of pathology, clinical oncology, biostatisticians and translational research experts conducted a systematic review of the literature. Panel members invited have had experience in TIL-assessment in phase III-trials or are involved in breast cancer translational research focused on the interactions between immunology and breast cancer. There are no existing guidelines on TIL-assessment in breast cancer available for comparison; neither is there proficiency testing data available from international organizations. No specific funding was obtained for this project. For details on prognosis, prospective-retrospective phase III trials were the main basis for these recommendations. However, we searched PubMed from Jan 1 2009 to April 30 2014 for full reports of studies involving clinical trial datasets or large institutional cohorts and evaluation of TILs, CD8+, CD3+ or immune gene signatures in primary breast cancer. Studies were not limited to randomized trials, but included also large consecutive and retrospective series. In-press publications were also taken into consideration. With regards to specific TILs pathological assessment, the panel undertook a formal expert consensus–based process by regular mail, teleconferences as well as two F2F-meetings by the writing committee (RS, SL, SdM, KWG, SA) to produce these recommendations. Draft manuscripts were circulated by email to all co-authors and the writing committee had responsibility for approving the final manuscript.
Introduction

A fundamental role of the immune system is maintenance of tissue homeostasis by continuous immunosurveillance and initiation of inflammatory reactions that involve the coordinated activation of innate and adaptive immune cells [1]. Neoplastic transformation alters the orderly structure of tissues and induces immune responses that can eliminate incipient tumors. In situations where elimination is incomplete neoplastic transformation of cells is able to escape immune control. This process has been best conceptualized by the cancer immunoediting theory, which is supported by a large body of experimental data and clinical evidence [2]. Immunoediting defines malignant progression on the basis of tumor and immune cell interactions in three phases: elimination, equilibrium and escape. While patients are most frequently diagnosed in the escape phase, this relationship between the tumor and host immunity continues to evolve and sometimes with it the magnitude of the anti-tumor immune response. Even at advanced disease stages, immune parameters have now been recognized as directly or indirectly influencing patient survival [3].

Recently, new therapies that reactivate anticancer immune-responses to cancer, for example in melanomas and lung cancer, have entered clinical practice and have improved outcome [47, 48]. Several recent clinical studies have evaluated the prognostic and predictive importance of tumor infiltrating lymphocytes (TILs) in breast cancer (BC). Some of these studies used similar methodological approaches for evaluating TILs, which allows for comparison of the results. The foreseen inclusion of TILs assessment in current and future clinical studies and diagnostic assessments necessitates a detailed description of a standardized methodology.

In December 2013, a group of investigators from around the world representing major breast cancer research and clinical teams convened to candidly discuss the important parameters to consider as well as methodological obstacles in evaluating TILs in breast cancer. The group
recognized the need to provide the breast cancer community with consensus recommendations for TILs-evaluation to foster their integration into future clinical trials, translational research and diagnostic practice. These efforts may evolve into the establishment of a breast cancer “immunological grade”, reflecting the strength of an individual patient’s anti-tumor immune response [4]. Here, we outline the current fundamental concepts for TILs-evaluation by pathologists to facilitate its widespread use at this stage in our understanding of its relevance for breast cancer. The recommendations focus on: 1) what areas to examine in the tumor, 2) how to score the TILs and 3) why TILs are clinically important.

**What is the composition and role of the immune infiltrate in human breast cancer?**

Immune cells infiltrating tumors are frequently observed, but the composition of cells involved in innate and adaptive immunity varies between tumor types or organ sites [5]. Cumulative data from murine and human studies have associated most leukocyte subsets with a predominant contribution to either pro- or anti-tumor activities (illustrated in Figure 1). Murine models have identified myeloid lineage leukocytes, including tumor-associated macrophages, dendritic cells and myeloid-derived suppressor cells as playing a central role in shaping the microenvironment via the factors they produce, towards either an immuno-stimulatory anti-tumor milieu or a wound healing tumor-promoting microenvironment. Anti-tumor T cells migrating into these contrasting settings can therefore either be activated or suppressed [6]. In turn, macrophage polarization towards pro-tumorigenic M2 or anti-tumor M1 functional phenotypes are regulated by T-lymphocytes [7], highlighting the importance of cellular cross-talk in shaping the tumor microenvironment.

Studies in humans have demonstrated a significant association between the presence of specific subsets of immune cells and clinical response in patients with a variety of solid tumors.
Furthermore, accumulating evidence suggests that adaptive immunity mediated by T and B lymphocytes provides the critical foundation for effective and sustained anti-tumor responses. In breast cancer, extensive tumor infiltration by cytotoxic CD8⁺ T cells was strongly associated with patient survival [8, 42] and response to therapy [9]. The presence of CD4⁺ regulatory T cells (Treg) has been associated with both good and bad [10, 11, 12]. Among the other CD4⁺ T cell subpopulations, Th1 cells (the principal cellular source of IFNγ) have been associated with favorable clinical outcomes [13], whereas Th2 cells have been reported to be associated with dampening of the anti-tumor response [14]. Th17 cells, producers of the pro-inflammatory IL-17 cytokine family, appear to have variable effects depending upon the surrounding cytokine milieu, which may in part be linked with the organ site and tumor type [15]. The presence of follicular helper (Tfh) cells, the newest CD4⁺ subset, was recently positively associated with patient outcome both in the adjuvant and neoadjuvant settings [13]. The precise role of tumor infiltrating B cells is currently not well defined and remains controversial [16, 17].

Given the functional heterogeneity of intratumoral lymphocytes it is intriguing that the degree of lymphocytic infiltration assessed by simple evaluation of H&E-stained tumor sections has been shown to have predictive and prognostic value in triple negative and HER2+ breast cancer despite a lack of detailed information on the immune subpopulations of the infiltrate [18-21]. A possible explanation is that negative immune regulators are present as part of a normal feedback loop reacting to an active and ongoing anti-tumor immune response, which therefore potentially defines tumors that are more immunogenic [22]. This consideration has several important implications. The first is that a focused evaluation of individual subsets may have limited value. For example, low or absent Treg infiltration may reflect tumors that are disregarded by the immune system while high Treg in tumors may signal an active, albeit unsuccessful, attempt at tumor rejection. Second, TIL-rich and TIL-poor breast cancers may
each reflect a distinct tumor cell biology that likely has markedly different susceptibility to immunotherapy. Finally, in moderate to extensively infiltrated tumors the presence of peritumoral or stromal TLS can be seen in some patients [13]. Thus, despite the inability of the immune system to reject a clinically detectable tumor, an organized immune response at the tumor site may signal the generation of immunological memory with the potential to effectively control residual disease. Variability has also been detected within individual tumors [23], suggesting that the nature of tumor-immune interactions may parallel tumor heterogeneity.

Cytotoxic treatments such as chemotherapy and radiotherapy may sometimes act to jump start the system [24-26]. In this context, a stronger anti-tumor immune response directed to a broader range of BC-antigens would potentially have a higher likelihood of controlling the heterogeneous malignant cell population present in large primary tumors and emerging metastases [23]. This hypothesis is supported by studies showing that the degree of lymphocyte infiltration is predictive of a better local response to neoadjuvant treatment and prognostic of long-term disease control [18, 19, 21].

**Current data on clinical validity and utility of TILs in breast cancer**

Examples of notable adjuvant and neoadjuvant studies that have assessed infiltrating lymphocytes are included in Table 1. In the majority of these studies, both intratumoral and stromal TILs have been assessed, with evaluation of the stromal compartment shown to be more reproducible between studies. Some studies focused on TILs using immunohistochemistry, while others evaluated molecular markers using immunohistochemistry and gene expression analysis.
Adjuvant studies

Triple Negative Breast Cancer (TNBC)

TILs have been assessed in full face sections of >1300 TNBC and > 3500 hormone-receptor positive breast cancers at diagnosis. TILs were found to be a positive prognostic biomarker in 297 TNBC but not in the luminal subtypes. This correlation was first reported using baseline samples from the BIG 2-98 trial [19] and subsequently independently confirmed in 481 TNBC sample prospectively collected during two phase III adjuvant randomized BC trials [United States Eastern Cooperative Oncology Group (ECOG) trials 2197 and 1199 [21]. Therefore, in TNBC, the more stromal TILs a patient has at diagnosis, the better their outcome after adjuvant anthracycline-based chemotherapy. Hence, according to Simon et al. [28], the results for the prognostic value of TILs in TNBC could be considered Level I evidence. However, given the lack of prognostic information for patients with primary TNBC not treated with chemotherapy, TILs should not be used as a biomarker for withholding chemotherapy.

HER2+ disease

Recent data from randomized clinical trials, evaluating TILs on full face sections suggest the importance of immunity in HER2+ disease [19, 20]. The FINHER study, where patients were randomized to receive received trastuzumab or no trastuzumab, reported that higher TILs in baseline samples resulted in higher responses to trastuzumab treatment. Recent data from the N9831 study [40] suggested that tumors that were "immune enriched", as defined using gene expression, had better outcomes if they received trastuzumab. While these findings will not affect the use of trastuzumab in newly-diagnosed HER2+ breast cancer, they suggest a potential mechanism of action for trastuzumab-based therapy. Based on current knowledge, however, TILs should not be used to either withhold or prescribe trastuzumab therapy. Given that trastuzumab with chemotherapy is the standard of care today, attention has turned to TILs as a
prognostic factor in HER2+ disease treated with anti-HER2 therapy [49]. Data suggest that high levels of TILs are also associated with excellent outcomes in HER2 disease treated with lapatinib as well as dual trastuzumab and lapatinib with chemotherapy (unpublished data).

Neoadjuvant studies

To date, core biopsies from more than 3000 patients have been assessed for correlation between immune markers and response to neoadjuvant chemotherapy, including institutional cohorts, but also biomaterials from clinical trials. An overview on the studies is given in Table 1. In summary, histological as well as molecular data indicate that immunological parameters, including stromal TILs are associated with higher rates of pCR, independent of other clinico-pathological prognostic factors or the chemotherapy regimen. Intriguingly, an interaction between stromal-TILs and a benefit to carboplatin added in the neoadjuvant setting has been reported [27], although the biological mechanism remains unclear.

Methodological recommendations for evaluating TILs in Breast Cancer

Prior to attending the December 2013 meeting, the participants with experience in evaluating TILs for phase III studies were asked to complete a questionnaire covering topics pertinent to their assessment in breast cancer. These questions are detailed in Supplementary information 1. The goal of this approach was to derive a consensus based on current experience within the group and use it as the foundation for this guideline. Based on these discussions, the working group participants made recommendations for harmonizing TILs-evaluation, which are summarized in Table 2. Additionally, a tutorial has been prepared and is included as a supplemental data file (Supplementary Tutorial (Supplementary information 2) “Standardized
evaluation of Tumor-Infiltrating Lymphocytes (TILs) in Breast Cancer for daily clinical and research practice or clinical trial setting”).

**Technical issues for evaluation of TILs in Breast Cancer**

1. Microscope magnification does not really make a difference, but usually a magnification of 200-400 x (ocular 10x, with an objective of 20x-40x) is recommended.

2. Slide thickness is not critical, with a standard thickness of 4-5 µm considered optimal. The majority of existing experience is based on scoring 4-5µm sections of FFPE-tissues, while the feasibility of TILs-evaluation on frozen sections is undocumented outside of a research setting and thus cannot be recommended for routine use at the present time.

3. TILs can be evaluated using core biopsies in the neoadjuvant setting as well as surgical specimens in the adjuvant setting. Considering all of the above, the scoring of one FFPE-block/patient is sufficient in the neoadjuvant and adjuvant setting. Some studies have assessed the prognostic or predictive importance of TILs on post-treatment tissues, but more studies are needed before formal recommendations can be made on the methodology of scoring TILs after neoadjuvant treatment [29].

4. Originally, Tissue Microarrays (TMAs) were not recommended for evaluating TILs, since there was no published evidence that TMAs mirror the potential heterogeneity of TILs, and the number of cores needed and the defined core-diameter, accurately reflecting TIL composition in a full section are unknown. However, recently published studies [30, 41, 42] using TMAs and well annotated clinical data sets show that results are concordant with other studies in the field using mostly biomarker-based determinations of TILs-subsets, not H&E determinations of general TILs. TMA’s may be a good option for future studies,
particularly for the rapid evaluation of large clinical cohorts. More investigation is needed before firm methodological recommendations can be offered.

5. All mononuclear cells including lymphocytes and plasma cells should be scored (granulocytes and other polymorphonuclear leukocytes are excluded). The quantitative assessment of other mononuclear cells such as dendritic cells and macrophages is currently not recommended, although there is increasing evidence that they may be functionally important since they are observed in TLS.

6. Several studies used immunohistochemistry to assess the clinical importance of subtyping lymphocytes. CD45, CD8, CD3 and various other markers expressed on lymphoid cells have been tested and while immunohistochemistry may improve accuracy, at the present time any added value from these markers is unclear. The TILs working group does not currently recommend that immunohistochemistry be used to detect specific subpopulations outside of the research setting, until further evidence is available.

7. Machine scoring approaches, while promising; have not been published in large series with consistent methodology. These approaches represent an important area for further study.

8. It is unknown if either RNA or protein classification of TILs by will reveal prognostic and predictive value beyond that achievable by simple morphology. New techniques, like CyTOF [44], can review protein based signatures of inflammatory infiltrates. While these are all still in the domain of research, pathologists should be aware of this potential. Clinical utility will drive the development of specific immune markers.
The concept of stromal vs. intratumoral TILs - focus on the stromal compartment

The initial studies of breast cancer TILs have evaluated stromal and intratumoral lymphocytes separately. Intratumoral TILs are defined as lymphocytes in tumor nests having cell-to-cell contact with no intervening stroma and directly interacting with carcinoma cells, while stromal TILs are located dispersed in the stroma between the carcinoma cells and do not directly contact carcinoma cells. Since both are localized in the region defined as tumor tissue, it should be emphasized that both categories represent true TILs. Furthermore, as TILs are able to move within a living tissue microenvironment, the distinction may be somewhat artificial and related to the static situation in histological slides that are used for diagnostic assessment. The original hypothesis was that lymphocytes directly interacting with carcinoma cells might be more relevant and therefore more useful for diagnostic assessment. While this hypothesis may still be biologically and/or clinically relevant, for diagnostic purposes on H&E-stained sections, most current studies have found stromal TILs to be a superior and more reproducible parameter. The main reasons are that intratumoral TILs are typically present in lower numbers and detected in fewer cases, they are more heterogeneous and are difficult to observe on H&E-stained slides (i.e. without using immunohistochemistry or immunofluorescence). Scoring intratumoral TILs does not add to the information provided by stromal TILs since they usually parallel stromal TILs. However, focusing on the stromal compartment (instead of the tumor as a whole) has a clear advantage because the density and growth pattern of carcinoma nests will not affect the TIL count because stromal TILs are measured only in the spaces between the carcinoma nests. Nevertheless, recent evidence indicates that in the neoadjuvant TNBC setting, and despite the methodological reasoning mentioned above, both stromal as well as intratumoral TILs are predictive of pathological response to neoadjuvant platinum-based chemotherapy [43]. Also, using CD3 or CD8-immunohistochemistry intratumoral TILs may potentially become as easy to detect as stromal TILs. Nevertheless, the TILs working group's current recommendation is to
evaluate stromal TILs as the principal parameter in future studies, allowing the straightforward evaluation of a single parameter. Additional parameters, including TLS in the peritumoral region, TILs at the invasive edge or intratumoral TILs can still be included for research purposes to further determine and/or confirm their potential clinical relevance.

In the tutorial, evaluations are based on stromal TILs, which are reported in an approximatively manner semi-quantitatively on a continuous scale as a percentage of stromal TILs. The working group’s consensus is that TILs may provide more biological relevant information when scored as a continuous variable, since this will allow more accurate statistical analyses, which can later be categorized around different thresholds. However, in daily practice most pathologists will rarely report for example 13.5% of TILs and will round up to the nearest 5-10%, in this example thus 15%. Pathologist should report their scores in as much detail as the pathologist feels comfortable with.

The original methodology for scoring TILs described by Denkert et al. in 2010 [18] was used in the majority of subsequently published studies, thereby providing sufficient data for this initial stage in developing a uniform methodology (based on the definitions in Figure 2). Interpersonal discussions among pathologists applying this approach since 2010 has slightly modified the original version [18] as shown in Figures 3 and 4). While some studies have scored TILs using other semi-quantitative approaches [13, 24, 30], the SABCS TILs working group considers the clinical validity of the modified Denkert et al. [18] approach described here to be superior at this time. This alternative assessment of TILs does of course not invalidate previously published findings using other methods of TILs assessment, but provides a framework for future standardization.

Using the recommendations in Denkert et al. [18], stromal TILs should be scored uniquely as a percentage of the stromal areas alone and areas occupied by carcinoma cells should not be
included in the total assessed surface area. This is an important point because otherwise the size of the epithelial cell nests as well as the tumor growth pattern could influence the stromal TILs value. For example, a score of 50% stromal TILs means that 50% of the stromal surface area and thus not stromal nuclei is occupied by TILs and also not 50% of the stroma plus epithelial cell area. For semiquantitative assessment of percentage values, the dissociated growth pattern of lymphocytes needs to be taken into account. Lymphocytes typically do not form solid cellular aggregates, therefore the designation “100% stromal TILs” would still allow some empty tissue space between the individual lymphocytes.

This recommendation is based on the methodology used in published phase III studies, implying that there is room for future refinement as evidence accumulates to show the validity of alternative parameters and/or methodologies that improve upon this practice. The current recommendations are illustrated in Figures 3 and 4 with the methodology fully explained in the Supplementary Information (Supplementary Information 2).

The statistical analysis as a non-continuous variable can also be considered as a secondary option. For this analysis, the term lymphocyte-predominant breast cancer (LPBC) has been coined, which can be used as a descriptive term for tumors that contain “more lymphocytes than carcinoma cells”. Typically the threshold of stromal lymphocytes for LPBC is around 50-60% of the stromal surface area. Note however that this term should not be used as a definition for a specific tumor type, but just as a descriptive term to facilitate discussions about lymphocyte-rich tumors. LPBC is not the same as medullary breast cancer, which has additional histological features. It is unclear if this cut-off will be used in the future as such dominant TILs infiltration in tumors has been found to be infrequent (~10%).
Selection of tumor areas for evaluation

For evaluating TILs, the boundaries of the invasive tumor should be identified with only TILs inside them evaluated. TILs in areas with crush artefacts, necrosis, and inflammation around biopsy sites or extensive central regressive hyalinization should not be scored. A necrotic biopsy is considered unscorable.

Most pathologists recognize that immune infiltrates can also be observed at some distance from the main tumor bed, surrounding extra- and intra-tumoral DCIS and also in adjacent normal lobules. These infiltrates outside of the tumor borders should not be included in the standardized stromal TILs-assessment, but they can be recorded as separate parameters for research purposes.

In areas surrounding the tumor, follicular aggregates, although rare can be observed, including TLS with germinal centers indicative of an active immune response. These aggregates should also not be included in the stromal TILs assessment; however, they can be evaluated separately as a research parameter, as they represent areas of T and B cell activation. TLS may become important in the future as more evidence emerges on their clinical relevance [13]. In the meantime, no formal recommendations for scoring TLS in daily practice can be put forward. TLS generally form in the peritumoral regions and therefore may be absent or underrepresented in cores or TMAs and heterogeneous in full sections. Furthermore, the criteria for unequivocal identification of TLS are currently unclear and the use of IHC may be required. Therefore, at the present time TLS should only be assessed in a research setting.
Intratumoral heterogeneity and evaluation of the invasive edge

Based on the collective experience within the TILs working group most tumors are not heterogeneous at the morphological level in their TILs content between FFPE-blocks of the same tumor, although there is currently no published evidence to support this statement. Nevertheless, heterogeneity in a single tissue section can be encountered. The most used methodology is global assessment of the slide by a trained pathologist, with a mean infiltrate score based on all available tissue being reported. Stromal TILs should be reported as a percentage. If the percentage of TILs is questionable, discuss the case with a second pathologist. In heterogeneous tumors, evaluate different regions and report the average. The working group does not recommend focusing on “hot spots”, defined as small areas with increased TILs. These small areas are often observed, and they should be included in the average TILs-assessment. There is no current evidence demonstrating whether the extent of heterogeneity might be clinically important. Since heterogeneity has not yet been investigated neither formally characterized either at the morphological nor functional level, the TILs working group’s current recommendation, although not formally supported by data is that if there is a choice between full sections and core biopsies, whole tissue sections are preferred over core biopsies.

There is no current evidence demonstrating that TILs at the invasive edge are functionally different from TILs in the center of the tumor. Based on this lack of knowledge, it was suggested that scoring TILs at the invasive edge as a separate parameter from TILs located in the inner stroma should at the present time merely be done in a research setting. In daily practice a distinction should not be made and all TILs within the tumor boundaries, including the invasive edge should be scored together as stromal TILs.
Inter- and intra-laboratory assessment, thresholds and machine learning algorithms

Most published or ongoing unpublished studies have not formally assessed intra-personal and inter-personal differences in scoring slides by pathologists. The accepted intra- and inter-personal discordance between pathologists depends on the clinical use/consequences of the measurements. The total allowable margin of error between pathologists will thus need to be determined in accordance with the clinical validity and utility of this methodology. It should be emphasized that if the primary purpose is to find an approach for daily practice, the impact on daily routine should be minimal without a significant increase in the pathologist’s time. A tutorial to help pathologists evaluate TILs accompanies this paper.

At present, there are no established thresholds for TILs. The consensus of the group was that a valid methodology was top priority and that thresholds for clinical decision can be determined once a solid methodology with clinical utility is in place. Therefore, there are currently no recommendations for the best threshold in clinical practice. We recommend that TILs be analyzed as a continuous variable unless it is clear that the prognostic information is not linearly associated with increasing levels of TILs. Further research will determine whether a threshold is required. We again emphasize that the level of TILs should not be used to withhold chemotherapy or trastuzumab therapy in TN and HER2+ BC, respectively.

The assessment of TILs by digital image analysis might be useful for standardization in the future, since this approach has the potential, for example, to determine the number of TILs per mm² stromal tissue as an exact measurement contrary to the approximate semi-quantitative evaluation suggested at this moment. Based on the standardized methodology recommended here (summarized in Table 2), an inter-laboratory Ring study will be initiated to assess the reproducibility and clinical validity of TILs assessment, including machine learning algorithms.
Future Directions

This article has focused on a standardized approach for measuring the percentage of stromal TILs in primary tumor specimens prior to therapy, using visual assessment of standard H&E-stained sections. Our goal is to facilitate the use of TILs as a biomarker in research and clinical trial settings (i.e. as a stratification or adjustment factor). This should also provide a platform for pathologists to further engage in an effort for the harmonization of the assay.

While it may be argued that stromal TILs are a robust prognostic factor in TNBC treated with standard adjuvant anthracycline-based chemotherapy, with three published prospective validation studies currently provide level I evidence for its clinical validity, we do not yet advocate that adjuvant treatment decisions be based on the level of TILs in the baseline TNBC neither on HER2+ cancer samples because the analytical validity and clinical utility of TILs in these subtypes remains to be firmly determined. We are yet to determine if TILs will be predictive of response to immunotherapeutic regimens, in particular T-cell checkpoint inhibition, which ultimately may be its clinical utility.

While TILs have been measured morphologically and have been shown to add predominantly prognostic information, methodological open questions in the morphological evaluation of TILs still remain (Supplementary Information 3 and Supplementary Figure 1). The measurement on H&E-stained slides most likely represents the beginning of the efforts to use infiltrating cell properties as companion diagnostic tests. The huge complexity of lymphocytes, both from the standpoint of cell type and activation suggests that molecular characterization of this infiltrate
may add both sensitivity and specificity to the predictive value of morphologically defined TILs [45, 46]. Thus, as a field, we should be open to the introduction of molecular methods, most likely in situ, that can classify the TILs-component. However, at this time, these molecular methods are still experimental and not sufficiently documented for introduction into standard practice.

Further scientific questions concerning the underlying breast cancer pathology associated with higher levels of TILs at diagnosis, the relevance of TILs subtyping and what can be done clinically to enhance host anti-tumor immune responses have not been addressed at present. As our understanding evolves and clinical evidence accumulates, some of the methodological statements provided in these 2014 guidelines will be updated in subsequent articles.

Acknowledgments:

No funding was received for this project. No medical writing was used for this manuscript.

Conflicts of Interest:

Carsten Denkert is shareholder of Sividon Diagnostics, Cologne and is a named co-inventor of patent application EP14153692.0. The other authors declared no conflicts of interest
References:


43. Vinayak S, Gray RJ, Adams S et al. Association of increased tumor-infiltrating lymphocytes (TILs) with immunomodulatory (IM) triple-negative breast cancer (TNBC) subtype and response to neoadjuvant platinum-based therapy in PrECOG0105. J Clin Oncol 32:5s, 2014 (suppl; abstr 1000)


Table and Figure Legends

Table 1
Adjuvant and neoadjuvant studies that have assessed TILs and prognosis are included in Table 1

Table 2
Recommendations for assessing tumor-infiltrating lymphocytes (TILs) in breast cancer

Figure 1
The cellular crosstalk between different leukocyte subsets and their predominant contribution to either pro- or anti-tumor activities, including myeloid lineage leukocytes, tumor-associated macrophages with either pro-tumorigenic (M2) or anti-tumorigenic (M1) properties, helper T cell subsets, cytotoxic T cells, regulatory T-cells, B cells, dendritic cells and myeloid-derived suppressor cells are shown. These cells play central roles in shaping the microenvironment via the factors they produce thereby driving either an immune mediated anti- or pro-tumor activities in the microenvironment.

Figure 2
Morphology, definitions, biological and diagnostic relevance of the different immune infiltrates found in breast cancer.

Figure 3
Standardized approach for TILs-evaluation in Breast Cancer
Figure 4

Standardisation and guidelines for TILs-assessment. Stromal TILs should be reported as a percentage (the schematic images might provide some guidance). If the percentage of TILs is questionable, discuss the case with a second pathologist. In heterogeneous tumors, evaluate different regions and report the average. For this standardized graphic, images were selected that are representative of different TILs- levels, based on the results of three pathologists as well as image analysis. The stromal area was marked in each image. The central images are digitally generated graphics showing the same ROI (Region Of Interest) and a similar density of TILs as the corresponding histological image. Please note that the central images contain idealized TILs generated graphically with comparably density, but not with the exact configuration and distribution as the TILs in the histological images.
Figure 2 Morphology, definitions, biological and diagnostic relevance of the different immune infiltrates found in breast cancer

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<th>Morphology</th>
<th>Definition and biological relevance</th>
<th>Diagnostic relevance</th>
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<td><strong>Lymphocyte-predominant breast cancer (LPBC)</strong></td>
<td>Working category to describe tumors with “more lymphocytes than tumor cells”.</td>
<td>Definitions vary across studies with stromal TILs of 50-60% used as a threshold. LPBC can be used for predefined subgroup analyses and for description of tumors with a particularly high immune infiltrate, however, keep in mind that TILs are a continuous parameter and the threshold for LPBC is still arbitrary.</td>
</tr>
<tr>
<td><strong>Stromal TILs</strong></td>
<td>Indicator of increased accumulation of immune-cells in tumor tissue</td>
<td>Stromal TILs have been shown to be predictive for increased response to neoadjuvant chemotherapy as well as improved outcome after adjuvant chemotherapy. Based on current data, this parameter is the best parameter for characterization of TILs.</td>
</tr>
<tr>
<td><strong>Intratumoral TILs</strong></td>
<td>TILs with direct cell-cell contact with carcinoma cells, might be an indicator of direct cell-based anti-tumor effects.</td>
<td>Several studies have shown that intratumoral TILs are more difficult to evaluate and do not provide additional predictive/prognostic information compared to stromal TILs.</td>
</tr>
<tr>
<td><strong>TILs at the invasive margin</strong></td>
<td>The localization of TILs are the invasive edge is included in the evaluation approach presented in this guideline.</td>
<td>For breast cancer there are no studies with a separate evaluation of TILs at the invasive edge. For practical purposes, the reliable evaluation of the invasive edge might be difficult when using core biopsies in the neoadjuvant setting.</td>
</tr>
<tr>
<td><strong>Tertiary lymphoid structures (TLS)</strong></td>
<td>Typically localized in the surrounding area of the tumor. TLS might be localized in normal tissue directly adjacent to the tumor, consisting of a T cell zone next to a B cell follicle, often with germinal centers.</td>
<td>While these structures may be important for the biology of tumor-immune reactions, they are not yet optimized for non-research based assessments. The main problem is that TLS have a spatial heterogeneity and are principally localized in areas surrounding the tumor. They might not be in the plane of the tissue section that is being evaluated, in particular when using core biopsies. Furthermore, it might be difficult to distinguish lymphoid aggregates from true TLS, in particular when the germinal center is not in the plane of the section.</td>
</tr>
</tbody>
</table>
Figure 3 Standardized approach for TILs-evaluation in Breast Cancer

Step 1: Select tumor area

Step 2: Define stromal area

Step 3: Scan at low magnification

Step 4: Determine type of inflammatory infiltrate

Step 5: Assess the percentage of stromal TILs
(examples of percentages shown in Figure 4)
Figure 4 Standardized evaluation of the percentage of stromal TILs
## Table 1 Overview of adjuvant and neoadjuvant studies

<table>
<thead>
<tr>
<th>Reference</th>
<th>Study (level of evidence if applicable according to Simon et al [28])</th>
<th>Regimen</th>
<th>Tumor tissue Assay</th>
<th>Sample size</th>
<th>Correlation with outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>[19]</td>
<td>BIG 02-98 (category B)</td>
<td>A→CMF or AC→CMF</td>
<td>Full section H&amp;E</td>
<td>2009 total</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>256 TNBC</td>
<td>Stromal TILs (sTIL)</td>
</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td>(continuous, per 10% increase) univariate:</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>HR 0.84 (p=0.02, DFS)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>HR 0.82 (p=0.02, OS)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
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<td></td>
<td>sTIL multivariate:</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>HR 0.85 (p=0.02, DFS)</td>
</tr>
<tr>
<td>Study Reference</td>
<td>Treatment</td>
<td>Biomarker</td>
<td>Stain</td>
<td>Sample Size</td>
<td>Univariate HR</td>
</tr>
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</tr>
<tr>
<td>[21]</td>
<td>E2197 E1199 (category B)</td>
<td>AC vs AC AC → Docetaxel or paclitaxel</td>
<td>Full section H&amp;E</td>
<td>481 TNBC</td>
<td>HR 0.86 (p=0.02, DFS)</td>
</tr>
</tbody>
</table>

**Notes:**
- **HR 0.83 (p=0.02, OS)**
- **297 HER2+** None
- **1078 HR+** None

sTIL (continuous, per 10% increase)

Univariate:
- HR 0.86 (p=0.02, DFS)
- HR 0.81 (p=0.01, OS)

Multivariate:
- HR 0.84 (p=0.005, DFS)
- HR 0.79 (p=0.003, OS)
<table>
<thead>
<tr>
<th><strong>[20]</strong></th>
<th>FINHER (category B)</th>
<th>Docetaxel or Vinorelbine → FEC With trastuzumab if HER2+</th>
<th>Full section H&amp;E</th>
<th>934 total</th>
<th>None</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>134 TNBC</td>
<td>sTIL (continuous, per 10% increase)</td>
</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td>Univariate:</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>HR 0.79 (p=0.03, DDFS)</td>
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<td></td>
<td>HR 0.80 (p=0.08, OS)</td>
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<td></td>
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<td></td>
<td>Multivariate:</td>
</tr>
<tr>
<td></td>
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<td></td>
<td></td>
<td></td>
<td>HR 0.77 (p=0.02, DDFS)</td>
</tr>
<tr>
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<td></td>
<td></td>
<td></td>
<td>HR 0.81 (p=0.14, OS)</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>209 HER2+</td>
<td>sTIL (continuous, per 10% increase) correlate with DDFS (HR 0.82,</td>
</tr>
<tr>
<td>Study Reference</td>
<td>Subjects</td>
<td>Treatment</td>
<td>Method</td>
<td>Tumor CD8+ T Cells</td>
<td>Description</td>
</tr>
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</tr>
<tr>
<td>[41]</td>
<td>591 HR+</td>
<td>None</td>
<td>TMA CD8, FOXP3 immunohistochemistry</td>
<td>12,439</td>
<td>CD8+ T Cells in tumor and stroma was associated with 28% and 21% reduced risk of BCSS. Greater benefit in ER-negative disease and ER=/HER2</td>
</tr>
</tbody>
</table>
| [8]             | Consecutive CMF | TMA CD8-immunohistochemistry | 1334 | Binary high versus low: Total CD8 correlates with BCSS (HR 0.55, p=0.001multivariate- }
<table>
<thead>
<tr>
<th>Study Reference</th>
<th>Study Design</th>
<th>Treatment</th>
<th>Immunohistochemistry</th>
<th>Number of Patients</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>[51]</td>
<td>Consecutive</td>
<td>MF, AC, FAC or no chemotherapy</td>
<td>TMA CD8- immunohistochemistry</td>
<td>1985 HR+, None</td>
<td>496 TNBC Binary any versus none: CD8 correlates with BCSS, multivariate iTIL (intratumoral TILs) HR 0.48, p&lt;0.001</td>
</tr>
<tr>
<td>[30]</td>
<td>Institutional</td>
<td>Varied chemotherapy not specified</td>
<td>PD-L1 mRNA TILs</td>
<td>636</td>
<td>Higher PD-L1 mRNA associated with better recurrence-free survival.</td>
</tr>
<tr>
<td>Reference</td>
<td>Study</td>
<td>Regimen</td>
<td>Tumor tissue Assay</td>
<td>Sample size</td>
<td>Correlation with outcome</td>
</tr>
<tr>
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</tr>
<tr>
<td>[31]</td>
<td>Institutional cohort</td>
<td>anthracycline-taxane-based regimens</td>
<td>CD3-immunohistochemistry</td>
<td>73</td>
<td>CD3 positively correlated with pCR</td>
</tr>
</tbody>
</table>

Consecutive CMF, AC, CEF or CAF TMA CD3-immunohistochemistry 255 Binary high versus low total CD3 correlates with DFS in anthracycline group (HR 0.25, p=0.0056)
<table>
<thead>
<tr>
<th>Reference</th>
<th>Study Description</th>
<th>Treatment</th>
<th>Biomarker</th>
<th>N</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>[18]</td>
<td>GeparDuo GeparTrio (category B)</td>
<td>EC-Doc (GeparDuo) TAC +/- Vinorelbine/Capecitabine (GeparTrio)</td>
<td>TILs in H&amp;E core biopsy</td>
<td>1058</td>
<td>Stromal TILs and LPBC associated ($p=0.001$) with pCR TILs significant in subgroups (HR+/-; HER2+/-)</td>
</tr>
<tr>
<td>[32]</td>
<td>Publicly available gene expression data from EORTC 10994/BIG 00-01</td>
<td>FEC vs TET Gene expression data</td>
<td>ER-</td>
<td>113</td>
<td>TILs correlate with pCR ($p=0.001$)</td>
</tr>
<tr>
<td>[33]</td>
<td>Institutional cohort</td>
<td>Neoadjuvant anthracycline-based; cyclophosphamide-based or taxane-based regimens</td>
<td>TILs in H&amp;E core biopsy</td>
<td>474 total 92 TNBC</td>
<td>TILs correlate with pCR in TNBC ($p=0.004$)</td>
</tr>
<tr>
<td>[34]</td>
<td>Institutional cohort</td>
<td>Neoadjuvant anthracycline-taxane-based regimens</td>
<td>TILs in H&amp;E core biopsy</td>
<td>68</td>
<td>TILs correlate with pCR ($p&lt;0.0001$)</td>
</tr>
<tr>
<td>[35]</td>
<td>Institutional cohort</td>
<td>Neoadjuvant paclitaxel FEC CD8, FOXP3, IL17F immunohistochemistry</td>
<td>180</td>
<td>CD8, FOXP3 positively correlated with pCR ($p&lt;0.001$)</td>
<td></td>
</tr>
<tr>
<td>[36]</td>
<td>Publicly available gene expression data (7 cohorts)</td>
<td>Anthracycline-based neoadjuvant therapy IGKC gene expression</td>
<td>845</td>
<td>IGKC predicted response to NACT ($p&lt;0.001$)</td>
<td></td>
</tr>
<tr>
<td>[37]</td>
<td>GeparQuinto Predict clinical study (category B)</td>
<td>EC-Doc Core biopsy H&amp;E</td>
<td>313</td>
<td>Stromal TILs and LPBC associated with pCR ($p&lt;0.001$)</td>
<td></td>
</tr>
<tr>
<td>Reference</td>
<td>Study Design</td>
<td>Treatment</td>
<td>Immunohistochemistry</td>
<td>Sample Size</td>
<td>Findings</td>
</tr>
<tr>
<td>-----------</td>
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</tr>
<tr>
<td>[9]</td>
<td>Institutional cohort</td>
<td>Neoadjuvant anthracycline-taxane-based regimens</td>
<td>CD8, CD4, FOXP3 immunohistochemistry</td>
<td>153</td>
<td>CD8, CD4, FOXP3 positively correlated with pCR ($p=0.003$, $p&lt;0.001$ and $p=0.001$)</td>
</tr>
<tr>
<td>[38]</td>
<td>Institutional cohort</td>
<td>Neoadjuvant anthracycline-taxane-based regimens</td>
<td>TILs by H&amp;E; CD3, CD8, FOXP3 immunohistochemistry</td>
<td>175</td>
<td>TILs, CD3, CD8, FOXP3 positively correlated with pCR</td>
</tr>
<tr>
<td>[39]</td>
<td>TVA neoadjuvant phase 2 study</td>
<td>4FEC100 +/- 4 docetaxel + panitumumab</td>
<td>CD8 immunohistochemistry</td>
<td>47 TNBC</td>
<td>High CD8/FoxP3 were predictive of pCR</td>
</tr>
<tr>
<td>[50]</td>
<td>Pooled analysis of publicly available gene expression data (8 cohorts)</td>
<td>Neoadjuvant anthracycline</td>
<td>STAT1 and immune response</td>
<td>996</td>
<td>High score of STAT1 and immune response</td>
</tr>
<tr>
<td>/ anthracycline + taxane based chemotherapy</td>
<td>response gene modules</td>
<td>gene modules is associated with increased pCR rates in all breast cancer subtypes based on ER and HER2 status</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table 2 Recommendations for assessing tumor-infiltrating lymphocytes (TILs) in breast cancer

1. TILs should be reported for the stromal compartment (= % stromal TILs). The denominator used to determine the % stromal TILs is the area of stromal tissue (i.e. area occupied by mononuclear inflammatory cells over total intratumoral stromal area), not the number of stromal cells (i.e. fraction of total stromal nuclei that represent mononuclear inflammatory cell nuclei).

2. TILs should be evaluated within the borders of the invasive tumor.

3. Exclude TILs outside of the tumor borders, e.g. around DCIS and normal lobules.

4. Exclude TILs in tumor zones with crush artefacts, necrosis, regressive hyalinization as well as in the previous core biopsy site.

5. All mononuclear cells (including lymphocytes and plasma cells) should be scored, but polymorphonuclear leukocytes are excluded.

6. 1 section (4-5 µm, magnification 200-400x) per patient is currently considered to be sufficient.

7. Full sections are preferred over biopsies whenever possible. Cores can be used in the pre-therapeutic neoadjuvant setting; currently no validated methodology has been developed to score TILs after neo-adjuvant treatment.

8. A full assessment of average TILs in the tumor area by the pathologist should be used. Do not focus on hotspots.

9. The working group’s consensus is that TILs may provide more biological relevant information when scored as a continuous variable, since this will allow more accurate statistical analyses, which can later be categorized around different thresholds. However, in daily practice most pathologists will rarely report for example 13.5% and will round up
to the nearest 5-10%, in this example thus 15%. Pathologist should report their scores in as much detail as the pathologist feels comfortable with.

10. TILs should be assessed as a continuous parameter. The percentage of stromal TILs is a semiquantitative parameter for this assessment, for example, 80% stromal TILs means that 80% of the stromal area shows a dense mononuclear infiltrate. For assessment of percentage values, the dissociated growth pattern of lymphocytes needs to be taken into account. Lymphocytes typically do not form solid cellular aggregates, therefore the designation “100% stromal TILs” would still allow some empty tissue space between the individual lymphocytes.

11. No formal recommendation for a clinically relevant TIL threshold(s) can be given at this stage. The consensus was that a valid methodology is currently more important than issues of thresholds for clinical use, which will be determined once a solid methodology is in place. LPBC (lymphocyte predominant breast cancer) can be used as a descriptive term for tumors that contain “more lymphocytes than tumor cells”. However, the thresholds vary between 50-60% stromal lymphocytes.