



the United Nations, 2005). Peppers are also an important part of Turkish cuisine and are consumed fresh, pickled, grilled, and stuffed as well as in salads and as a component of cooked dishes. In the present study, genotypic variation for antioxidants in pepper was studied by analyzing total water-soluble antioxidant capacity, phenolic content, and vitamin C content in 43 pepper cultivars, including 29 Turkish accessions. Fourteen non-Turkish cultivars grown in Turkey or worldwide were also analyzed. These foreign cultivars included standard, widely grown varieties such as ‘California Wonder’ and ‘Yolo Wonder’. Only water-soluble antioxidant capacity was measured because previous studies indicated that, compared with hydrophilic antioxidants, lipophilic antioxidants contribute very little to the total antioxidant capacity of pepper. For example, Wu et al. (2004) found that antioxidant capacity of the lipophilic fraction represented only 4.6% of the total antioxidant capacity. Similarly, Navarro et al. (2006) found that the antioxidant capacity of the lipophilic fraction of green fruits was less than 1% of that for the hydrophilic fraction. Significant genetic diversity was detected in the water-soluble antioxidant capacities and components of the tested pepper accessions and candidate lines were identified for future breeding programs. Breeding of higher antioxidant pepper cultivars could help improve human health because a diet rich in fruits and vegetables is considered to be the most important protection against many types of diseases (Ferrari and Torres, 2003).

## Materials and Methods

**Plant material.** Seeds for the Turkish cultivars were obtained from the Turkish National Germplasm Collection at the Aegean Agricultural Research Institute (AARI, Izmir, Turkey) and from the Atatürk Central Horticultural Research Institute (Yalova, Turkey). Seeds for the other cultivars were obtained from the Center for Genetic Resources, The Netherlands, and from seed distributors in Turkey and the United States (Table 1). Seeds were planted in a climate-controlled greenhouse in Apr. 2006 and three to five replicate plants were grown for each accession. Although some cultivars are consumed when they are red, to standardize the results, all fruits were harvested at the mature green stage in July and August and samples were stored at  $-20\text{ }^{\circ}\text{C}$  until assays were performed. All assays were completed within 1 month of harvest.

**Determination of antioxidant capacity.** For the extraction of antioxidants, a 150-g sample was taken from at least four individual peppers and was homogenized with 150 mL cold distilled water for 2 min at low speed in a Waring blender (Model HGB2WTS3; Waring Corp., Torrington, CT) equipped with a 1-L double-walled stainless steel jar chilled by circulating water at  $4\text{ }^{\circ}\text{C}$ . The samples were deseeded before homogenization with the exception of two very small-

Table 1. Description of pepper cultivars used for antioxidant trait assays.

Cultivar (origin)	Accession number (source <sup>2</sup> )	Country of origin	Type	Pungency	Color <sup>3</sup>
333 Biber	NA <sup>4</sup> (1)	Turkey	Çarliston	Sweet	Yellow
Acı Biber (Gaziantep)	TR47780 (1)	Turkey	Dolmalık	Hot	Green
Acı Sivri Biber (Bursa)	TR66271 (1)	Turkey	Sivri	Hot	Green
Apollo F1	NA (2)	Hungary	Dolmalık	Sweet	Yellow
Arnavut Biber	TR66272 (1)	Turkey	Süs	Hot	Green
Arnavut Biber, sivri	TR66299 (1)	Turkey	Süs	Hot	Green
Ayaş	NA (1)	Turkey	Sivri	Sweet	Green
California Wonder	NA (3)	USA	Dolmalık	Sweet	Green
Çarliston Biber (Bursa)	TR66275 (1)	Turkey	Çarliston	Sweet	Yellow
Carolina Wonder	NA (2)	USA	Dolmalık	Sweet	Green
Cecil RZ F1	NA (2)	Hungary	Dolmalık	Sweet	Yellow
Charleston Belle	NA (2)	USA	Dolmalık	Sweet	Green
Cherry Pick	NA (2)	USA	Süs	Sweet	Green
Chile Negro	NA (3)	Mexico	Süs	Hot	Dark green
Cuma Ovası	NA (1)	Turkey	Sivri	Hot	Light green
Dolmalık	TR70630 (1)	Turkey	Dolmalık	Hot	Green
Dolmalık Yeşil (Bursa)	TR66270 (1)	Turkey	Dolmalık	Sweet	Green
Domat Biberi (Bursa)	TR66393 (1)	Turkey	Dolmalık	Hot	Light green
Düğme Biber (Bursa)	TR66316 (1)	Turkey	Süs	Hot	Green
Edison	NA (2)	Netherlands	Dolmalık	Sweet	Green
Ege-91	NA (1)	Turkey	Sivri	Sweet	Light green
Farya	NA (2)	USA	Çarliston	Sweet	Yellow
Fiesta	NA (2)	Netherlands	Dolmalık	Sweet	Green
Finli Biber	TR66380 (1)	Turkey	Sivri	Hot	Light green
Kale	NA (1)	Turkey	Dolmalık	Hot	Light green
Kandil Dolma Biber	NA (1)	Turkey	Dolmalık	Sweet	Light green
Menderes	NA (1)	Turkey	Sivri	Hot	Light green
Raspires F1	NA (2)	Hungary	Çarliston	Hot	Yellow
Şahnalı Biber	NA (1)	Turkey	Sivri	Hot	Green
Salçalık Biber	TR66259 (1)	Turkey	Salçalık	Sweet	Red
Salçalık Biber (Bursa)	TR66389 (1)	Turkey	Salçalık	Sweet	Red
Salçalık Biber (Gaziantep)	TR48614 (1)	Turkey	Salçalık	Sweet	Red
Sera Demre	NA (1)	Turkey	Sivri	Sweet	Green
Sweet Long Slim Red	NA (3)	USA	Sivri	Sweet	Red
Tatlı Kıvrıkcık Biber	TR66305 (1)	Turkey	Sivri	Sweet	Light green
Variogated Flash	NA (3)	USA	Süs	Hot	Purple
Yağlık Biber	TR66378 (1)	Turkey	Salçalık	Sweet	Red
Yağlık Biber (Bursa)	TR66384 (1)	Turkey	Salçalık	Sweet	Red
Yalova Biber	NA (4)	Turkey	Sivri	Sweet	Yellow
Yalova Çarliston 341	NA (4)	Turkey	Çarliston	Sweet	Yellow
Yalova Tatlı Sivri Biber	NA (4)	Turkey	Sivri	Sweet	Light green
Yalova Yağlık	NA (4)	Turkey	Salçalık	Sweet	Red
Yolo Wonder 31–22	NA (3)	USA	Dolmalık	Sweet	Green

<sup>2</sup>1 = Aegean Agricultural Research Institute, Izmir, Turkey; 2 = purchased from Turkish or U.S. distributor; 3 = Center for Genetic Resources, The Netherlands; 4 = Atatürk Central Horticultural Research Institute, Yalova, Turkey.

<sup>3</sup>Color of fruits at normal stage of consumption.

<sup>4</sup>NA = no accession number.

sized peppers (‘Arnavut Biber’ and ‘Variogated Flash’), which are consumed with seeds. For cultivars with small fruits, a 50-g sample was homogenized with 50 mL distilled water using a 200-mL jar and the same homogenization conditions. A 20-g sample of fruit pulp was then filtered through four layers of cheesecloth. The filtrate was further clarified by centrifugation at  $3000 \times g$  for 10 min at  $4\text{ }^{\circ}\text{C}$ . The clear supernatant was used for the determination of antioxidant capacity according to the method of Re et al. (1999). In this method, ABTS [2,2'-azinobis-(3-ethyl-benzothiazoline-6-sulfonic acid)] radical cation decolorization caused by the test samples was monitored by spectrophotometer (Model 1700; Shimadzu, Kyoto, Japan) at 734 nm. The reaction mixture contained 2 mL potassium persulfate oxidized ABTS radical solution in phosphate-buffered saline at pH 7.4 and 2.5, 5, or 7.5  $\mu\text{L}$  of extract [or 20  $\mu\text{L}$  of Trolox (0.0045–0.03  $\mu\text{mol}$  in

reaction mixture) to prepare the standard curves]. The decrease in absorbance of each sample was monitored for 6 min and tests were conducted three times at each sample volume for each supernatant. Percent inhibitions at 1, 3, and 6 min were then plotted against sample volume. The slope for each line (1, 3, and 6 min), which indicated the percent inhibition of the sample per microliter, was determined and graphed against time using KaleidaGraph (Synergy Software, Reading, PA). This software was also used to calculate the area under the curve (AUC). This value and the AUC for the Trolox standard curve were used to calculate antioxidant capacity values, which were expressed as mmol Trolox/kg fresh weight (FW) of peppers.

**Determination of total phenolic content.** For determination of total phenolic content, homogenates were prepared as described for antioxidant capacity determination. After

centrifugation, the clear supernatant was used for the determination of total phenolic content according to the method of Singleton and Rossi (1965) using Folin-Ciocalteu as the reactive reagent and gallic acid as the standard. Briefly, 2 mL of sample extract was mixed with 10 mL 2 N (10%) Folin-Ciocalteu's reagent. After 3 min, 8 mL 0.7 M sodium carbonate was added. The reaction mixture was incubated for 2 h at room temperature and absorbance measured at 765 nm in a spectrophotometer. Three replicates were measured for each supernatant sample. The results were expressed as milligrams gallic acid equivalents/kg FW of peppers.

**Determination of vitamin C content.** Vitamin C content was determined by the AOAC 967.21 titrimetric method (Augustin, 1994) using 2,6-dichloroindophenol as the reactive substance and L-(+)-ascorbic acid for calibration. The extractions were conducted by homogenization of 100 g peppers without seeds (taken from at least four individual fruit) with 115 mL acetic acid–metaphosphoric acid extraction solution for 2 min at low speed in a Waring blender at 4 °C. A 35-g sample of each homogenate was diluted with extraction solution to a final volume of 100 mL. This dilution was filtered and used in titration. For each diluted pepper extract, the vitamin C content of three replicate samples was measured. The results were expressed as milligrams vitamin C/kg FW of peppers.

**Statistical analyses.** Total water-soluble antioxidant capacity, total phenolic content, and vitamin C content of the pepper fruits were analyzed using analysis of variance and Fisher's protected least significant difference. Analyses were performed across all cultivars and also across cultivars grouped by morphology/use type as explained in the "Results and Discussion."

## Results and Discussion

The 43 pepper accessions analyzed in this work included both Turkish and non-Turkish cultivars (Table 1). Most (25 of 29) of the Turkish lines were obtained from the National Germplasm Collection at AARI representing the diversity of pepper accessions grown in the country. These lines included varieties such as 'Ayaş' and 'Kale', which are grown throughout Turkey as well as regional cultivars that are grown only in specific areas. An example of such a cultivar is the accession 'Acı Biber' (meaning literally, hot pepper) from Gaziantep in southeastern Anatolia, a region that is famous for its hot peppers. Also included were 14 non-Turkish varieties, including F<sub>1</sub> hybrids such as 'Apollo' and 'Cecil' that are grown in Turkey and standard cultivars that are grown throughout the world. For comparison, the peppers were classified into five groups based on their morphology or primary use. These classes were: bell-type (Dolmalık) peppers that are used for stuffing; long, pointed (Sivri) peppers and long, blunt-ended Charleston-type (Çarliston) peppers that are often consumed raw; small-fruited "fancy" (Süs) peppers that are eaten fresh or

pickled; and paste (Salçalık) peppers that are processed into paste. However, these classes cannot be considered definitive because each pepper type is not used exclusively for only one purpose. For example, paste peppers can also be stuffed. Representative fruits of each type are shown in Figure 1. Most classes included both pungent (hot) and sweet pepper accessions with 15 hot and 28 sweet cultivars (Table 1).

**Total water-soluble antioxidant capacity.** Significant variation in total water-soluble antioxidant capacity was observed in the analyzed pepper cultivars. Values ranged from 2.57 to 18.96 mmol Trolox/kg, a 7.4-fold difference (Table 2). The five cultivars with highest antioxidant activities were all Turkish cultivars: Ege-91, Yalova Tatlı Sivri Biber, Domat Biberi, Finli Biber, and Ayaş.



Fig. 1. Five types of peppers commonly grown in Turkey. From left to right, fruits of Salçalık, Çarliston, Sivri, Süs (top), and Dolmalık (bottom) -type cultivars.

Table 2. Antioxidant capacity, total phenolic content, and vitamin C content in pepper cultivars.<sup>2</sup>

Cultivar (location)	Antioxidant capacity (mmol Trolox/kg) ± SE		Phenolic content (mg·kg <sup>-1</sup> ) ± SE		Vitamin C content (mg·kg <sup>-1</sup> ) ± SE	
	Rank	Rank	Rank	Rank	Rank	Rank
Ege-91	18.96 ± 0.24 a <sup>1</sup>	1	2,724 ± 4.9 a	1	1,519 ± 2.7 b	2
Yalova Tatlı Sivri Biber	17.65 ± 0.25 b	2	1,220 ± 4.5 no	21	1,502 ± 2.7 b	4
Domat Biberi (Bursa)	14.60 ± 0.64 c	3	1,796 ± 2.1 f	6	1,177 ± 7.6 gh	12
Finli Biber	13.40 ± 0.12 d	4	2,239 ± 8.9 c	3	1,276 ± 18.4 e	7
Ayaş	12.67 ± 0.10 e	5	1,730 ± 3.7 g	9	964 ± 12.4 l	21
Çarliston Biber (Bursa)	12.64 ± 0.11 e	6	1,782 ± 19 f	8	1,140 ± 48.1 i	15
Düğme Biber (Bursa)	10.25 ± 0.58 f	7	1,094 ± 8.6 q	26	1,257 ± 4.4 ef	8
Arnavut Biber, sivri	9.99 ± 0.15 fg	8	2,185 ± 12.9 d	4	1,098 ± 8.5 j	16
Menderes	9.88 ± 0.06 fg	9	1,925 ± 8.9 e	5	1,164 ± 13.5 ghi	13
Arnavut Biber	9.51 ± 0.18 gh	10	1,440 ± 12.4 k	15	1,631 ± 9.7 a	1
Sera Demre	9.46 ± 0.11 gh	11	946 ± 1.2 vw	35	926 ± 4.4 mno	25
Variegated Flash	9.13 ± 0.26 hi	12	2,311 ± 11.3 b	2	ND <sup>3</sup>	
Cecil RZ F1	8.89 ± 0.06 ij	13	988 ± 7.5 u	33	522 ± 4.7 w	42
Şahmalı Biber	8.81 ± 0.16 ij	14	1,578 ± 11.9 i	11	778 ± 24.3 s	32
Tatlı Kıvrıkcık Biber	8.78 ± 0.14 ij	15	1,394 ± 4.5 l	17	1,198 ± 24.7 g	10
Acı Sivri Biber (Bursa)	8.59 ± 0.14 j	16	1,476 ± 7.5 j	13	1,376 ± 11.6 d	6
Chile Negro	8.52 ± 0.36 j	17	1,790 ± 3.3 f	7	1,088 ± 5.0 j	17
Yalova Biber	8.37 ± 0.13 j	18	1,691 ± 10.6 h	10	916 ± 8.8 nopq	27
Cherry Pick	7.52 ± 0.10 k	19	1,482 ± 9.8 j	12	1,436 ± 20.0 c	5
Yolo Wonder 31–22	7.34 ± 0.08 k	20	1,232 ± 8.1 n	20	1,519 ± 3.0 b	3
Cuma Ovası	6.99 ± 0.08 k	21	1,440 ± 13.8 k	14	943 ± 6.1 lmn	23
Charleston Belle	6.30 ± 0.28 l	22	756 ± 9.8 z	41	778 ± 6.1 s	34
Apollo F1	6.11 ± 0.09 lm	23	1,110 ± 9.7 pq	25	778 ± 3.0 s	33
California Wonder	6.07 ± 0.11 lm	24	764 ± 12.2 z	40	1,153 ± 1.7 hi	14
Kandırlı Dolma Biber	5.62 ± 0.10 m	25	896 ± 13 x	38	974 ± 5.9 l	20
Dolmalık	5.62 ± 0.04 mn	26	1,052 ± 7.7 r	27	627 ± 0.0 u	38
Acı Biber (Gaziantep)	5.48 ± 0.16 no	27	1,411 ± 7.4 l	16	1,234 ± 22.9 f	9
Dolmalık Yeşil (Bursa)	5.43 ± 0.13 no	28	1,014 ± 6.2 st	31	945 ± 11.6 lm	22
Raspires F1	5.31 ± 0.07 no	29	1,233 ± 11.9 n	19	939 ± 4.5 lmno	24
Salçalık Biber (Gaziantep)	5.03 ± 0.12 op	30	1,204 ± 6.4 o	22	905 ± 4.8 opqr	28
Sweet Long Slim Red	4.59 ± 0.13 pq	31	1,202 ± 4.5 o	23	1,178 ± 21.8 gh	11
Edison	4.44 ± 0.01 q	32	925 ± 2.5 w	37	766 ± 2.6 s	35
Carolina Wonder	4.44 ± 0.11 q	33	607 ± 3.8 β	43	649 ± 6.9 u	37
Yağlık Biber (Bursa)	4.42 ± 0.02 q	34	1,324 ± 6.1 m	18	921 ± 5.8 nop	26
333 Biber	4.17 ± 0.04 qr	35	1,024 ± 1.2 st	30	568 ± 10.3 v	39
Farya	3.87 ± 0.13 rs	36	1,052 ± 11.3 r	28	561 ± 6.8 v	40
Salçalık Biber	3.86 ± 0.07 rs	37	956 ± 8.6 v	34	1,075 ± 13.1 j	18
Yağlık Biber	3.67 ± 0.17 rst	38	1,118 ± 9.8 p	24	872 ± 16.1 r	31
Kale	3.53 ± 0.27 st	39	1,037 ± 7.7 rs	29	883 ± 2.5 qr	30
Fiesta	3.14 ± 0.06 tu	40	649 ± 5.4 α	42	885 ± 3.0 pqr	29
Salçalık Biber (Bursa)	2.94 ± 0.05 uv	41	1,011 ± 4.3 tu	32	714 ± 7.6 t	36
Yalova Yağlık	2.72 ± 0.02 uv	42	926 ± 2.1 w	36	539 ± 4.6 vw	41
Yalova Çarliston 341	2.57 ± 0.09 v	43	852 ± 9.8 y	39	1,024 ± 2.8 k	19

<sup>2</sup>Cultivars are ranked by total antioxidant capacity. Rankings for phenolic and vitamin C content are also included.

<sup>3</sup>Values followed by different letters are significantly different at  $P < 0.05$  as determined by Fisher's protected least significant difference.

<sup>4</sup>Vitamin C content for Variegated Flash could not be determined (ND) because the purple-colored fruit extract prevented detection of color change during titration.

All of these are Sivri types with the exception of 'Domat Biber', which is a stuffing pepper. Mean antioxidant capacity for all lines was  $7.47 \pm 0.59$  (SE) mmol Trolox/kg. Because many different methods are used to determine total antioxidant capacity of fruits and vegetables, direct comparison of the results of the present study with those of other researchers is difficult. However, using a similar method, Pellegrini et al. (2003) found that green chili peppers had a Trolox-equivalent antioxidant capacity of  $7.62 \text{ mmol}\cdot\text{kg}^{-1}$ , a value that is similar to the mean water-soluble antioxidant capacity ( $7.47 \text{ mmol}\cdot\text{kg}^{-1}$ ) of the cultivars used in this work.

When peppers were grouped by type, it was clear that some types had significantly higher antioxidant activities (Table 3). Sivri types had the highest mean antioxidant capacity closely followed by Ss types. Dolmalık and arliston types had intermediate levels, whereas Salalik types had the lowest mean level of antioxidant capacity, which was 2.8-fold lower than the mean for Sivri types. It must be noted, however, that Salalik types are usually consumed when they are red. At this stage of maturity, these peppers may have considerable lipophilic antioxidant capacity because of their high carotenoid content. Some types of peppers showed more variation for antioxidant capacity among accessions (Fig. 2). Although only five arliston-type cultivars were tested, this type showed the most variation with a 4.9-fold difference between the cultivars with the highest (arliston) and lowest ('Yalova arliston 341') activities. Similarly, Dolmalık and Sivri types showed 4.6- and 4.1-fold differences in total antioxidant capacity, respectively. In contrast, Salalik and Ss pepper types were more uniform having only 1.8- and 1.4-fold differences in capacity, respectively.

In comparison with F<sub>1</sub> hybrids and standard varieties, some Turkish cultivars showed dramatically higher antioxidant activities. For example, 'arliston Biber' (Bursa) had at least 2.4-fold greater antioxidant capacity than the non-Turkish arliston types (Fig. 2). The three Turkish Ss pepper types also had significantly higher antioxidant activities than the other three Ss cultivars.

**Total phenolic content.** Total phenolic content in the pepper cultivars ranged from 607 to 2724 mg·kg<sup>-1</sup>, a 4.5-fold difference in content (Table 2). This range of phenolic

content was similar to that reported by other researchers (Antonious et al., 2006; Chassy et al., 2006). The five cultivars with the highest phenolic content included four Turkish cultivars [Ege-91, Finli Biber, Arnavut Biber (sivri), and Menderes] and one non-Turkish cultivar (Variegated Flash). All of these peppers except for 'Variegated Flash' are Sivri types. 'Variegated Flash' is a Ss type and was one of the two cultivars for which the fruit extract contained seeds. It was reported that seeds are a source of phenolic compounds in pepper (Velioglu et al., 1998). Therefore, the high phenolic content of 'Variegated Flash' may be attributable to its seeds. However, the other cultivar that had seeds in its extract ('Arnavut Biber') did not have especially high phenolic content. Mean phenolic content for all lines was  $1316 \pm 72$  (SE) mg·kg<sup>-1</sup>. Ss and Sivri types had significantly higher mean phenolic content than the other three types of pepper (Table 3). Dolmalık and Sivri types showed the most variation in phenolic content with  $\approx$ 3-fold variation in these cultivars (Fig. 3). The least variation was seen in Salalik types.

As with antioxidant capacity, some Turkish lines had significantly higher phenolic content than the non-Turkish cultivars. For example, the Dolmalık types, 'Domat' and 'Acı Biber' (Gaziantep), had significantly higher phenolic content than 'Yolo Wonder' and 'Apollo F<sub>1</sub>' (Fig. 3). 'arliston Biber' also had significantly higher phenolic content than the F<sub>1</sub> hybrid 'Raspire's' and cultivar 'Farya'.

The total phenolic content of pepper as measured by the Folin-Ciocalteu assay encompasses a wide diversity of compounds, including simple phenols, phenolic acids, flavonoids, lignin precursors, capsaicinoids, and reducing sugars (Howard et al., 2000). Individual flavonoids, including luteolin, quercetin, and kaempferol (Chassy et al., 2006; Howard et al., 2000), were measured in pepper and some recent studies provided detailed qualitative and quantitative characterization of pepper phenolic compounds (Marin et al., 2004; Materska and Perucka, 2005).

**Vitamin C content.** Vitamin C content in the peppers ranged from 522 to 1631 mg·kg<sup>-1</sup>, a 3.1-fold difference in content (Table 2). This range of vitamin C content was similar to that seen in other studies (Antonious et al., 2006; Chassy et al., 2006; Deepa et al., 2006; Howard et al., 2000; Marin et al., 2004). A

notable exception is the work of Guil-Guerrero et al. (2006), which reported vitamin C contents of 100 to 380 mg/100 g for 10 pepper cultivars grown in Spain. The five cultivars with highest vitamin C content included three Turkish and two non-Turkish cultivars. These lines were 'Arnavut Biber', 'Ege-91', 'Yolo Wonder', 'Yalova Tatlı Sivri Biber', and 'Cherry Pick' with vitamin C content averaging  $990 \pm 47$  (SE) mg·kg<sup>-1</sup>. Interestingly, 100-g serving sizes of all but four of the cultivars assayed in this work supply 100% of the daily Recommended Dietary Allowance of vitamin C, 60 mg (Table 2). Similarly, the majority of the cultivars (67%) meet the more recently devised Dietary Reference Intake for vitamin C, which averages between 75 and 90 mg for adult women and men, respectively (International Food Information Council, 2002). Sivri and Ss types had the highest mean vitamin C content (Table 3). Dolmalık types showed the most variation in vitamin C content with a 2.9-fold range in concentration (Fig. 4). The other pepper types had 1.5- to 2.0-fold variation in vitamin C content.

Turkish Sivri, Ss, and arliston-type pepper lines had significantly higher vitamin C content than non-Turkish cultivars. However, 'Yolo Wonder', a non-Turkish cultivar, had the highest vitamin C content of the Dolmalık types (Fig. 4).

**Relationship between pungency and antioxidant capacity.** To determine the relationship between pepper pungency and antioxidant capacity, mean values for hot and sweet types were compared (Table 3). Hot types had higher total antioxidant capacity, phenolic content, and vitamin C content; however, the difference between means was only statistically significant ( $P < 0.05$ ) for phenolic content (Table 3). This was not unexpected because capsaicin, the compound that gives peppers their pungency, is a capsaicinoid, a type of phenolic compound (Estrada et al., 2002).

**Correlation between antioxidant parameters.** All three antioxidant parameters showed statistically significant ( $P < 0.05$ ) correlations between each other. The strongest correlation was between total antioxidant capacity and phenolic content ( $r = 0.71$ ). There were also significant positive but weaker correlations between total antioxidant capacity and vitamin C content ( $r = 0.51$ ) and between vitamin C and phenolic content ( $r = 0.31$ ). Other researchers observed significant correlations between total antioxidant capacity and its components. Significant positive correlations were seen between total antioxidant capacity and phenolic content in pepper (Deepa et al., 2007), tomato (Hanson et al., 2004), eggplant (Hanson et al., 2006), cranberry (Wang and Stretch, 2001), and blueberry (Howard et al., 2003). Antonious et al. (2006) reported a much stronger correlation between phenolic and vitamin C content ( $r = 0.97$ ) than that reported in the present study. The correlations between the different antioxidant components were also apparent when the pepper cultivars were ranked by the value for each measured parameter (Table 2). Thus, 'Ege-91',

Table 3. Mean values for antioxidants in pepper cultivars grouped by type and pungency.

Pepper type	Number of cultivars	Mean antioxidant capacity (mmol Trolox/kg) $\pm$ SE	Mean phenolic content (mg·kg <sup>-1</sup> ) $\pm$ SE	Mean vitamin C content (mg·kg <sup>-1</sup> ) $\pm$ SE
Sivri	12	10.68 $\pm$ 1.23 a <sup>z</sup>	1,630 $\pm$ 140 a	1,145 $\pm$ 70.2 a
Dolmalık	14	6.21 $\pm$ 0.76 bc	1,017 $\pm$ 84 b	921 $\pm$ 72.8 ab
Ss	6	9.15 $\pm$ 0.41 ab	1,717 $\pm$ 191 a	1,117 $\pm$ 204 ab
arliston	5	5.71 $\pm$ 1.79 bc	1,188 $\pm$ 160 b	846 $\pm$ 119 ab
Salalik	6	3.77 $\pm$ 0.36 c	1,090 $\pm$ 63 b	838 $\pm$ 76.0 b
Hot	15	8.64 $\pm$ 0.77 a	1,600 $\pm$ 110 a	1,044 $\pm$ 88 a
Sweet	28	6.84 $\pm$ 0.80 a	1,163 $\pm$ 81 b	962 $\pm$ 55 a

<sup>z</sup>Within each column and grouping, values followed by different letters are significantly different at  $P < 0.05$  as determined by Fisher's protected least significant difference.

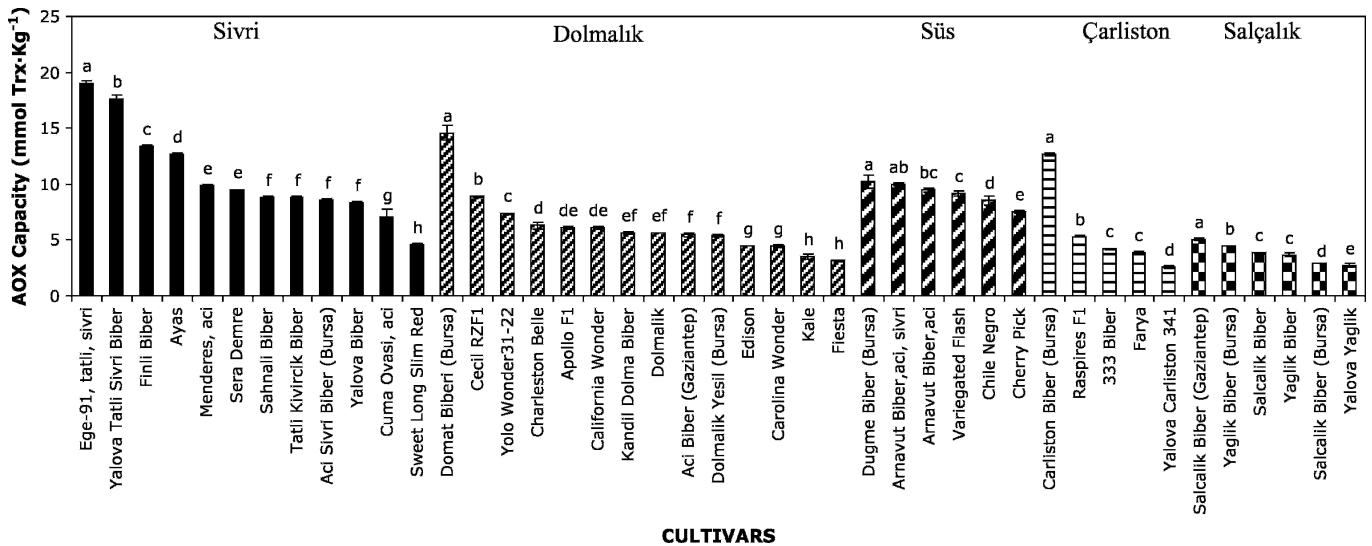


Fig. 2. Antioxidant capacities of the pepper cultivars grouped by type. Within each type, columns labeled with different letters are significantly different at  $P < 0.05$  as determined by Fisher's protected least significant difference.

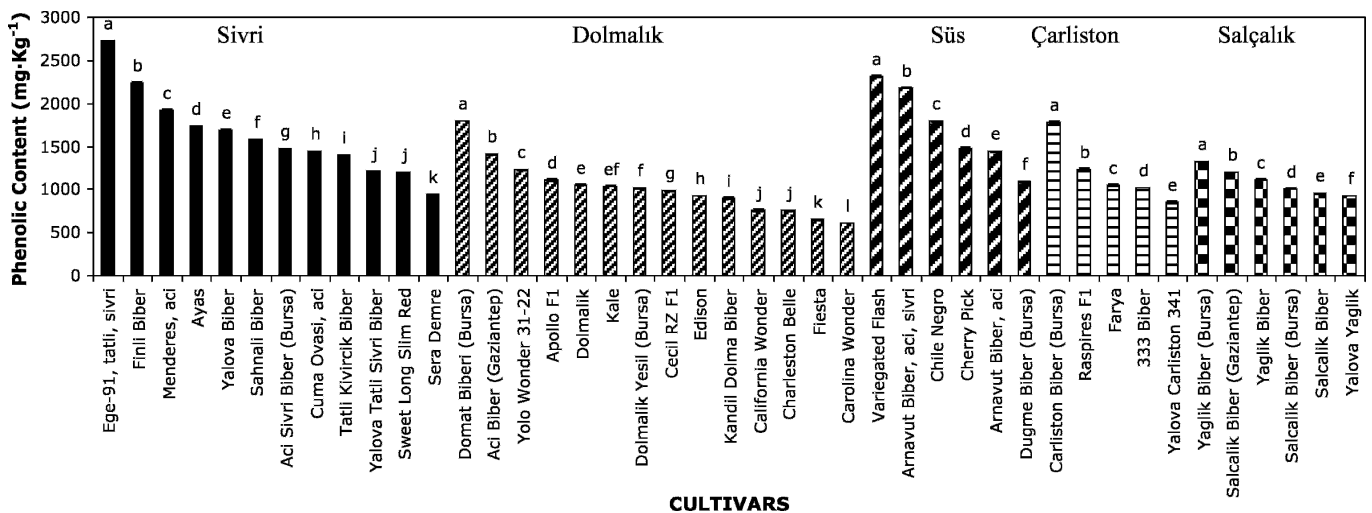


Fig. 3. Total phenolic content of the pepper cultivars grouped by type. Within each type, columns labeled with different letters are significantly different at  $P < 0.05$  as determined by Fisher's protected least significant difference.

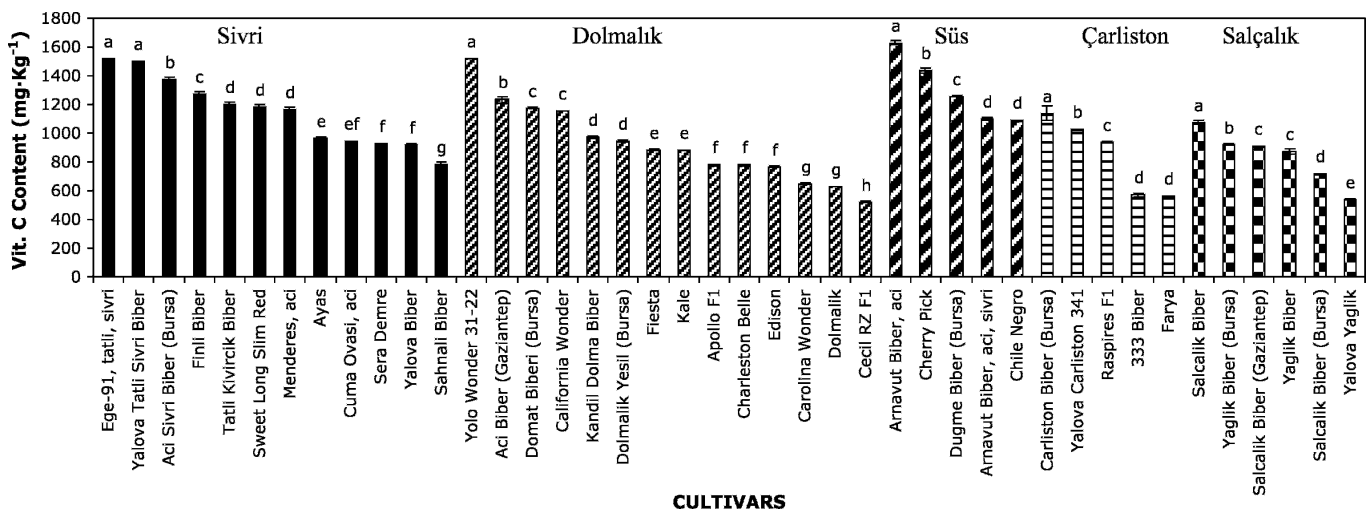


Fig. 4. Vitamin C content of the pepper cultivars grouped by type. Within each type, columns labeled with different letters are significantly different at  $P < 0.05$  as determined by Fisher's protected least significant difference.

which ranked first for total antioxidant capacity, also ranked first for phenolic content and second for vitamin C content. The correlations were especially obvious when cultivars were ranked within each type (Sivri, Dolmalik, and so on, Figs. 2–4). Within their type categories, ‘Ege-91’ (Sivri) and ‘Çarliston Biber’ ranked first for all three parameters. ‘Domat Biberi’ ranked first for total antioxidant capacity and phenolic content and third for vitamin C content in the Dolmalik types. Similarly, ‘Salçalık Biber’ from Gaziantep ranked first, second, and third for total antioxidant capacity, phenolic content, and vitamin C content, respectively. Such correlations were expected because the total antioxidant capacity assay measured the activity of all water-soluble antioxidants, including phenolics and vitamin C.

### Conclusions

The genetic diversity present in Turkish pepper germplasm for total water-soluble antioxidant capacity, phenolic content, and vitamin C content can be exploited for the development of populations for identification and genetic mapping of the loci controlling these traits in pepper and for the breeding of cultivars with improved antioxidant capacity. ‘Ege-91’ was the best cultivar for all three antioxidant parameters and is a good candidate for improvement of antioxidants in Turkish peppers, especially in Sivri types. Similarly, ‘Domat Biber’ would be a good candidate for improvement of total antioxidant capacity and phenolic content of Dolmalik-type peppers. Development and consumption of pepper cultivars with high antioxidant activity may help decrease the incidence of certain types of diseases in humans. It will also be interesting to see if these improved cultivars have increased tolerance to biotic and abiotic stress.

### Literature Cited

- Antonious, G.F., T.S. Kochhar, R.L. Jarret, and J.C. Snyder. 2006. Antioxidants in hot pepper: Variation among accessions. *J. Environ. Sci. Heal. B* 41:1237–1243.
- Augustin, J. 1994. Vitamin analysis, p. 249–260. In: Nielsen, S.S. (ed.). *Introduction to the chemical analysis of food*. Jones & Bartlett, Boston, MA.
- Bloch, A. and C.A. Thomson. 1995. Position of the American Dietetic Association: Phytochemicals and functional foods. *J. Amer. Diet. Assoc.* 95:493–496.
- Cao, G., E. Sofic, and R.L. Prior. 1996. Antioxidant capacity of tea and common vegetables. *J. Agr. Food Chem.* 44:3426–3431.
- Chassy, A.W., L. Bui, E.N.C. Renaud, M. Van Horn, and A.E. Mitchell. 2006. Three-year comparison of the content of antioxidant microconstituents and several quality characteristics in organic and conventionally managed tomatoes and bell peppers. *J. Agr. Food Chem.* 54:8244–8252.
- Chu, Y.-F., J. Sun, X. Wu, and R.H. Liu. 2002. Antioxidant and antiproliferative activities of common vegetables. *J. Agr. Food Chem.* 50:6910–6916.
- Davey, M.W., M. van Montagu, D. Inze, M. Sanmartin, A. Kanellis, N. Smirnoff, I.J.J. Benzie, J.J. Strain, D. Favell, and J. Fletcher. 2000. Plant L-ascorbic acid: Chemistry, function, metabolism, bioavailability and effects of processing. *J. Sci. Food Agr.* 80:825–860.
- Deepa, N., C. Kaur, B. George, B. Singh, and H.C. Kapoor. 2007. Antioxidant constituents in some sweet pepper (*Capsicum annuum* L.) genotypes during maturity. *LWT-Food Sci. Technol.* 40:121–129.
- Deepa, N., C. Kaur, B. Singh, and H.C. Kapoor. 2006. Antioxidant activity in some red sweet pepper cultivars. *J. Food Compos. Anal.* 19:572–578.
- Estrada, B., M.A. Bernal, J. Diaz, F. Pomar, and F. Merino. 2002. Capsaicinoids in vegetative organs of *Capsicum annuum* L. in relation to fruiting. *J. Agr. Food Chem.* 50:1188–1191.
- Ferrari, C.K.B. and E.A.F.S. Torres. 2003. Biochemical pharmacology of functional foods and prevention of chronic diseases of aging. *Biomed. Pharmacother.* 57:251–260.
- Food and Agriculture Organization of the United Nations. 2005. Agricultural data FAOSTAT. 1 May 2007. <<http://faostat.fao.org/site/340/default.aspx>>.
- Gnayfeed, M.H., H.G. Daoood, P.A. Biacs, and C.F. Alcaraz. 2001. Content of bioactive compounds in pungent spice red pepper (paprika) as affected by ripening and genotype. *J. Sci. Food Agr.* 81:1580–1585.
- Guil-Guerrero, J.L., C. Martinez-Guirado, M. Reboloso-Fuentes, and A. Carrique-Perez. 2006. Nutrient composition and antioxidant activity of 10 pepper (*Capsicum annuum* L.) varieties. *Eur. Food Res. Technol.* 224:1–9.
- Halliwell, B. 2006. Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. *Plant Physiol.* 141:312–322.
- Halvorsen, B.L., K. Holte, M.C.W. Myhrstad, I. Barikmo, E. Hvattum, S.F. Remberg, A.-B. Wold, K. Haffner, H. Baugerod, L.F. Andersen, J.O. Moskaug, D.R. Jacobs, and R. Blomhoff. 2002. A systematic screening of total antioxidants in dietary plants. *J. Nutr.* 132:461–471.
- Hanson, P.M., R.-Y. Yang, S.C.S. Tsou, D. Ledesma, L. Engle, and T.-C. Lee. 2006. Diversity in eggplant for superoxide scavenging activity, total phenolics and ascorbic acid. *J. Food Comp. Anal.* 19:594–600.
- Hanson, P.M., R.-Y. Yang, J. Wu, J.-T. Chen, D. Ledesma, and S.C.S. Tsou. 2004. Variation for antioxidant activity and antioxidants in tomato. *J. Amer. Soc. Hort. Sci.* 129:704–711.
- Howard, L.R., J.R. Clark, and C. Brownmiller. 2003. Antioxidant capacity and phenolic content in blueberries as affected by genotype and growing season. *J. Sci. Food Agr.* 83:1238–1247.
- Howard, L.R., S.T. Talcott, C.H. Brenes, and B. Villalon. 2000. Changes in phytochemical and antioxidant activity of selected pepper cultivars (*Capsicum* species) as influenced by maturity. *J. Agr. Food Chem.* 48:1713–1720.
- International Food Information Council. 2002. Dietary reference intakes: An update. 1 May 2007. <<http://ific.org/publications/other/driupdateom.cfm?renderforprint=1>>.
- Marin, A., F. Ferreres, F.A. Tomas-Barberan, and M.I. Gil. 2004. Characterization and quantitation of antioxidant constituents of sweet pepper (*Capsicum annuum* L.). *J. Agr. Food Chem.* 52:3861–3869.
- Materska, M. and I. Perucka. 2005. Antioxidant activity of the main phenolic compounds isolated from hot pepper fruit (*Capsicum annuum* L.). *J. Agr. Food Chem.* 53:1750–1756.
- Navarro, J.M., P. Flores, C. Garrido, and V. Martinez. 2006. Changes in the contents of antioxidant compounds in pepper fruits at different ripening stages, as affected by salinity. *Food Chem.* 96:66–73.
- Ou, B., D. Huang, M. Hampsch-Woodill, J.A. Flanagan, and E.K. Deemer. 2002. Analysis of antioxidant activities of common vegetables employing oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) assays: A comparative study. *J. Agr. Food Chem.* 50:3122–3128.
- Pellegrini, N., M. Serafini, B. Colombi, D. Del Rio, S. Salvatore, M. Bianchi, and F. Brighenti. 2003. Total antioxidant capacity of plant foods, beverages and oils consumed in Italy assessed by three different in vitro assays. *J. Nutr.* 133:2812–2819.
- Percival, M. 1998. Antioxidants. *Clinical nutrition insights*. 1:1–5. 1 May 2007. <<http://acudoc.com/Antioxidants.PDF>>.
- Podsedek, A. 2007. Natural antioxidants and antioxidant capacity of *Brassica* vegetables: A review. *LWT-Food Sci. Technol.* 40:1–11.
- Re, R., N. Pellegrini, A. Progettante, A. Pannala, M. Yang, and C. Rice-Evans. 1999. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Bio. Med.* 26:1231–1237.
- Sakihama, Y., M.F. Cohen, S.C. Grace, and H. Yamasaki. 2002. Plant phenolic antioxidant and prooxidant activities: Phenolics-induced oxidative damage mediated by metals in plants. *Toxicology* 177:67–80.
- Singleton, V.L. and J.A. Rossi, Jr. 1965. Colorimetry of total phenolics with phosphomolybdic phosphotungstic acid reagents. *Amer. J. Enol. Viticult.* 16:144–158.
- Slater, A., N.W. Scott, and M. Fowler. 2003. *Plant biotechnology: The genetic manipulation of plants*. Oxford Univ. Press, Oxford, UK.
- Surh, Y.J. and S.K. Seoul. 2002. Anti-tumor promoting potential of selected spice ingredients with oxidative and anti-inflammatory activities. *Food Chem. Toxicol.* 40:1091–1097.
- USDA Nutrient Data Laboratory. 2007. USDA national nutrient database for standard reference. 1 May 2007. <<http://www.nal.usda.gov/fnic/foodcomp/search>>.
- Velioglu, Y.S., G. Mazza, L. Gao, and B.D. Oomah. 1998. Antioxidant activity and total phenolics in selected fruits, vegetables and grain products. *J. Agr. Food Chem.* 46:4113–4117.
- Wang, S.Y. and A.W. Stretch. 2001. Antioxidant capacity in cranberry is influenced by cultivar and storage temperature. *J. Agr. Food Chem.* 49:969–997.
- Wu, X., G.R. Beecher, J.M. Holden, D.B. Haytowitz, S.E. Gebhardt, and R.L. Prior. 2004. Lipophilic and hydrophilic antioxidant capacities of common foods in the United States. *J. Agr. Food Chem.* 52:4026–4037.