Review

Eosinophilic esophagitis is characterized by a non-IgEmediated food hypersensitivity

Review initiated by the EAACI Eosinophilic Esophagitis Interest Group

D. Simon^{*,1}, A. Cianferoni^{*,2,3}, J.M Spergel^{2,3}, S. Aceves⁴, M. Holbreich⁵, C. Venter^{6,7}, M. E. Rothenberg⁶, I. Terreehorst⁸, A. Muraro⁹, A. J. Lucendo ¹⁰, A. Schoepfer¹¹, A. Straumann¹² & H.-U. Simon¹³

¹Department of Dermatology, Inselspital, Bern University Hospital, University of Bern, Bern, Switzerland ²Division of Allergy and Immunology, Children's Hospital Philadelphia, University of Pennsylvania, Philadelphia, USA ³Perelman School of Medicine, University of Pennsylvania, Philadelphia, USA ⁴Department of Pediatrics and Medicine, Division of Allergy and Immunology, Center for Infection, Inflammation, and Immunology, La Jolla, USA ⁵Allergy and Asthma Consultants, Indianapolis, USA ⁶Division of Allergy and Immunology, Cincinnati Children's Hospital Medical Center, Cincinnati, USA ⁷School of Health Sciences and Social Work, University of Portsmouth, Portsmouth, UK ⁸Department of ENT and Pediatrics, AMC, Amsterdam, Netherlands ⁹Food Allergy Referral Centre Veneto Region, Padua General University Hospital, Padua, Italy ¹⁰Department of Gastroenterology, Hospital General de Tomelloso, Tomelloso, Spain ¹¹Division of Gastroenterology and Hepatology, Centre Hospitalier Universitaire Vaudois/CHUV, Lausanne, Switzerland ¹²Swiss EoE Research Network, Olten, Switzerland ¹³Institute of Pharmacology, University of Bern, Bern, Switzerland

*These authors contributed equally to this paper.

Correspondence: Dagmar Simon, MD; Department of Dermatology, Inselspital, Bern University Hospital, CH-3010 Bern, Switzerland; Tel.: +41 31 632 2278; E-mail: dagmar.simon@insel.ch

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Key words: eosinophilic esophagitis, food allergy, immunoglobulin E, epithelial barrier, immune response, microbiota, eosinophils

Abbreviations used in this article: ACD, allergic contact dermatitis; AD, atopic dermatitis; APT, atopy patch test; DHR, drug hypersensitivity reaction; EoE, eosinophilic esophagitis; FPIES, food protein-induced enterocolitis; GERD, gastro-esophageal reflux disease; GI, gastrointestinal; Ig, immunoglobulin; IBD, inflammatory bowel disease; OAS, oral allergy syndrome; PPI-REE, proton pump inhibitor-responsive esophageal eosinophilia; SFED, six food elimination diet; SPT, skin prick test; TCR, T cell receptor; TNF, tumor necrosis factor.

Abstract

Eosinophilic esophagitis (EoE) is a chronic disease characterized clinically by symptoms of esophageal dysfunction and histologically by eosinophil-predominant inflammation. EoE is frequently associated with concomitant atopic diseases and immunoglobulin E (IgE) sensitization to food allergens in children as well as to aeroallergens and crossreactive plant allergen components in adults. Patients with EoE respond well to elemental and empirical food elimination diets. Recent research has, however, indicated that the pathogenesis of EoE is distinct from IgE-mediated food allergy. In this review, we discuss the individual roles of epithelial barrier defects, dysregulated innate and adaptive immune responses, and of microbiota in the pathogenesis of EoE. Although food has been recognized as a trigger factor of EoE, the mechanism by which it initiates or facilitates eosinophilic inflammation appears to be largely independent of IgE and needs to be further investigated. Understanding the pathogenic role of food in EoE is a prerequisite for the development of specific diagnostic tools and targeted therapeutic procedures.

Current definition of EoE

As a consequence of intense research in the field of esophageal eosinophilia, our understanding of eosinophilic esophagitis (EoE) has developed from strict clinicpathologic criteria leading towards a conceptual definition which includes pathogenic aspects (1-3). According to current recommendations, EoE represents a chronic, immune/antigen-mediated esophageal disease characterized clinically by symptoms of esophageal dysfunction and histologically by eosinophil-predominant inflammation (2). EoE has been recognized as having a spectrum of clinical signs and symptoms, endoscopic findings as well as pathologic features. However, the term "immune/antigen-mediated" does not address the question where EoE should be positioned in the wide range between autoimmune and allergic diseases. In addition, there are likely subgroups of patients who do not meet this strict definition; for example, some have less than 15 eosinophils per high power field (hpf), but otherwise fulfill the criteria of EoE (2).

The two most common causes of eosinophilia in the esophagus, normally devoid of eosinophils in healthy humans, are gastroesophageal reflux (GERD) and EoE (2). However, yet another form of esophageal eosinophilia has recently emerged having clinical manifestations and histological features indistinguishable from EoE, but distinct from GERD, apart from the fact that it is responsive to high-dose of PPI whereas EoE is histologically refractory to PPI. Hence, it is called PPI-responsive esophageal eosinophilia (PPI-REE) (4,5). Patients with PPI-REE frequently exhibit environmental and/or food allergen sensitizations like patients with EoE, whereas the atopy rate in patients with GERD is similar to that of the general population. Moreover, the inflammatory markers of PPI-REE are more similar to those of EoE than of GERD: positive for factors involved in eosinophil chemotaxis (eotaxin 3, CCL26), barrier integrity (desmoglein 1, DSG1), tissue remodeling (periostin, POSTN), and mast cell specific activity (carboxypeptidase A, CPA3) (4). The molecular signature typical of PPI-REE and EoE could be reversed by PPI therapy only in PPI-REE (4), suggesting the molecular signature is either a sign of disease or marker of eosinophilic inflammation. Mechanisms proposed to explain the PPI response include an acid-independent, anti-inflammatory action of PPIs on the one hand, or a PPI-induced restoration of esophageal barrier function on the other (6). In summary,

it is possible that PPI-REE and EoE are the consequence of the same underlying immunologic mechanism, but additional research is required to confirm this concept.

Already early reports on EoE mentioned concomitant allergic diseases and elevated total serum immunoglobulin E (IgE) levels in about 70% of the patients (7,8). After receiving elemental formulas, children with esophageal eosinophilia not responding to pharmacological and/or surgical anti-reflux therapy, showed marked improvements (9). This observation suggested that EoE could represent an allergic disease in which food proteins play an important role. However, further research revealed that EoE seems not to be simply an IgE-mediated food allergy. What, then, are the underlying causes of EoE and what role might food and/or other antigens play in the pathogenesis? In this review initiated by the EAACI Eosinophilic Esophagitis Interest Group, we will discuss recently published work on EoE in the context of an immune/antigen-mediated disease.

EoE-associated IgE-sensitization to food and aeroallergens

EoE is associated with elevated total IgE levels as well as IgE-sensitization to food and aeroallergens (10). In a pediatric EoE cohort, sensitizations to food and environmental allergens have been observed in 75% and 79%, respectively (11). Skin prick testing in children with EoE revealed increasing reactivity with inhalant allergen with age, while the reactivity to foods decreased (12). Children with EoE were mainly sensitized to milk, eggs, soy, wheat/rye, beef and peanuts (13). In adult EoE patients, specific IgEs to food and inhalant allergen components have been detected in 91% (14). These patients were mainly sensitized to pollens, in particular cross-reactive plant allergen components such as profilins and pathogenesis-related (PR)10 proteins (14). Noteworthy is the observation of local immunoglobulin class switching and production of IgE in the esophageal mucosa of pediatric EoE patients (15). Considering all these findings, EoE was initially suspected of being an IgE-mediated allergy to food and cross-reactive plant allergens.

On the other hand, clinical trials of targeted food elimination diets, as well as of IgE blocking, failed to show an IgE-mediated mechanism. Measuring specific IgE levels and/or skin prick testing were not sufficient to clearly identify causative food allergens (13,14,16,17). Moreover, elimination diets based solely on IgE-sensitization to food allergens as determined by skin prick tests (SPT) and/or specific IgE determinations could

not improve EoE in a significant number of patients (16,18,19). The positive predictive values for causative food identified by SPT ranged from 26% to 96%, with an average of 47% (16). Based on the assumption that IgE plays a key role in pathogenesis, a therapy with an anti-IgE antibody for 12 weeks in pediatric and adult EoE patients was initiated in a non-placebo controlled study resulting in a remission rate of only 33% despite an effective reduction of IgE levels observed in the esophageal tissue (20). In a double-blind placebo-controlled study anti-IgE treatment was not better than placebo in inducing EoE remission (21). Taken together, recent clinical and research data lead us to conclude that EoE, while often associated with IgE sensitization, is not simply an IgE-mediated food allergy.

EoE exhibits features of a Th2 predominant inflammation

The inflammation of EoE is predominantly eosinophilic, but is also characterized by increased numbers of T cells and mast cells infiltrating the esophageal mucosa, as well as high expression levels of IL-5 and TNF- α (Figure 1) (22). Transcriptome analysis of EoE tissue showed a distinct Th2 pattern with significantly elevated mRNA levels of eotaxin-3, IL-5, IL-5 receptor α -chain and IL-13 (23,24). In experimental models, both eotaxin and IL-5 were essential for eosinophil recruitment, accumulation and activation in the esophagus as well as for epithelial hyperplasia and remodeling (25-28). Moreover, IL-13 can induce eotaxin-3 production by esophageal epithelial cells (29). In addition to the Th2 cytokines, EoE patients show elevated blood levels of IL-1 α , IL-6 and IL-8, but lower levels of IL-12, IL-17 and CD40L as compared with healthy controls, while the gene expression of receptors for IL-1, IL-9 and IL-17 is also upregulated in EoE lesions (23,24).

Treatment with corticosteroids resulted in a reduced expression of eotaxin-3, IL-5 and IL-13 and was followed by a decrease of eosinophil numbers in the esophagus of EoE patients (29). Although reducing eosinophil inflammation in the esophagus, blocking IL-5 or IL-13 with therapeutic antibodies has yet to be proven to be clinically useful, although trends have been seen in preliminary studies (30,31). In summary, Th2 immune responses are a striking feature and most likely contribute to the pathogenesis of EoE, but are not the sole players as pro-inflammatory cytokines are also expressed that may regulate additional responses.

Lessons learnt from hypersensitivity reactions of the skin

EoE shares many similarities with dermatoses that are due to T cell responses of the skin independent of IgE. Therefore, it appears logical to consider antigen-triggered T cell-mediated mechanisms for the pathogenesis of EoE (Figure 1).

T cell responses in allergic contact dermatitis

In allergic contact dermatitis (ACD), chemical allergens penetrate into the skin where they form complexes or bind covalently to proteins of immune and structural cells in the skin and, thus, may induce innate immune responses as well as generate T cell epitopes (32). Contact allergens, e.g. nickel, are recognized by pattern recognition receptors (PRR) resulting in the production of pro-inflammatory cytokines such as IL-1 and IL-18. This irritant effect of contact allergens is essential for the subsequent activation of the adaptive immune system leading to a Tc1/Th1 and Tc17/Th17 effector/memory T cell response (33). Contact hypersensitivity is dependent on T cell-mediated cytotoxicity via FAS/FASL and perforin pathways (34). In ACD, Th1/Th17 cells may amplify the cytotoxic cascade as they increase T cell–keratinocyte adhesiveness and promote ICAM-1–dependent non-antigen-specific keratinocyte killing by T lymphocytes (35). However, there is little evidence for an IL-17-mediated process in EoE (36).

T cells in drug hypersensitivity

While immediate allergic drug hypersensitivity reactions (DHR) are mediated by specific IgE bound to mast cells and basophils, delayed (non-immediate) allergic DHR are T cellmediated. Analogous to haptens, drugs are presented either covalently bound to peptides in the binding grove of MHC molecules on antigen-presenting cells or complexed to amino acids in MHC molecules and TCR (37). Recently, a concept for the pharmacological interaction of drugs with immune receptor (p-i concept) has been proposed, suggesting a non-covalent binding enabling a direct interaction with immunological receptors such as MHC and TCR (38,39). Thus, the antigen might bind either to the MHC complex, thereby modifying the structure that is recognized by the TCR leading to a specific T cell activation or directly to a specific TCR requiring additional MHC interaction for full T cell activation (38). In cutaneous reactions, drug-specific cytotoxic T cells have been demonstrated that can contribute to tissue damage via perforin/granzyme B or FAS/FASL mechanisms (40,41). In DRESS, an oligoclonal expansion of activated CD8+ T cells directed against viral antigens derived from *Herpes* viruses, whose replication is enhanced by the culprit drug, has been observed in the skin and visceral organs (42).

Food-specific T cell responses in the skin

Over 80% of patients with atopic dermatitis (AD) have increased IgEs to foods and inhalant allergens in the peripheral blood (43). However, the positive predictive value of IgE specific to food allergens is low (44). Interestingly, in 45% of patients reacting upon food allergen challenge, eczematous reactions with or without prior immediate reactions have been observed, suggesting the occurrence of late, most likely T cell-mediated reactions against foods (44). Indeed, in patients with food-triggered AD exacerbations, relevant food allergen specific T cells have been detected in the peripheral blood as well as the skin (45,46). Moreover, positive atopy patch test (APT) reactions to inhalant and food allergens can be detected in the absence of corresponding IgE-responses (47). Although widely used, the APT has limited value in the diagnosis of food allergy in EoE (16) perhaps owing to the fact that here the skin and not the esophagus is tested. Upon food allergen, but not nonspecific stimulation, peripheral blood mononuclear cells from EoE patients with or without allergen-specific IgE produce significant amounts of IL-5 (48). In peanut-allergic children, skin- and gut-homing T cells expressing Th2 and Th9 genes as well as IL-9 and IL-5 production by distinct T helper cell populations have been reported (49).

To date, the presence of food allergen-specific T cells in EoE has not been demonstrated. Furthermore, it remains uncertain when and where the sensitization to food allergens occurs. In adult EoE patients, airway allergy precedes EoE (50). Recent data suggest that an epicutaneous sensitization with ovalbumin may result in an antigen-induced gastrointestinal food allergy via the TSLP-basophil axis or in an IL-17-mediated response depending on the animal model (51,52). Furthermore, filaggrin mutations as risk factors for eczema, the atopic march and peanut allergy have been reported,

indicating that an impaired epithelial barrier function may predispose to allergen sensitization and atopy (53,54).

Epithelial barrier and innate immune responses in EoE

There is increasing evidence that EoE is associated with a dysfunction at the epithelial barrier followed by an eosinophilic inflammation similar to AD which is concomitant in over half of EoE patients (Figure 1). In esophageal epithelial cells, the expression of epidermal differentiation complex (EDC) genes, e.g. filaggrin, SPRR3 and keratins, is downregulated in response to IL-13 and in active EoE, where it could be only partially normalized upon therapy (55,56). Desmoglein (DSG)-1, an intercellular adhesion molecule responsible for epithelial integrity and barrier function was one of the most strongly downregulated genes in EoE (29). A downregulation of DSG-1 gene, e.g. by IL-13, was shown to result in the separation of epithelial cells (spongiosis) followed by impaired barrier function as well as by periostin induction further potentiating inflammation (57). Ultrastructural analysis revealed a significantly decreased number of desmosomes per cell in EoE biopsies as compared to healthy controls, which was reversible after treatment (58). Furthermore, the expression of filaggrin and the tight junction proteins zonula occludens (ZO)-3 and claudin-1 is decreased in EoE, correlating with spongiosis (59). Consistent with this finding, mutations in filaggrin are over-represented in EoE patients (55) and homozygous mutations of DSG1 cause a severe atopy syndrome which includes EoE (60).

In stratified epithelia, the activity of proteases is tightly regulated by protease inhibitors. The loss of inhibition results in cleavage of desmosomal proteins and loss of barrier integrity, facilitating the penetration of allergens and microbes as well as the subsequent generation of danger signals and protease activated receptor (PAR)-2 activation (61). In active EoE, a significantly decreased expression of the protease inhibitor LEKTI has been observed (36).

TSLP that is produced by epithelial cells in response to PAR-2, toll-like receptor (TLR) stimulation or mechanical injury, strongly induces Th2 immune responses by stimulating dendritic cells, T cells, eosinophils, mast cells and basophils (62). Upon stimulation with TSLP, eosinophils that bear the TSLP receptor on their surface generate

extracellular DNA traps associated with granule proteins that are able to kill bacteria (63). Interestingly, the expression of TSLP is increased in EoE and correlates with the number of eosinophils generating eosinophil extracellular traps (36). Genetic variants of TSLP and its receptor have been associated with an increased susceptibility to EoE overall, and in males, respectively (64,65). Furthermore, the gene of esophageal selective calpain (CAPN) 14, a member of the calpain protease family involved in the cleavage of inflammatory mediators such as IL-33, was upregulated in active EoE, while the calpain inhibitor CAST was downregulated (66). In line with these findings, genetic variants in the CAPN 14 gene locus are linked with EoE susceptibility (67) and increased expression of innate cytokines including IL-33 by epithelial cells has been detected in EoE (36).

Immense efforts have been undertaken to identify the role of the microbiota in the immune system, in particular in association with immune-mediated diseases. Microbiota research aims at elucidating their role in initiating and perpetuating inflammation and, conversely, the effect of diseases and treatment procedures on the microbiota. Compared to healthy controls, the bacterial load of the esophagus is increased in EoE patients regardless of treatment and disease activity, with a relative abundance of gram-negative bacteria in active EoE (68,69). Recently, IgE-sensitization to *Candida albicans* has been reported in pediatric and adult EoE patients (14,70). Whether an esophageal colonization with *Candida albicans* and later sensitization is owing to EoE inflammation or corticosteroid therapy remains to be investigated. Furthermore, any potential role of IgE specific for *Candida albicans* in the pathogenesis of EoE is uncertain.

Taken together, recent research suggests that impaired epithelial barrier function plays a major role in initiating and perpetuating EoE inflammation as it facilitates the penetration of allergens and microbes and generates danger signals leading to an activation of epithelial cells as well as innate and adaptive immune cells with subsequent chemokine and cytokine production resulting in Th2 immune responses. There is evidence of a dysbiosis of microbiota in EoE, however, the consequences in terms of microbial-triggered eosinophilic inflammation and the particular role of diet on the microbiome in the esophagus remain to be investigated.

Similarities and differences between EoE and IBD

With inflammatory bowel diseases (IBD), such as Crohn's disease (CD) and ulcerative colitis (UC), the pathogenesis is determined by genetic factors, environmental and microbial factors together with an epithelial barrier dysfunction and subsequent innate and adaptive immune responses (71). The susceptibility to IBD is determined by genetic variants related to innate immunity, autophagy and phagocytosis in CD and to barrier function in UC (72). Due to an increased intestinal epithelial permeability, food antigens and microbes may activate pattern recognition receptors on epithelial cells resulting in a release of pro-inflammatory cytokines such as TNF- α , IL-1, IL-18 and IL-33 (Figure 1) (71). In contrast to EoE, predominantly Th1 cells and the IL-23/Th17 axis are activated in IBD (71). It has been hypothesized that due to a dysregulated innate intestinal immunity and barrier function, affecting both the diversity and composition of the microbiota, the immune response is initiated to eliminate invading antigens (e.g. microbes, food) and to restore epithelial barrier integrity, but may later turn into a chronic inflammation leading to the clinical manifestations of IBD (71,73).

Thus, the principal pathomechanisms of IBD seem congruent with those of EoE, although it is currently not clear which tissue-specific characteristics, including immune responses, environmental factors such as microbiota and food, as well as genetic predispositions favor a chronic Th1/Th17 inflammation as in IBD or a Th2 predominant inflammation as in EoE with corresponding clinical phenotypes. While both diseases have common mechanisms, the upstream events are likely to be different as EoE is associated with unique genetic susceptibility (TSLP and CAPN14) and atopy; whereas IBD is more related to innate immunity to microbial flora.

EoE is distinct from IgE-mediated food allergies

If one were to consider EoE as a kind of food allergy, how would its symptoms agree with the current concept of gastrointestinal (GI) allergies? A food allergy is defined as an abnormal immunologic response to a food substance occurring in a susceptible host and causing some type of GI inflammation. The vast majority of food allergies affecting the GI tract are characterized by a Th2 inflammation with predominant Th2 cytokine expression (that is IL-4, IL-13, and IL-5). Th2 inflammation can cause B cells to produce IgE antibodies specific to certain foods or can lead to a chronic cellular inflammation frequently characterized by the presence of Th2 cell and eosinophils (74).

According to the immunological mechanism elicited, food allergies can be classified into: (a) IgE-mediated, which are immediate, short-lived reactions mediated by antibodies belonging to the IgE class, (b) cell-mediated, which usually have a delayed/chronic course, typically involving the GI tract and the cell component of the immune system responsible for inflammation, or (c) mixed, IgE- and cell-mediated (75). IgE-mediated reactions to foods are acute and highly reproducible. They are initiated by the cross linking of two or more allergen-specific IgE antibodies bound to their high-affinity receptor (FccRI) expressed on mast cells and basophils as a result of a specific food allergen engagement. Such cross-linking determines the release of preformed mediators, in particular, histamine, that cause vasodilatation, angioedema, smooth muscle constriction, and increased mucus production (76).

Examples of typical IgE-mediated allergic reactions affecting the GI tract are the oral allergy syndrome (OAS) and the more severe GI food allergy, also known as "gastrointestinal anaphylaxis". When comparing IgE-mediated OAS and GI food allergies with EoE, the following differences become evident: EoE symptoms might be instant, but they are not transient, EoE inflammation is chronic, anaphylaxis is not a feature of EoE, and pollen-associated food allergens are not a typical trigger of EoE. It should be noted, however, that EoE patients can concurrently suffer from OAS and/or a GI food allergy.

Food protein-induced enterocolitis (FPIES), an increasingly recognized form of non-IgE mediated food hypersensitivity, is characterized by a delayed onset of vomiting with or without diarrhea, typically occurring in infants and toddlers from 2 to 6 h post-ingestion of the trigger food (77,78). FPIES is usually a transient disease which starts at 4 to 9 months of life or when solid foods are first introduced, and resolves by age 2 to 5 years (77). The foods most commonly involved in FPIES are milk, soy, rice, oats and eggs. IgEs specific to the trigger foods are usually not detectable (77,79). Although FPIES and EoE seem to share some clinical (symptoms, age of onset) and pathogenic (causative food triggers, increased TNF- α , epithelial barrier defects) features (80), other characteristics such as disease course, endoscopic and histologic findings discriminate FPIES from EoE.

Experience with omalizumab: Its lack of clinical efficacy in EoE

Omalizumab is an anti-IgE humanized monoclonal antibody that binds to the fragment crystallizable (Fc) region of the IgE molecule and thus prevents its binding to the highaffinity IgE receptor (Fc epsilon RI, FccRI). In the only published prospective, randomized, double-blind, placebo-controlled study in 30 adult EoE patients (16 treated with omalizumab and 14 with placebo) omalizumab was given every 2-4 weeks for 16 weeks, based on weight and serum level of IgE. Before starting the treatment and at the end of the trial (16 weeks of treatment) symptoms evaluation, EGD and histological assessment of the eosinophil density (peak eos/hpf) in esophageal biopsies were performed. Patients treated with omalizumab had neither a significant improvement in symptoms nor a decrease of the eosinophil infiltration of the esophageal mucosa compared with placebo (21). This study confirmed anecdotal data from clinical cases reported in which omalizumab had been considered to improve IgE-mediated symptoms of food allergy, but not of EoE (81). Overall these data support the notion that EoE is not IgE-mediated. Clayton et al. speculated that IgG4 antibodies specific for a food allergen are blocking IgE responses (21). Indeed, in allergic diseases, an IgG4 response follows an IgE-mediated response and does block IgE-mediated mast cell activation (21). In EoE, extracellular granular deposits of IgG4 and abundant IgG4-containing plasma cells in the tissue, as well as increased serum levels of IgG4 reactive with specific foods have been observed, suggesting that in adults, EoE might be an IgG4- and IgE-associated disease and perhaps the balance between the two antibodies could be a key determinant (21). However, B cell-deficient mice also develop typical EoE, suggesting that antibodies may simply be non-pathogenic (82). Moreover, the anti-food IgG4 levels did not correlate with the age and duration of disease symptoms (21). Further studies will be necessary to really understand the pathogenic role of IgG4 in EoE.

EoE is characterized by a non-IgE-mediated food hypersensitivity

Since the first description of a series of clinical cases of EoE, food allergies have appeared to play a major role in causing a severe esophageal eosinophilia that resolved

on elemental diet, but not on aggressive GERD treatment, including Nissen fundoplication (6). In view of this, food allergens have been identified as triggers of EoE in most children and adults (6,16,82,83).

Thus, food as a trigger of EoE fulfills Koch's postulates since addition or subtraction of foods can cause disease or eliminate disease in EoE in nearly all patients. The most effective treatment in patients with EoE is an elemental diet that induces histological and clinical resolution in over 95% of pediatric and adult patients (83-86). Noteworthy is that IBD may also resolve upon elemental diet (87) with a mechanism that involves both bowel rest and a change in microbiome. So far, the explanation for remission of EoE on an elemental diet has always been linked to the avoidance of food allergens, rather than bowel rest/change in microbiome, but this possibility needs to be investigated further. This presumption was supported by the fact that elimination diets based on removal of the six most common food allergens (SFED-six food elimination diet) (82) or of the foods to which patients were sensitized (targeted elimination diets) have been shown to induce and maintain EoE remission in 72% and 45% of EoE patients. respectively (16,88). According to biopsy confirmation, the most common food proteins causing EoE are milk, followed by wheat, eggs, beef, soy and legumes, and chicken (16,83,89,90). Interestingly, peanuts, tree nuts, fish and shellfish are rare as causes for EoE despite being common causes of IgE-mediated reactions in adults.

The evidence that EoE is generally non-IgE-mediated is based on both clinical and research findings:

(1) Despite the fact that the majority of patients with EoE have specific IgEs to food allergens and/or aeroallergens, the detection of specific IgEs for food allergens, either by SPT or specific sera IgE (sIgE), has not proven successful for the identification of causative foods in EoE (84,85). Indeed, removal of SPT or sIgE positive foods is not superior to SFED (2,16,17,83,91). Moreover, it has been reported that the introduction of skin test negative foods into the diet sometimes induces clinical disease (6,16).

(2) Clinical trials and case series have shown that therapy with omalizumab is not effective in inducing remission of EoE (21,81).

(3) Oral immunotherapy, which has been used successfully in IgE-mediated food allergy, is associated with an increased risk of developing EoE (e.g. in 2 to 10% of treated patients) (92-94).

(4) Children who outgrow IgE-mediated food allergy and therefore are able to reintroduce these foods in their diet, can later develop EoE to the same food (95).

(5) In experimental models in which food allergens are able to induce an EoE-like disease, mice with depleted IgE and devoid of mast cells still could develop esophageal inflammation and consequent food impaction similar to the wild-type mice (96,97).

Confirmation of EoE diagnosis and the practical search for offending foods

EoE is a clinico-pathological diagnosis. However, EoE and GERD have a substantial overlap of clinical and of histological features. For instance, the presence of heartburn and marked esophageal eosinophilia might be fairly common in both entities (2). In order to solve this diagnostic conundrum, updated consensus recommendations for diagnosis and management of EoE advocate performing a PPI trial in patients having symptoms suggestive of EoE and esophageal eosinophilia (2). Accordingly, a diagnosis of GERD was recommended for those patients responding to PPI therapy, whereas patients whose symptoms and inflammation persist were regarded as having EoE (2). Unfortunately, this diagnostic PPI trial did not fulfil the expectation of differentiating EoE from GERD, but unexpectedly uncovered a third category of patients, called PPI-REE, presenting with symptoms of EoE, but responding to PPI (5). With the exception of the responsiveness to PPI, PPI-REE, and EoE have common clinical, endoscopic, histological and molecular features.

EoE is a chronic and progressive disease. If left untreated complications, such as food impaction, esophageal stricture, narrow-caliber esophagus, and esophageal perforation are common (98,99). Therefore, once the diagnosis is confirmed, it is important to treat the eosinophilic inflammation not only to control the presenting symptoms, but also to preserve the morphological and functional integrity of the esophagus (2,10,87,99). Beside medications, diets avoiding culprit foods are an important therapeutic option (100). Of note, before an elimination diet can be established, it is necessary to identify the triggering foods, ideally with the help of a dietitian specialized in

15

dealing with this disease. Currently culprit foods are identified by demonstrating histological and clinical remission of EoE after the establishment of an elimination diet. In practice, after avoidance and after re-introduction of any food category, the effect must be controlled endoscopically and histologically (82,83). Serial endoscopies are therefore required to figure out an individual elimination diet. This approach is time consuming, inconvenient for patients, expensive, and affects the quality of life (101). Therefore, there is a need to develop non-invasive methods for the identification of the offending foods. The determination of food-specific IgG4 in the serum is a method currently under evaluation. Further phenotyping patients based on their esophageal gene expression, using a 94 gene transcript profile, is promising to be helpful (102).

Conclusions

There is strong evidence that foods, most likely food-proteins, are triggers of EoE, since elimination of culprit food categories as well as protein-free elemental diets result in an improvement of histological and endoscopic signs as well as of symptoms. Furthermore, the observation that the eosinophilic inflammation and the Th2 inflammation pattern reappear rapidly after re-introduction of the culprit foods are strong arguments that EoE is likely a food-driven disorder with features of food allergy. However, the spectrum of clinical presentations of EoE, the results of IgE-based diagnostic procedures, as well as the lack of efficacy of anti-IgE treatment suggests that EoE cannot be regarded as an IgE-mediated food allergy.

The mechanism by which food elicits EoE is not yet understood. It seems likely that a cellular mechanism similar to contact allergy of the skin or drug hypersensitivity plays a role. IgG4 formed against foods has been suspected of playing a role in EoE, perhaps as a blocking antibody analogous to AD and IBD, an impairment of the epithelial barrier, alterations of the microbiota and subsequent chronic inflammation might be the underlying pathogenic factors for EoE. Given this scenario, food might interfere either as an irritant, modulator of the microbiota or as an antigen/allergen to initiate and perpetuate inflammation (Figure 1). The identification of offending foods by empirical elimination diets and controlled re-introduction of foods is inconvenient for patients, time-consuming, and in the clinical routine hardly applicable. Nevertheless, this procedure is currently the only

reliable method to identify food triggers in EoE patients. Elucidating the exact mechanism of how foods affect EoE would allow the development of novel diagnostic tests. For instance, the determination of food-specific markers including T cell responses to specific foods could possibly overcome the limitations of SPT, APT, and empirical diets. As in IBD and atopic diseases, EoE should be considered as a complex disease with a disordered interplay between the epithelial barrier, innate and adaptive immune responses together with the composition of the microbiota.

References

- Furuta GT, Liacouras CA, Collins MH, Gupta SK, Justinich C, Putnam PE et al. First International Gastrointestinal Eosinophil Research Symposium (FIGERS) Subcommittees. Eosinophilic esophagitis in children and adults: a systematic review and consensus recommendations for diagnosis and treatment. *Gastroenterology* 2007;**133**:1342-1363.
- Liacouras CA, Furuta GT, Hirano I, Atkins D, Attwood SE, Bonis PA, Burks AW, Chehade M, Collins MH, Dellon ES, Dohil R, Falk GW, Gonsalves N, Gupta SK, Katzka DA, Lucendo AJ, Markowitz JE, Noel RJ, Odze RD, Putnam PE, Richter JE, Romero Y, Ruchelli E, Sampson HA, Schoepfer A, Shaheen NJ, Sicherer SH, Spechler S, Spergel JM, Straumann A, Wershil BK, Rothenberg ME, Aceves SS. Eosinophilic esophagitis: updated consensus recommendations for children and adults. *J Allergy Clin Immunol* 2011;**128**:3-20.
- Furuta GT, Katzka DA. Eosinophilic esophagitis. N Engl J Med 2015;373:1640-1648.
- Wen T, Dellon ES, Moawad FJ, Furuta GT, Aceves SS, Rothenberg ME. Transcriptome analysis of proton pump inhibitor-responsive esophageal eosinophilia reveals proton pump inhibitor-reversible allergic inflammation. J Allergy Clin Immunol 2015;135:187-197.
- Molina-Infante J, Bredenoord AJ, Cheng E, Dellon ES, Furuta GT, Gupta SK, Hirano I, Katzka DA, Moawad FJ, Rothenberg ME, Schoepfer A, Spechler S, Wen T, Straumann A, Lucendo AJ. Proton pump inhibitor-responsive oesophageal eosinophilia: an entity challenging current diagnostic criteria for eosinophilic oesophagitis. *Gut* 2016, doi: 10.1136/gutjnl-2015-310991 (E-pub ahead of print).
- Eluri S, Dellon ES. Proton pump inhibitor-responsive oesophageal eosinophilia and eosinophilic oesophagitis: more similarities than differences. *Curr Opin Gastroenterol* 2015;**31**:309-315.
- 7. Attwood SE, Smyrk TC, Demeester TR, Jones JB. Esophageal eosinophilia with dysphagia, a distinct clinicopathologic syndrome. *Dig. Dis. Sci.* 1993;**38:**109-116.

- 8. Straumann A, Spichtin HP, Bernoulli R, Loosli J, Vögtlin J. Idiopathic eosinophilic esophagitis: a frequently overlooked disease with typical clinical aspects and discrete endoscopic findings. *Schweiz Med Wochenschr* 1994;**124**:1419-1429.
- Kelly KJ, Lazenby AJ, Rowe PC, Yardley JH, Perman JA, Sampson HA.
 Eosinophilic esophagitis attributed to gastroesophageal reflux: improvement with an amino acid-based formula. *Gastroenterology* 1995;**109**:1503-1512.
- Straumann A, Aceves SS, Blanchard C, Collins MH, Furuta GT, Hirano I, Schoepfer AM, Simon D, Simon HU. Pediatric and adult eosinophilic esophagitis: similarities and differences. *Allergy* 2012;67:477-490.
- Assa'ad AH, Putnam PE, Collins MH, Akers RM, Jameson SC, Kirby CL, Buckmeier BK, Bullock JZ, Collier AR, Konikoff MR, Noel RJ, Guajardo JR, Rothenberg ME. Pediatric patients with eosinophilic esophagitis: an 8-year follow-up. *J Allergy Clin Immunol* 2007;**119**:731-738.
- Sugnanam KK, Collins JT, Smith PK, Connor F, Lewindon P, Cleghorn G, Withers G. Dichotomy of food and inhalant allergen sensitization in eosinophilic esophagitis. *Allergy* 2007;**62**:1257-1260.
- Spergel JM, Beausoleil JL, Mascarenhas M, Liacouras CA. The use of skin prick tests and patch tests to identify causative foods in eosinophilic esophagitis. J Allergy Clin Immunol 2002;109:363-368.
- Simon D, Straumann A, Dahinden C, Simon HU. Frequent sensitization to Candida albicans and profilins in adult eosinophilic esophagitis. *Allergy* 2013;68:945-948.
- Vicario M, Blanchard C, Stringer KF, Collins MH, Mingler MK, Ahrens A, Putnam PE, Abonia JP, Santos J, Rothenberg ME. Local B cells and IgE production in the oesophageal mucosa in eosinophilic oesophagitis. *Gut* 2010;**59**:12-20.
- Spergel JM, Brown-Whitehorn TF, Cianferoni A, Shuker M, Wang ML, Verma R, Liacouras CA. Identification of causative foods in children with eosinophilic esophagitis treated with an elimination diet. *J Allergy Clin Immunol* 2012;**130**:461-467.

- Henderson CJ, Abonia JP, King EC, Putnam PE, Collins MH, Franciosi JP, Rothenberg ME. Comparative dietary therapy effectiveness in remission of pediatric eosinophilic esophagitis. *J Allergy Clin Immunol* 2012;**129**:1570-1578.
- Simon D, Straumann A, Wenk A, Spichtin H, Simon HU, Braathen LR.
 Eosinophilic esophagitis in adults--no clinical relevance of wheat and rye sensitizations. *Allergy* 2006;61:1480-1483.
- Spergel JM, Andrews T, Brown-Whitehorn TF, Beausoleil JL, Liacouras CA. Treatment of eosinophilic esophagitis with specific food elimination diet directed by a combination of skin prick and patch tests. *Ann Allergy Asthma Immunol* 2005;**95**:336-343.
- Loizou D, Enav B, Komlodi-Pasztor E, Hider P, Kim-Chang J, Noonan L, Taber T, Kaushal S, Limgala R, Brown M, Gupta R, Balba N, Goker-Alpan O, Khojah A, Alpan O. A pilot study of omalizumab in eosinophilic esophagitis. *PLoS One* 2015;**10**:e0113483.
- Clayton F, Fang JC, Gleich GJ, Lucendo AJ, Olalla JM, Vinson LA, Lowichik A, Chen X, Emerson L, Cox K, O'Gorman MA, Peterson K. Eosinophilic esophagitis in adults is associated with IgG4 and not mediated by IgE. *Gastroenterology* 2014;**147**: 602-609.
- Straumann A, Bauer M, Fischer B, Blaser K, Simon HU. Idiopathic eosinophilic esophagitis is associated with a T(H)2-type allergic inflammatory response. J Allergy Clin Immunol 2001;108:954-961.
- Blanchard C, Wang N, Stringer KF, Mishra A, Fulkerson PC, Abonia JP, Jameson SC, Kirby C, Konikoff MR, Collins MH, Cohen MB, Akers R, Hogan SP, Assa'ad AH, Putnam PE, Aronow BJ, Rothenberg ME. Eotaxin-3 and a uniquely conserved gene-expression profile in eosinophilic esophagitis. *J Clin Invest* 2006;**116**:536-547.
- 24. Blanchard C, Stucke EM, Rodriguez-Jimenez B, Burwinkel K, Collins MH, Ahrens A, Alexander ES, Butz BK, Jameson SC, Kaul A, Franciosi JP, Kushner JP, Putnam PE, Abonia JP, Rothenberg ME. A striking local esophageal cytokine expression profile in eosinophilic esophagitis. *J Allergy Clin Immunol* 2011;**127**:208-217.

- Mishra A, Hogan SP, Brandt EB, Rothenberg ME. An etiological role for aeroallergens and eosinophils in experimental esophagitis. *J Clin Invest* 2001;**107**:83-90.
- Mishra A, Rothenberg ME. Intratracheal IL-13 induces eosinophilic esophagitis by an IL-5, eotaxin-1, and STAT6-dependent mechanism. *Gastroenterology* 2003;**125**:1419-1427.
- Kohan M, Puxeddu I, Reich R, Levi-Schaffer F, Berkman N. Eotaxin-2/CCL24 and eotaxin-3/CCL26 exert differential profibrogenic effects on human lung fibroblasts. *Ann Allergy Asthma Immunol* 2010;**104**:66-72.
- Zuo L, Fulkerson PC, Finkelman FD, Mingler M, Fischetti CA, Blanchard C, Rothenberg ME. IL-13 induces esophageal remodeling and gene expression by an eosinophil-independent, IL-13R alpha 2-inhibited pathway. *J Immunol* 2010;**185**:660-669.
- 29. Blanchard C, Mingler MK, Vicario M, Abonia JP, Wu YY, Lu TX, Collins MH, Putnam PE, Wells SI, Rothenberg ME. IL-13 involvement in eosinophilic esophagitis: transcriptome analysis and reversibility with glucocorticoids. *J Allergy Clin Immunol* 2007;**120**:1292-300.
- Straumann A1, Conus S, Grzonka P, Kita H, Kephart G, Bussmann C, Beglinger C, Smith DA, Patel J, Byrne M, Simon HU. Anti-interleukin-5 antibody treatment (mepolizumab) in active eosinophilic oesophagitis: a randomised, placebo-controlled, double-blind trial. *Gut* 2010;**59**:21-30.
- Rothenberg ME, Wen T, Greenberg A, Alpan O, Enav B, Hirano I, Nadeau K, Kaiser S, Peters T, Perez A, Jones I, Arm JP, Strieter RM, Sabo R, Gunawardena KA. Intravenous anti-IL-13 mAb QAX576 for the treatment of eosinophilic esophagitis. *J Allergy Clin Immunol* 2015;**135**:500-507
- 32. Martin SF. Allergic contact dermatitis: xenoinflammation of the skin. *Curr Opin Immunol* 2012;**24**:720-729.
- 33. Martin SF. New concepts in cutaneous allergy. *Contact Dermatitis* 2015;**72**:2-10.
- Kehren J, Desvignes C, Krasteva M, Ducluzeau MT, Assossou O, Horand F,
 Hahne M, Kägi D, Kaiserlian D, Nicolas JF. Cytotoxicity is mandatory for CD8(+)
 T cell-mediated contact hypersensitivity. *J Exp Med* 1999;**189**:779-786.

- 35. Pennino D, Eyerich K, Scarponi C, Carbone T, Eyerich S, Nasorri F, Garcovich S, Traidl-Hoffmann C, Albanesi C, Cavani A. IL-17 amplifies human contact hypersensitivity by licensing hapten nonspecific Th1 cells to kill autologous keratinocytes. *J Immunol* 2010;**184**:4880-4888.
- 36. Simon D, Radonjic-Hösli S, Straumann A, Yousefi S, Simon HU. Active eosinophilic esophagitis is characterized by epithelial barrier defects and eosinophil extracellular trap formation. *Allergy* 2015;**70**:443-452.
- 37. Martin SF, Esser PR, Schmucker S, Dietz L, Naisbitt DJ, Park BK, Vocanson M, Nicolas JF, Keller M, Pichler WJ, Peiser M, Luch A, Wanner R, Maggi E, Cavani A, Rustemeyer T, Richter A, Thierse HJ, Sallusto F. T-cell recognition of chemicals, protein allergens and drugs: towards the development of in vitro assays. *Cell Mol Life Sci* 2010;**67**:4171-4184.
- 38. Pichler WJ, Naisbitt DJ, Park BK. Immune pathomechanism of drug hypersensitivity reactions. *J Allergy Clin Immunol* 2011;**127**:74-81.
- Demoly P, Adkinson NF, Brockow K, Castells M, Chiriac AM, Greenberger PA, Khan DA, Lang DM, Park HS, Pichler W, Sanchez-Borges M, Shiohara T, Thong BY. International Consensus on drug allergy. *Allergy* 2014;69:420-437.
- Yawalkar N, Egli F, Hari Y, Nievergelt H, Braathen LR, Pichler WJ. Infiltration of cytotoxic T cells in drug-induced cutaneous eruptions. *Clin Exp Allergy* 2000;**30**:847-855.
- Schmid S, Kuechler PC, Britschgi M, Steiner UC, Yawalkar N, Limat A, Baltensperger K, Braathen L, Pichler WJ. Acute generalized exanthematous pustulosis: role of cytotoxic T cells in pustule formation. *Am J Pathol* 2002;**161**:2079-2086.
- 42. Picard D, Janela B, Descamps V, D'Incan M, Courville P, Jacquot S, Rogez S, Mardivirin L, Moins-Teisserenc H, Toubert A, Benichou J, Joly P, Musette P. Drug reaction with eosinophilia and systemic symptoms (DRESS): a multiorgan antiviral T cell response. *Sci Transl Med* 2010;**2**:46-62.
- 43. Akdis CA, Akdis M, Simon D, Dibbert B, Weber M, Gratzl S, Kreyden O, Disch R, Wüthrich B, Blaser K, Simon HU. T cells and T cell-derived cytokines as

pathogenic factors in the nonallergic form of atopic dermatitis. *J Invest Dermatol* 1999;**113**:628-634.

- Breuer K, Heratizadeh A, Wulf A, Baumann U, Constien A, Tetau D, Kapp A, Werfel T. Late eczematous reactions to food in children with atopic dermatitis. *Clin Exp Allergy* 2004;**34**:817-824.
- 45. Reekers R, Beyer K, Niggemann B, Wahn U, Freihorst J, Kapp A, Werfel T. The role of circulating food antigen-specific lymphocytes in food allergic children with atopic dermatitis. *Br J Dermatol* 1996;**135**:935-941.
- 46. Reekers R, Busche M, Wittmann M, Kapp A, Werfel T. Birch pollen-related foods trigger atopic dermatitis in patients with specific cutaneous T-cell responses to birch pollen antigens. *J Allergy Clin Immunol* 1999;**104**:466-472.
- 47. Darsow U, Laifaoui J, Kerschenlohr K, Wollenberg A, Przybilla B, Wüthrich B, Borelli S Jr, Giusti F, Seidenari S, Drzimalla K, Simon D, Disch R, Borelli S, Devillers AC, Oranje AP, De Raeve L, Hachem JP, Dangoisse C, Blondeel A, Song M, Breuer K, Wulf A, Werfel T, Roul S, Taieb A, Bolhaar S, Bruijnzeel-Koomen C, Brönnimann M, Braathen LR, Didierlaurent A, André C, Ring J. The prevalence of positive reactions in the atopy patch test with aeroallergens and food allergens in subjects with atopic eczema: a European multicenter study. *Allergy* 2004;**59**:1318-1325.
- 48. Yamazaki K, Murray JA, Arora AS, Alexander JA, Smyrk TC, Butterfield JH, Kita H. Allergen-specific in vitro cytokine production in adult patients with eosinophilic esophagitis. *Dig Dis Sci* 2006;**51**:1934-1941.
- Brough HA, Cousins DJ, Munteanu A, Wong YF, Sudra A, Makinson K, Stephens AC, Arno M, Ciortuz L, Lack G, Turcanu V. IL-9 is a key component of memory Th cell peanut-specific responses from children with peanut allergy. *J Allergy Clin Immunol* 2014;**134**:1329-1338.
- Simon D, Marti H, Heer P, Simon HU, Braathen LR, Straumann A. Eosinophilic esophagitis is frequently associated with IgE-mediated allergic airway diseases. J Allergy Clin Immunol 2005;115:1090-1092.
- 51. Noti M, Kim BS, Siracusa MC, Rak GD, Kubo M, Moghaddam AE, Sattentau QA, Comeau MR, Spergel JM, Artis D. Exposure to food allergens through inflamed

skin promotes intestinal food allergy through the thymic stromal lymphopoietinbasophil axis. *J Allergy Clin Immunol* 2014;**133**:1390-1399.

- 52. He R, Kim HY, Yoon J, Oyoshi MK, MacGinnitie A, Goya S, Freyschmidt EJ, Bryce P, McKenzie AN, Umetsu DT, Oettgen HC, Geha RS. Exaggerated IL-17 response to epicutaneous sensitization mediates airway inflammation in the absence of IL-4 and IL-13. *J Allergy Clin Immunol* 2009;124:761-770
- 53. Marenholz I, Nickel R, Rüschendorf F, Schulz F, Esparza-Gordillo J, Kerscher T, Grüber C, Lau S, Worm M, Keil T, Kurek M, Zaluga E, Wahn U, Lee YA. Filaggrin loss-of-function mutations predispose to phenotypes involved in the atopic march. J Allergy Clin Immunol 2006;**118**:866-871.
- 54. Brown SJ, Asai Y, Cordell HJ, Campbell LE, Zhao Y, Liao H, Northstone K, Henderson J, Alizadehfar R, Ben-Shoshan M, Morgan K, Roberts G, Masthoff LJ, Pasmans SG, van den Akker PC, Wijmenga C, Hourihane JO, Palmer CN, Lack G, Clarke A, Hull PR, Irvine AD, McLean WH. Loss-of-function variants in the filaggrin gene are a significant risk factor for peanut allergy. *J Allergy Clin Immunol* 2011;**127**:661-667.
- 55. Blanchard C, Stucke EM, Burwinkel K, Caldwell JM, Collins MH, Ahrens A, Buckmeier BK, Jameson SC, Greenberg A, Kaul A, Franciosi JP, Kushner JP, Martin LJ, Putnam PE, Abonia JP, Wells SI, Rothenberg ME. Coordinate interaction between IL-13 and epithelial differentiation cluster genes in eosinophilic esophagitis. *J Immunol* 2010;**184**:4033-4041.
- 56. Kiran KC, Rothenberg ME, Sherrill JD. In vitro model for studying esophageal epithelial differentiation and allergic inflammatory responses identifies keratin involvement in eosinophilic esophagitis. *PLoS One* 2015;**10**:e0127755.
- 57. Sherrill JD, Kc K, Wu D, Djukic Z, Caldwell JM, Stucke EM, Kemme KA, Costello MS, Mingler MK, Blanchard C, Collins MH, Abonia JP, Putnam PE, Dellon ES, Orlando RC, Hogan SP, Rothenberg ME. Desmoglein-1 regulates esophageal epithelial barrier function and immune responses in eosinophilic esophagitis. *Mucosal Immunol* 2014;**7**:718-729.

- Capocelli KE, Fernando SD, Menard-Katcher C, Furuta GT, Masterson JC, Wartchow EP. Ultrastructural features of eosinophilic oesophagitis: impact of treatment on desmosomes. *J Clin Pathol* 2015;68:51-56.
- 59. Katzka DA, Tadi R, Smyrk TC, Katarya E, Sharma A, Geno DM, Camilleri M, Iyer PG, Alexander JA, Buttar NS. Effects of topical steroids on tight junction proteins and spongiosis in esophageal epithelia of patients with eosinophilic esophagitis. *Clin Gastroenterol Hepatol* 2014;**12**:1824-1829.
- Samuelov L, Sarig O, Harmon RM, Rapaport D, Ishida-Yamamoto A, Isakov O, Koetsier JL, Gat A, Goldberg I, Bergman R, Spiegel R, Eytan O, Geller S, Peleg S, Shomron N, Goh CS, Wilson NJ, Smith FJ, Pohler E, Simpson MA, McLean WH, Irvine AD, Horowitz M, McGrath JA, Green KJ, Sprecher E. Desmoglein 1 deficiency results in severe dermatitis, multiple allergies and metabolic wasting. *Nat Genet* 2013;45:1244-1248.
- 61. Hovnanian A. Netherton syndrome: skin inflammation and allergy by loss of protease inhibition. *Cell Tissue Res* 2013;**351**:289-300.
- Roan F, Bell BD, Stoklasek TA, Kitajima M, Han H, Ziegler SF. The multiple facets of thymic stromal lymphopoietin (TSLP) during allergic inflammation and beyond. *J Leukoc Biol* 2012;**91**:877-886.
- Morshed M, Yousefi S, Stöckle C, Simon HU, Simon D. Thymic stromal lymphopoietin stimulates the formation of eosinophil extracellular traps. *Allergy* 2012;67:1127-1137.
- 64. Rothenberg ME, Spergel JM, Sherrill JD, Annaiah K, Martin LJ, Cianferoni A, Gober L, Kim C, Glessner J, Frackelton E, Thomas K, Blanchard C, Liacouras C, Verma R, Aceves S, Collins MH, Brown-Whitehorn T, Putnam PE, Franciosi JP, Chiavacci RM, Grant SF, Abonia JP, Sleiman PM, Hakonarson H. Common variants at 5q22 associate with pediatric eosinophilic esophagitis. *Nat Genet* 2010;**42**:289-291.
- Sherrill JD, Gao PS, Stucke EM, Blanchard C, Collins MH, Putnam PE, Franciosi JP, Kushner JP, Abonia JP, Assa'ad AH, Kovacic MB, Biagini Myers JM, Bochner BS, He H, Hershey GK, Martin LJ, Rothenberg ME. Variants of thymic

stromal lymphopoietin and its receptor associate with eosinophilic esophagitis. *J Allergy Clin Immunol* 2010;**126**:160-165.

- 66. Kottyan LC, Davis BP, Sherrill JD, Liu K, Rochman M, Kaufman K, Weirauch MT, Vaughn S, Lazaro S, Rupert AM, Kohram M, Stucke EM, Kemme KA, Magnusen A, He H, Dexheimer P, Chehade M, Wood RA, Pesek RD, Vickery BP, Fleischer DM, Lindbad R, Sampson HA, Mukkada VA, Putnam PE, Abonia JP, Martin LJ, Harley JB, Rothenberg ME. Genome-wide association analysis of eosinophilic esophagitis provides insight into the tissue specificity of this allergic disease. *Nat Genet* 2014;**46**:895-900.
- Sleiman PM, Wang ML, Cianferoni A, Aceves S, Gonsalves N, Nadeau K, Bredenoord AJ, Furuta GT, Spergel JM, Hakonarson H. GWAS identifies four novel eosinophilic esophagitis loci. *Nat Commun* 2014;**5**:5593.
- Harris JK, Fang R, Wagner BD, Choe HN, Kelly CJ, Schroeder S, Moore W, Stevens MJ, Yeckes A, Amsden K, Kagalwalla AF, Zalewski A, Hirano I, Gonsalves N, Henry LN, Masterson JC, Robertson CE, Leung DY, Pace NR, Ackerman SJ, Furuta GT, Fillon SA. Esophageal microbiome in eosinophilic esophagitis. *PLoS One* 2015;**10**:e0128346.
- Benitez AJ, Hoffmann C, Muir AB, Dods KK, Spergel JM, Bushman FD, Wang ML. Inflammation-associated microbiota in pediatric eosinophilic esophagitis. *Microbiome* 2015;3:23.
- 70. Erwin EA, James HR, Gutekunst HM, Russo JM, Kelleher KJ, Platts-Mills TA. Serum IgE measurement and detection of food allergy in pediatric patients with eosinophilic esophagitis. Ann Allergy Asthma Immunol. 2010 Jun;104(6):496-502
- 71. Corridoni D, Arseneau KO, Cominelli F. Inflammatory bowel disease. *Immunol Lett* 2014;**161**:231-235.
- 72. Xavier RJ, Podolsky DK. Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 2007;**448**:427-434.
- 73. Huttenhower C, Kostic AD, Xavier RJ. Inflammatory bowel disease as a model for translating the microbiome. *Immunity* 2014;**40**:843-854.
- 74. Cianferoni A, Spergel JM. Food allergy: review, classification and diagnosis. *Allergol Int* 2009;**58**:457-466.

- 75. Nowak-Wegrzyn A, Sampson HA. Adverse reactions to foods. *Med Clin North Am* 2006;**90**:97-127.
- 76. Simons FE. Anaphylaxis. *J Allergy Clin Immunol* 2010;**125**:161-181.
- 77. Nowak-Wegrzyn A, Muraro A. Food protein-induced enterocolitis syndrome. *Curr Opin Allergy Clin Immunol* 2009;**9**:371-377.
- 78. Venter C, Groetch M. Nutritional management of food protein-induced enterocolitis syndrome. *Curr Opin Allergy Clin Immunol* 2014;**14**:255-262.
- Ruffner MA, Ruymann K, Barni S, Cianferoni A, Brown-Whitehorn T, Spergel JM (2013). Food protein-induced enterocolitis syndrome: insights from review of a large referral population. *J Allergy Clin Immunol Pract* 2013;1:343-349.
- 80. Berin MC. Immunopathophysiology of food protein-induced enterocolitis syndrome. *J. Allergy Clin Immunol* 2015;**135**:1108-1113.
- 81. Rocha R, Vitor AB, Trindade E, Lima R, Tavares M, Lopes J, Dias JA.
 Omalizumab in the treatment of eosinophilic esophagitis and food allergy. *Eur J Pediatr* 2011;**170**:1471-1474.
- Kagalwalla AF, Sentongo TA, Ritz S, Hess T, Nelson SP, Emerick KM, Melin-Aldana H, Li BU. Effect of six-food elimination diet on clinical and histologic outcomes in eosinophilic esophagitis. *Clin Gastroenterol Hepatol* 2006;**4**:1097-1102.
- Gonsalves N, Yang GY, Doerfler B, Ritz S, Ditto AM, Hirano I. Elimination diet effectively treats eosinophilic esophagitis in adults; food reintroduction identifies causative factors. *Gastroenterology* 2012;**142**:1451-1459.
- Liacouras CA, Spergel JM, Ruchelli E, Verma R, Mascarenhas M, Semeao E, Flick J, Kelly J, Brown-Whitehorn T, Mamula P, Markowitz JE. Eosinophilic esophagitis: a 10-year experience in 381 children. *Clin Gastroenterol Hepatol* 2005;**3**:1198-1206.
- Spergel JM, Brown-Whitehorn TF, Beausoleil JL, Franciosi J, Shuker M, VermaR, Liacouras CA. 14 years of eosinophilic esophagitis: clinical features and prognosis. *J Pediatr Gastroenterol Nutr* 2009;48:30-36.
- 86. Arias A, Gonzalez-Cervera J, Tenias JM, Lucendo AJ. Efficacy of dietary interventions for inducing histologic remission in patients with eosinophilic

esophagitis: a systematic review and meta-analysis. *Gastroenterology* 2014;**146**:1639-1648.

- Lee D, Albenberg L, Compher C, Baldassano R, Piccoli D, Lewis JD, Wu GD.
 Diet in the pathogenesis and treatment of inflammatory bowel diseases.
 Gastroenterology 2015;**148**:1087-1106.
- Spergel JM, Beausoleil JL, Mascarenhas M, Liacouras CA (2002). The use of skin prick tests and patch tests to identify causative foods in eosinophilic esophagitis. *J Allergy Clin Immunol* 2002;**109**:363-368.
- Kagalwalla AF, Shah A, Li BU, Sentongo TA, Ritz S, Manuel-Rubio M, Jacques K, Wang D, Melin-Aldana H, Nelson SP. Identification of specific foods responsible for inflammation in children with eosinophilic esophagitis successfully treated with empiric elimination diet. *J Pediatr Gastroenterol Nutr* 2011;**53**:145-149.
- 90. Molina-Infante J, Arias A, Barrio J, Rodríguez-Sánchez J, Sanchez-Cazalilla M, Lucendo AJ. Four-food group elimination diet for adult eosinophilic esophagitis: A prospective multicenter study. *J Allergy Clin Immunol* 2014;**134**:1093-1099
- 91. van Rhijn BD, Vlieg-Boerstra BJ, Versteeg SA, Akkerdaas JH, van Ree R, Terreehorst I, Sprikkelman AB, Verheij J, Smout AJ, Bredenoord AJ. Evaluation of allergen-microarray-guided dietary intervention as treatment of eosinophilic esophagitis. J Allergy Clin Immunol 2015;**136**:1095-1097.
- 92. Lucendo AJ, Arias A, Tenias JM. Relation between eosinophilic esophagitis and oral immunotherapy for food allergy: a systematic review with meta-analysis. *Ann Allergy Asthma Immunol* 2014;**113**:624-629.
- 93. Ridolo E, De Angelis GL, Dall'aglio P. Eosinophilic esophagitis after specific oral tolerance induction for egg protein. *Ann Allergy Asthma Immunol* 2011;**106**:73-74.
- Miehlke S, Alpan O, Schroeder S, Straumann A. Induction of eosinophilic esophagitis by sublingual pollen immunotherapy. *Case Rep Gastroenterol* 2013;**7:**363-368.
- 95. Maggadottir SM, Hill DA, Ruymann K, Brown-Whitehorn TF, Cianferoni A, Shuker M, Wang ML, Chikwava K, Verma R, Liacouras CA, Spergel JM.

Resolution of acute IgE-mediated allergy with development of eosinophilic esophagitis triggered by the same food. *J Allergy Clin Immunol* 2014;**133**:1487-1489.

- 96. Niranjan R, Mavi P, Rayapudi M, Dynda S, Mishra A. Pathogenic role of mast cells in experimental eosinophilic esophagitis. *Am J Physiol Gastrointest Liver Physiol* 2013;**304**:1087-1094.
- 97. Noti M, Wojno ED, Kim BS, Siracusa MC, Giacomin PR, Nair MG, Benitez AJ, Ruymann KR, Muir AB, Hill DA, Chikwava KR, Moghaddam AE, Sattentau QJ, Alex A, Zhou C, Yearley JH, Menard-Katcher P, Kubo M, Obata-Ninomiya K, Karasuyama H, Comeau MR, Brown-Whitehorn T, de Waal Malefyt R, Sleiman PM, Hakonarson H, Cianferoni A, Falk GW, Wang ML, Spergel JM, Artis D. Thymic stromal lymphopoietin-elicited basophil responses promote eosinophilic esophagitis. *Nat Med* 2013;**19**:1005-1013.
- Schoepfer AM, Safroneeva E, Bussmann C, Kuchen T, Portmann S, Simon HU, Straumann A. Delay in diagnosis of eosinophilic esophagitis increases risk for stricture formation in a time-dependent manner. *Gastroenterology* 2013;**145**:1230-1236.
- 99. Dellon ES, Kim HP, Sperry SL, Rybnicek DA, Woosley JT, Shaheen NJ. A phenotypic analysis shows that eosinophilic esophagitis is a progressive fibrostenotic disease. *Gastrointest Endosc* 2014;**79**:577-585.
- 100. Straumann A. Treatment of eosinophilic esophagitis: diet, drugs, or dilation? *Gastroenterology* 2012;**142:**1409-1411.
- 101. Lucendo AJ, Arias A, Gonzalez-Cervera J, Yague-Compadre JL, Guagnozzi D, Angueira T, Jimenez-Contreras S, Gonzalez-Castillo S, Rodriguez-Domingez B, De Rezende LC, Tenias JM. Empiric 6-food elimination diet induced and maintained prolonged remission in patients with adult eosinophilic esophagitis: a prospective study in the food cause of the disease. *J Allergy Clin Immunol* 2013;**131**:797-804.
- 102. Wen T, Stucke EM, Grotjan TM, Kemme KA, Abonia JP, Putnam PE, Franciosi JP, Garza JM, Kaul A, King EC, Collins MH, Kushner JP, Rothenberg ME.

Molecular diagnosis of eosinophilic esophagitis by gene expression profiling. *Gastroenterology* 2013;**145**:1289-1299.

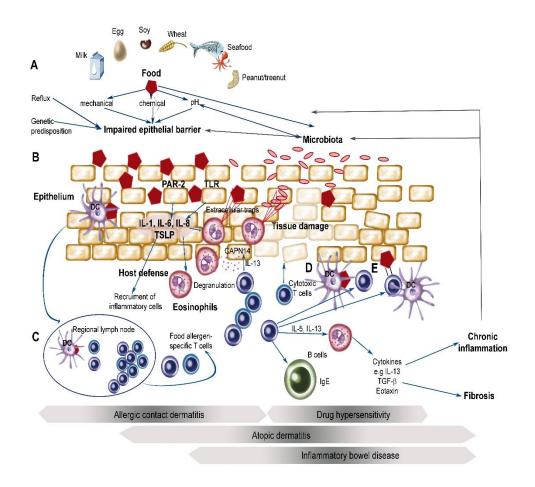


Figure 1 Food as a trigger in the pathogenesis of EoE. (A) In addition to the presence of a genetic predisposition or reflux disease, a food allergy would further disrupt the epithelial barrier and affect the microbiota. (B) Food allergens could then penetrate also in the skin, bind to pathogen-related receptors and activate epithelial cells to produce proinflammatory cytokines responsible for the recruitment and activation of inflammatory cells including eosinophils. (C) Antigen-presenting cells capturing food antigens, would migrate to the regional lymph nodes where they stimulate food-specific T cells. Food proteins may induce T cell responses either as a consequence of antigen presentation by dendritic cells (D) or directly (E) with subsequent eosinophil activation. By releasing toxic granule proteins and cytokines, eosinophils defend against invading pathogens, but cause tissue damage, stimulating fibrosis and perpetuating inflammation. The pathomechanisms of EoE overlapping with other diseases are indicated.