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Published in:
Frontiers in Plant Science

DOI:
[10.3389/fpls.2014.00655](https://doi.org/10.3389/fpls.2014.00655)

Publication date:
2014

Document Version
Publisher's PDF, also known as Version of record

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Wiesel, L., Newton, A. C., Elliott, I., Booty, D., Gilroy, E. M., Birch, P. R. J., & Hein, I. (2014). Molecular effects of resistance elicitors from biological origin and their potential for crop protection. *Frontiers in Plant Science*, 5, [655]. [10.3389/fpls.2014.00655](https://doi.org/10.3389/fpls.2014.00655)

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Molecular effects of resistance elicitors from biological origin and their potential for crop protection

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Plants contain a sophisticated innate immune network to prevent pathogenic microbes from gaining access to nutrients and from colonizing internal structures. The first layer of inducible response is governed by the plant following the perception of microbe- or modified plant-derived molecules. As the perception of these molecules results in a plant response that can provide efficient resistance toward non-adapted pathogens they can also be described as “defense elicitors.” In compatible plant/microbe interactions, adapted microorganisms have means to avoid or disable this resistance response and promote virulence. However, this requires a detailed spatial and temporal response from the invading pathogens. In agricultural practice, treating plants with isolated defense elicitors in the absence of pathogens can promote plant resistance by uncoupling defense activation from the effects of pathogen virulence determinants. The plant responses to plant, bacterial, oomycete, or fungal-derived elicitors are not, in all cases, universal and need elucidating prior to the application in agriculture. This review provides an overview of currently known elicitors of biological rather than synthetic origin and places their activity into a molecular context.

Keywords: crop protection, elicitors, pathogen effectors, priming, disease resistance

THE ROLE OF DEFENSE ELICITORS IN PLANT IMMUNITY

Plants are under constant threat of microbial pathogen attack. Plant cell walls, cuticles and phytoanticipins are preformed, physical and chemical barriers that limit access of microbes to plant cells (Underwood, 2012; Newman et al., 2013). In addition to these non-inducible defenses, plants recognize and respond to defense elicitors which are signal-inducing compounds perceived by the innate immune system that prime and/or induce defense responses (Henry et al., 2012; Maffei et al., 2012; Newman et al., 2013). Elicitor compounds can be biological in origin, derived from either the plant or the microbe, or can be synthetically generated (Walters et al., 2013). We will focus on elicitors from biological origin (**Table 1**) rather than synthetic analogs of known signaling or defense molecules such as Bion, acibenzolar-S-methyl (ASM), beta-amino-butyric acid (BABA), and cis-jasmone. Elicitor activity has, for example, been shown for plant-derived cell wall components such as oligogalacturonides (Ferrari et al., 2013), proteinaceous pathogen molecules such as bacterial flagellin (Gomez-Gomez and Boller, 2002), oomycete-derived elicitin INF1 (reviewed in Hein et al., 2009) and non-proteinaceous molecules such as lipopolysaccharides (Erbs and Newman, 2012). However, intact plant- or microbe-derived structures as well as highly-polymerized molecules often tend to result in few recognition responses. In contrast, leakage of metabolites or even minor or partial breakdown of complex host or pathogen molecules leads to

the production of eliciting components that are biologically active.

The co-evolution between plants and potential microbial pathogens has been described as a zigzag model by Jones and Dangl (2006) and can also be applied to deducing the biological activity of elicitors (**Figure 1**). According to the zigzag model, the first inducible responses are a consequence of the perception of chemical elicitors, microbe-associated molecular patterns (MAMPs), pathogen-associated molecular patterns (PAMPs) and/or damage-associated molecular patterns (DAMPs). The latter are also known as danger-associated molecular patterns (Mazzotta and Kemmerling, 2011). MAMPs describe general microbe-derived molecules including those originating from beneficial microbes whereas PAMPs specifically describe molecules from pathogenic microbes such as fungi, oomycetes, and bacteria (Henry et al., 2012; Newman et al., 2013). Thus, PAMPs are a subgroup of MAMPs (Maffei et al., 2012). In contrast, DAMPs are typically plant-derived and are produced after, for example, wounding by insects or herbivores as well as degradation or perturbation of host molecules by microbes (Henry et al., 2012; Newman et al., 2013). All of these molecules, which could universally be described as “patterns that elicit immunity” (PEIs), are often recognized by transmembrane pattern recognition receptors (PRRs) in plant cells (Jones and Dangl, 2006; Maffei et al., 2012; Newman et al., 2013). Upon recognition of MAMP- or DAMP-derived patterns, PTI (PAMP- or

Table 1 | List of plant-, bacterial-, oomycete-, and fungal-derived elicitor compounds, their activity against pathogens and effectiveness in plants.

Origin	Elicitor compound	Effective toward	Plants effects shown in	References
Plant	Oligogalacturonides	<i>Botrytis cinerea</i> , <i>Blumeria graminis</i>	Several	Aziz et al., 2007; Randoux et al., 2010; Galletti et al., 2011
	Milsana (giant knotweed)	<i>Botrytis cinerea</i> , <i>Leveillula taurica</i>	Cucumber, tomato	Daayf et al., 1997, 2000; Konstantinidou-Doltsinis et al., 2006
	Burdock fructooligosaccharide	<i>Colletotrichum lagenarium</i> , <i>Botrytis cinerea</i> , TMV	Cucumber, tobacco, tomato	Wang et al., 2009; Guo et al., 2012
	Elicitor peptide 1 (Pep1)	<i>Cochliobolus heterostrophus</i> , <i>Colletotrichum graminicola</i>	Maize	Huffaker et al., 2011
	Carrageenans	<i>Sclerotinia sclerotiorum</i> , TMV	<i>A. thaliana</i> , tobacco	Sangha et al., 2010; Vera et al., 2011
	Fucans	TMV	Tobacco	Vera et al., 2011
	Ulvans	Several	Several	Jaulneau et al., 2011; Vera et al., 2011
	Laminarin	<i>Erwinia carotovora</i> , <i>Plasmopara viticola</i> , <i>Botrytis cinerea</i> , <i>Fusarium solani</i>	Beans, grapevine, tobacco	Craigie, 2011; Vera et al., 2011
Bacteria	Harpin	<i>Xanthomonas oryzae</i>	Rice	Lee et al., 2001; Li et al., 2012
	Lipopeptides	<i>Botrytis cinerea</i>	Tomato	Henry et al., 2012
	Dimethylsulfide	<i>Cochliobolus heterostrophus</i> , <i>Botrytis cinerea</i>	Maize, tobacco	Huang et al., 2012
	Pseudobactin	<i>Botrytis cinerea</i> , <i>Erwinia carotovora</i>	Several	De Vleeschauwer and Höfte, 2009
Oomycetes	CBEL	<i>Phytophthora parasitica</i>	<i>A. thaliana</i> , tobacco	Mateos et al., 1997; Khatib et al., 2004
	Cryptogein	<i>Phytophthora parasitica</i> , <i>Sclerotinia sclerotiorum</i>	Tobacco	Bonnet et al., 1996
	Eicosapentaenoic acid	<i>Phytophthora infestans</i>	Potato	Henriquez et al., 2012
	Pep-13	<i>Phytophthora</i> spp.	Parsley, potato	Nürnberg et al., 1994; Brunner et al., 2002; Parker, 2003
	INF1	<i>Phytophthora infestans</i>	Tobacco	Takahashi et al., 2007; Hein et al., 2009; Kawamura et al., 2009
Fungi	β -glucans	Several	Several	Hahn and Albersheim, 1978; Fu et al., 2011; Falcón-Rodríguez et al., 2012; Henriquez et al., 2012
	Chitosan	Several	Several	Kishimoto et al., 2010; Kombrink et al., 2011
	Chitin	Several	Several	El Ghaouth et al., 1994; Copping and Duke, 2007; El Hadrami et al., 2012
	Ergosterol	<i>Botrytis cinerea</i>	Grapevine, tobacco	Laquitaine et al., 2006; Vatsa et al., 2011
	<i>Trichoderma</i> species: xylanases, peptaibol, cerato-platanin family	<i>Pseudomonas syringae</i> , <i>Botrytis cinerea</i> , <i>Colletotrichum graminicola</i>	<i>A. thaliana</i> , cotton, maize	Ron and Avni, 2004; Djonović et al., 2007; Viterbo et al., 2007; Yang et al., 2009; de Oliveira et al., 2011
	Cerebrosides	<i>Fusarium</i> spp.	Several	Umemura et al., 2004
	HR-inducing protein	<i>Magnaporthe oryzae</i>	Rice	Chen et al., 2012; Kulye et al., 2012

(Continued)

Table 1 | Continued

Origin	Elicitor compound	Effective toward	Plants effects shown in	References
	PeaT1	TMV	Tobacco	Zhang et al., 2010, 2011b
	PebC1	<i>Botrytis cinerea</i>	Tomato	Zhang et al., 2010
	PevD1	TMV	Tobacco	Wang et al., 2012a,b
	PemG1	<i>Pseudomonas syringae</i> , <i>Xanthomonas oryzae</i>	<i>A. thaliana</i> , rice	Qiu et al., 2009; Peng et al., 2011

pattern-triggered immunity) is activated in the plant and the perceived molecules could be described as immune elicitors. This defense reaction aims to restrict the growth of the intruder and can lead to systemic induced resistance leaving the plant less susceptible to subsequent pathogen attack (Henry et al., 2012).

Systemic induced resistance can be divided into systemic acquired resistance (SAR) or induced systemic resistance (ISR). Systemic acquired resistance is often characterized by localized necrosis, expression of pathogenesis related (PR) protein genes, and involves the salicylic acid (SA) pathway whereas ISR is often triggered by plant growth-promoting rhizobacteria (PGPR) (Walters et al., 2013), is not associated with necrosis and involves the jasmonic acid (JA) and ethylene (ET) pathways (Walters and Heil, 2007; Henry et al., 2012). Typical responses of PTI include cell wall alterations and the production of reactive oxygen species (ROS) which can be directly cytotoxic but also play a role in signaling. Other responses comprise the production of phytoalexins, expression of PR proteins, activation of mitogen activated protein kinase (MAPK) pathways, and defense signaling involving calcium (Ca^{2+}) influx from extracellular spaces and changes in free cytosolic Ca^{2+} concentrations (Garcion et al., 2007). To counteract the initial plant defense reaction, successful microbes have evolved specialized effectors that perturb recognition of defense elicitors or subsequent plant defense mechanisms to promote effector-triggered susceptibility (ETS). However, if these pathogen effectors are in turn recognized by cognate plant resistance (R) proteins, the second layer of inducible response, effector-triggered immunity (ETI), is initiated that often yields a hypersensitive resistance response (HR) (Jones and Dangl, 2006; Deslandes and Rivas, 2012).

The outcome of plant/microbe interactions can result in symbiosis, disease or disease resistance and is governed by further levels of sophisticated co-evolution. Indeed, it must be recognized that pathogen colonization of plants can generate dynamic pathogenic, mutualistic or parasitic interactions of varying magnitude and specificity. Furthermore, organisms recognized as pathogens in, for example, a crop context, could be benign or even beneficial in another context such as a different host or environment (Newton et al., 2010). It is thus essential for the plant to evaluate the scale of threat and to mount appropriate and proportionate responses. These may range from priming, being ready to respond faster to actual attack, or expression of PTI-based defense mechanisms to yield incompatibility if the microbe/pathogen is unable to suppress these responses. The use of elicitors in agriculture holds the potential to decrease the need

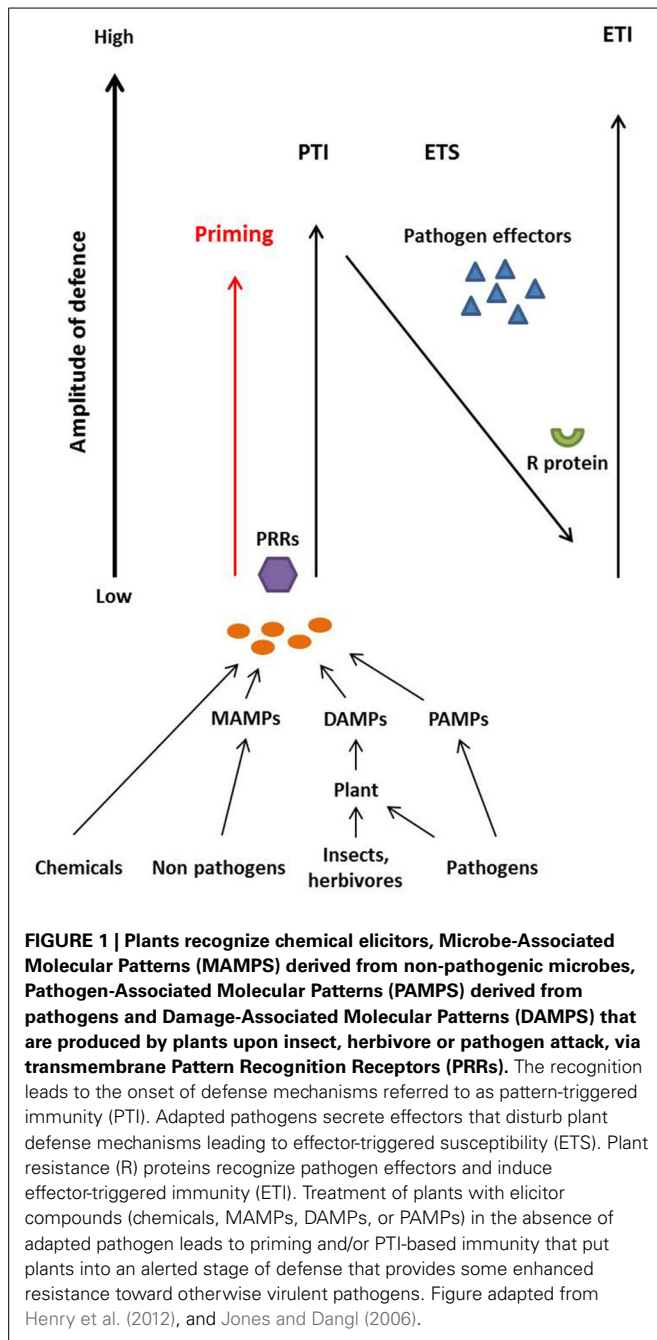
for pesticide application by using the plant's own defense system. However, there is a need to understand this process on a molecular level to maximize the efficacy of the treatments.

INDUCIBLE DEFENSE RESPONSE IN THE ABSENCE OF PATHOGENS

In agricultural practice, elicitor treatments of plants in the absence of virulent pathogens yields a defense response such as priming and/or PTI that is uncoupled from ETS and can provide some protection to subsequent pathogen challenges. Priming is defined as a physiological status of plants leading to faster and stronger activation of defense responses to subsequent biotic and abiotic stresses (reviewed in Conrath et al., 2006; Conrath, 2011; Pastor et al., 2013). Crucially, this is distinct from the level of resistance induction in response to the recognition of true pathogens that are potentially capable of causing disease and where recognition would cause resistance mechanism expression that is more costly to the plant but still proportionate to potential disease cost (Walters and Heil, 2007).

In primed plants, chromatin modifications in the form of methylation and acetylation of histones take place that impact on the interaction of DNA with histones and/or open binding sites for transcriptional co-activators such as WRKY22 and WRKY29 (Eulgem, 2005; Conrath, 2011; Po-Wen et al., 2013). These chromatin modifications in primed plants have been shown to lead to increased expression of transcription factors WRKY6, WRKY29, and WRKY53 after stress exposure (Jaskiewicz et al., 2011). In *Arabidopsis thaliana*, mRNA and inactive MPK3 and MPK6 accumulate in cells of elicitor-treated plants. Upon exposure to *Pseudomonas syringae* both MAP kinases are more strongly activated in primed plants than in non-primed plants (Beckers et al., 2009).

When PTI-associated mechanisms are primed by elicitor treatments plants often accumulate ROS and produce a stronger, secondary oxidative burst following pathogen challenge, activate MPKs and stimulate SA-, JA-, and abscisic acid (ABA)-pathways (Beckers et al., 2009; Pastor et al., 2013). Callose deposition, which is potentially also linked to the ABA-pathway, can be enhanced in elicitor-treated plants (Kohler et al., 2002; Flors et al., 2005; Pastor et al., 2013) and elicitor treatment often induces expression of phenylalanine ammonia lyase (PAL) which is required for the production of SA precursors (Chen et al., 2009). In line with SA involvement, pathogenesis-related genes such as PR-1, PR-2, and PR-5 have been implicated with elicitor treatments (Kohler et al., 2002; Conrath et al., 2006). Both priming and the activation



of defense mechanisms due to elicitor treatment can lead to a reduction of disease severity when subsequent pathogen attack occurs. Biologically active defense elicitors that either prime plant defenses or initiate PTI responses have been identified from diverse sources. Molecular studies have provided clues to their mechanism and to the processes that govern specificity.

A MOLECULAR PERSPECTIVE OF ELICITOR ACTIVITY IN PLANT IMMUNITY

Several studies have shown that elicitor-treated plants show lower infection rates following inoculation with virulent pathogens but responses can vary between plant species (Table 1). In addition

to the observed disease reduction, molecular studies are revealing how the elicitor compounds affect gene expression levels in plants and therefore impact on defense responses (Section Plant Genes and Pathways Involved in Elicitor Recognition). Similarly, the diverse mechanisms by which pathogen effectors suppress PTI responses are emerging but, due to the complexity of this research, only selected examples are highlighted in this review.

PLANT-DERIVED ELICITORS

Plant cell walls are composed of cellulose, hemicellulose (cross-linking glycans), pectic polysaccharides, protein, lignin, and a variety of lipids (Wei et al., 2009). Bacteria and fungi can produce cellulases, xylanases, and lignin peroxidases that break down plant cell wall components and common products are β -glucans, xylose, and phenylpropanoid-containing compounds. These break-down products function as plant-derived elicitors and several examples of disease reduction due to the application of plant-derived elicitors exist (Table 1). Well studied plant-derived elicitors include oligogalacturonides (OGs), which are structural components of plant cell walls and are released upon partial degradation of homogalacturonan by microbial polygalacturonases during infection or by plant polygalacturonases induced upon wounding (Ferrari et al., 2013). Plant cell wall-derived OGs are recognized by wall-associated kinase 1 (WAK1) and subsequent signaling is JA-, SA-, and ET-independent (Brutus et al., 2010; Ferrari et al., 2013). A MAP kinase cascade is triggered upon OG perception in *A. thaliana*, and MPK3 and MPK6 are phosphorylated. However, the importance of these signaling events remains elusive and it has been shown, for example, that lack of MPK3 increases basal susceptibility to *Botrytis cinerea* but elicitor-induced resistances are not affected (Galletti et al., 2011). In contrast, MPK6 is necessary for OG-induced resistance but does not play a role in basal resistance toward *B. cinerea* (Galletti et al., 2011).

BACTERIAL-DERIVED ELICITORS

In addition to plant-derived elicitors, the application of bacterial-derived elicitors has also been shown to reduce pathogen infection in plants (Table 1). Extracellular polysaccharides (EPS) produced by the bacterial wilt causing pathogen *Ralstonia solanacearum* have been shown to induce defense responses in tomato (Milling et al., 2011) and lipopolysaccharides (LPS) from Gram-negative bacteria also trigger induced resistance in several other plant species (Dow et al., 2000; Gerber et al., 2004; Desaki et al., 2006; Erbs and Newman, 2012). PGPRs (plant growth-promoting rhizobacteria) can induce resistance in plants by exuding elicitors (De Vleeschauwer and Höfte, 2009 and references therein) and filtrates from cultures of bacteria such as *Bacillus subtilis* can also elicit crop protection effects (Schönbeck et al., 1980, 1982), though these may be a combination of direct toxicity and/or elicitor recognition events.

The molecular background to bacterial MAMPS, effectors and their plant targets has been reviewed recently (Deslandes and Rivas, 2012) and two well-studied bacterial MAMPS are flagellin and the elongation factor Tu (EF-Tu). Flagellin is recognized in a variety of plant species whereas EF-Tu, one of the most abundant proteins in bacterial cells, and bacterial cold-shock proteins seem

to be specifically recognized in *Brassicaceae* and *Solanaceae* plants, respectively (Gomez-Gomez and Boller, 2002; Felix and Boller, 2003; Bittel and Robatzek, 2007; Jeworutzki et al., 2010). In both proteins, the N-terminus contains the eliciting site which, for flagellin, can often be described as a 22 amino acid long epitope (flg22), whereas that for EF-Tu is 18 amino acids (Elf18). Flagellin and EF-Tu are recognized by two distinct plant receptors (Gomez-Gomez and Boller, 2002; Kunze et al., 2004). Flagellin is recognized by FLAGELLIN-SENSING 2 (FLS2) whereas EF-Tu is recognized by EF-Tu RECEPTOR (EFR) which has only been found in *Brassicaceae* (Gomez-Gomez and Boller, 2002; Zipfel et al., 2006). This provides molecular insight into the specificity of elicitors and emphasizes the need to assess candidate defense eliciting compounds in a diverse range of plant species. It is interesting to note that heterologous expression of EFR in *Solanaceae* plants provides some resistance to bacteria that express EF-Tu (Lacombe et al., 2010) which suggests that downstream signaling cascades could be conserved for different PRRs and in different plant species. Indeed, both FLS2 and EFR are leucine rich repeat receptor like kinases (LRR-RLK) and both interact with BRI1-associated receptor kinase 1 (BAK1) triggering SA-, JA-, and ET-independent signaling (Zipfel et al., 2004, 2006; Chinchilla et al., 2007). Recognition of flg22 and Elf18 leads to an increase in cytosolic Ca²⁺ and it has been shown that early signaling is BAK1-dependent and involves calcium associated plasma membrane anion channel opening (Jeworutzki et al., 2010). Subsequently, a MAP kinase cascade involving MPK3, MPK4, MPK6 and MPK11, and other genes such as Ca²⁺-dependent proteinase kinases are activated to establish PTI (Zipfel et al., 2004, 2006; Chinchilla et al., 2007; Bethke et al., 2012).

Pathogenic bacteria secrete, amongst others, type III effectors into plant cells to suppress PTI and this mechanism has been well studied in the plant pathogen *P. syringae* (reviewed by Block and Alfano, 2011; Deslandes and Rivas, 2012). These bacterial effectors target a variety of plant genes and metabolites including plasma membrane components like RPM1-interacting protein 4 (RIN4) in *A. thaliana* (Day et al., 2006). Similarly, host nuclear components are, for example, perturbed by effectors such as PopP2 as well as by transcription-activator like (TAL) type III effectors from *Xanthomonas* that directly bind to plant DNA and thereby activate gene expression changes that promote virulence and pathogen colonization (Boch et al., 2009; Deslandes and Rivas, 2012; Coll and Valls, 2013). Other examples include the effector HopA1 that is widely conserved in bacterial plant pathogens and interferes with the MAPK signaling genes MPK3 and MPK6 to suppress PTI (Zhang et al., 2007). Furthermore, chloroplast components are also modified by bacterial effectors such as HopI1 that causes remodeling of the chloroplast thylakoid structure and interferes with SA accumulation (Jelenska et al., 2007). As mentioned above, plants have a variety of R genes, the products of which, directly or indirectly, recognize some of these bacterial effectors to elicit ETI (reviewed by Block and Alfano, 2011; Deslandes and Rivas, 2012).

OOMYCETE-DERIVED ELICITORS

Oomycetes are taxonomically and structurally distinct from both plants and fungi. Several oomycetes are plant pathogenic and

include those from the genus *Phytophthora* that are responsible for substantial yield losses in crops. Oomycete cell walls consist of cellulose, glycan, and hydroxyproline-rich proteins and several oomycete elicitors have been described (Table 1). For example, necrosis and ethylene-inducing peptide 1 (Nep1)-like proteins (NLP) are recognized in dicots and it has been shown that these proteins trigger a variety of defense responses in *A. thaliana* (Qutob et al., 2006). Similarly, *P. infestans* INF1 elicitor causes an HR response in *Nicotiana benthamiana* (Kamoun et al., 1998) that is dependent on the receptor-like kinase SERK3/BAK1 which, as a central regulator of innate immunity in plants, is required for multiple resistance responses, including those mediated through FLS2 (Heese et al., 2007). Other PTI eliciting molecules from *Phytophthora* include GP42, a member of the transglutaminase family, and for which the active peptide has been described as Pep-13 (Nürnberg et al., 1994; Brunner et al., 2002), as well as the cellulose binding elicitor lectin (CBEL) that is associated with adhesion to the plant cell (Gaulin et al., 2006; reviewed in Hein et al., 2009).

To suppress PTI during infection, *Phytophthora*, like other plant pathogens, secretes extracellular and intracellular effectors into plants. Some extracellular effectors encode protease or glucanase inhibitors to prevent, respectively, host protease or glucanase activity in the apoplast (reviewed in Hein et al., 2009; Schornack et al., 2009). Some intracellular effectors contain the canonical RXLR motif and contain an N-terminal signal peptide and a C-terminal effector activity site (Birch et al., 2009).

The modes of action of RXLR effectors in promoting virulence are diverse. For example, it has recently been shown that the *P. infestans* RXLR effector PexRD2 interacts with the kinase domain of the host MAPKKKε to perturb PTI signaling pathways and to yield ETS responses (King et al., 2014). The RXLR effector PITG_03192, on the other hand, targets two membrane-associated NAC transcription factors that rapidly accumulate following PTI elicitation (McLellan et al., 2013). The effector prevents the release of these NAC transcription factors from the endoplasmic reticulum and subsequent accumulation in the plant nucleus that is typically observed as part of a PTI response. In contrast, the *P. infestans* RXLR effector Avrblb2 prevents secretion of an immune-associated protease (Bozkurt et al., 2011), whereas two *P. sojae* RXLRs have been shown to act as silencing suppressors (Qiao et al., 2013). One of the best-characterized intracellular RXLR effectors is Avr3a from *P. infestans*. Avr3a interacts with and stabilizes the potato E3 ubiquitin ligase CMPG1 and thus perturbs cell death responses triggered by INF1 (Bos et al., 2010) and a range of other pathogen elicitors (Gilroy et al., 2011). Avr3a exists in two forms that both suppress INF1 responses but differ in two amino acids that determine recognition by the potato R gene R3 that subsequently triggers ETI (Armstrong et al., 2005). Finally, several RXLRs from *P. infestans* act redundantly to suppress flg22-mediated signal transduction and early transcriptional changes (Zheng et al., 2014).

FUNGAL-DERIVED ELICITORS

As with plant and oomycete cell walls, break-down products from fungal cell walls, which contain chitin, mannoproteins, and β-glucans, can elicit a range of defense responses as signals

of potential colonization (Table 1). Yeast extracts, for example, have widely been used to study defense responses in plants (e.g., Hahn and Albersheim, 1978; Reglinski et al., 1994b, 1995; Suzuki et al., 2005; Khokon et al., 2010). Ergosterol, a fungal cell membrane component, induces defense responses in tobacco, and *Cladosporium fulvum* host and non-host plant necrosis inducer 1 (CFHNNI1), which shows high homologies to genes encoding bZIP transcription factors, has been shown to induce resistance in tomato and tobacco (Takken et al., 2000; Xu et al., 2012). Similarly, a proteinaceous elicitor called SCLEROTINIA CULTURE FILTRATE ELICITOR1 (SCFE1) has recently been identified from the necrotrophic fungal pathogen *Sclerotinia sclerotiorum* that induces BAK1-dependent PTI responses upon recognition by the *A. thaliana* RECEPTOR-LIKE PROTEIN30 (RLP30) (Zhang et al., 2013).

Two of the best studied fungal-derived elicitors are chitin and chitosan, a deacetylated derivative of chitin. Both have been well described as active components that increase resistance to bacterial and fungal pathogens in several plant species including crop plants (El Ghaouth et al., 1994; Copping and Duke, 2007; Kishimoto et al., 2010; El Hadrami et al., 2012; Kombrink et al., 2011). Chitin is detected in plants by a chitin elicitor receptor kinase (CERK1) which is also known as LysMRLK1 (Wan et al., 2008; Kombrink et al., 2011). In *A. thaliana*, chitin-induced dimerization of AtCERK1 has shown to be necessary for activation of PTI (Liu et al., 2012). In rice, OsCERK1 forms a complex with chitin elicitor binding protein (CEBiP) upon chitin perception and both proteins are critical for chitin-induced signaling (Shimizu et al., 2010). A homolog of OsCEBiP has been identified in barley and HvCEBiP has also been shown to play a role in responses to *Magnaporthe oryzae* (Tanaka et al., 2010). In contrast to rice, the homolog in *A. thaliana*, AtCEBiP, binds chitin but does not seem to be required for chitin-induced signaling (Shinya et al., 2012). Chitin-induced PTI is JA-, SA-, and ET-independent but a RING zinc-finger like protein (ATL9) has shown to be induced upon chitin treatment (Berrocal-Lobo et al., 2010).

To suppress these responses, the fungal pathogen *C. fulvum* has developed two distinct effectors that suppress chitin-induced PTI leading to ETS *in planta* (de Jonge and Thomma, 2009; de Jonge et al., 2010; Kombrink et al., 2011). The fungal chitin-binding protein Avr4 specifically binds chitin in fungal cell walls and thereby prevents the chitin from degradation by plant chitinases (van den Burg et al., 2006; Wan et al., 2008). Furthermore, the extracellular protein 6 (Ecp6), an effector protein with 3 LysM domains, binds chitin competitively to prevent recognition of chitin by CEBiP (de Jonge and Thomma, 2009; de Jonge et al., 2010). Homologs of Avr4 have been identified in fungi belonging to the class of *Dothideomycetes* and Ecp6-like genes are widespread within the fungal kingdom (Kombrink et al., 2011). The plant receptor Cf4 is a receptor-like protein (RLP) without kinase activity that recognizes Avr4 (Thomas et al., 1997) and it has recently been shown that SOBIR1, a receptor-like kinase (RLK) from tomato interacts with Cf4 and might be required for Cf4-mediated resistance (Liebrand et al., 2013).

Compared with bacterial and oomycete effectors, the biological function and the targets of fungal effectors remain more elusive (Rafiqi et al., 2012; Liu et al., 2013). This has partly been

attributed to the fact that fungal effectors do not seem to have canonical amino acid domains that enable a rapid candidate effector discovery (Rafiqi et al., 2012). Recently, Doehlemann and Hemetsberger (2013) reviewed the current knowledge of effectors from filamentous plant pathogens and compiled a list of known apoplastic effectors and their function. Most fungal effectors are secreted through the fungal endoplasmic reticulum (ER) secretory pathway but the way by which cytoplasmic effector proteins enter the host cells remains unknown (Rafiqi et al., 2012). In the genome of *Blumeria graminis*, 491 potential effector proteins have been identified but their biological function remains unknown (Pedersen et al., 2012). Similarly, in *M. oryzae* 15 candidate effector proteins have been identified so far (Liu et al., 2013). An effector protein from *Ustilago maydis* has been identified as a chorismate mutase, Cmu1, which is required for full virulence. Cmu1 functions by diverting metabolic precursors of the shikimate pathway toward production of aromatic amino acids, and away from the production of SA (Djamei and Kahmann, 2012).

PLANT GENES AND PATHWAYS INVOLVED IN ELICITOR RECOGNITION

GENES UP-REGULATED DUE TO ELICITOR TREATMENTS

In the elicitor research field, the response of plant genes to elicitor treatment is of great interest and several gene expression studies have been conducted. More recently, several microarray studies have been performed in different plant species to gain greater knowledge of the diversity of genes responsive to elicitors (e.g., Medeiros et al., 2009; Kano et al., 2011; Povero et al., 2011; Amelot et al., 2012), albeit knowledge on plant gene expression in response to elicitors has mainly been focused on *A. thaliana*. As mentioned previously, the presence of the cognate receptors can determine responsiveness to elicitors (Lacombe et al., 2010) and it is thus essential to investigate elicitor effects in diverse crop plants (Nguyen et al., 2010).

As part of this review, we aim to provide an overview of the current knowledge of differentially expressed plant genes following elicitor treatments and identify typically affected plant processes. This will facilitate identification of responses to elicitor application such as plant growth or nutrient metabolisms that are not directly linked to defense but impact on agriculture. For this we have combined over 50 publications to create a list of plant genes that are differentially expressed following the recognition of elicitors (Table S1). Reciprocal BLAST (Basic Local Alignment Search Tool; Altschul et al., 1990) has been used to identify the *A. thaliana* homologs when the original experiment was performed in a different plant species (>70% identity of nucleotide sequences, *E*-value < 0.0001). In addition to the publications, PathoPlant®, a database featuring compiled expression data and components of signal transduction pathways related to plant pathogenesis, has been used (Bülow et al., 2004, 2007). This database enables querying differential plant gene expression following diverse pathogen stimuli which, for this study, include *Botrytis cinerea*, chitin, *Erysiphe orontii*, *Phytophthora infestans*, *Pseudomonas syringae* pv. Maculicola, and *Pseudomonas syringae* pv. Phaseolicola.

A total of 1592 plant genes that were activated by the recognition of elicitors have been identified (Table S1). Ontological

analysis was performed using agriGo (Du et al., 2010). In this analysis, genes of interest are grouped by gene ontology (GO) terms describing biological processes, molecular functions and cellular components (Ashburner et al., 2000). The list of genes of interest is compared to a defined background gene list which, in this study, included the whole genome of *A. thaliana* as provided by The Arabidopsis Information Resource (TAIR) (Lamesch et al., 2011) to identify GO terms that are significantly over-represented. A total of 762 shared GO terms were identified, comprising 474 biological processes, 206 molecular functions and 82 cellular components (Table S2).

The three most over-represented biological processes were “response to stimulus” (GO:0050896), “multi-organism process” (GO:0051704), and “immune system process” (GO:0002376). The GO term “response to stimulus” contained 36.5% of the elicitor responsive genes in comparison to 10.7% representation in the whole *A. thaliana* genome. The GO term “multi-organism process” contained 11.1% of the genes in the target gene list, compared to 2.1% of the whole genome, and 6.6% instead of 1% were classed in the GO term “immune system process” (Figure 2).

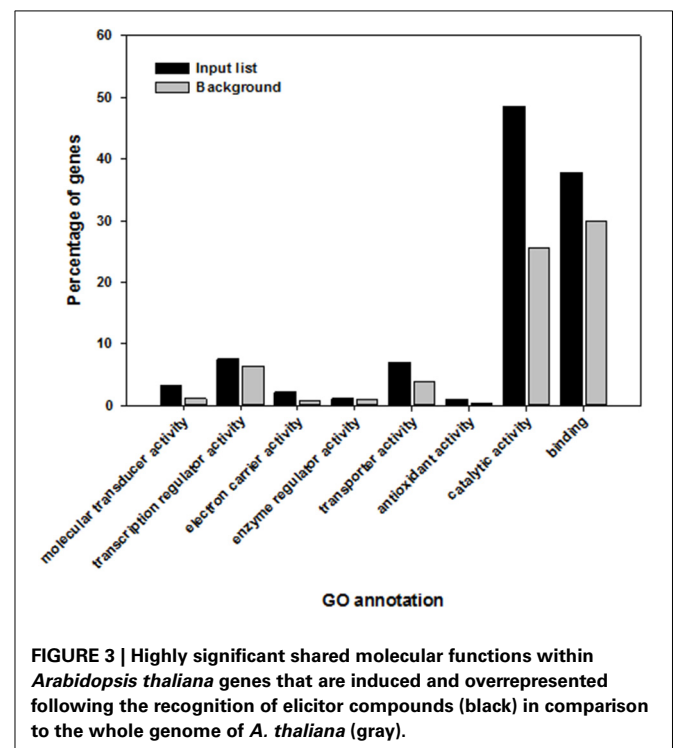
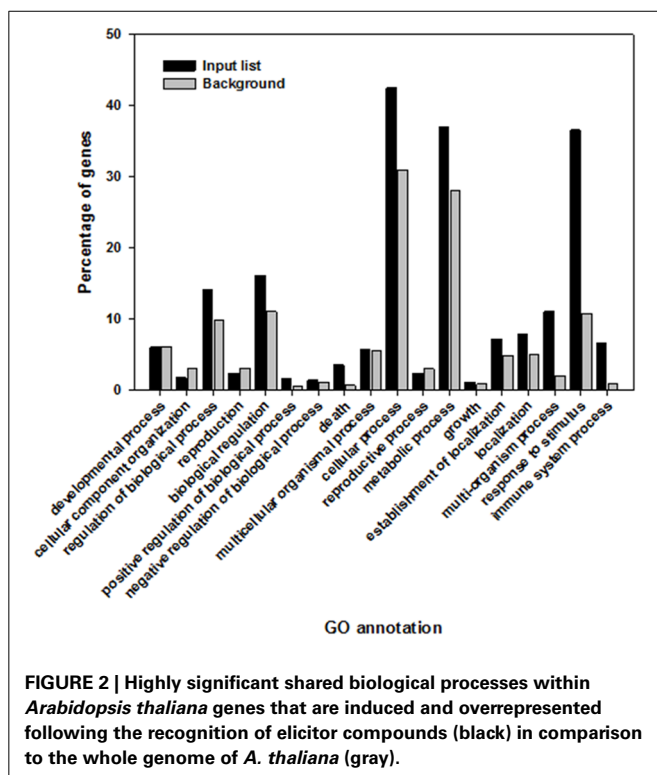
Three of the over-represented molecular functions were “catalytic activity” (GO:0003824), “binding” (GO:0005488), and “molecular transducer activity” (GO:0060089). The GO term “catalytic activity” featured in 48.5% of the elicitor responsive genes in comparison to 25.5% of the whole *A. thaliana* genome. The GO term “binding” contained 37.8% of the genes in the target gene list in comparison to 29.8% of the whole genome and 3.1% instead of 1.1% were classed in the GO term “molecular transducer activity” (Figure 3).

The relationships of all over-represented genes in the classification “biological processes” are shown in Figure S1. These include metabolic processes such as amine-, phosphate-, and phytoalexin metabolism; immune system processes and cell death, including regulation of defense response; plant-type hypersensitive response and apoptosis; response to stimuli including JA and SA; systemic acquired resistance and defense responses to fungi and bacteria (Figure S1).

The relationship of all genes over-represented in the GO term “molecular functions” are shown in Figure S2. These functions contain catalytic activity including oxidoreductase, lyase, and kinase activities; and binding activities including ATP and sugar binding (Figure S2). The relationships of all over-represented genes in the classification “cellular components” are shown in Figure S3. All cell parts are involved but the involvements of cell wall and plasma membranes are highly significant (Figure S3).

TARGETS OF PATHOGEN EFFECTORS

As noted above, pathogens produce effector molecules to interfere with plant defense responses. An analysis of plant-pathogen protein-protein interactions using *A. thaliana* and two pathogens, *P. syringae* and the obligate biotrophic oomycete *Hyaloperonospora arabidopsidis* revealed 137 *A. thaliana* proteins that were potentially targeted by pathogen effectors (Mukhtar et al., 2011). A recent review on bacterial effectors listed an additional 22 plant proteins targeted by several bacterial effectors (Deslandes and Rivas, 2012). In the analysis here, genes encoding these 159 proteins were used to search for overlap with the plant genes differentially induced upon elicitor recognition. A total of 23 genes were identified that are both induced by elicitors and targeted by pathogen effectors (Table S3). These comprise



receptors such as FLS2 and EFR, genes involved in MAPK cascades like RIPK, MPK3, MPK4, and MPK6, R protein-guarded host proteins such as RIN4 and genes involved in L-phenylalanine biosynthetic process like ADT4 and ADT5 (Table S3). These overlapping genes are grouped into several biological processes, molecular functions and cell parts and their involvements are highlighted with stars in Figures S1–S3. This analysis highlights the complexity of priming and plant immune responses, and the sophisticated interactions with pathogen effectors. It shows that the plant response to elicitor compounds does not only involve genes that are annotated as defense-related but that other metabolic pathways are also involved. We know that elicitor treatments result in positive and negative trade-offs (Walters and Heil, 2007) and therefore expression profiling of some of these genes should result in a better understanding of these responses and how they might be exploited.

“NON-DEFENSE” EFFECTS OF ELICITORS

As mentioned above, for successful use of elicitors in agriculture it is important to understand their effects not only on plant defense but also on other aspects of plant development and environmental responses. The activation of defense pathways as part of PTI can be very costly to plants but should be less than the potential loss caused by disease if no defense was mounted. However, in the context of crop protection, such costs are unlikely to be acceptable in the absence of known pathogen challenges of a high order. Crop protectants are preferred that enhance the efficacy of PTI assisting a quicker and more effective response when an actual pathogen challenge occurs and therefore is more efficient in its use of resources. This can be achieved through priming if priming is either not costly to the plant or its costs are mitigated by other beneficial means.

Some of the known priming genes are generally regulatory but not necessarily restricted to defense pathways. They regulate signal transduction events, particularly those identified in stress responses or in the GO terms “response to stimulus,” “multi-organism processes,” and “immune system processes,” i.e., the genes disproportionately up-regulated by elicitors highlighted above (see Figures S1, S2). Thus, many non-defense mechanism processes will be affected in their expression by elicitors. These might result in additional costs above those incurred by defense gene expression, but they may also have benefits. An example of a non-defense effect of an elicitor is reduced water use of pepper plants upon treatment with chitosan (Bittelli et al., 2001). Chitosan was also found to affect the net photosynthetic rate of soybean and maize after application (Khan et al., 2002). More general effects on yield, not directly attributable to disease control, were also found from applications of *B. subtilis* culture filtrates (Dehne et al., 1984; Steiner et al., 1988). Similar effects were recorded for some treatments of yeast cell wall-derived extracts (Reglinski et al., 1994a).

Indeed, on a molecular level, there is evidence of cross talk between the MAPKs involved in PTI and abiotic stress responses. For example, the transcription of MEKK1 is induced by diverse stresses including cold, salt, drought and wounding (Mizoguchi et al., 1998). Conversely, the activation of EDS1/PAD4-dependent signaling during ETI responses can rapidly antagonize ABA signal

transduction at the level of Ca²⁺ signaling (Kim et al., 2011). The overexpression of the gene ACTIVATED DISEASE RESISTANCE1 (ADS1) in *A. thaliana*, a member of the nucleotide-binding (NB) and leucine-rich repeat (LRR) containing NB-LRR genes, confers both disease resistance (Grant et al., 2003) and drought tolerance, requiring SA, EDS1 and ABA-INSENSITIVE1 (ABI1) (Chini et al., 2004).

Much depends on the basis of determination of costs as we tend to calculate these from an end-user yield perspective. We should also recognize that each of these pathways, whether defense-related or not, is in a complex expression and metabolic network of cross-talk and feedback mechanisms and thus affected by many environmental factors. Those that can be manipulated beneficially and perhaps synergistically fall in the category of nutrition (Walters and Bingham, 2007). Primed plants showed considerably higher fitness than non-primed plants when they were challenged by pathogens without major trade-off effects on growth and seed set (Conrath et al., 2006; van Hulten et al., 2006). Correlation of priming benefits with gene expression profiles may lead to very practical means for developing elicitor-based crop protectants that either off-set any direct costs, increase some aspect of resource use efficiency or specifically enhance other processes beneficial to yield or quality.

ACKNOWLEDGMENTS

The authors are grateful for financial support for this work in part from the Rural & Environment Science & Analytical Services Division of the Scottish Government. Lea Wiesel is supported by the Technology Strategy Board and its Knowledge Transfer Partnership grant KTP008814 with Omex Agriculture Limited.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://www.frontiersin.org/journal/10.3389/fpls.2014.00655/abstract>

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 12 September 2014; accepted: 04 November 2014; published online: 21 November 2014.

Citation: Wiesel L, Newton AC, Elliott I, Booty D, Gilroy EM, Birch PRJ and Hein I (2014) Molecular effects of resistance elicitors from biological origin and their potential for crop protection. *Front. Plant Sci.* 5:655. doi: 10.3389/fpls.2014.00655

This article was submitted to *Plant-Microbe Interaction*, a section of the journal *Frontiers in Plant Science*.

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