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# Microalgae of the Chlorophyceae Class: Potential Nutraceuticals Reducing Oxidative Stress Intensity and Cellular Damage

Blas-Valdivia Vanessa, Ortiz-Butron Rocio, Rodriguez-Sanchez Ruth, Torres-Manzo Paola, Hernandez-Garcia Adelaida and Cano-Europa Edgar Escuela Nacional de Ciencias Biológicas, Instituto Politécnico Nacional Departamento de Fisiología, México

#### 1. Introduction

Nutraceutical is a term combining the words nutrition and pharmaceutical. It is a food or food product that provides health and medical benefits, including the prevention and treatment of disease. A nutraceutical has beneficial effects because it possesses many compounds with antioxidant and intracellular signalling-pathway modulator effects. In recent years, it has been demonstrated that microalgae of the Chlorophyceae class could be excellent nutraceuticals because they contain polyphenols, chlorophyll, β-carotene, ascorbic acid, lycopene, α-tocopherol, xanthophylls, and PUFAs. For this reason, some research groups, including ours, have studied the nutraceutical properties of the genera *Dunalliela*, *Haematococcus*, and *Chlorella*. However, our research group has put special emphasis on the genera *Chlorella* and *Chlamydomonas*. For these genera, we present new results that reveal antioxidant effects in different models of oxidative stress and cell damage

## 2. Nutraceuticals

For a long time, natural products obtained from plants have been used as prominent sources of prophylactic agents for the prevention and treatment of disease in humans, animals, and in plants. Hippocrates (460-370 BC) started "let food be your medicine and medicine be your food". Now, the relationship between food and drugs is getting closer.

As we enter the third millennium, with increased life expectancy and greater media coverage of the health care issue, consumers are understandably more interested in the potential benefits of nutritional support for disease control or prevention. A recent survey in Europe concluded that diet is rated more highly by consumers than exercise or the hereditary factor for achieving good health (Hardy, 2000). For that reason, many entrepreneurs seek to introduce different products into the health and nutritional market. Marketing strategies have exploited the words "functional food" and "nutraceuticals" in their advertisements. Nutraceuticals and functional foods are the fastest growing segment of today's food industry, although nutraceuticals should be treated as pharmaceutical

products as we will detail. Nutraceuticals and functional foods are a market estimated at between \$6 billion US and \$60 billion US and it is growing at 5% per annum. Unfortunately, entrepreneurs in an effort to make money attract, as irresponsible market entrants, products that do not comply with biosafety tests. This is because there are few laws that regulate the production and sale of such products. Because the products are not submitted for standardized toxicology testing, sometimes they may be toxic for human consumption. There are no specific regulation in any country to control nutraceuticals, and they need to be established and should be considered under the same laws that regulate pharmaceuticals and food (Bernal et al., 2011). For our purposes, we will first define "nutraceuticals" and "functional foods" and how the microalgae could be excellent nutraceuticals.

The term nutraceutical was first mentioned in 1989 to describe the union between nutrition and pharmaceuticals, both key contributors to human wellness. Stephen DeFelice MD is the founder and chairman of the Foundation for Innovation in Medicine (FIM) and he defined a nutraceutical as a food (or part of the food) that provides medicinal health benefits, including the prevention or treatment of a disease. It was proposed that a nutraceutical is not a drug, which is a pharmacologically active substance that potentiates, antagonizes, or otherwise modifies any physiological function. A nutraceutical may be a single natural nutrient in powder, tablet, capsule, or liquid form. It is not necessarily a complete food but equally not a drug (Hardy, 2000). Also, it was proposed that a nutraceutical is a product that delivers a concentrated form of a presumed bioactive agent from a food, presented in a nonfood matrix, and it is used with the purpose of enhancing health in a dosage that exceeds those that could be obtained from normal food (Zeisel, 1999).

Functional food and nutraceutical are terms used incorrectly and indiscriminately for nutrients or nutrient-enriched food that can prevent or treat disease. Functional food is a product that resembles traditional food but it possesses demonstrated physiological benefits (Shahidi, 2009). For example a functional food could be a lutein-rich food as chicken, spinach, tomatoes, or oranges, or the omega-3 fatty acids found in fish oil. All functional foods are processed and consumed as food. A nutraceutical is not a nutritional supplement because the latter are nutrients that are added to the diet to correct or prevent deficiencies of vitamins, minerals, and proteins, and often used in the recovery of a patient suffering an illness or has undergone surgery, and also taken to improve overall health (Mandel et al., 2005). The beneficial effects of nutraceuticals and functional foods have been attributed to their components, such as polyphenols, polyunsatured fatty acids (PUFAs), terpenes, chlorophyll, and accessory pigments of the photosynthetic apparatus in cyanobacteria such as *Spirullina*. In general these compounds are antioxidants that reduce intensity of oxidative stress or modulate intracellular communication

#### 3. Nutraceutical effects of polyphenols, particularly flavonoids

The polyphenols are compounds characterized by a benzene ring bearing one or more hydroxyl groups attached to the ring. They are ubiquitous in the plants, vegetables, fruit, vines, tea, coffee and microalgae. The polyphenols in food originate from one of the main classes of secondary metabolites in plants. They are involved in the growth and reproduction and are produced as a response to defend injured plants against pathogens, and to participate in the defense mechanism against ultraviolet radiation (Biesalski, 2007). Polyphenols have different nutraceutical properties, such as an antioxidant, antiinflammatory (Biesalski, 2007), anticancer (Oz & Ebersole, 2010), antibacterial (Du et al.,

2011), antiatherogenic, and antiangiogenic (Rimbach et al., 2009). There are now polyphenols with therapeutic properties for which the mechanism of action at the molecular level has been discovered and they are used in clinical trials, e.g. flavonoids.

Flavonoids comprise the most common group of polyphenols and provide much of the flavor and color to fruit and vegetables. More than 6000 different flavonoids have been described and it is estimated that humans consume about 1 g/day.

The structure of flavonoids is C6-C3-C6 and they consist of two aromatic rings linked through three carbons usually forming an oxygenated heterocycle nucleus, named the flavan nucleus, and shown in figure 1. In general, the flavonoids are classified into six groups (Grassi et al., 2009).

- 1. **Flavones**: These kinds of flavonoids are used by angiosperms to color their flowers. Natural flavones include apigenin (4',5,7-trihydroxyflavone), (3',4',5,7-tetrahydroxyflavone), (4',5,6,7,8-pentamethoxyflavone), chrysin (5,7-dihydroxyflavone), baicalein (5,6,7-trihydroxyflavone), scutellarein (5,6,7,4'-tetrahydroxyflavone), wogonin (5,7-Dihydroxy-8-methoxyflavone). There are synthetic flavones such as diosmin and flavoxate.
- 2. **Flavonols**: These compounds are used by organisms to protect them from UV radiation. Their diversity stems from the different positions of the hydroxyl groups on the benzene rings (show figure 1). There are flavonols as kaempferol (3,4',5,7-tetrahydroxy-2-phenylchromen-4-one), quercetin (3,3',4',5,7-pentahydroxy-2-phenylchromen-4-one), myricetin (3,3',4',5',5,7-hexahydroxy-2-phenylchromen-4-one), galangin (3,5,7-trihydroxy-2-phenylchromen-4-one), and morin (2-(2,4-dihydroxyphenyl)-3,5,7-trihydroxychromen-4-one). **Flavanones**: These flavonoids are the direct precursors of the vast majority of flavonoids. Some examples of flavanones are: naringenin (4',5,7-trihydroxyflavanone) and butin (7,3',4'-trihydroxyflavanone).
- 3. **Catechin or flavanols**: These flavonoids have two chiral centers on the molecule on carbons 2 and 3, yielding four diastereoisomers. Two of the isomers are in the *trans* configuration and are called catechins and the other two are in the *cis* configuration and are called epicatechins. These flavonoids are present in food as a complexs or oligomerics and polymerics as procyanidins or proantocyanidins. The catechins are found in different fruits, i.e. apples, apricots, blackberries, and grapes. Catechins are also in red wine, but black tea and cocoa are the richest sources. The flavanols in finished food products depend on the cultivar type, geographical origin, agriculture practice, postharvesting handling, and food processing (Scalbert et al., 2005).
- 4. **Antocyanidins**: Antocyanidins are a large group of natural colorants. The color of most fruits, flowers, and berries are made from a combination of anthocyanins and anthocyanidins. Anthocyanins always contain a carbohydrate molecule, whereas anthocyanidins do not. Examples of antocyanidins are cyanidin (3,3',4',5,7-pentahydroxyflavylium chloride), pelargonidin (3,5,7-trihydroxy-2-(4-hydroxyphenyl) benzopyrylium chloride), and malvidin (3,5,7,4'-tetrahydroxy-3',5'-dimethoxyflavylium)
- 5. **Isoflavones**: This group is a class of organic compounds that sometimes act as phytoestrogens in mammals and are called antioxidants because of their ability to trap a singlet oxygen. Genistein (4',5,7-trihydroxyisoflavone) and daidzein (4',7-dihydroxyisoflavone) are two examples of isoflavones.

Some authors have proposed that aurones are another flavonoid group, however we consider that aurones are derived from chalcones (Fowler & Koffas, 2009).

The flavonoid synthesis is shown in figure 1. It begins when a cell transforms phenylananine or tyrosine into phenylpropanoic acid or cinnamic acid by phenylalanine-tyrosine ammonia lyase (PAL; EC 4.3.1.25/TAL; EC 4.3.1.25). Then cytochrome-P450 cinnamate 4-hydroxylase (C4H; EC 1.14.13.11) adds a 4'-hydroxyl group to form p-coumaric acid. The CoA esters are subsequently synthetized from cinnamic acid, caffeic acid, or p-coumaric acid by 4coumaryl:CoA ligase (4CL; EC 6.2.1.12). The type III polyketide chalcone synthase (CHS; EC 2.3.1.74) catalyzes the sequential condensation of three malonyl-CoA moieties with one CoA-ester molecule to form chalcones. The flavanones are formed when chalcones are isomerized into (2S)-flavanones by chalcone isomerase (CHI; EC 5.5.1.6). Many enzymes can modify the flavanones. For example the flavanones could be reduced to form isoflavones by isoflavone synthase (IFS; EC 1.14.13.86). After that, isoflavones are modified by different enzymatic systems to produce hydroxylation, reduction, alkylation, oxidation, and glucosylation alone or in combination in the three-ring phenylpropanoid core. Enzymes such as O-methyltransferse (IOMT, EC 2.1.1.150), isoflavone 2'-reductase (I2'R; EC 1.3.1.45), andisoflavone reductase (IFR; EC 1.3.1.45) can yield over 8000 different chemical structures from isoflavone (Winkel-Shirley, 2001; Fowler & Koffas, 2009). Another branch of the biosynthetic pathway of flavonoids is the flavones that are synthesized from flavanones through the action of the flavone synthase type I and II (FSI; EC 1.14.11.22). Flavonones are hydroxylated and then with flavonol synthase (FLS; EC 1.14.11.23) form flavonols. These compounds are the precursors of anthocyanins.

The beneficial effects can be divided into

- 1. **Antioxidants**: Flavonoids suppress the formation of reactive oxygen species (ROS) either by inhibiting enzymes or chelating trace elements involved in free radical production. Thus flavonoids help maintain an ROS steady state in the case of physical and chemical injury of the cell (Corradini et al., 2011). Not all flavonoids are ROS scavengers because some flavonoids, as nucleophiles, trap electrons from the ROS and become a free radical themselves, which then propagate a chain reaction causing a deleterious effect in the cell (Grassi et al., 2009).
- 2. **Modulators of intracellular communication**: The flavonoids and their metabolites act in the phosphoinositide 3-kinase (PI3K), Akt-protein kinase B (Akt-PKB), tyrosine kinase, and protein kinase C (PKC) signalling cascade. The inhibition or activation of these cascades modifies cellular function by altering the phosphorylation state of target molecules that modulate the expression of genes. This can explain the anticancer and neuroprotector flavonoid activities (Williams et al., 2004).
- 3. **Enzyme activity modulator**: Flavonoids offer cardiovascular protection because of their indirect inhibition of the angiotensin-converting enzyme (ACE; EC 3.4.15.1) (Actis-Goretta et al., 2006). Other enzymes inhibited by flavonoids are aromatase (EC 1.14.14.1) and  $\alpha$ -amylase (EC 3.2.2.1) (Hargrove et al., 2011). The inhibition of enzymes that have a Fe-S cluster has been demonstrated (Mena et al., 2011).

In general, flavonoids are molecules responsible of some of the beneficial effect of nutraceuticals and functional foods. The different effects of flavonoids are described in table 1.

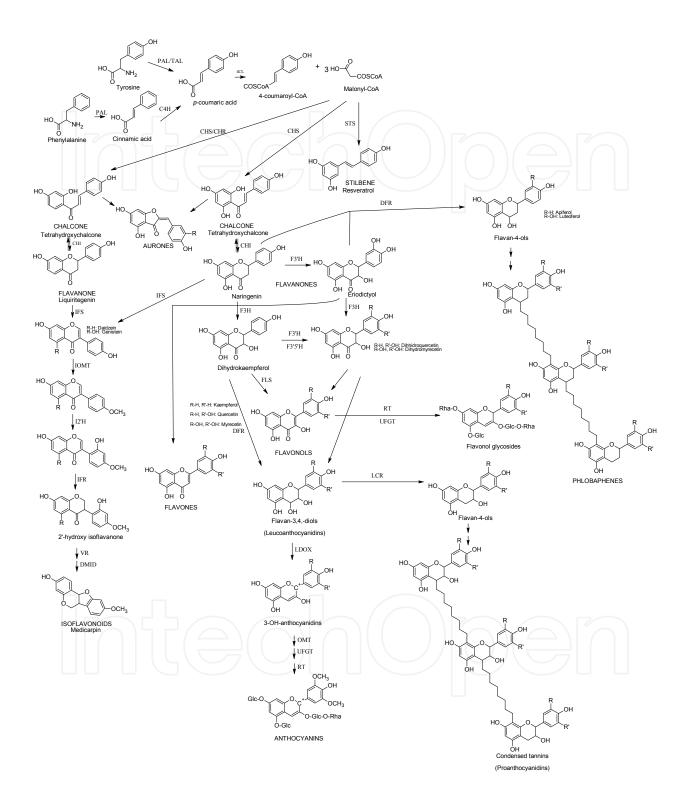


Fig. 1. Scheme of the major branch pathways of flavonoid biosynthesis, starting with general phenylpropanoid metabolism and leading to the nine major subgroups; chalcones, aurones, isoflavonoids, flavones, flavonols, flavandiols, anthocyanins, condensed tannins, and phlobaphene pigments

Flavonoid	Nutraceutic application	Reference
	LDL oxidation (atherosclerosis)	(Miranda et al., 2000)
	Cyclooxygenase inhibitory ability	(Kinghorn et al., 2004)
	(cancer)	(Kumar et al., 2003)
	Malaria chemotherapy (malaria)	(Kontogiorgis et al., 2008)
	Inflammation response trigger (reduce	(Mojzis et al., 2008)
	inflammation)	(Qin et al., 2011)
Flavanones	Antiangiogenic effect	(Prabu et al., 2011)
	Reduce lung metastases	(Celiz et al., 2011)
	Hepatoprotective action	(Orsolic et al., 2011)
	Antibacterial action	(Chao et al., 2010)
	Genoprotective action	(Sabarinathan et al., 2010)
	Inhibitors of NOS and COX in microglia	
	Promote apoptosis in C6 glioma cells	
	GLUT inhibitors (diabetes)	(Kwon et al., 2007)
	Cyclooxygenase inhibitory ability	(Kinghorn et al., 2004)
	(cancer)	(Balk, 2011; Polier et al.,
	Antitumoral activity	2011)
Flavones	Pancreatic cholesterol esterase inhibitor	(Peng et al., 2011)
	Reduce neurodegeneration	(Gasiorowski et al., 2011)
	Produce apoptosis in melanoma cells	(Mohan et al., 2011)
	Colitis treatment	(Ganjare et al., 2011)
	Antiinflammatory effect	(Funakoshi-Tago et al., 2011)
	GLUT inhibitors (diabetes)	
	Pancreatic lipase inhibitors (diabetes)	(Kwon et al., 2007; Park &
	Inhibitors of cell cycle control kinases	Levine, 2000)
	(cancer)	(Nakai et al., 2005)
	Regulate lipid profile in diabetic rats	(Hsu & Yen, 2006)
	Regulate serum glucose	(Liu et al., 2012)
Flavonols	Reduce apoptosis in cell culture	(Fontana Pereira et al., 2011)
	Hepatoprotective action	(Jang et al., 2011)
	Promote new bone formation	(Singab et al., 2010)
	Anti-inflammatory effect	(Yang et al., 2010)
	Reduce neuronal damage	(Mahat et al., 2010)
		(Lagoa et al., 2009)
		(Hirose et al., 2009)
	Alpha-glucosidase inhibitor (diabetes)	
	GLUT inhibitor (diabetes)	(Kim et al., 2000)
	Improves cholesterol regulation	(Kwon et al., 2007; Song et
	(diabetes)	al., 2002)
Isoflavonoids	Inhibitor of tyrosine kinase and	(Lee, 2006)
	antiinflammatory effect in kidney	(Elmarakby et al., 2011)
	Induce apoptosis in leukemia	(Li et al., 2011a)
	Neuroprotective action	(Xi et al., 2011)
	Antiinflammatory action	(Neelakandan et al., 2011)
Anthocyanins	Pancreatic lipase and glucosidase	(Kim et al., 2000)
	5	

Flavonoid	Nutraceutic application	Reference
	inhibitor (diabetes)	(Tsuda, 2008)
	Regulate adipocyte function (Obesity)	(Wolfram et al., 2006)
	Improves glucose and lipid metabolism	(Sternberg et al., 2008)
	(diabetes)	(Tokimitsu, 2004)
	Modulate blood hormone levels	(Mirshekar et al., 2010)
	(multiple sclerosis)	(Wang et al., 2010)
	Suppress body fat accumulation(obesity)	(Roghani et al., 2010)
	Reduce neuropathic hyperalgesia in	(Cvorovic et al., 2010)
	diabetic rats	
	Antioxidant effect	
	Neuroprotective action	
	produce cytotoxicity in colon cancer cells	

Table 1. Nutraceutical applications of flavonoids.

The mechanism of bioavailability and metabolism of particular flavonoids has been demonstrated in mammals. In general it has been shown that flavonoid absorption and metabolism occurs in a common pathway and it begins in the stomach and intestinal tract. In the small intestine flavonoids pass into the bloodstream in the form of glycosides, though esters or polymers cannot be absorbed. Some intestine cell enzymes or microorganisms of microflora hydrolyze them to be absorbed. In the bloodstream there are different thermodynamic pathways. They could interact with cells to modify intracellular communication. The polyphenols can be conjugated in the intestine or liver to form methylated, glucuronidated, or sulphated metabolites that reach the body via urinary and biliary excretion. The microflora also metabolizes some metabolized flavonoids that are secreted in the bile into the small intestine. Thus, there is a recycling of polyphenols that allow them more time in the plasma (Erdman et al., 2007; Manach et al., 2004). In general, the microalgae produce low quantities of polyphenols. For this reason, in the following parts of this chapter we give special attention to pigments and PUFAs.

# 4. Nutraceutical effects of terpenes

The terpenes are other secondary metabolites that have nutraceutical properties. The terpenes are not only the largest group of plant natural products, comprising at least 30,000 compounds, but also contain the widest assortment of structural types. Hundreds of different monoterpene (C10), sesquiterpene (C15), diterpene (C20), and triterpene (C30) carbon skeletons are known. The wealth of terpene carbon skeletons can be attributed to an enzyme class known as the terpene synthases (EC 4.2.3.20). These catalysts convert the acyclic prenyl diphosphates and squalene into a multitude of cyclic and acyclic forms. The chief causes of terpene diversity are the large number of different terpene synthases and that some terpene synthases produce multiple products. An excellent review of terpene synthase and the diversity of products were published by Degenhard and coworkers (Degenhardt et al., 2009). Microalgae produce terpenes in the form of carotenoids. These compounds offer therapeutic effects. Carotenoids are tetraterpenoid organic pigments that are naturally occurring in the chloroplasts and chromoplasts of photosynthetic organisms. The use of carotenoids by animals is because they cannot synthetize them. Animals obtain carotenoids in their diets, and they may employ them in various ways in their metabolism.

There are over 600 known carotenoids and they are divided into two classes, xanthophylls (that contain oxygen) and carotenes (that are purely hydrocarbons and contain no oxygen). Carotenoids in general absorb blue light. They serve two key roles in plants and algae; they absorb light energy for use in photosynthesis and they protect chlorophyll from photodamage (Armstrong & Hearst, 1996).

The biosynthesis of carotenes is explained in figure 2. The carotenogenesis differ somewhat among organisms and the current knowledge on the biosynthesis of carotenoids has been gained mainly from studies of bacteria and vascular plants (Armstrong & Hearst, 1996). In Figure 2, we proposed the model of Lohr for the carotenogenesis in *Chlamydomonas*. This is probably related to other microalgae of Chlorophyceae class (Lohr et al., 2005; Lohr, 2008). There are other major divisions in different organisms, such as diatoms (Bertrand, 2010) or plants (Cazzonelli & Pogson, 2010; Zhu et al., 2010), which references the readers can check to deepen their knowledge in this area.

There has been much interest in carotenoids, especially their effect on human health, because they have a market value of several hundred million Euros. Their chemical synthesis is still a demanding challenge for chemists. The major dietary source of vitamin A for mammals, including humans, is derived from carotenoids. Vitamin A is an essential micronutrient for cell growth, embryonic development, vision, and the function of the immune system (Jackson et al., 2008).

In general carotenoids exert their mechanism on health via an antioxidant pathway or by modulating intracellular communication.

- 1. **Antioxidant properties:** This property of carotenoids was characterized by the ability to quench singlet oxygen, the inhibition of peroxide formation, and the correlation of antioxidant dependency with oxygen partial pressures. The ketocarotenoids, such as astaxanthin and canthaxanthin, were the best radical scavengers that did not contain conjugated terminal carbonyl functions (see figure 2). These findings suggest that the keto function in conjugation with the polyene backbone is able to stabilize carboncentered radicals more effectively than the polyene backbone alone (Jackson et al., 2008).
- 2. **Modulation of intracellular communication:** Carotenes modulate the intracellular communication because they or their metabolites interact with nuclear receptors like the pregnant-X-receptor (PXR) or retinoic acid receptor (RAR). For PXR it has been postulated that β-carotene activated the PXR more than its metabolites. Following this pathway, the β-carotene-PXR enhanced the metabolism of xenobiotics, bile acids, and retinoids (Ruhl, 2005). The carotenoids can be converted into two molecules of 9-cisretinal, which is oxidized to 9-cis-retinoic acid. The RXR binds the 9-cis-retinoic acid with high affinity to modulate cell functions (Heyman et al., 1992). Carotenoids like lycopene modulate mevalonate and Ras pathways to modify cell growth inhibition of cancerous cells (Palozza et al., 2010), and it changes Wnt and hedgehog proteins in those cells (Sarkar et al., 2010). The PI3K-Akt and MAPK pathways are stimulated in kidney by lycopene (Chan et al., 2009).

In table 2, are some nutraceuticals of the most used carotenoids.

Fig. 2. Putative pathways of carotenoid biosynthesis in Chlamydomonas. Hypothetical zygospore-specific pathways are indicated by dotted arrows. For the enzymes of the pathways only abbreviations are given. DXS (1-deoxy-D-xylulose-5-phosphate synthetase, EC 2.2.1.7). DXR (1-deoxy-D-xylulose-5-phosphate reductoisomerase, EC 1.1.1.267). CMS (4diphosphocytidil-2-C-methyl-D-erythriol synthase, EC 2.7.7.60). CMK (4-diphosphocytidil-2-C-methyl-D-erythriol kinase, EC 2.7.1.14.8). MCS (2-C-methyl-D-erythritol-2,4cyclophosphate synthase, EC 4.6.1.12). HDS (4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase, EC 1.17.7.1). IDS (isopentenyl dimethylallyl diphosphate synthase, EC 1.17.1.2.3). IDI (isopentenyl-diphosphate delta-isomerase, EC 5.3.3.2). GGPPS (geranylgeranyldiphosphate synthase, EC 2.5.1.81). PSY (phytoene synthase, EC 2.5.1.32). PDS (phytoene desaturase, EC 1.3.5.5). Z-ISO ( $\zeta$ -Carotene isomerase); ZDS ( $\zeta$ -carotene desaturase, EC 1.3.5.6). CRTISO (carotenoid isomerase, EC 5.2.1.13). LCYB (lycopene-β-cyclase). LCYE (lycopene-ε-cyclase), CHYB (carotene-β-hydroxylase, EC 1.14.13.-). CYP97A5 (carotene-βhydroxylase, EC 1.14.13.129). CYP97C3 (carotene-ε-hydroxylase, EC 1.14.99.45). ZEP (zeaxanthin epoxidase, EC 1.14.13.90). VDE (violaxanthin epoxidase, EC 1.10.99.3). NSY (neoxanthin synthase, EC 5.3.99.9). LSY (loroxanthin synthase), and BKT (carotene-βketolase).

Carotenoid	Nutracetical effect	Reference
	Antimutagenic effect	(Polivkova et al., 2010)
	Neuroprotective action	(Sandhir et al., 2010)
	Nephroprotective action	(Sahin et al., 2010)
	Prevent preclampsia	(Banerjee et al., 2009)
	Reduce risk of hip fracture	(Sahni et al., 2009)
	Antioxidant effect	(Erdman et al., 2009)
O O	Reduce eosinophil influx in asthma	(Wood et al., 2008)
_	Cardioprotective effect against doxorubicin-	(Anjos Ferreira et al.,
a –	caused damage	2007)
Q	Reduce inflammatory cytokines expression in pancreatitis	(Kim, 2011a; Kim, 2011b)
Lyco	Inhibit the growth and progression of colon cancer	(Tang et al., 2011)
L \	Enhanced antioxidant enzymes and immunity function in gastric cancer	(Luo & Wu, 2011)
	Inhibit NFκB-modulated IL-8 expression in macrophages-cigarette activated	(Simone et al., 2011)
	Reduced oxidative stress in allergic rhinitis	(Li et al., 2011b)
	Attenuated endothelial dysfunction in	(Zhu et al., 2011)
	diabetes	,
	Reduce cognitive decline in Parkinson's disease	(Kaur et al., 2011)
	Reduces LDC cholesterol and systolic blood	(Ried & Fakler, 2011)
	Pressure Reduces endothelial dysfunction in diabetic	(Zhao et al., 2011)z
٢	rats	(Zhao et al., 2011)Z
	Produces anxiolytic-like effects in mice	(Nishioka et al., 2011)
n t h i	Reduces oxidative stress and mitochondrial	(Lee et al., 2011)
Ţ	dysfunction in brain due MPTP/MPTP+	(Eee et al., 2011)
a Z	Reduce IL-6 microglia production	(Kim et al., 2010)
×	Reduce blood pressure in hypertensive rats	(Monroy-Ruiz et al., 2011)
o Z	Neuroprotective action against focal ischemia	(Lu et al., 2010)
<b>-</b>	Attenuate thrombosis	(Khan et al., 2010)
s t	Reduce retinal injury in elevated intraocular	(Cort et al., 2010)
Ϋ́	pressure	
•	Reduce UVA – induced skins photoaging	(Suganuma et al., 2010)
	Hepatoprotective action	(Curek et al., 2010)

Table 2. Nutraceutical application of lycopene and astaxanthin.

Carotenoids are lipid soluble and in general they follow the same absorption pathway as lipids, however other mechanisms of absorption have been proposed. To learn more, read the review of Kotake-Nara and Nagao (Kotake-Nara & Nagao, 2011). Once in the bloodstream, carotenes are fundamentally ligated to low density lipoprotein (LDL) whereas the xanthophylls are more evenly distributed between high density lipoproteins (HDL) and

low density lipoproteins (LDL). Nonpolar carotenoids (lycopene,  $\alpha$ -carotene,  $\beta$ -carotene) are located in the hydrophobic core and the polar (xanthophylls) would be, at least in part, on the surface of lipoproteins (Furr & Clark, 1997). For the microalgae, carotenoids are synthesized in high concentrations under several different environmental conditions, and humans exploited these as nutraceuticals in food.

## 5. Nutraceutical effects of chlorophylls, PUFA and other vitamins

There are other components in microalgae that could modulate redox environment to prevent oxidative stress and can affect intracellular communication. These components are chlorophyll, PUFAs, and vitamins such as vitamin A, B, C, and E.

Microalgae, like all chloroplast-containing photosynthetic eukaryotes, synthesize chlorophyll pigments. In Chlorophyceae chorophylls a and b are the most predominant. The chlorophylls have a porphyrin ring structure similar to heme, but with a central nonreactive magnesium ion instead of iron. To review chlorophyll biosynthesis in microalgae, read the chapter of Beale (Beale, 2008). The information about the biological activities of chlorophyll as nutraceuticals is scarce. They do have antipoliferative (Wu et al., 2010) and antioxidant (Serpeloni et al., 2011) activities. The chlorophyllin-cooper complex, a water-soluble commercial version of chlorophyll, possesses antimutagenic (Chernomorsky et al., 1997) and anticancer activities (Chernomorsky et al., 1997). The other components of microalgae; PUFAs, and vitamins A, B, C, and E, could be a nutraceutical because there is much evidence of how they modulate intracellular signals and act as antioxidants.

#### 6. Chlorella genus as nutraceutic

Chlorella species are encountered in all water habitats having cosmopolitan occurrences. The species of this genus have a simple form, a unicellular green alga belonging to the Chlorophyceae family. The *Chlorella* sp. is morphologically classified into four types; a) spherical cells (ratio of the two axes equals one), b) ellipsoidal cells (ratio of the longest axis to the shorter axis 1.45 to 1.60), spherical or ellipsoidal cells, and globular to subspherical cells. Their reproduction is asexual. Each mature cell divides usually producing four or eight (and more rarely 16) autospores, which are freed by rupture or dissolution of the parental walls.

Our research group has used *Chlorella vulgaris* as nutraceutical, particularly against mercury-caused oxidative stress and renal damage. For that we used male mice that were assigned into six groups; 1) a control group that received 100 mM phosphate buffer (PB) ig and 0.9% saline ip, 2) PB + HgCl<sub>2</sub> 5 mg/kg ip, 3) PB + 1000 mg/kg *Chlorella vulgaris* ig, and three groups receiving HgCl<sub>2</sub> + 250, 500, or 1000 mg/kg *Chlorella vulgaris* ig. The administration of the microalgae or PB was made 30 min before saline or HgCl<sub>2</sub> for 5 days. Our results demonstrated that HgCl<sub>2</sub> caused oxidative stress and cellular damage, whereas *Chlorella vulgaris* administration prevents oxidative stress (figure 3) and cellular damage (figure 4) in the kidney (Blas-Valdivia et al., 2011). We proposed that *Chlorella vulgaris*'s carotenes play an important role in preventing HgCl<sub>2</sub>-caused lipid peroxidation. Carotenes have a wide pharmacological spectrum of effects. The inhibition of lipid peroxidation may

be caused by the free radical scavenging property of these compounds (Miranda et al., 2001). Carotenes can scavenge singlet oxygen and they terminate peroxides by their redox potential because of the hydroxyl group in its structure. Thus, the ROS-steady state is maintained in the kidney damage lower than in animals with mercury intoxication. The biochemical behavior of this microalgae against mercury-caused oxidative stress is similar to the purified component of cyanobacteria such as *Pseudoanabaena tenuis* (Cano-Europa et al., 2010) or *Spirulina maxima* (Sharma et al., 2007).

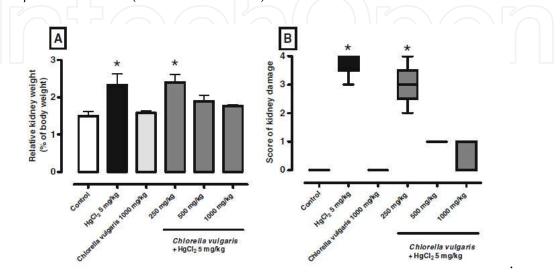


Fig. 3. Quantification of relative kidney weight (A) and the score of kidney damage (B) of mice treated with  $HgCl_2$  and *Chlorella vulgaris*. In A each bar represents the mean  $\pm$  S.E.M. In B each box represents the median  $\pm$  intercuartilic space.\* P < 0.05 vs. control. Author right permission. Springer ©.

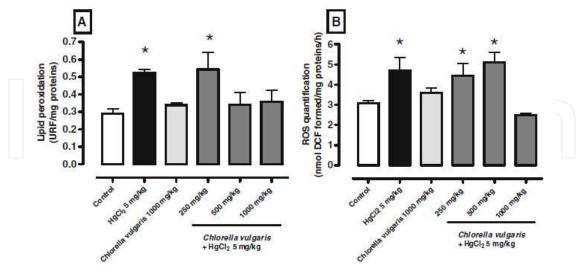


Fig. 4. Quantification of lipid peroxidation (A) and reactive oxygen species in the kidneys of mice treated with HgCl<sub>2</sub> and *Chlorella vulgaris*. Bar represents the mean  $\pm$  S.E.M.\* P < 0.05 vs. control. Author right permission. Springer ©.

Here are some experiments that demonstrated the nutraceutical use of *Chlorella* (Table 3).

Study	Evidences
The administration of <i>Chlorella</i> sp.	Chlorella administration inhibits bacterial
reduces endotoxemia, intestinal	culture and it avoids oxidative stress.
oxidative stress and bacterial traslocation	
in experimental biliary obstruction	
(Bedirli et al., 2009)	
Hot water extract of <i>Chlorella vulgaris</i>	The extract of <i>Chlorella vulgaris</i> inhibited
induced DNA damage and apoptosis	DNA synthesis, causing apoptosis and it
(Yusof et al., 2010)	increases p53, caspase-3, and Bax expression
	in hepatoma cells (HEpG2)
Attenuating effect of Chlorella	Chlorella supplementation decreases the
supplementation on oxidative stress and	NFκB activation and superoxide anion
NFκB. Activation in peritoneal	production and because it increases SOD
macrophages and liver of C57BL/6 mice	and catalase activity
fed on atherogenic diet (Lee et al., 2003)	
Chlorella accelerates dioxin excretion in	Chlorella enhanced dioxin metabolism and
rats (Morita et al., 1999)	excretion by feces
Effect of <i>Chlorella</i> and its fractions on	A Chlorella supplemented diet decreases
blood pressure, cerebral stroke lesions,	blood pressure and the incidence rate of
and life-span in stroke-prone	cerebral stroke in SHRSP.
spontaneously hypertensive rats	
(Sansawa et al., 2006)	
Hypocholesterolemic mechanism of	Chlorella powder increases the expression of
Chlorella: Chlorella and its indigestible	CYP7A1, a limiting enzyme of the main
fraction enhance hepatic cholesterol 7α-	pathway of the cholesterol catabolism,
hydroxylase in rats (Shibata et al., 2007)	lowering the concentration of LDL in plasma
Chlorella vulgaris triggers apoptosis in	Chlorella vulgaris inhibits the anti-apoptotic
hepatocarcinogenesis-induced rats	protein Bcl-2
(Mohd Azamai et al., 2009)	
Effect of Chlorella vulgaris on lipid	Chlorella vulgaris decreases HDL cholesterol
metabolism in Wistar rats fed high fat	concentration by a reduction in the intestinal
diet (Lee et al., 2008)	absortion
Antioxidant effect of the marine algae	Chlorella vulgaris inhibits production of free
Chlorella vulgaris against naphthalene-	radicals, decreasing lipoperoxidation, and
induced oxidative stress in the albino rats	increasing the activity of antioxidant
(Vijayavel et al., 2007)	enzymes as SOD, catalase, GPX and reduced
	glutathione, preventing from the toxicity of
	naftalene
Six-week supplementation with <i>Chlorella</i>	Chlorella supplement exhibits antioxidant
has favorable impact on antioxidant	activity decreasing ROS and increasing the
status in Korean male smokers (Lee et al.,	activity of SOD and catalase
2010)	
Chlorella pyrenoidosa supplementation	Chlorella pyrenoidosa exhibits an
reduces the risk of anemia, proteinuria	antiinflammatory activity regulated by
and edema in pregnant women (Nakano	cytokine. It increased the production of IL-10
et al., 2010)	

Study	Evidences
Effect of Chlorella intake on cadmium	Chlorella inhibits cadmium absorption and it
metabolism in rats (Shim et al., 2009)	promotes the excretion through the feces.
	Also, it stimulates the production of
	metallothionein in the small intestine.
Isolation of phophorylated	The Chlorella polysaccharides increases the
polysaccharides from algae: the	production of NO in macrophages
inmmunostimulatory principle of	enhancing the innate immune response,
Chlorella pyrenoidosa (Suarez et al., 2010)	mediated by Toll-like receptors (TLR-4)
Influence of Chlorella powder intake	Chlorella vulgaris exhibits an antioxidant
during swimming stress in mice	activity, reducing the lipoperoxidation,
(Mizoguchi et al., 2011)	avoiding the DNA damage. However it does
	not show hypoglycemic activity

Table 3. Nutraceutical evidences of Chlorella.

# 7. Chlamydomonas genus as nutraceutic

Chlamydomonas spp. are unicellular algae with cell walls and with either two or four flagella. The genus Chlamydomonas is of worldwide distribution and is found in a diversity of habitats including temperate, tropical, and polar regions. Chlamydomonas species have been isolated from freshwater ponds and lakes, sewage ponds, marine and brackish waters, snow, garden and agricultural soil, forests, deserts, peat bogs, damp walls, sap on a wounded elm tree, an artificial pond on a volcanic island, mattress dust in the Netherlands, roof tiles in India, and a Nicaraguan hog wallow. These algae belong to the family Chlamydomonadaceae that consists of approximately 30 genera. DNA sequence analysis clearly demonstrates, however, that this family is composed of multiple phylogenetic lineages that do not correspond to the morphologically defined genera. Although the identities of the species are uncertain, it is noteworthy that the traits in which they differed included body shape, thickness of the cell wall, presence or absence of the apical papilla, lateral vs. basal position of the chloroplast, chloroplast position, and shape of the eyespot, all of which were later used as criteria to separate species. Although cell-body shape and size vary among Chlamydomonas species (as defined by morphological criteria), the overall polar structure, with paired apical flagella and basal chloroplast surrounding one or more pyrenoids, is constant. Cells are usually free-swimming in liquid media but on solid substrata may be nonflagellated, and are often seen in gelatinous masses similar to those of the algae Palmella or Gloeocystis in the order Tetrasporales. This condition has been referred to as a palmelloid state. Some species may also form asexual resting spores, or akinetes, in which the original vegetative cell wall becomes much thicker, and carotenoids, starch, and lipids may accumulate (Harris et al., 2008).

Our group has studied the nutraceutical properties of *Chlamydomonas gloeopara*, a microalgae collected from a eutrophic reservoir (La Piedad Lake) in Cuautitlan Izcalli, Mexico. That reservoir is located at 19°39′N (latitude) and 99°14′W (longitude). Our research group has used *Chlamydomonas gloeopara* as a nutraceutical, particularly against mercury-caused oxidative stress and renal damage. For that we used male mice that were assigned into six groups; 1) a control group that received 100 mM phosphate buffer (PB) ig and 0.9% saline ip, 2) PB + HgCl<sub>2</sub> 5 mg/kg ip, 3) PB + 1000 mg/kg *Chlamydomonas gloeopara* ig, and three

groups receiving HgCl<sub>2</sub> + 250, 500, or 1000 mg/kg *Chlamydomonas gloeopara* ig. The administration of the microalgae or PB was made 30 min before saline or HgCl<sub>2</sub> for 5 days. Our results demonstrated that *Chlamydomonas gloeopara* as well as *Chlorella* prevents renal damage (figure 5, panel A-F) by reducing the oxidative stress of lipid peroxidation (figure 5, panel G).

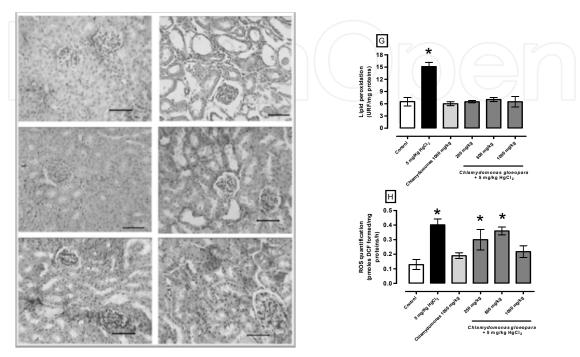


Fig. 5. Effect on *Chlamydomonas gloeopara* administation on  $HgCl_2$ -caused renal damage (panel A-F) and oxidative stress (panel G and H). Photomicrographs of renal cortex . Panel A shows control group. Panel B shows group treated with  $HgCl_2$ . Panel C shows group treated with *Chlamydomonas gloeopara* 1000 mg/kg . Panels D, E and F show groups treated with *Chlamydomonas gloeopara* 250, 500 and 1000 mg/kg plus  $HgCl_2$ . The tissue was stained by hematoxylin-eosin. Treatment with  $HgCl_2$  causes cell atrophy, hyperchromatic nuclei, and edema. Histological alterations were partially ameliorated in groups treated with *Chlamydomonas gloeopara*. *Chlamydomonas gloeopara* administration reduced lipid peroxidation (G) and reactive oxygen species (H) in the kidneys of mice treated with  $HgCl_2$  and *Chlorella vulgaris*. Bar is the mean  $\pm$  SE \* P < 0.05 vs. control.

#### 8. Haematococcus genus as nutraceutic

Haematococcus are green microalgae; single-celled aquatic organisms. It is known that Haematococcus is the primarily source of astaxanthin, a ketocarotenoid that is a natural nutritional component. In the marine environment, astaxanthin is biosynthesized in the food chain, within the microalgae or phytoplankton, at the primary production level. When these algae are exposed to harsh environmental conditions and ultraviolet light, they accumulate the highest level of astaxanthin and in this process, the algae become blood red. Astaxanthin accumulates 2% to 3% of dry weight and constitutes 85% to 88% of the total carotenoids. Chemically it is a ketocarotenoid (3,3′-dihydroxy- $\beta$ ,  $\beta$ -carotene-4,4′dione) and is the principal pigment of salmonoids and shrimp. Astaxanthin has a higher antioxidant activity

than lutein, lycopene,  $\alpha$  or  $\beta$ -carotene, and  $\alpha$ -tocopherol. Astaxanthin has 100 times and 10 times greater antioxidant activity than vitamin E and  $\beta$ -carotene (Guerin, 2003).

Morphological studies have shown that the algae have a life cycle. The, green vegetative cells with two flagellae grow autotrophycally in the light and heterotrophically in the dark. In culture, H. pluvialis has the typical characteristics of a motile stage, with biflagellate spherical cells. The growth in a bioreactor, with mechanical stirring, favors the occurrence of more or less mature aplanospores. This stage becomes dominant together with the evolution of growth. The aplanospore color turns gradually red, because of the accumulation of carotenoids in the chloroplast, and especially outside of them in lipid globules (astaxanthin). The red aplanospores are known as haematocysts. This stage may appear under stress conditions caused by light, high temperature, increased salinity, nutritional limitation, or change of carbon source. During the growth stage, the cells with a diameter of 30 µm were spherical to ellipsoid and enclosed by a cell wall. The cells had 2 flagellae of equal length emerging form an anterior papilla. As they age, the cells ceased to be mobile, yet the cellular structure remained the same without the flagellae. Under stress conditions, the volume of the cells increased to a diameter of  $> 40 \mu m$  and the cell wall became resistant. The maturation of the cyst cells was accompanied by enhanced carotenoid biosynthesis and a gradual change in cell color to red. When the cystic cells were transferred to optimal growth conditions, daughter cells were released from the cystic cells, and then vegetative cells regenerated from daughter cells (Cysewski & Todd Lorenz, 2004).

*Haematococcus* has the potential as a nutraceutical because there is various evidence of this. In table 4, we show some articles that employed *Haematococcus* or its astaxanthin.

Study	Evidences
Haematococcus astaxanthin: applications	This is a review about the uses of astaxantin
for human health and nutrition (Guerin,	from Haematococcus in health
2003)	
Optimization of microwave-assisted	The extracts have a high antioxidant
extraction of astaxanthin from	capacity, inhibit peroxidation of linoleic acid,
Haematococcus pluvialis by response	and neutralize free radicals
surface methodology and antioxidant	
activities of the extracts (Zhao et al., 2009)	
Cardioprotection and myocardial salvage	The astaxanthin is an antioxidant,
by a disodium disuccinate astaxanthin	antiinflammatory, and cardioprotective.
derivative (Cardax <sup>TM</sup> ) (Gross &	reducer of levels of nitric oxide, tumor
Lockwood, 2004)	necrosis factor alpha, and prostaglandin E2
Ulcer preventive and antioxidative	The astaxanthin exerts its gastroprotection of
properties of astaxanthin from	gastric ulceration by activation of
Haematococcus pluvialis (Kamath et al.,	antioxidant enzyme such as catalase,
2008)	superoxide dismutase, and glutathione
	peroxidase. It inhibits the activity pump Na-
	K ATPase
Safety assessment of astaxanthin-rich	The administration of astaxanthin has no
microalgae biomass: acute and	adverse effects
subchronic toxicity studies in	

Study	Evidences
rats(Stewart et al., 2008)	
Astaxanthin, a carotenoid with potential	The antihypertensive and neuroprotective
in human health and nutrition (Hussein	potentials of the compound
et al., 2006).	
Protective effects of Haematococcus	The results suggest that ASX
astaxanthin on oxidative stress in healthy	supplementation might prevent oxidative
smokers (Kim et al., 2011).	damage in smokers by suppressing lipid
	peroxidation and stimulating the activity of
	the antioxidant system in smokers
Astaxanthin-rich extract from the green	It results suggest that supplementation of
alga Haematococcus pluvialis lowers	astaxanthin-rich Haematococcus extract
plasma lipid concentrations and enhances	improves cholesterol and lipid metabolism
antioxidant defense in apolipoprotein E	as well as antioxidant defense mechanisms,
knockout mice (Yang et al., 2011)	all of which could help mitigate the
	progression of atherosclerosis.

Table 4. Nutraceutical evidences of *Haematococcus*.

## 9. Dunaliella genus as nutreutic

Dunaliella salina is a unicelular green alga belonging to the Chlorophyceae family. Dunaliella cells are ovoid, spherical, pyriform, fusiform, or ellipsoid with sizes varying from 5 to 25 µm in length and from 3 to 13 μm in width. The cells also contain a single chloroplast, which mostly has a central pyrenoid surrounded by starch granules. Dunaliella multiplies by lengthwise division, but sexual reproduction does occur rarely by isogametes with a conjugation process. It proliferates in extremely varied salinities from 0.5 to 5.0 M NaCl. The alga cells do not contain a rigid cell wall; instead a thin elastic membrane surrounds them. It is known to accumulate carotenoids under various stress conditions. It possesses a remarkable degree of environmental adaptation by producing an excess of β-carotene and glycerol to maintain its osmotic balance.  $\beta$ -carotene occurs naturally as its isomers, namely, all-trans, 9-cis, 13-cis, and 15-cis forms and functions as an accessory light harvesting pigment, thereby protecting the photosynthetic apparatus against photo damage in all green plants including algae. β-carotene, a component of the photosynthetic reaction center is accumulated as lipid globules in the interthylakoid spaces of the chloroplasts of Dunaliella. They protect the algae from damage obtained during excessive irradiance by preventing the formation of reactive oxygen species, by quenching the triplet-state chlorophyll, or by reacting with singlet oxygen (1O<sub>2</sub>) and also act as a light filter (Ben-Amotz, 2004). Dunaliella nutraceutical properties are shown in table

Study	Conclusion
<i>In vivo</i> antioxidant activity of carotenoids	Carotenoids provide protection against
from Dunaliella salina a green microalga	CCl <sub>4</sub> -caused hepatic damage by restoring
(Chidambara-Murthy et al., 2005)	the activity of hepatic enzymes like
	peroxidase, super oxide dismutase, and
	catalase, which reduce ROS and lipid
	peroxidation.

Study	Conclusion
9-cis β-carotene-rich powder of the alga	Dunaliella treatment increases plasma
Dunaliella bardawil increases plasma HDL-	HDL-cholesterol and lower plasma
cholesterol in fibrate-treated patients	triglyceride levels
(Shaish et al., 2006)	
Ethanol extract of Dunaliella salina induces	Ethanol extract of <i>Dunaliella salina</i> inhibits
cell cycle arrest and apoptosis in A545	cell proliferation and causes apoptosis
human non-small cell lung cancer cells	possibly via p53 and p21 promoting the
(Sheu et al., 2008)	protein expression of Fas and FasL
Protective effects of Dunaliella salina against	The <i>chlorophyta</i> has a protective effect
experimental induced fibrosarcoma on	against experimentally caused
Wistar rats (Raja et al., 2007).	fibrosarcoma
Bioavailability of the isomer mixture of	9-cis phytoene has a stronger antioxidative
phytoene and phytofluene-rich alga	effect than the all trans isomer
Dunaliella bardawil in rat plasma and tissues	
(Werman et al., 2002).	
Hypercholesterolemia induced oxidative	Dunaliella salina components inhibit lipid
stress is reduced in rats with diets enriched	peroxidation and also increases Type1 5'-
with supplement from Dunaliella salina	iodothyronine deiodinase (5´-DI)
algae (Bansal & Sapna, 2011).	expression, which leads to a T <sub>3</sub> level
	increase
Evaluation of carotenoid extract from	Carotenoid extract of Dunaliella salina
Dunaliella salina against cadmium-induced	contains abundant <i>cis</i> and <i>trans</i> β-
cytotoxicity and transforming growth	carotenes. These antioxidants decrease the
factor β1 induced expression of smooth	lipid peroxidation and also inhibit
muscle α-actin with rat liver cell lines (Jau-	activation of hepatic stellate cells (HSCs).
Tien et al., 2011).	
Protective effects of Dunaliella salina- a	Carotenoids of <i>D. salina</i> inhibit the lipid
carotenoids-rich alga, against carbon	peroxidation and increases the antioxidant
tetrachloride-induced hepatotoxicity in	enzyme activity
mice (Hsu et al., 2008).	

Table 5. Nutraceutical evidences of Dunaliella.

#### 10. Final remarks

The functional food and nutraceutical market is growing. However, to promote health the active compounds must be ingested in high concentration. This is a great problem because sometimes the components such as carotenoids, polyphenols, and chlorophylls are extracted from vegetables or plants. In their production, we are modifying the environment, thus the use of biotechnology of microalgae or other microorganisms like bacteria or fungus could be an alternative because they may be environmentally friendly. The sun can be used as energy source and the medium could be fresh or sea water, with the carbon source as CO<sub>2</sub> and other inorganic or organic sources. In this chapter we show the evidence of some genera, particularly of Chlorophyceae class as *Chlorella*, *Chlamydomonas*, *Haematococcus*, and *Dunaliella*. It is evident that their components modulate intracellular communication and they act as antioxidants.

There are many microalgae never used as nutraceuticals that could be used for human or animal health, such as the microalgae used in aquaculture to fed shrimp and fish. Examples of those kinds of microalgi are *Pavlova* and *Tetraselmis* that produce high concentration of PUFAs.

## 11. Acknowledgement

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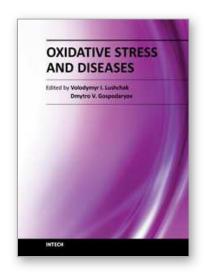
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#### **Oxidative Stress and Diseases**

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The development of hypothesis of oxidative stress in the 1980s stimulated the interest of biological and biomedical sciences that extends to this day. The contributions in this book provide the reader with the knowledge accumulated to date on the involvement of reactive oxygen species in different pathologies in humans and animals. The chapters are organized into sections based on specific groups of pathologies such as cardiovascular diseases, diabetes, cancer, neuronal, hormonal, and systemic ones. A special section highlights potential of antioxidants to protect organisms against deleterious effects of reactive species. This book should appeal to many researchers, who should find its information useful for advancing their fields.

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# InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447

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#### InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元

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