

1 **Association of apolipoprotein E gene polymorphisms with blood lipids and their**  
2 **interaction with dietary factors**

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41 **Abstract**

42 **Background:** Several candidate genes have been identified in relation to lipid  
43 metabolism, and among these, lipoprotein lipase (*LPL*) and apolipoprotein E (*APOE*)  
44 gene polymorphisms are major sources of genetically determined variation in lipid  
45 concentrations. This study investigated the association of two single nucleotide  
46 polymorphisms (SNPs) at *LPL*, seven tagging SNPs at the *APOE* gene, and a common  
47 *APOE* haplotype (two SNPs) with blood lipids, and examined the interaction of these  
48 SNPs with dietary factors.

49 **Methods:** The population studied for this investigation included 660 individuals from the  
50 Prevention of Cancer by Intervention with Selenium (PRECISE) study who supplied  
51 baseline data. The findings of the PRECISE study were further replicated using 1,238  
52 individuals from the Caerphilly Prospective cohort (CaPS). Dietary intake was assessed  
53 using a validated food-frequency questionnaire (FFQ) in PRECISE and a validated semi-  
54 quantitative FFQ in the CaPS. Interaction analyses were performed by including the  
55 interaction term in the linear regression model adjusted for age, body mass index, sex and  
56 country.

57 **Results:** There was no association between dietary factors and blood lipids after  
58 Bonferroni correction and adjustment for confounding factors in either cohort. In the  
59 PRECISE study, after correction for multiple testing, there was a statistically significant  
60 association of the *APOE* haplotype (rs7412 and rs429358; E2, E3, and E4) and *APOE*  
61 tagSNP rs445925 with total cholesterol ( $P=4 \times 10^{-4}$  and  $P=0.003$ , respectively). Carriers of  
62 the E2 allele had lower total cholesterol concentration ( $5.54 \pm 0.97$  mmol/L) than those  
63 with the E3 ( $5.98 \pm 1.05$  mmol/L) ( $P=0.001$ ) and E4 ( $6.09 \pm 1.06$  mmol/L) ( $P=2 \times 10^{-4}$ )  
64 alleles. The association of *APOE* haplotype (E2, E3, and E4) and *APOE* SNP rs445925

65 with total cholesterol ( $P=2 \times 10^{-6}$  and  $P=3 \times 10^{-4}$ , respectively) was further replicated in the  
66 CaPS. Additionally, significant association was found between *APOE* haplotype and  
67 *APOE* SNP rs445925 with low density lipoprotein cholesterol in CaPS ( $P=4 \times 10^{-4}$  and  
68  $P=0.001$ , respectively). After Bonferroni correction, none of the cohorts showed a  
69 statistically significant SNP-diet interaction with lipid outcomes.

70 **Conclusion:** In summary, our findings from the two cohorts confirm that genetic  
71 variations at the *APOE* locus influence plasma total cholesterol concentrations, however,  
72 the gene-diet interactions on lipids require further investigation in larger cohorts.

73

74 **Keywords:** *APOE* gene, total cholesterol, LDL-C, PRECISE, Caerphilly Prospective  
75 studies

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77

78 **Background**

79 Cardiovascular diseases (CVD) are common multifactorial conditions  
80 characterized by dyslipidaemia, type 2 diabetes and hypertension [1, 2]. Elevated  
81 triacylglycerol (TAG) and reduced high density lipoprotein cholesterol (HDL-C)  
82 concentrations are associated with an increased risk of developing CVD [3-5].  
83 Furthermore, several studies have reported that certain genetic variants influence  
84 susceptibility to altered circulating lipid concentrations, leading to an increased risk of  
85 CVD events [6-8]. Genetic variations have been shown to be associated with lipid  
86 outcomes, while dietary factors appear to modulate the effect of such genes on lipid  
87 concentrations [9, 10]. Previous studies have shown that single nucleotide  
88 polymorphisms (SNPs) of the apolipoprotein E (*APOE*) [6, 11] and lipoprotein lipase  
89 (*LPL*) [12-14] genes contribute to significant variation in lipid concentrations.

90 The *APOE* protein plays a key role in the transport and metabolism of cholesterol  
91 and TAG containing particles by serving as a receptor-binding ligand that mediates the  
92 clearance of dietary derived chylomicrons, and hepatically derived very low density  
93 lipoprotein (VLDL) and their remnants from the circulation [6]. The three most  
94 recognized alleles of the *APOE* gene are E2, E3 and E4, with carriage of E4 associated  
95 with CVD risk factors and increased low density lipoprotein cholesterol (LDL-C)  
96 concentrations [11, 15, 16], and hence increased CVD risk [17, 18].

97 Genetic variations in the *LPL* gene have been reported to be involved with lipid  
98 metabolism and partly explain the phenotypic variation in blood lipid levels [19]. *LPL* is  
99 a lipolytic enzyme that catalyses hydrolysis of TAG in all of the major classes of TAG-

100 rich lipoproteins [20]. High enzyme activity is associated with favourable lipid levels,  
101 including relatively low TAG concentrations [21]. The two most widely studied *LPL*  
102 SNPs, rs328 (S447X) and rs320 (HindIII) [22, 23]. The ‘G’ minor alleles of both the  
103 SNPs, rs328 and rs320, are associated with decreased TAG concentrations and increased  
104 HDL-C concentrations, whereas the opposite association was found for the ‘C’ allele and  
105 ‘T’ allele respectively [24-26].

106 Data from several studies supports the role of genetic factors in lipid metabolism  
107 [27]; however, only a few studies have examined the effects of lifestyle factors such as  
108 diet on the association of polymorphisms with lipid-related outcomes [10, 28, 29].  
109 Therefore, the present study aimed to investigate the effect of seven *APOE* tagSNPs  
110 (rs405509, rs769450, rs439401, rs445925, rs405697, rs1160985, and rs1064725), one  
111 *APOE* haplotype (rs7412 and rs429358), and two commonly studied *LPL* SNPs (rs328  
112 and rs320) on blood lipid profile in 660 participants (baseline data) from the Prevention  
113 of Cancer by Intervention with Selenium (PRECISE) study. As diet type and intake is  
114 also known to modify lipid levels [30-32], the potential impact of the interaction between  
115 these SNPs and dietary factors on lipid levels was also investigated. To confirm the  
116 findings, the Caerphilly Prospective Study (CaPS; n=1,238) was used as a replication  
117 cohort.

118

## 119 **Material and methods**

### 120 *PRECISE cohort*

#### 121 *Participants and methods*

122 Baseline data of 660 individuals from the PRECISE study, conducted in two  
123 populations [UK (n=468) and Denmark (n=192)] were used for the analysis [33, 34].  
124 Briefly, study participants were selected from four general practices (study centres) in  
125 various areas of the UK that were affiliated with the Medical Research Council General  
126 Practice Research Framework (MRC GPRF). Between June 2000 and July 2001, research  
127 nurses recruited similar numbers of men and women from each of three age groups: 60–  
128 64, 65–69 and 70–74 years. The Danish participants were men and women recruited from  
129 the same three age groups from the County of Funen in Denmark.

130 The UK study obtained approval from the appropriate UK Local Research Ethics  
131 Committees [South Tees (ref: 99/69), Worcestershire Health Authority (ref: LREC  
132 74/99), Norwich District (ref: LREC 99/ 141), Great Yarmouth and Waveney (under  
133 reciprocal arrangements with Norwich District LREC)], and the participants provided  
134 written informed consent. The regional Danish Data Protection Agency and Scientific  
135 Ethical Committees of Vejle and Funen counties approved the Danish study (Journal  
136 number. 19980186).

### 137 *Dietary information*

138 Information about each participant's usual dietary intake was obtained using  
139 validated EPIC food frequency questionnaires (FFQ) [35]. Total energy intake and  
140 macronutrient composition were analysed using the FETA software program [36].

### 141 *Anthropometric measurements and biochemical analysis*

142 Body mass index (BMI) was calculated as body weight in kilograms divided by

143 height in square metres ( $\text{kg}/\text{m}^2$ ). Participants provided non-fasting blood samples for  
144 biochemical analysis and these samples were stored at  $-80^\circ\text{C}$ . Total cholesterol and  
145 HDL-C concentrations in lithium-heparin plasma were measured using an Architect  
146 c16000 analyser (Abbott) with dedicated reagents. Measurements were performed by  
147 enzymatic colorimetric analysis. Traceability for total cholesterol and HDL-C was  
148 ensured through participation in the National Reference System for Cholesterol  
149 (NRS/CHOL), as established by the Clinical and Laboratory Standards Institute, with  
150 isotope dilution-MS used as the reference method, and reference material taken from the  
151 National Institute of Standard and Technology. Evidence of equivalence in the analytical  
152 performance of the cholesterol-oxidase assays performed in the UK and Denmark from a  
153 comparison of total cholesterol on forty-four serum samples which produced a limit of  
154 variation of 2% [33].

155 *SNP selection:*

156 The *APOE* gene is located on chromosome 19q13.32. It comprises four exons,  
157 which are transcribed into the *APOE* mRNA which is 1,180 nucleotides long. The seven  
158 tagSNPs for the *APOE* gene were chosen based on International HapMap Phase II  
159 collected from individuals of Northern and Western European ancestry (CEU) (HapMap  
160 Data release 27 Phase 2+3, Feb 09, NCBI B36 assembly, dbSNP b126). The Haploview  
161 software V3.3 (<http://www.broadinstitute.org/haploview/haploview-downloads>) was used  
162 to assess the linkage disequilibrium between SNPs. Tagger software was used to select  
163 tagSNPs with the ‘pairwise tagging only’ option. Two criteria were used to filter the  
164 SNPs included in the analysis, minor allele frequency  $\geq 5\%$  and Hardy–Weinberg  
165 equilibrium P-value  $> 0.01$ . In total, seven tagSNPs [rs405509 (G>T), rs1160985 (C>T),



166 rs769450 (G>A), rs439401 (C>T), rs445925 (G>A), rs405697 (G>A), and rs1064725  
167 (T>G)] representing the entire common genetic variations across the *APOE* gene were  
168 selected for the study. The *APOE* haplotype/SNPs [6, 11, 37-44] and *LPL* [12, 13] SNPs  
169 were chosen based on their previous association with various lipid outcomes.

#### 170 *DNA isolation and genotyping*

171 The genotyping for the selected SNPs using a KASP assay with a competitive  
172 allele-specific PCR assay® was performed on DNA samples by LGC Genomics  
173 (Hoddesdon, Herts, UK). The eleven SNPs were in Hardy Weinberg Equilibrium (HWE)  
174 ( $P>0.05$  for all comparisons) (**Supplementary Table 1**).

175

#### 176 *Caerphilly Prospective Study (CaPS)*

##### 177 *Participants and methods*

178 The CaPS was used to replicate the findings from the PRECISE study. The phase  
179 1 (July 1979 to September 1983) recruitment for the CaPS included 2,512 men aged 45-  
180 59 years who were living in the town of Caerphilly and five of its adjacent villages in the  
181 UK; these participants were followed up at regular intervals [45, 46]. The follow-up data  
182 collection included periods from 1984 to 1988 (phase 2), from 1989 to 1993 (phase 3),  
183 from 1993 to 1997 (phase 4), and from 2002 to 2005 (phase 5). For the current study, the  
184 data analysed were taken from phase 3 (n=1,238), which had the maximum number of  
185 samples and variables appropriate to this analysis (total cholesterol and dietary  
186 information), and from phase 5 (n=529) (HDL-C and LDL-C). Ethical approval was  
187 obtained from the South Wales Research Ethics Committee D, and each subject provided

188 written informed consent.

189 *Dietary information*

190 Participants completed validated semi-quantitative FFQ in phase 3 [47, 48]. The  
191 FFQ included 50 typical food items in the British diet in order to estimate the mean daily  
192 energy intake and macronutrients and micronutrients consumption.

193 *Anthropometric measurements and biochemical analysis*

194 Height and weight was recorded in order to calculate the BMI. Height was  
195 measured on a stadiometer and weight was measured on a beam balance. Plasma prepared  
196 from blood samples taken after an overnight fast were transported at 4°C to the  
197 laboratories on the day of venepuncture. Total cholesterol and HDL-C, LDL-C  
198 concentrations were measured using enzymatic procedures [49]. and the LDL-C levels  
199 were calculated using the Friedewald Formula [50].

200 *DNA isolation and genotyping*

201 DNA was extracted from blood samples collected during the period 1992–1994.  
202 SNP information was obtained from the Illumina Cardio Metabochip, which includes  
203 data on 200,000 SNPs from regions previously identified for associations with risk  
204 factors for cardiometabolic disease [51]. Imputation was conducted against the 1000-  
205 genomes reference panel, providing information on approximately two million typed or  
206 imputed SNPs. Duplicate samples were genotyped to compute the error rate. Quality  
207 control on genotyped samples has been previously reported [52] and the SNPs had a call  
208 rate of >98%. The SNPs were in HWE ( $P > 0.05$ ) (**Supplementary Table 1**).

209

210 *Statistical analysis*

211 Statistical analysis was performed using the SPSS software package, version 22.0. The  
212 data were presented as mean  $\pm$  standard deviation (SD) in Tables 1 and 3 and beta  
213 regression coefficients and standard error (SE) were presented in Tables 2, 4, and 5.  
214 Independent t-test was used to compare means between men and women at baseline in  
215 the PRECISE cohort (Table 1). Univariate linear regression analysis was applied to test  
216 for association of the SNPs with total cholesterol and HDL-C, controlling for age, sex,  
217 BMI and country. SNP-diet interactions on total cholesterol and HDL-C were  
218 investigated using a univariate general linear model. In this model, total cholesterol and  
219 HDL-C were the dependent variables, SNPs were fixed factors, and dietary factors (fat  
220 energy %, protein energy %, carbohydrate energy %), sex, age BMI, and country were  
221 covariates. The dominant model was applied for all SNPs with minor allele frequency  
222  $\leq 0.3$  and the additive model applied for SNPs with minor allele frequency  $\geq 0.4$ . For  
223 analytical purposes, the six *APOE* genotype groups (E2/E2, E2/E3, E3/E3, E3/E4, E4/E4,  
224 and E2/E4) were classified into three groups. The E3/E3 genotype was classified as a  
225 group as it occurs at high frequency in the population (wild type). The E2/E2 and E2/E3  
226 genotypes were combined and presented as E2 carriers. The E3/E4 and E4/E4 genotypes  
227 were also combined, and presented as E4 carriers [29]. Previous studies have shown that  
228 the impact of the E2 allele on serum lipids is greater than that of the E4 allele [17],  
229 therefore, the E2/E4 genotype was excluded from the analysis. The Bonferroni correction  
230 was applied separately for association and interaction analyses. For association between  
231 phenotypic and dietary factors, the Bonferroni-corrected P value was 0.008 (2 lipid

232 outcomes\* 3 dietary factors) for the PRECISE study and P value was 0.01 for CaPS (total  
233 cholesterol was the only variable available). For association between SNPs and lipids  
234 (PRECISE study), the Bonferroni corrected P value was 0.003 (10 SNPs\*2 lipid  
235 outcomes = 20 tests). For interactions (PRECISE study), the Bonferroni corrected P value  
236 was 0.001 (10 SNPs\*2 lipid outcomes\*3 dietary factors = 60 tests). In the replication  
237 analysis (CaPS cohort), the Bonferroni corrected P value for association was 0.002 (10  
238 SNPs\*3 lipid outcomes = 30 tests), while for interactions it was 0.001 (10 SNPs\*1 lipid  
239 outcome\* 3 dietary factors = 30 tests).

240

## 241 **Results**

### 242 *Participant characteristics*

243 The general characteristics of the participants by sex are presented in **Table 1**. In  
244 the PRECISE study, women were found to have significantly higher total cholesterol and  
245 HDL-C concentrations than men ( $P=2.31 \times 10^{-10}$  and  $P= 2.71 \times 10^{-16}$ , respectively). The  
246 consumption of carbohydrates ( $P=1.42 \times 10^{-9}$ ) and protein (energy %) ( $P=5 \times 10^{-5}$ ) were  
247 higher in women than in men, whereas the consumption of fat (energy %) and total  
248 energy intake were lower in women than in men ( $P=0.01$ ). Characteristics of the  
249 individuals from CaPS are given in **Table 1**. Elevated total cholesterol levels were  
250 observed among men at phase 3. Dietary-pattern data showed higher consumption of  
251 energy from total fat.

### 252 *Association between dietary factors and blood lipids*

253 In both the PRECISE and CaPS, there was no association between the dietary

254 factors and total cholesterol or high-density lipoprotein after Bonferroni correction and  
 255 adjustment for confounding factors (**Table 2**).

256

257 **Table 2: Association between dietary factors and lipids in PRECISE and Caerphilly**  
 258 **Prospective studies**

<b>PRECISE study</b>		
<i>Association between dietary factors and total cholesterol</i>		
Fat total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>	Protein total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>	Carbohydrate total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>
0.01 (0.01) 0.47	-0.01 (0.01) 0.13	-0.004 (0.01) 0.40
<i>Association between three dietary factors and HDL-C high density lipoprotein</i>		
Fat total energy % intake	Protein total energy % intake	Carbohydrate total energy % intake
-0.002 (0.002) 0.29	-0.002 (0.004) 0.59	-0.004 (0.002) 0.02
<b>Caerphilly Prospective study</b>		
<i>Association between three dietary factors and total cholesterol</i>		
Fat total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>	Protein total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>	Carbohydrate total energy % intake Beta ( $\pm$ S.E), P <sub>association</sub>
0.01 (0.004) 0.06	-0.01 (0.01) 0.26	-0.01 (0.004) 0.17

259 HDL-C, high density lipoprotein cholesterol.  
 260 P values were obtained using linear regression adjusted for age, sex, body mass index and  
 261 country.

262

263

264 *Genotypes and serum lipid levels in the PRECISE study*

265 As shown in **Table 3**, of the seven tagSNPs at *APOE*, tagSNP rs445925 was  
 266 significantly associated with total cholesterol (P=0.003) after correction for multiple  
 267 testing. The ‘A’ allele carriers (5.65 $\pm$  0.98 mmol/L) had 5% lower levels of total

268 cholesterol than GG homozygotes ( $5.99 \pm 1.06$  mmol/L).

269 The levels of HDL-C were significantly different among the *LPL* SNP genotypes,  
270 rs328 ( $P=0.04$ ) and rs320 ( $P=0.02$ ), where the carriers of the ‘G’ minor allele of both  
271 SNPs had higher levels of HDL-C ( $1.68 \pm 0.41$  mmol/L for rs328 and  $1.66 \pm 0.40$  mmol/L  
272 for rs320) than CC homozygotes (rs328) and TT homozygotes (rs320) ( $1.61 \pm 0.38$  and  
273  $1.60 \pm 0.39$  mmol/L) respectively. However, these associations were not statistically  
274 significant after Bonferroni correction.

#### 275 *APOE* haplotype and serum lipid levels in the PRECISE study

276 The effects of *APOE* haplotypes (E2, E3, and E4) on serum lipids are shown in  
277 **Table 3**. These haplotypes (E2, E3, and E4) were significantly associated with total  
278 cholesterol ( $P=4 \times 10^{-4}$ ) after correction for multiple testing. The carriers of the E2 allele  
279 ( $5.54 \pm 0.97$  mmol/L) had lower total cholesterol concentrations than the carriers of the  
280 E3 ( $P=0.001$ ) ( $5.98 \pm 1.05$  mmol/L) and E4 alleles ( $6.09 \pm 1.06$  mmol/L) ( $P=2 \times 10^{-4}$ ).

#### 281 *Interactions between genotypes and dietary factors on serum lipid in the PRECISE study*

282 None of the dietary factors significantly interacted with the *APOE* SNPs,  
283 haplotypes and *LPL* SNPs with plasma lipids after correction for multiple testing ( $P$   
284  $>0.001$ ) (**Table 4**).

#### 285 *Replication analysis: Effect of SNPs at APOE and LPL on serum lipids in the CaPS*

286 The associations of *APOE* and *LPL* SNPs with blood lipids in the CaPS are  
287 presented in **Table 3**. The association of *APOE* haplotype (E2, E3, and E4) and *APOE*

288 SNP rs445925 with total cholesterol ( $P=2 \times 10^{-6}$  and  $P=3 \times 10^{-4}$ , respectively) was  
289 replicated. The 'A' allele carriers of *APOE* SNP rs445925 had lower total cholesterol  
290 ( $5.96 \pm 1.24$  mmol/l) than 'GG' genotypes ( $6.24 \pm 1.08$  mmol/L). In the *APOE* haplotype  
291 analysis, the carriers of the E2 allele had 5% and 14% lower total cholesterol than carriers  
292 of the E3 ( $P=4 \times 10^{-4}$ ) and E4 alleles ( $P=3 \times 10^{-6}$ ), respectively. Additionally, significant  
293 association was seen between *APOE* haplotypes (E2, E3, and E4) and *APOE* SNP  
294 rs445925 and LDL-C ( $P=4 \times 10^{-4}$ , 0.001, respectively).

295         There was an interaction between fat (% energy) and *APOE* haplotype (E2, E3,  
296 and E4) on total cholesterol ( $P=0.038$ ) in CaPS. However, after correction for multiple  
297 testing, all the SNP-diet interactions were consistent with chance variation (**Table 5**).

298

## 299 **Discussion**

300         Our findings demonstrated significant associations between the *APOE* haplotype  
301 (E2, E3, and E4) and *APOE* SNP rs445925 with total plasma cholesterol and LDL-C  
302 (only CaPS) concentration, which were further replicated in an independent UK  
303 Caucasian cohort. The levels of total cholesterol were significantly lower in carriers of  
304 the *APOE* E2 allele and the 'A' allele of the SNP rs445925 than carriers of E3, E4 and  
305 'GG' genotype of the *APOE* SNP rs445925, respectively. Given that our findings confirm  
306 that genetic polymorphisms of *APOE* influence the inter-individual variation in total  
307 plasma cholesterol, a marker of dyslipidemia, changes in dietary consumption to reduce  
308 disease susceptibility could be implemented for individuals at genetic risk.

309         The effects of *APOE* polymorphisms on lipid concentrations have previously been  
310 investigated in different ethnic groups [11, 53, 54] and studies have shown that the *APOE*

311 gene variants contributed to 7% variability in total cholesterol [55]. The results of the  
312 current study were in line with previously reported findings that *APOE* haplotypes (E2,  
313 E3, and E4) are associated with serum total cholesterol and LDL-C, with E4 carriers  
314 associated with increased concentrations compared with E3/E3 wildtype and particularly  
315 E2 carriers [16, 53, 56]. One of the primary roles of APOE is binding the low density  
316 lipoprotein receptor (LDLR) and the LDLR-related protein, to facilitate cellular uptake of  
317 lipoprotein particles [57]. The three alleles, E2, E3, and E4, differ in their amino-acid  
318 sequences, resulting in functional differences in receptors-binding affinity. Amino-acid  
319 sequences of the E2 allele have lower binding affinity than those of the E3 and E4 alleles,  
320 causing decreased hepatic VLDL and chylomicron remnants clearance, thus reducing the  
321 uptake of postprandial lipoprotein particles [57]. Furthermore, it could be postulated that  
322 increase in apoE TAG-rich lipoproteins in E4 carriers could possibly increase the affinity  
323 to bind LDL-receptors resulting in decreased uptake of LDL and increased circulating  
324 plasma cholesterol [58]. E2 carriers also have an impaired conversion of the VLDL  
325 particles to LDL-C compared to E4 carriers [59], who have a higher rate of VLDL  
326 catabolism [60], which explains in part the lower total cholesterol and LDL-C in E2 allele  
327 carriers.

328         Furthermore, our study highlights an association between *APOE* SNP rs445925,  
329 which is one of the selected tagSNPs within the *APOE* gene, and total cholesterol. The  
330 SNP rs445925 has not been extensively studied, however, a genome-wide association  
331 study showed a significant association between SNP rs445925 and LDL-C levels in 3,644  
332 black and white individuals from the US and Europe [61]. In addition, previous genome-  
333 wide linkage and association studies have shown linkage disequilibrium (LD) between



334 *APOE* SNPs rs7412 and rs445925 [62] and between ‘A’ allele carriers at SNP rs445925  
335 and E2 haplotype [63], respectively, which could explain in part a similar function in  
336 cholesterol synthesis. It is also possible that A’ allele carriers of the SNP rs445925 might  
337 exhibit lower conversion of the VLDL particles to LDL-C which could have resulted in  
338 the decreased rate of LDL formation and hence lowered the total cholesterol  
339 concentrations [63].

340 Besides genetic associations, our study also identified an interaction of *APOE*  
341 haplotypes (E2, E3, and E4) with intake from fat (%) on total cholesterol in the CaPS,  
342 where, among those who consumed a low-fat diet (%), individuals carrying the E2 allele  
343 had significantly lower total cholesterol concentrations than to E4 allele carriers.  
344 However, this interaction was not statistically significant after correction for multiple  
345 testing. A previous study has examined the response of *APOE* genotype to fat intake in  
346 45 individuals using a prospective design, where after consumption of a lower-fat-  
347 cholesterol diet (34% fat, 265 mg/day) according to modified National Cholesterol  
348 Education program there was a significant reduction in total cholesterol by 14%, 9%, and  
349 4% in E4/E4, E3/E4, and E3/E3 genotypes, respectively [64]. Another study showed that  
350 the response to a diet high in cholesterol increases total cholesterol in E3 and E4  
351 compared to E2 allele carriers in a study comprising 29 healthy men [65]. By contrast, a  
352 cross sectional study in European Caucasians (n=996) reported that E2 allele carriers had  
353 lower total cholesterol levels, but there were no reported between interactions between  
354 saturated fatty acids and total cholesterol [66]. Given that the previous studies have given  
355 inconsistent results and have used various types of fatty acids, replication of our gene-diet

356 interaction finding in a large well-designed randomized controlled trial is highly  
357 warranted.

358 Previous studies have shown that the minor allele of *LPL* SNP rs328 enhance  
359 lipolytic activity [12]. Increased activity of LPL results in enhance clearance of TAG  
360 from the circulation, and associated with higher HDL-C concentrations [67]. The *LPL*  
361 SNP rs320 (HindIII) is in LD with rs328 (S447X) and they have been shown to have  
362 similar effects on HDL-C, where minor allele was reported to increase HDL-C [24, 68].  
363 In our study, in accordance with findings from other studies, there were associations  
364 between *LPL* SNPs, rs320 and rs328, and HDL-C concentrations, where common  
365 homozygotes of both SNPs had lower HDL-C [22-24, 26]. However, in our study, these  
366 associations were no longer statistically significant after Bonferroni correction.  
367 Furthermore, there were no significant *LPL* SNP-diet interactions with HDL-C or total  
368 cholesterol concentrations in either cohort. To date, there has only been one study that  
369 has shown an interaction between *LPL* rs328 and total fat intake on HDL-C in 8,764  
370 individuals from the US population, where high fat intake associated with increase HDL-  
371 C in CC homozygotes and CG heterozygotes carriers [28]. One of the main reasons we  
372 did not identify a significant interaction may be our small sample size; however, we  
373 cannot rule out an effect of differences in dietary fat sources between European and the  
374 US population.

375 The present study has some limitations. Importantly, some lipid-related outcomes,  
376 such as LDL-C and TAG concentrations, were not measured in the PRECISE study. The  
377 PRECISE study was also conducted in two populations, a UK cohort and a Danish  
378 cohort, which used different food frequency questionnaires and this might have

379 introduced measurement bias, even though the current results were adjusted for country  
380 in the regression analysis to avoid confounding. Another possible limitation is the use of  
381 a cross-sectional design (in both studies) to investigate genetic effects at a single point in  
382 time, whereas a longitudinal analysis design would have captured the genetic effects on  
383 lipid outcomes over a specific time period. The effect-size of the minor allele of some of  
384 the studied SNPs was relatively small, and hence a large sample size is required to detect  
385 reliably detect any interaction between SNPs and dietary factors. Despite the fact that this  
386 study was not adequately powered to detect such an interaction, it was sufficiently  
387 powered to detect the main effects (i.e., associations). Significant gene-diet interactions  
388 were identified, however these did not reach the Bonferroni-corrected P value ( $P=0.001$ )  
389 and hence need to be confirmed in larger cohorts. This study is strengthened by the fact  
390 that it is the first study to investigate the role of tagSNPs at the *APOE* gene in relation to  
391 dietary factors and lipid outcomes. The fact that genetic associations from the PRECISE  
392 study were replicated in another Caucasian cohort (CaPS) confirms the validity of our  
393 findings. Additionally, CaPS was based on a cohort with a very high response rate, and is  
394 therefore closely representative of the general population.

## 395 **Conclusion**

396 Our study, carried out in two Caucasian populations, confirmed that genetic  
397 variations at the *APOE* gene locus influence plasma lipid concentrations. Thus, our  
398 results suggest that *APOE* gene variants affect risk of dyslipidemia in individuals who  
399 carry the E4 risk allele and GG genotype at SNP rs445925. Future studies with a larger  
400 sample size examining tagSNPs at *APOE*, particularly prospectively genotyped dietary  
401 intervention studies are required to confirm the gene-diet interactions identified in our

402 study.

**Abbreviations:** *APOE*: apolipoprotein E; *LPL*: Lipoprotein lipase; PRECISE: Prevention of Cancer by Intervention with Selenium; CVD: cardiovascular disease; SNPs; single nucleide polymorphisms; TAG: triacylglycerol; HDL-C: high density lipoprotein cholesterol; FFQ: food frequency questionnaire; LDL-C: low density lipoprotein cholesterol; VLDL: very low density lipoprotein; HWE: Hardy Weinberg Equilibrium; CaPS: Caerphilly Prospective cohort; LD: linkage disequilibrium.

## **Declarations**

**Ethics approval and consent to participate:** Written informed consent was obtained from each study participant, and the study was approved by the regional Danish Data Protection Agency and Scientific Ethical Committees of Vejle and Funen counties approved the Danish study (PRECISE), the appropriate UK Local Research Ethics Committees [South Tees (ref: 99/69), Worcestershire Health Authority (ref: LREC 74/99), Norwich District (ref: LREC 99/ 141), Great Yarmouth and Waveney (under reciprocal arrangements with Norwich District LREC)] (PRECISE), the South Wales Research Ethics Committee D (CaPS).

**Consent for publication:** Written informed consent for publication was obtained from all the study participants.

**Availability of data and material:** Not applicable

**Competing interests:** None

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### **Author contributions**

IMS performed the statistical analysis and drafted the manuscript; KSV conceived and designed the nutrigenetics study; KW and MR designed and conducted the PRECISE study; PE designed and led the conduct of the Caerphilly Prospective study and YBS was involved in the design and conduct of phase V as well as obtaining funding for genetic analysis. JAL, BE, KW, MR, YBS, PE, IG, and KSV critically reviewed the manuscript. All authors contributed to and approved the final version of the manuscript.

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**Figure legend:**

**Figure 1** Association of *APOE* haplotypes (E2, E3, and E4) with total cholesterol concentrations in the Prevention of Cancer by Intervention with Selenium (PRECISE) study and Caerphilly Prospective study (CaPS). E2 allele carriers have significantly lower levels of total cholesterol than E3 ( $P=0.001$  and  $P=4 \times 10^{-4}$  in the PRECISE and CaPS, respectively) and E4 ( $P=2 \times 10^{-4}$  and  $P=3 \times 10^{-6}$  in the PRECISE and CaPS, respectively) allele carriers.

**Table 1: Baseline characteristics of the PRECISE and Caerphilly Prospective study participants**

Characteristics	PRECISE study			Caerphilly Prospective study (CaPS)
	Men (N=248 UK, 95 Danish)	Women (N=220 UK, 97 Danish)	P value	Men (N=1,238)
Age (years)	67 ± 4	67± 4	0.12	62± 4
Body mass index (kg/m <sup>2</sup> )	27.2± 4.9	27.3± 4.9	0.82	26.8± 3.7
Total Cholesterol (mmol/L)	5.6± 0.9	6.2± 1.1	2.31x10 <sup>-10</sup>	6.1± 1.1
High density lipoprotein cholesterol (mmol/L)*	1.5± 0.3	1.7± 0.4	2.71x10 <sup>-16</sup>	1.3± 0.3
Protein intake (total energy %)	17.6± 3.7	18.8± 3.7	5X10 <sup>-5</sup>	14.9± 2.7
Carbohydrate intake (total energy %)	42.8± 13.3	48.2± 8.7	1.42x10 <sup>-9</sup>	48.4± 7.5
Fat intake (total energy %)	35.3± 7.1	33.9± 6.9	0.01	36.5± 6.9
Total energy intake (kcal)	2256 ± 658	1992± 613	2.63x10 <sup>-7</sup>	1964 ± 625
Total energy intake (MJ)	9.4± 2.7	8.3± 2.6	2.63x10 <sup>-7</sup>	8.2± 2.6

Data shown are represented as means ± SD, wherever appropriate. P values are for the differences in the means between men and women. P values were calculated by using independent t-test.

\*For CaPS, HDL-C levels were obtained from phase 5 while all other variables were obtained from phase 3.