Spheroid Culture for Enhanced Differentiation of Human Embryonic Stem Cells to Hepatocyte-Like Cells

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Stem cell-derived hepatocyte-like cells hold great potential for the treatment of liver disease and for drug toxicity screening. The success of these applications hinges on the generation of differentiated cells with high liver specific activities. Many protocols have been developed to guide human embryonic stem cells (hESCs) to differentiate to the hepatic lineage. Here we report cultivation of hESCs as three-dimensional aggregates that enhances their differentiation to hepatocyte-like cells. Differentiation was first carried out in monolayer culture for 20 days. Subsequently cells were allowed to self-aggregate into spheroids. Significantly higher expression of liver-specific transcripts and proteins, including Albumin, phosphoenolpyruvate carboxykinase, and asialoglycoprotein receptor 1 was observed. The differentiated phenotype was sustained for more than 2 weeks in the three-dimensional spheroid culture system, significantly longer than in monolayer culture. Cells in spheroids exhibit morphological and ultrastructural characteristics of primary hepatocytes by scanning and transmission electron microscopy in addition to mature functions, such as biliary excretion of metabolic products and cytochrome P450 activities. This three-dimensional spheroid culture system may be appropriate for generating high quality, functional hepatocyte-like cells from ESCs.

Introduction

Treatments for end stage liver failure are largely dependent on liver organ or hepatocyte cell transplantation, which are limited by the availability of donor organs or cells [1–3]. Severe end-stage liver disease may benefit from an extracorporeal bioartificial liver (BAL) device as a bridge to liver transplantation or even regeneration [4,5]. However, such BAL devices for extracorporeal liver support have to resort to xenogeneic cell sources or tumorigenic cell lines because of the lack of human hepatocytes. A renewable cell source, especially stem cell-derived human hepatocytes, will greatly enhance the prospect of liver cell based therapy [6]. Stem cell-derived hepatocytes may also find applications in drug metabolism and toxicity studies [7–9]. Successful derivation of hepatocytes from pluripotent human stem cells will therefore, ensure virtually unlimited cell sources for discovery and clinical applications.

Human embryonic stem cells (hESCs) cultured under low adhesive conditions form aggregates called embryoid bodies (EBs). They have a propensity to spontaneously differentiate to multiple cell lineages, including the hepatic endoderm, but at a rather low efficiency [10]. In contrast directed and controlled differentiation to hepatic lineage has had more success in many laboratories [11–14]. These directed differentiation protocols entail plating of hESC on extracellular matrices and treatment with a series of cocktails of cytokines and growth factors to promote hepatic differentiation as evidenced by the presence of liver markers [15].

We recently described a 20-day, four stage protocol for differentiation of ES and induced pluripotent stem cells (iPS), from both human and mouse, to cells of hepatic lineage [11]. Cells grown to confluency on matrigel were guided toward definitive endoderm with Activin-A and Wnt3a, followed by specification to hepatic endoderm by treatment with bone morphogenetic protein (BMP)4 and basic fibroblast growth factor (bFGF), to bipotential hepatoblasts using aFGF, FGF4b, and FGF8, and finally toward a hepatocyte-like cell state by treatment with hepatocyte growth factor (HGF) and Follistatin. This stepwise treatment results in ~10%–20% of the cell population demonstrating a fetal hepatocyte-like with some characteristics of adult hepatocytes.

Others have used similar sets of growth factors invariably encompassing combinations of Activin A, Wnt3, bFGF, BMP-4, HGF, Oncostatin M, and/or Dexamethasone (Dex) [16,17]. The number of stages, in which different combinations of growth factors are used, differ somewhat, in various protocols, ranging from three to five.

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The current directed differentiation protocols give rise to heterogeneous populations. The transcript levels of many key hepatic genes are still relatively low. The yields of hepatocyte-like cells, as well as their functional maturity, need to be further enhanced. Furthermore, the differentiated hepatic functions in those cells need to be sustained for a longer period for most applications.

Primary hepatocytes, when cultured on a surface with a decreased adhesiveness and at a subconfluent density, self-assemble into spheroids that exhibit enhanced levels of a variety of hepatic functions. They sustain hepatic functions over a longer period in culture than the monolayer culture [18,19].

We previously applied our differentiation protocol to direct the differentiation of rat multipotent adult progenitor cells cultivated as 3D aggregates to hepatic lineage. The resulting hepatocyte-like cells had higher liver specific functions, including albumin (ALB) and urea secretion and cytochrome P450 (CYP450) activity, than those obtained from monolayer culture [20]. Here we examined the effect of cultivation of hESCs in 3D spheroids on the maturation of the hepatocyte-like progeny. As hESCs differentiate spontaneously when cultivated as EBs, they were thus, first differentiated toward hepatic lineage in monolayer cultures for 20 days, and then formed spheroids. We report here the enhanced maturation and liver functions for hESC-derived hepatocyte like cells cultured as spheroids.

Materials and Methods

HESC culture

The hESC line HSF6 was cultured as previously described (1). Briefly, hESCs were maintained in Dulbecco’s modified Eagle’s medium (DMEM) (Gibco/BRL), 20% Knockout Serum Replacement (Gibco), 2 mM glutamine, 0.1 mM nonessential amino acids, 0.1 mM β-mercaptoethanol, and bFGF (R&D) (4 ng/mL) on mouse embryonic fibroblasts (MEFs). MEFs were derived from E13–E14 CF-1 mice (Charles River Laboratories) and mitotically inactivated using gamma radiation (3,000 rads). hESCs were plated on the MEFs and cultured in hESC medium at 37°C in 10% CO2. The cells were routinely passaged using 0.1% (w/v) Collagenase Type IV (Gibco) in ES basal media every 3–5 days.

Hepatocyte differentiation

HSF6 cells were plated in 12 well plates, precoated with 2% matrigel (BD Biosciences) in MEF-conditioned media (CM) until 50% confluency was reached. To initiate differentiation, the CM was aspirated, the cells were washed once with PBS, and differentiation medium was added. The differentiation basal medium consisted of a 60/40 (v/v) mixture of low glucose DMEM (Gibco) and MCDB-201 (Sigma), and supplemented with 0.026 μg/mL ascorbic acid 3-phosphate (Sigma), linoleic acid bovine serum albumin (LA-BSA; Sigma) (1 mg/mL BSA and 8.13 μg/mL LA), insulin-transferrin-selenium (Sigma) (2.5 μg/mL insulin, 1.38 μg/mL transferrin, 1.25 ng/mL sodium selenite), 0.4 μg/mL Dex (Sigma), 4.3 μg/mL β-mercaptoethanol (Hyclone), 100 IU/mL penicillin and 100 μg/mL streptomycin (Gibco) and qualified fetal bovine serum (2% (v/v) for the first 6 days and 0.5% (v/v) for the subsequent time period).

Cytokines and growth factor were supplemented as follows: (1) D0: Activin A (100 ng/mL) and Wnt3a (50 ng/mL) (2) D6: bFGF (10 ng/mL) and BMP4 (50 ng/mL) (3) D10: FGF8b (25 ng/mL), aFGF (50 ng/mL) and FGF4 (10 ng/mL) (4) D14: HGF (20 ng/mL) and Follistatin (100 ng/mL). 50% media change was performed every 2 days using medium of the corresponding differentiation stage. On D0, D6, D10, and D14, medium was completely replaced with fresh medium of the ensuing differentiation stage after a brief rinse with PBS.

Formation of hepatic spheroids

Cells were detached after treatment with 0.1% (w/v) collagenase (type IV) (Invitrogen) for 15–20 min at 37°C. After centrifugation (1,000 rpm 5 min) the cell pellet was treated with 0.05% (w/v) trypsin with 2% (v/v) chicken serum (37°C, 10 min). The cell suspension was then passed through a 100 μm nylon mesh to obtain a single cell suspension.

Approximately 8,000 cells in 100 μL of stage IV differentiation medium was added to each well of an ultra-low attachment (ULA) round bottomed 96 well plate (Corning). After centrifugation for 4 min at 1,500 rpm (400g), cells settled to the bottom of the well to form spheroids in the following 2 to 3 days.

Characterization of spheroids

Quantitative real time PCR was used to measure the transcript levels of hepatic genes. Flow cytometry was used to quantify the cell fraction exhibiting mature hepatic markers using antibodies for asialoglycoprotein receptor (ASGPR; Thermoscientific, 1:10 dilution) and phosphoenolpyruvate carboxykinase (PEPCK; Santa-Cruz, H-300, 1:2,000 dilution).

The secondary antibodies used were anti-mouse IgG1 A488 labeled (Molecular Probes, 1:500 dilution) and anti-rabbit IgG A488 (Molecular Probes, 1:500 dilution). The secretion of ALB was measured by Enzyme-linked immunosorbent assay and CYP450 activities were imaged and quantified by biotransformation of fluorogenic substrate [21]. Biliary secretion was monitored through fluorescein activity. Ultrastructural features were assessed using scanning and transmission electron microscopy (TEM). Details of these assays can be found in Supplementary Material and Methods section; Supplementary Data are available online at www.liebertpub.com/scd

Results

Three-dimensional spheroid formation

The hESC line, HSF6, was differentiated towards the hepatic lineage as described [11]. On D20, cells were detached for formation of three-dimensional spheroids. Typically, we obtained 4 × 10⁷ cells/cm² from the monolayer culture with high viability (>90%). Approximately 8,000 cells were added to each well of a U-bottom ULA culture plates. Over time ~40% of the cells in each well form multiple small clusters, which progressively agglomerate into larger clusters in 2 days. The viability immediately after seeding was very high (Fig. 1). At 24h, cells within the clusters remained viable, while individual cells not in clusters were no longer viable. Subsequently cells in the cluster became more compact and the size of the aggregate decreases somewhat (Fig. 1a). Six days after the initiation of 3D formation (D26) the surface of
Expression of hepatic genes in 3D spheroids

The expression of several hepatic-specific genes in spheroid culture was evaluated and compared to cells maintained in monolayer culture until D26. Results are presented as ratio of transcripts of D26 cells to D20 cells in monolayer culture (Fig. 2). The transcript levels for all liver specific genes tested increased in D26 spheroids from D20 and are higher compared to D26 monolayer cultures. Most notably the transcript level of UDP glucuronosyltransferase 1A1 (UGT1A1), a gene encoding an enzyme involved in bilirubin conjugation in the liver, increased about 60-fold in spheroids comparing to about 2-fold increase in monolayer culture. The level of ALB transcript increased by about 16-fold over the 6 days in spheroids, but remained relatively unchanged in monolayer culture. The level of increase was more moderate for other genes, including arginase 1 (ARG1), an enzyme involved in the urea cycle, hepatocyte nuclear factor 4a (HNF4a), a liver enriched transcription factor, Factor VII (FVII), a member of coagulation factor family, and connexin 32 (CX32), a gap junction protein. Nevertheless, the levels of transcripts of those genes were all higher than the levels in cells maintained in monolayer culture.

The expression level of most mature hepatic transcripts, including ALB, ARG1, UGT1A1, HNF4a, and FVII were maintained till D35 in the 3D spheroid culture (Fig. 3). In comparison, the expression levels of these genes in the monolayer culture either remained at a constant but lower level than the spheroid culture or decreased in the same period.

Expression of hepatic proteins in spheroids

The differentiation of ESCs to the hepatic lineage yields heterogeneous progeny containing cells with different degrees of hepatocyte maturation, as well as other less well-identified cells. We examined the expression of two mature liver specific proteins; ASGPR-1 and PEPCK (Fig. 3g, h) in the spheroids by flow cytometry. ASGPR-1 plays an important role in the uptake and degradation of glycoproteins with exposed terminal galactose or N-acetylgalatosamine, while PEPCK is a pivotal enzyme in gluconeogenesis in liver. On D20, ~16% of the hESC hepatic progeny were ASGPR-1 positive. The fraction of ASGPR-1 positive cells increased to 60% on D26, after spheroid formation. A similar increase with the fraction of cells staining positive for PEPCK was seen, from 17% on D20 to 70% on D26, and about 80% on D32. In both cases, the percentage of cells expressing the hepatic-specific proteins increased significantly in spheroid cultures. The presence of PEPCK protein was confirmed by immunostaining after cell permeabilization (Fig. 3i). Further immunohistochemistry on D32 spheroids for alpha-fetoprotein (AFP), ALB, CK8, and CK18 is shown in Supplementary Fig. S3.
Structural characteristics of spheroids

Under scanning electron microscopy (SEM), the spheroids appear densely packed; the boundaries of individual cells were nearly indistinguishable (Fig. 4a, b). A micrograph with a higher magnification is shown in Fig. 4b. Typical hepatocyte morphology can be seen on cell clusters on the exterior of the aggregate (Fig. 4c) with some polygonal cells exhibiting abundant microvilli on the cell surface (Fig. 4d).

TEM revealed that cells in the spheroids on D30 displayed the typical polygonal morphology with numerous microvilli.
Intercellular bile-canaliculi delimited by tight junctions were also observed; Golgi apparatus, plentiful mitochondria, lipid droplets, and rough endoplasmic reticulum were all seen. Lysosomes and inter-cellular coated vesicles involved in degrading biological materials were also visible.

Hepatocytes exhibit polarized surface domains even in culture. In vivo they excrete bile acid into the bile canaliculi. In culture, they accumulate excreted chemical species in the space in between cells. Incubation with a dilute solution of fluorescein diacetate, which forms fluorescent fluorescein upon intracellular cleavage by esterase, results in an accumulation of excreted fluorescein in the intercellular space. This has been used as an indication of polarization of cellular surface domains [22,23]. Spheroids at D32 were incubated in 3 μg/mL of fluorescein diacetate, and the accumulation of fluorescein in the intercellular space was visible by confocal imaging (Fig. 4j), indicating cell polarization.

**Functional activity of HESC derived spheroids**

We next evaluated a number of hepatic activities in the hESC derived hepatocyte-like cells and compared the level in spheroids to that in cells maintained as monolayer. The cell numbers of spheroids in wells were enumerated and the average ALB secretion rate was estimated. ALB secretion rate, initially at 1.83 pg/cell/day on D20 before spheroid formation increased to 4.52 and 12.05 pg/cell/day on D26 and D32 respectively in spheroid culture, but subsided to 0.45 and 0.53 pg/cell/day in monolayer culture on D26 and D32 (Fig. 3j). Thus, spheroid formation resulted in a substantial increase in the ALB secretion rate, and the higher rate was sustained over the cultivation period.

We also assessed Cyp1A1/2 and Cyp2B6/7 activity by ethoxyresorufin-O-dealkylation (EROD) and pentoxyresorufin-O-dealkylation (PROD) reactions. The intracellular accumulation of the fluorescent resorufin after the cleavage of ethoxyresorufin or pentoxyresorufin, indicative of EROD

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**FIG. 4.** Images of hESC derived hepatic-like cell spheroids. Scanning electron microscopy images of hESC derived spheroids on D30 at (a) 70×(scale bar, 500 μm), (b) 200×(scale bar, 200 μm), (c) 1,000×(scale bar, 50 μm), (d) 7,000×(scale bar, 5 μm). (e) Transmission electron microscopic micrographs of hESC-derived spheroids on D30. Labels: nucleus (N); microvilli (MV); bile-canaliculi (BC); tight junctions (TJ); golgi body (GB); mitochondria (M); lipid droplets (L); and rough endoplasmic reticulum (rER) (scale bars: upper left panel, 10 μm; lower left panel, 500 nm; all others 1 μm). (f, g) Accumulation of fluorescent resorufin from Ethoxyresorufin O-dealkylation reaction in D32 spheroids without (f) or with (g) 24 h of β-naphthoflavone treatment. (h, i) Accumulation of fluorescent resorufin from Pentoxyresorufin O-dealkylation reaction in D32 spheroids, without (h) or with (i) 24 h of phenobarbital treatment. Insets show transmitted light images of the spheroids (scale bars, 50 μm). (j) Polarized accumulation of fluorescein in hESC derived spheroids (scale bar, 50 μm). Color images available online at www.liebertpub.com/scd
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(Fig. 4g) and PROD (Fig. 4i) activities was seen in spheroids under confocal microscopy. The spheroids were then treated with β-naphthoflavone and Phenobarbital, inducers of Cyp1A1 and Cyp2B6/7, respectively. A modest increase in EROD activity after β-naphthoflavone treatment (Fig. 4g) was observed, but little change was observed in PROD activity after Phenobarbital treatment (Fig. 4i).

EROD and PROD activities were also quantified by measuring the rate of resorufin product release into the media (Supplementary Fig. S4a, b). Resorufin release increased nearly five-fold after β-naphthoflavone induction but not after Rifampicin treatment (EROD assay). Likewise, resorufin release increased marginally after treatment with Phenobarbital but not Rifampicin (PROD assay).

To further interrogate the phenomenon of specific CYP450 activity, the expression of the relevant CYP450 gene products were evaluated by quantitative PCR with or without drug treatment (Supplementary Fig. S4c, d). Cyp1a1 transcript expression increased 24-fold after 24 h of β-naphthoflavone induction, while Rifampicin treatment resulted in minimal change in Cyp1a1 expression. Thus, consistent results were seen for intracellular accumulation and excretion of resorufin and transcript levels for the specific Cyp450: induction of Cyp1A1/2 expression and function by β-naphthoflavone, but no increase in expression and function of Cyp2B6/7 by Phenobarbital.

Discussion

In this study, we extended our monolayer hESC differentiation into three-dimensional spheroid culture. After a 20-day differentiation that directed cells to the hepatic lineage, cultivation of the cells as spheroids resulted in enhanced expression of many hepatic lineage markers. This was illustrated by the increased gene expression at the transcript level for a number of liver specific genes, including UGT1A1 and ARG1 (Fig. 2) and protein levels for the intracellular enzyme PEPCK and cell surface protein ASGPR-1, and secreted ALB (Fig. 3).

The 3D spheroid culture system also allowed maintenance of the differentiation markers for an extended period. During extended culture for 15 days in the spheroid system, expression levels of hepatic-specific transcripts were maintained at the level achieved after 6 days of spheroid culture (Fig. 3). In contrast, the expression levels of hepatic transcripts in cells that were maintained in monolayer culture significantly decreased between D26 and D35. The results suggest that the observed enhanced differentiation was not merely an outcome of the extended culture period. It is more likely other cell types present in the differentiated culture, as similar multistage differentiations from hESC and human iPSC cells toward hepatic lineages all yield cells expressing apparently similar transient levels of AFP and ALB, with levels of ALB expression similar to or exceeding expression in primary hepatocytes [16,17,38]. AFP is expressed in fetal liver cells but not in adult hepatocytes. In our case, the persistent
expression of AFP, thus, suggests the presence of immature fetal hepatocyte-like cells in the spheroids. Furthermore, some nonhepatic cells are likely still present within the spheroids and contribute to the heterogeneity of the cell population as seen in flow cytometric assays.

Flow cytometry for ASGPR-1 clearly demonstrated a definite enrichment of ASGPR-1 expressing cells on Day 26 of the 3D spheroid culture as compared to the Day 20 2D culture. Likewise, the enrichment in the fraction of PEPCK expressing cells from ~17% in the Day 20 samples to about 70% in Day 32 samples was also observed. Such increase in the fraction of more mature hepatocyte-like cells could be the results of spheroid formation. Another possibility of the enrichment of more mature cells in the initial phase of cell agglomeration is through specific cell-cell interactions.

The enhancement of both the fraction of differentiated cells and the liver specific gene expression through spheroid formation, although significant, still falls short of reaching the state of mature hepatocytes. This has been seen in all reports of hepatic lineage differentiation from embryonic and other stem cells. Recently, it was shown that overexpression of exogenous FOXA2 and HNF1a via adenoviral vectors in differentiating hESC or hiPSC increases the percentage of cells reaching a more mature liver phenotype [24]. ALB expression in the differentiated progeny suggested that a very high percentage of cells exhibit hepatocyte characteristics with slight enhancement of CYP450 activities. Thus, enhancing functional maturity through overexpression of hepatic transcription factors or genes may be worth exploring.

In summary, we have developed a simple and reproducible differentiation method consisting of step-wise monolayer based directed differentiation to the hepatic endoderm for 20 days, followed by a process of spontaneous self-assembly of cells into three-dimensional spheroids that can be maintained for at least an additional 15 days. This culture system not only improves the phenotypic and functional maturity of the differentiated cell population, but also allows for a more efficient maintenance of the cells for an extended culture time. This approach may thus, find application in high-throughput drug screening, in BAL support devices, as well as in hepatocyte transplantation.

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Author Disclosure Statement

No competing financial interests exist.

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