

# Body Fluid Cytokine Levels in Mild Cognitive Impairment and Alzheimer's Disease: a Comparative Overview

Frederic Brosseron · Marius Krauthausen ·  
Markus Kummer · Michael T. Heneka

Received: 8 November 2013 / Accepted: 4 February 2014 / Published online: 25 February 2014  
© The Author(s) 2014. This article is published with open access at Springerlink.com

**Abstract** This article gives a comprehensive overview of cytokine and other inflammation associated protein levels in plasma, serum and cerebrospinal fluid (CSF) of patients with Alzheimer's disease (AD) and mild cognitive impairment (MCI). We reviewed 118 research articles published between 1989 and 2013 to compare the reported levels of 66 cytokines and other proteins related to regulation and signaling in inflammation in the blood or CSF obtained from MCI and AD patients. Several cytokines are evidently regulated in (neuro-) inflammatory processes associated with neurodegenerative disorders. Others do not display changes in the blood or CSF during disease progression. However, many reports on cytokine levels in MCI or AD are controversial or inconclusive, particularly those which provide data on frequently investigated cytokines like tumor necrosis factor alpha (TNF- $\alpha$ ) or interleukin-6 (IL-6). The levels of several cytokines are possible indicators of neuroinflammation in AD. Some of them might increase steadily during disease progression or temporarily at the time of MCI to AD conversion. Furthermore, elevated body fluid cytokine levels may correlate with an

increased risk of conversion from MCI to AD. Yet, research results are conflicting. To overcome interindividual variances and to obtain a more definite description of cytokine regulation and function in neurodegeneration, a high degree of methodical standardization and patients collective characterization, together with longitudinal sampling over years is essential.

**Keywords** Neuroinflammation · Cytokines · Serum · Cerebrospinal fluid · Mild cognitive impairment · Alzheimer's disease

## Introduction

Cytokines are small signaling proteins with a large spectrum of functions in inflammatory processes and immune system regulation [1]. Therefore, they have been investigated in the context of neuroinflammation, a process accompanying and probably contributing to pathology in several neurodegenerative diseases including Alzheimer's disease (AD) or Parkinson's disease (PD) [2–5]. One key feature of neuroinflammation is activation of microglia, which includes local changes of cytokine expression [2, 3]. Additionally, systemic levels of cytokines may rise in response to aging and stress, known risk factors for neurodegeneration [6–8]. Susceptibility for inflammation rises with age and might be enhanced by each inflammatory event [9]. Furthermore, chronic inflammation and the delirium accompanying severe systemic infection have been shown to be risk factors for AD in the elderly, and vice versa, several risk factors for AD are also inducers of systemic inflammation [10–13]. As a consequence, levels of cytokines, their receptors and other proteins associated with immune responses in blood and CSF of AD patients have been frequently investigated to uncover mechanisms of neuroinflammation in dementia or in the context of biomarker research. However, much of the data obtained from different

---

**Electronic supplementary material** The online version of this article (doi:10.1007/s12035-014-8657-1) contains supplementary material, which is available to authorized users.

---

F. Brosseron · M. T. Heneka  
German Center for Neurodegenerative Diseases (DZNE), Bonn,  
Germany

M. Krauthausen · M. Kummer · M. T. Heneka  
Clinic and Polyclinic for Neurology, Clinical Neuroscience Unit,  
University Hospital Bonn, Bonn, Germany

M. T. Heneka (✉)  
German Center for Neurodegenerative Diseases (DZNE), Clinical  
Neuroscience Unit, Clinic and Polyclinic for Neurology,  
Sigmund-Freud-Str. 25, 53127 Bonn, Germany  
e-mail: michael.heneka@ukb.uni-bonn.de  
URL: [www.henekalab.com](http://www.henekalab.com)

studies is controversial. Here, we give a comprehensive overview of published research in this field and discuss possible reasons behind the conflicting observations.

## Results

### Literature Overview

We included 118 PubMed-listed articles providing data explicitly on levels of immune signaling proteins—primarily cytokines and their receptors—in serum, plasma or CSF of patients with diagnosed MCI or AD in comparison to unaffected control groups. We excluded studies on cytokine levels in human or murine brain tissue, cytokine production by lymphocytes, cytokine polymorphisms or cytokine levels in other neurodegenerative diseases, like PD or frontotemporal dementia. In total, the 118 articles reported data on 66 cytokines, cytokine receptors and other proteins induced by cytokines or otherwise associated with inflammatory signaling and regulation.

Table 1 gives a short summary of literature features: In general, about one third of the articles investigated MCI or other dementia types additional to AD. Plasma, serum and CSF were used in equal terms, and the most frequent method for cytokine determination was singleplex enzyme-linked immunosorbent assay (ELISA). By the last decade, multiplex assays and cytokine arrays were used with increasing frequency. A variety of cognition testing methods and diagnostic criteria were used in the different studies, although most articles noted the use of the National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA) criteria and mini-mental state examination (MMSE) for patient characterization [14, 15]. Supplementary 1 contains a more detailed description of the reviewed articles contents, investigated proteins and used methods.

### General Observations

A brief overview of the described regulations of different cytokines and inflammation associated proteins is given in Table 2. A list of observed effects and used methods for each protein is given in Supplementary 2. Above all, there is a tendency that with growing number of research papers on a particular cytokine there is also an increase in contradictions. For instance, the most frequently investigated cytokines, tumor necrosis factor alpha (TNF- $\alpha$ ) and interleukin-6 (IL-6), described in 20–25 % of articles, are reported as upregulated, not regulated or downregulated in the blood or CSF of AD patients (see below, Table 2 and Supplementary 2).

One explanation for the conflicting results could be differences between the technical approaches of the studies.

**Table 1** Characteristics of reviewed articles on cytokine levels in AD and MCI. The table lists investigated disease type, diagnostic criteria/tests, sample types and methods of the reviewed 118 articles (Supplementary 1). Note that some articles investigated more than one disease or body fluid or used more than a single method. Roughly 1/3 of articles investigated MCI or other dementia types additionally to AD. Most studies reported use of at least one class of diagnostic criteria and one type of cognitive testing. Plasma, serum and CSF were used in equal terms, and the most frequent method was singleplex ELISA, followed especially in the last decade by multiplex assays and cytokine arrays. The percentages reflect the respective proportion assessing the respective features

Disease type	96 %	Alzheimer's disease
	38 %	Mild cognitive impairment
	27 %	Other dementia or neurological disease
Diagnostic criteria / tests	76 %	NINCDS-ADRDA
	73 %	MMSE
	37 %	DSM-IV
	16 %	DSM-III-R
	12 %	CDR
	33 %	Other
Sample type	38 %	Plasma
	40 %	Serum
	37 %	CSF
Methods	77 %	ELISA (singleplex)
	8 %	Multiplex assay
	4 %	Cytokine array
	3 %	Western blot
	4 %	Cell-based bioassays
	6 %	Immunodiffusion (solemnly for quantification of ACT)
4 %	Other methods (radioimmunoassay, immunophelometry, qRT-PCR)	

However, methodological differences alone may not be the solemn source of the variances, as many of the studies used comparable methods: Over 75 % of reviewed articles obtained results from singleplex ELISA using recombinant protein standards for absolute quantification of cytokines (Table 1). It is possible that different ELISA kits do not give identical absolute values of the same analyte [16]. Yet, this cannot explain why different studies reported cytokine levels in AD patients to be higher, unchanged or lower when comparing to control groups. Further, studies using cytokine array technology in a methodological comparable manner also did not provide reproducible results, which indicates that not only technical differences cause conflicting results in cytokine analysis [17–19]. The same applies for other multiplex platforms (e.g., Luminex<sup>®</sup> platform): Despite high methodical similarities, there were considerable differences in sensitivity, specificity and composition of biofluids-based multianalyte patterns for differentiation between MCI and AD patients and controls [20–25].

**Table 2** Regulation of cytokines and inflammation associated proteins in serum/plasma and CSF of AD and MCI patients

Described regulation	Serum/plasma		CSF	
	MCI	AD	MCI	AD
↑ Upregulation	BDNF, IL-1β, MIF, MIP-4, RANTES	CTACK, FGF1, MIF, MIG, sCD40, SCF, VEGF	IL-8, IL-10, MIF, MIG, MIP-4α <sup>a</sup> , sTNF-RII	FGF1, IL-11, IL-18
↑ Upregulation + → No regulation	ICAM-1, IFN-α, TNF-α	ACT, ANG-2, IFN-α, IFN-γ, IL-1β, IL-10, IL-11, IL-18 <sup>a</sup> , MIP-1α, sTNF-RI, VCAM-1	MCP-1 <sup>a</sup>	ACT, IL-1β, IL-1RII, IL-8, IP-10, MCP-1 <sup>a</sup> , VEGF
→ No regulation	ACT, ANG-2, β-NGF, CD40L, CTACK, EGF, G-CSF, Eotaxin, GDNF, GRO-α, HGF, IL-1α, IL-1RII, IL-2R, IL-3, IL-6, IL-10, IL-11, IL-12, IL-16, IL-18, IP-10, LIF, M-CSF, MCP-1, MCP-3, MCP-4, MIG, MIP-1δ, MIP-4α, PDGF-BB, sCD40 <sup>a</sup> , SCF, SCGF, SDF-1α, sTNF-RI <sup>a</sup> , sTNF-RII, TRAIL, VCAM-1	β-NGF, E-Selectin, GM-CSF, GRO-α, HGF, IGFBP-6, IL-1RA, IL-1RII, IL-2, IL-2R, IL-7, IL-12 <sup>a</sup> , IL-16 <sup>a</sup> , IP-10, LIF, MIP-4, sTNF-RII, TRAIL, TRAIL-R4	BDNF, Eotaxin, IL-1β, IL-1RII, MCP-4	β-NGF, FGF2, GDNF, GM-CSF, HGF, IFN-γ, IL-1RA, IL-2, IL-2R, IL-10, IL-12 <sup>a</sup> , M-CSF, MIP-1α, SDF-1α, sTNF-RI, sTNF-RII
→ No regulation + ↓ Downregulation	IL-8	G-CSF, IL-1α, IL-6R, MCP-3, SDF-1α		BDNF, IL-6R
↓ Downregulation	IGF-1	G-CSF, IL-1α, IL-6R, MCP-3, P-selectin, SDF-1α	IL-7, M-CSF, TNF-α, TGF-β, VEGF	
↑ Upregulation + → No regulation + ↓ Downregulation		BDNF <sup>a</sup> , CRP, EGF, GDNF, ICAM-1, IL-3, IL-6 <sup>a</sup> , IL-8, M-CSF, MCP-1, PDGF-BB, RANTES, TNF-α, TGF-β <sup>a</sup>		IL-6, TNF-α, TGF-β

Overview of the results of the reviewed articles, separated by observed protein expression regulations for serum/plasma and CSF as well as MCI and AD. For several investigated proteins, multiple directions of regulation are described in different articles. For details on synonyms, frequency of effect observation and used methods, see Supplementary 2

<sup>a</sup> Proteins for which disease progression-dependent regulation is described

Taken together, these observations point to other critical factors, like patient collective composition and patient characterization. For example, it has been shown that cytokine profiles correlate to amyloid burden or APOE genotype, which might be of particular importance for the investigation of such proteins in AD [21, 26]. In this context, it is interesting that in some articles AD patient collectives were subdivided by severity of disease. These reports found differences in cytokine levels between mild, modest or severe AD, e.g., studies by Motta et al., Baranowska-Bik et al., Galimberti et al. [27–29]. Other studies outlined correlations between cytokine levels and disease risk, progression or MCI to AD conversion [27, 29–48]. Yet, a recent meta-analysis of Koyama et al. came to the conclusion that elevation of peripheral cytokine levels is a modest risk factor for neurodegeneration in general, but unspecific for AD [49].

In many studies, strongest upregulation of cytokines was observed in patients with mild AD indicating that cytokine signaling might primarily play a role in the intermediate stages of the disease. On the contrary, patients with advanced AD showed less strong upregulation of cytokines or no differences

compared to controls. This might explain why in AD patient collectives, which did not discriminate for disease progression state, no differences to controls or simply higher variances in the AD cohort were observed. Unfortunately, only few studies provide data on disease duration, disease severity or results of neuropsychological examinations like MMSE, which makes it difficult to compare these studies.

Another interesting observation is that some cytokines, especially those apparently not regulated in AD (e.g., interleukin-2, IL-2) where less controversial between studies than cytokines frequently reported to be regulated in any direction (like TNF-α, see Supplementary 2). Thus, the latter still provide interesting research targets, especially under the consideration that subgrouping of patients might provide improved insights into cytokine regulation in AD. In the following, we will give a more detailed description of the regulation of selected cytokines:

#### TNF-α

TNF-α is one of the most frequently investigated cytokines. From the 118 articles included, 13 articles describe

upregulation, 5 downregulation, and 15 no regulation of TNF- $\alpha$  levels in plasma or serum of AD patients in comparison to control groups [2, 17–19, 28, 32, 33, 35, 50–73]. In an attempt to reduce these variances, we focused on ten articles which report absolute values of plasma or serum TNF- $\alpha$  concentration as obtained by ELISA, include patient group sizes of  $n > 10$  and use the MMSE as an estimate of disease severity [18, 28, 33, 35, 50, 52, 64, 68, 69, 74]. Among these ten studies, six report no regulation and four modest upregulation of TNF- $\alpha$  in blood, the latter mostly in patients with severe AD. This might point to disease-state-dependent changes of TNF- $\alpha$  blood levels. Furthermore, the mean values for TNF- $\alpha$  in blood of controls range from 0.7 to 23.0 pg/ml between the studies, pointing towards interassay variances. Also, all studies show high interindividual variances and overlaps between patient and control groups.

Studies analyzing TNF- $\alpha$  in the CSF of AD patients are smaller in number but reflect the same picture: three studies report upregulation of TNF- $\alpha$ , one downregulation and five no regulation [2, 3, 58, 65, 75–79]. In MCI patients, two studies report upregulation and three no regulation in plasma or serum, whereas one study reports downregulation in CSF [3, 19, 32, 51, 52, 63]. The variances between the studies are therefore not limited to blood values.

As studies which reported increased TNF- $\alpha$ -levels often investigated patients with severe AD, it is possible that the levels of this cytokine increase slightly but continuously over the time course of the disease. It is also possible that TNF- $\alpha$  is only upregulated in subgroups of patients which have yet to be defined, e.g., patients suffering from neuroinflammation in addition to a neurodegenerative process.

#### TNF Receptors

A different picture is drawn for soluble variants of the TNF receptors (sTNF-RI and sTNF-RII). The levels of both receptors are mostly reported as unchanged in the blood or CSF of AD patients in comparison to controls [32, 33, 50, 65, 75, 80, 81]. For MCI patients, however, data are controversial [32, 80, 82]. Follow up-studies show correlations of TNF receptor levels with risk of MCI to AD conversion [32, 83]. It is possible that individuals with TNF receptor expression in the upper tertile are at increased risk of developing AD. Yet, the observed differences are too small to be used as reliable biomarkers.

#### Soluble CD40 and CD40 Ligand

Another member of the TNF receptor superfamily, soluble CD40 (sCD40), is reported to be regulated in AD in a remarkably congruent manner: Three studies describe the elevation of sCD40 plasma levels in AD patients [84–86]. A fourth article by Buchhave et al. reports that levels of sCD40 positively correlate to risk of MCI to AD conversion [38]. Despite

variances in effect strength between the studies, sCD40 might be an interesting target for biomarker research, especially since it has not been investigated in CSF of AD or MCI patients. Plasma levels of its binding partner CD40 ligand (CD40L) are described as not regulated in MCI patients and as upregulated in AD patients and might therefore represent another biomarker candidate [38, 84].

#### IL-1 $\beta$

IL-1 $\beta$  is another frequently investigated target in AD, whereas only few reports describe levels in MCI. Interestingly, IL-1 $\beta$  is mostly described as not regulated in CSF of AD patients, while approximately 50 % of reports on serum or plasma levels describe upregulation [2, 33, 35, 50, 53, 54, 56, 58, 65, 67, 71, 75, 76, 87–94]. The other 50 % of the studies on IL-1 $\beta$  plasma levels in AD show slightly increased values in patients, which are yet not statistically significant due to high interindividual variances and overlaps between patients and controls. Furthermore, no study reports downregulation of IL-1 $\beta$ . Similar to TNF- $\alpha$ , it can be hypothesized that IL-1 $\beta$  is only elevated in subgroups of patients or during certain disease stages. Also, peripheral IL-1 $\beta$  might increase slowly during the time course of the disease. However, even if one of these hypotheses is correct, the effects visible in the periphery are probably small, as reflected by the large number of studies showing no significant changes between AD patients and controls. Therefore, it would be interesting to follow IL-1 $\beta$ -levels in AD patients' blood and CSF longitudinally.

#### IL-6

IL-6 has been examined in AD with similar frequency as TNF- $\alpha$ , and with similar contradictory results [2, 28, 31, 34, 52–54, 58, 60, 61, 65, 67–69, 71, 75–77, 79, 87–89, 91, 92, 95–111]. We focused on articles reporting absolute concentrations in collectives of at least 20 individuals. Both criteria were fulfilled by 18 publications [28, 30, 31, 34, 52, 58, 68, 75, 79, 91, 94, 95, 102, 103, 107, 108, 112]. Most of the studies show either upregulation or no regulation of IL-6 in blood or CSF derived from AD patients. Noteworthy is that only 2 of 18 studies report downregulation of IL-6 [75, 113]. These findings are similar between blood and CSF. Only one study analyzed IL-6 levels in the blood of MCI patients but reports no regulation [52].

When comparing the data, we made two observations which might explain the conflicts: First, all included articles showed large interindividual variances of IL-6 levels, sometimes ranging from 50–100 % of the reported mean values. As a consequence, there is a high probability that comparisons in small patient cohorts produce misleading data, as it is highly probable that some individuals will show higher or lower cytokine levels than others just by chance.



Further, patients with severe AD showed higher plasma levels of IL-6 than patients with less severe disease or healthy controls. This could be interpreted in the way that peripheral levels of IL-6 slightly increase over the time course of AD, as shown by Kalman et al [31].

These observations much resemble those made for IL-1 $\beta$  and TNF- $\alpha$ , and as before, intraindividual data over the time course of disease would be the most promising way to obtain a clearer picture regarding IL-6 levels.

#### IL-6 Receptors

Levels of soluble IL-6 receptor (sIL-6R) have been analyzed in seven of the reviewed articles which investigated AD patients, but not in MCI cases [75, 81, 101, 105, 109, 112, 114]. Each of these articles report either no regulation or downregulation of sIL-6R in blood or CSF of AD patients. Absolute values are relatively consistent between the studies, ranging from approx. 20–220 ng/ml in serum and 0.5–1.6 ng/ml in CSF. Similar to the cytokines described above, high interindividual variances and a large overlap between controls and patients were observed in all studies on sIL-6R. Although the tendency to reduction of sIL-6R levels in AD is apparently weak, none of the reviewed studies reported upregulation of this cytokine receptor. This is especially interesting as IL-6 levels appear to increase slightly during AD. To our knowledge, no study so far has analyzed the ratio of IL-6 to sIL-6R in AD or changes of this ratio over the time course of disease. Again, it seems also possible that so far only uncharacterized subgroups of AD patients display lower sIL-6R levels compared to others.

#### IL-18

So far, all cytokines described in this review appear to increase slowly with disease progression, while the respective receptors might be decreased. Nevertheless, some cytokines present a different picture. IL-18 has mostly been investigated in the plasma and with at first glance contradictory findings: several studies report no significant changes in IL-18 blood levels of both MCI- and AD patients, although always with a tendency to elevated levels [71, 115–117]. Two other studies show elevation of blood levels in AD [118, 119]. Most of these studies differed in the used ELISA kit and/or in patient cohort characterization, which might be one reason for the observed differences. Yet, there may be another possibility: In a study of Motta et al., the patient cohort was divided according to MMSE into mild, modest, and severe AD subgroups. These authors showed that IL-18 levels were elevated in the early stages of the disease, but later dropped again to levels equal to those of controls [27]. After the initial rise, the following decline of IL-18 levels occurred in a disease progression-dependent manner. In other words, IL-18 levels reached a

peak in mild AD patients and correlated positively with the MMSE afterwards. These findings would fit to several other studies (e.g., [117, 119]) and support the concept of analyzing AD subgroups. They also support the theory of neuroinflammation as an early event in AD [120]. In this context, it is interesting to note that no study analyzing IL-18 reports effects in the plasma of MCI patients [71, 116, 117]. Together with the results of Motta et al., these findings may indicate that IL-18 levels are elevated in the early phases of AD, possibly during the turnover from “normal” MCI to AD. To our knowledge, only one study analyzed IL-18 levels in CSF of AD patients and found elevated levels of this cytokine [115]. It should further be mentioned that IL-18-binding protein (IL-18BP), a regulator of IL-18 function, has been described as downregulated in AD, indicating that the ratio of IL-18 and IL-18BP is influenced by regulation of both proteins [119]. Summarized, IL-18—and possibly its regulator IL-18BP—represent interesting candidates to be analyzed in plasma and especially CSF of well-characterized MCI and AD patients.

#### CCL2/MCP-1

MCP-1 has been analyzed in plasma and CSF of AD and MCI patients. Although results were again controversial, several studies find MCP-1 to be upregulated in the CSF of AD and also MCI patients [121–123]. In plasma, most articles report no regulation of MCP-1 [51, 70, 116, 121]. Only one study conducted by Galimberti et al. investigated patients divided in MCI, mild-modest AD and severe AD groups and revealed elevated levels in MCI and mild-modest AD patients, while subjects with severe AD showed lower levels [29]. The effect strength was statistically significant, yet modest in size and there were large overlaps between the groups. However, MCP-1 levels correlated to MMSE after onset of MCI. This induction pattern is highly similar to the one described by Motta et al. for other cytokines and might be the result of innate immune activation in the early stages of AD, as mirrored by central and peripheral cytokine levels [27, 124].

#### CXCL10/IP-10

The 10-kDa interferon gamma-induced protein (IP-10) is reported to be elevated in the CSF of MCI patients [82, 122]. After conversion from MCI to AD, CSF levels drop again and correlate over the time course of disease with MMSE scores and cognitive decline [82, 122, 123]. In contrast, plasma levels are uniformly reported to be unchanged in AD [70, 116, 121]. Therefore, IP-10 might resemble MCP-1 or IL-18 by showing a peak of CSF levels only in early disease stages.

## TGF- $\beta$

One of the cytokines showing the most inconsistent data is TGF- $\beta$  [27, 61, 78, 85, 94, 118, 125–131]. It has been primarily investigated in AD and is described as not regulated, upregulated, downregulated and regulated dependent on disease state. We focused on eight articles which used ELISA for detection, but still found high variances in patient characterization and results [27, 78, 94, 127–131]. The mean values for healthy controls ranged from 10 pg/ml to 60 ng/ml, most likely derived from the lack of technical standardization. Still, as mentioned above, this does not explain the different directions of regulation between the reports. In contrast to IL-18, it was not possible to explain these different results based on a disease progression-dependent regulation of TGF- $\beta$ .

## Cytokines with No or Marginal Changes in AD

Several cytokines have been intensively investigated in AD patients without finding an induction or regulation in blood or CSF. A good representative for this group is interleukin-2 (IL-2), which was analyzed in three studies on CSF and seven studies on plasma of AD patients [2, 33, 54, 67, 68, 71, 79, 89]. As all of these studies uniformly reported no changes in CSF or plasma levels compared to controls, IL-2 is probably not regulated in AD. Similar findings have also been documented for its receptor IL-2R and some other cytokines like GM-CSF, IFN- $\gamma$ , IL-1 $\alpha$ , IL-1RA, and IL-3 (Supplementary 2). Still, some of these factors have barely been investigated in the CSF of AD or MCI patients and it cannot be excluded that changes might be visible in CSF which are undetectable in peripheral blood.

## Other Inflammation Associated Proteins

Together with cytokines, several other proteins induced by cytokines or otherwise involved in or associated with inflammatory processes, like growth factors, selectins or acute phase proteins have been investigated (Supplementary 2). The resulting findings were often as contradictory as for cytokines, although available data might sometimes be too scarce for final conclusions. Two frequently analyzed examples are alpha-1-antichymotrypsin (ACT) and brain-derived neurotrophic factor (BDNF):

ACT has been extensively studied in AD patients using the methods of immunodiffusion and ELISA [30, 65, 71, 72, 87, 88, 95–100, 132–135]. Data on ACT levels in MCI, on the other hand, are scarce. Approximately 50 % of the articles on ACT describe modest upregulation in AD, while the other half does not find differences in serum or CSF. It has been stated that ACT levels might show a weak positive correlation with disease

progression in AD, which might explain the differences between the reports [95]. When evaluated as a biomarker, ACT levels were insufficient to discriminate AD from other dementias, whereas elevated levels in other diseases lead to a high false-positive rate [132–134].

The effects reported for BDNF were mostly modest whereas interindividual differences were high and overlapping between the groups [35–37, 78, 136–139]. The largest study by O'Bryant et al. investigating nearly 100 individuals showed no differences between AD patients and controls [140]. Therefore, smaller collectives might provide misleading results due to the high interindividual variances, and BDNF levels might in reality be unchanged in AD.

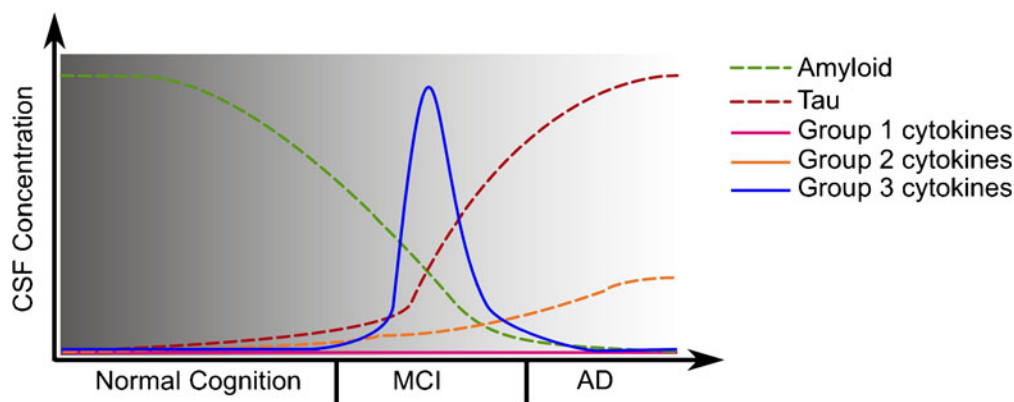
## Conclusions

Studies on proteins involved in immune signaling and regulation often present a heterogeneous picture. Methodical variances caused by use of different ELISA kits, might be one contributing factor to the observed discrepancies. Despite from various diluents and detection methods, capture or detection antibodies might recognize different antigens, resulting in the quantification of various protein isoforms. Comparative studies between numerous antibody-based single- and multiplex approaches for cytokine quantification and a better characterization of the epitopes recognized by the respective antibodies might therefore be desirable. As recently pointed out, use of serum or plasma biobanking conditions and sample handling may significantly affect the results of cytokine detection, which is why improvement of standardization between research groups should also be considered [141].

Further differences might be based on patient collective characterization, especially in terms of disease progression, as several studies discuss correlations of cytokine expression to disease state [27, 29–32, 36–38, 47]. As a possible guideline for future studies, levels of cytokines, other immune signaling related regulators and their receptors in blood or CSF of MCI and AD patients can be divided into five groups by involvement into disease, available information and consequences for research (Fig. 1):

The first group contains cytokines like IL-2 or IL-1 $\alpha$  which are frequently and uniformly reported as unchanged during disease progression, especially in regard of blood levels. Of note, this does not exclude any intra- and intercellular function of these cytokines, but makes them less promising targets for biomarker research.

The second group includes cytokines like IL-1 $\beta$ , IL-6, and TNF- $\alpha$  which seem to increase slightly but steadily



**Fig. 1** Hypothetical time course of CSF cytokine expression in AD. Graphs display the estimated CSF concentration changes of amyloid and tau protein during the development of AD, as described by others [142]. As different cytokines and other inflammatory proteins appear to display different changes in CSF levels during disease development, they might be divided into groups: First, cytokines like IL-1 $\alpha$  or IL-2 which might remain unchanged in AD; Second, cytokines like IL-1 $\beta$ , IL-6 or

TNF- $\alpha$  which might increase slowly during disease progression; third, cytokines like IL-18, MCP-1 or IP-10 which might show a peak at certain disease stages, especially at time of MCI to AD conversion. However, data becomes scarce for early disease stages. To test this hypothesis and the grouping of cytokines, longitudinal CSF sampling from individuals at risk of dementia over years would be the most efficient way

over the time during the course of AD, not only in the CSF but also in blood. Members of this group often show effects which are too small to be used as reliable biomarkers. Aside from steady increase, there are the possibilities that individuals with elevated levels of these cytokines are at higher risk to develop AD or that subgroups of AD patients display elevated levels.

The third group includes cytokines for which a peak in mild AD or around the conversion from MCI to AD has been documented. A longitudinal validation of these observations seems to be a promising target for biomarker research. Likewise, cytokines from the second group may be successfully attributed to a distinct time point of disease and thus allow for further functional insight.

The fourth group comprises the less frequently analyzed cytokines and cytokine receptors, like CD40, which were only investigated in a limited amount of studies and require further validation. Studies of such cytokines, especially from CSF samples, could be a useful addition to the large number of already existing analyses.

The last group includes cytokines like TGF- $\beta$ , for which the documented data are just too inconsistent to allow for any interpretation. For the latter, it would helpful to optimize the characterization of the patient collective and to standardize the detection methods. When picking candidates from these groups, it should be noted that pairs of cytokines and the respective receptors or binding partners (like TNF- $\alpha$  and TNF receptor, IL-6 and IL-6 receptor or IL-18 and IL-18BP) often showed coregulation or inverse regulation. This observation could be useful to create ratios between cytokines and their receptors or binding partners. Such ratios could represent more valid and reliable biomarkers than each cytokine level alone.

Overall, there is a substantial lack of longitudinal data of cytokine expression, which might account for much of the contradictory results. Studies analyzing large cohorts of elderly and MCI patients over several years are missing to date. Ideally, such studies would continuously collect blood and CSF samples according to a predefined schedule. At the same time, clinical evaluations and cerebral imaging, along with the detection of the classical CSF biomarkers, amyloid, tau and phospho-tau, should be assessed and related to inflammatory mediators. Once such studies are performed, they will provide important information and allow for a more solid picture of the role of cytokine expression during AD development.

**Funding** This work was funded by the German Center for Neurodegenerative Diseases (DZNE e.V.) within the Helmholtz Association, by the Deutsche Forschungsgemeinschaft (DFG, Klinische Forschergruppe 177, TP4) and the EU-FP7 consortium INMIND.

**Open Access** This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

## References

- Dinarelli CA (2007) Historical insights into cytokines. *Eur J Immunol* 37(Suppl 1):S34–S45
- Rao JS et al (2012) Neuroinflammation and synaptic loss. *Neurochem Res* 37(5):903–910
- Tarkowski E et al (2003) Intrathecal inflammation precedes development of Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 74(9):1200–1205

4. Lee KS et al (2009) Peripheral cytokines and chemokines in Alzheimer's disease. *Dement Geriatr Cogn Disord* 28(4):281–287
5. Swardfager W et al (2010) A meta-analysis of cytokines in Alzheimer's disease. *Biol Psychiatry* 68(10):930–941
6. Inadera H et al (1999) Increase in circulating levels of monocyte chemoattractant protein-1 with aging. *J Interferon Cytokine Res* 19(10):1179–1182
7. Bruunsgaard H et al (2003) Elevated levels of tumor necrosis factor alpha and mortality in centenarians. *Am J Med* 115(4):278–283
8. Kiecolt-Glaser JK et al (2003) Chronic stress and age-related increases in the proinflammatory cytokine IL-6. *Proc Natl Acad Sci U S A* 100(15):9090–9095
9. Kale SS, Yende S (2011) Effects of aging on inflammation and hemostasis through the continuum of critical illness. *Aging Dis* 2(6):501–511
10. Perry VH (2010) Contribution of systemic inflammation to chronic neurodegeneration. *Acta Neuropathol* 120(3):277–286
11. Jackson JC et al (2004) The association between delirium and cognitive decline: a review of the empirical literature. *Neuropsychol Rev* 14(2):87–98
12. Blennow K, de Leon MJ, Zetterberg H (2006) Alzheimer's disease. *Lancet* 368(9533):387–403
13. Bettcher BM, Kramer JH (2012) Inflammation and clinical presentation in neurodegenerative disease: a volatile relationship. *Neurocase* 19(2):182–200
14. McKhann G et al (1984) Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA work group under the auspices of department of health and human services task force on Alzheimer's disease. *Neurology* 34(7):939–944
15. Folstein MF, Folstein SE, McHugh PR (1975) Mini-mental state. A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res* 12(3):189–198
16. Zasada AA et al (2013) Comparison of seven commercial enzyme-linked immunosorbent assays for the detection of antiphosphorylated tau antibodies. *Eur J Clin Microbiol Infect Dis* 32(7):891–897
17. Ray S et al (2007) Classification and prediction of clinical Alzheimer's diagnosis based on plasma signaling proteins. *Nat Med* 13(11):1359–1362
18. Bjorkqvist M et al (2012) Evaluation of a previously suggested plasma biomarker panel to identify Alzheimer's disease. *PLoS One* 7(1):e29868
19. Marksteiner J et al (2011) Five out of 16 plasma signaling proteins are enhanced in plasma of patients with mild cognitive impairment and Alzheimer's disease. *Neurobiol Aging* 32(3):539–540
20. Martins TB (2002) Development of internal controls for the Luminex instrument as part of a multiplex seven-analyte viral respiratory antibody profile. *Clin Diagn Lab Immunol* 9(1):41–45
21. Burnham SC et al (2013) A blood-based predictor for neocortical Aβ burden in Alzheimer's disease: results from the AIBL study. *Mol Psychiatry*, pp 1–8
22. Doecke JD et al (2012) Blood-based protein biomarkers for diagnosis of Alzheimer disease. *Arch Neurol* 69(10):1318–1325
23. Johnstone D et al (2012) Multivariate protein signatures of pre-clinical Alzheimer's disease in the Alzheimer's disease neuroimaging initiative (ADNI) plasma proteome dataset. *PLoS One* 7(4):e34341
24. Soares HD et al (2009) Identifying early markers of Alzheimer's disease using quantitative multiplex proteomic immunoassay panels. *Ann N Y Acad Sci* 1180:56–67
25. Soares HD et al (2012) Plasma biomarkers associated with the apolipoprotein E genotype and Alzheimer disease. *Arch Neurol* 69(10):1310–1317
26. Ringman JM et al (2012) Plasma signaling proteins in persons at genetic risk for Alzheimer disease: influence of APOE genotype. *Arch Neurol* 69(6):757–764
27. Motta M et al (2007) Altered plasma cytokine levels in Alzheimer's disease: correlation with the disease progression. *Immunol Lett* 114(1):46–51
28. Baranowska-Bik A et al (2008) Plasma beta amyloid and cytokine profile in women with Alzheimer's disease. *Neuroendocrinol Lett* 29(1):75–79
29. Galimberti D et al (2006) Serum MCP-1 levels are increased in mild cognitive impairment and mild Alzheimer's disease. *Neurobiol Aging* 27(12):1763–1768
30. Vom Berg J et al (2012) Inhibition of IL-12/IL-23 signaling reduces Alzheimer's disease-like pathology and cognitive decline. *Nat Med* 18(12):1812–1819
31. Kalman J et al (1997) Serum interleukin-6 levels correlate with the severity of dementia in Down syndrome and in Alzheimer's disease. *Acta Neurol Scand* 96(4):236–240
32. Diniz BS et al (2010) Higher serum sTNFR1 level predicts conversion from mild cognitive impairment to Alzheimer's disease. *J Alzheimers Dis* 22(4):1305–1311
33. De Luigi A et al (2002) Peripheral inflammatory response in Alzheimer's disease and multiinfarct dementia. *Neurobiol Dis* 11(2):308–314
34. Galimberti D et al (2008) Intrathecal levels of IL-6, IL-11 and LIF in Alzheimer's disease and frontotemporal lobar degeneration. *J Neurol* 255(4):539–544
35. Yasutake C et al (2006) Serum BDNF, TNF-α and IL-1β levels in dementia patients: comparison between Alzheimer's disease and vascular dementia. *Eur Arch Psychiatry Clin Neurosci* 256(7):402–406
36. Laske C et al (2011) Higher BDNF serum levels predict slower cognitive decline in Alzheimer's disease patients. *Int J Neuropsychopharmacol* 14(3):399–404
37. Laske C et al (2006) Stage-dependent BDNF serum concentrations in Alzheimer's disease. *J Neural Transm* 113(9):1217–1224
38. Buchhave P et al (2009) Elevated plasma levels of soluble CD40 in incipient Alzheimer's disease. *Neurosci Lett* 450(1):56–59
39. Yaffe K et al (2003) Inflammatory markers and cognition in well-functioning African-American and white elders. *Neurology* 61(1):76–80
40. Dik MG et al (2005) Serum inflammatory proteins and cognitive decline in older persons. *Neurology* 64(8):1371–1377
41. Schmidt R et al (2002) Early inflammation and dementia: a 25-year follow-up of the Honolulu-Asia aging study. *Ann Neurol* 52(2):168–174
42. Weaver JD et al (2002) Interleukin-6 and risk of cognitive decline: MacArthur studies of successful aging. *Neurology* 59(3):371–378
43. Engelhart MJ et al (2004) Inflammatory proteins in plasma and the risk of dementia: the Rotterdam study. *Arch Neurol* 61(5):668–672
44. Tan ZS et al (2007) Inflammatory markers and the risk of Alzheimer disease: the Framingham study. *Neurology* 68(22):1902–1908
45. Holmes C et al (2009) Systemic inflammation and disease progression in Alzheimer disease. *Neurology* 73(10):768–774
46. Locascio JJ et al (2008) Plasma amyloid beta-protein and C-reactive protein in relation to the rate of progression of Alzheimer disease. *Arch Neurol* 65(6):776–785
47. Westin K et al (2012) CCL2 is associated with a faster rate of cognitive decline during early stages of Alzheimer's disease. *PLoS One* 7(1):e30525
48. Honma T et al (2013) Increased systemic inflammatory interleukin-1ss and interleukin-6 during agitation as predictors of Alzheimer's disease. *Int J Geriatr Psychiatry* 28(3):233–241
49. Koyama A et al (2013) The role of peripheral inflammatory markers in dementia and Alzheimer's disease: a meta-analysis. *J Gerontol A Biol Sci Med Sci* 68(4):433–440



50. De Luigi A et al (2001) Inflammatory markers in Alzheimer's disease and multi-infarct dementia. *Mech Ageing Dev* 122(16):1985–1995
51. Kim SM et al (2011) Identification of peripheral inflammatory markers between normal control and Alzheimer's disease. *BMC Neurol* 11:51
52. Bermejo P et al (2008) Differences of peripheral inflammatory markers between mild cognitive impairment and Alzheimer's disease. *Immunol Lett* 117(2):198–202
53. Zuliani G et al (2007) High interleukin-6 plasma levels are associated with functional impairment in older patients with vascular dementia. *Int J Geriatr Psychiatry* 22(4):305–311
54. Corsi MM et al (2011) Reduced plasma levels of P-selectin and L-selectin in a pilot study from Alzheimer disease: relationship with neuro-degeneration. *Biogerontology* 12(5):451–454
55. Kassner SS et al (2008) Novel systemic markers for patients with Alzheimer disease?—A pilot study. *Curr Alzheimer Res* 5(4):358–366
56. Alvarez XA et al (1996) Blood levels of histamine, IL-1 beta, and TNF-alpha in patients with mild to moderate Alzheimer disease. *Mol Chem Neuropathol* 29(2–3):237–252
57. Bruunsgaard H et al (1999) A high plasma concentration of TNF-alpha is associated with dementia in centenarians. *J Gerontol A Biol Sci Med Sci* 54(7):M357–M364
58. Tarkowski E et al (1999) Intracerebral production of tumor necrosis factor-alpha, a local neuroprotective agent, in Alzheimer disease and vascular dementia. *J Clin Immunol* 19(4):223–230
59. Chen R et al (2012) Elevation of serum TNF-alpha levels in mild and moderate Alzheimer patients with daytime sleepiness. *J Neuroimmunol* 244(1–2):97–102
60. Maes M et al (1999) Inflammatory markers in younger vs elderly normal volunteers and in patients with Alzheimer's disease. *J Psychiatr Res* 33(5):397–405
61. Chao CC et al (1994) Serum cytokine levels in patients with Alzheimer's disease. *Clin Diagn Lab Immunol* 1(4):433–436
62. Kester MI et al (2012) Decreased mRNA expression of CCL5 [RANTES] in Alzheimer's disease blood samples. *Clin Chem Lab Med* 50(1):61–65
63. Alvarez A et al (2007) Serum TNF-alpha levels are increased and correlate negatively with free IGF-I in Alzheimer disease. *Neurobiol Aging* 28(4):533–536
64. Solerte SB et al (2000) Overproduction of IFN-gamma and TNF-alpha from natural killer (NK) cells is associated with abnormal NK reactivity and cognitive derangement in Alzheimer's disease. *Ann N Y Acad Sci* 917:331–340
65. Lanzrein AS et al (1998) Longitudinal study of inflammatory factors in serum, cerebrospinal fluid, and brain tissue in Alzheimer disease: interleukin-1beta, interleukin-6, interleukin-1 receptor antagonist, tumor necrosis factor-alpha, the soluble tumor necrosis factor receptors I and II, and alpha1-antichymotrypsin. *Alzheimer Dis Assoc Disord* 12(3):215–227
66. Fillit H et al (1991) Elevated circulating tumor necrosis factor levels in Alzheimer's disease. *Neurosci Lett* 129(2):318–320
67. Angelopoulos P et al (2008) Cytokines in Alzheimer's disease and vascular dementia. *Int J Neurosci* 118(12):1659–1672
68. Bonotis K et al (2008) Systemic immune aberrations in Alzheimer's disease patients. *J Neuroimmunol* 193(1–2):183–187
69. Zuliani G et al (2008) Markers of endothelial dysfunction in older subjects with late onset Alzheimer's disease or vascular dementia. *J Neurol Sci* 272(1–2):164–170
70. Choi C et al (2008) Multiplex analysis of cytokines in the serum and cerebrospinal fluid of patients with Alzheimer's disease by color-coded bead technology. *J Clin Neurol* 4(2):84–88
71. Ozturk C et al (2007) The diagnostic role of serum inflammatory and soluble proteins on dementia subtypes: correlation with cognitive and functional decline. *Behav Neurol* 18(4):207–215
72. Matsubara E et al (1989) Serum concentration of alpha 1-antichymotrypsin is elevated in patients with senile dementia of the Alzheimer type. *Prog Clin Biol Res* 317:707–714
73. Cacabelos R et al (1994) Serum tumor necrosis factor (TNF) in Alzheimer's disease and multi-infarct dementia. *Methods Find Exp Clin Pharmacol* 16(1):29–35
74. Llano DA et al (2012) Cerebrospinal fluid cytokine dynamics differ between Alzheimer disease patients and elderly controls. *Alzheimer Dis Assoc Disord* 26(4):322–328
75. Richartz E et al (2005) Decline of immune responsiveness: a pathogenetic factor in Alzheimer's disease? *J Psychiatr Res* 39(5):535–543
76. Engelborghs S et al (1999) Unchanged levels of interleukins, neopterin, interferon-gamma and tumor necrosis factor-alpha in cerebrospinal fluid of patients with dementia of the Alzheimer type. *Neurochem Int* 34(6):523–530
77. Garlind A et al (1999) Soluble interleukin-1 receptor type II levels are elevated in cerebrospinal fluid in Alzheimer's disease patients. *Brain Res* 826(1):112–116
78. Blasko I et al (2006) Measurement of thirteen biological markers in CSF of patients with Alzheimer's disease and other dementias. *Dement Geriatr Cogn Disord* 21(1):9–15
79. Jia JP et al (2005) Cerebrospinal fluid tau, Abeta1-42 and inflammatory cytokines in patients with Alzheimer's disease and vascular dementia. *Neurosci Lett* 383(1–2):12–16
80. Hernanz A et al (2007) Plasma aminothiols compounds, but not serum tumor necrosis factor receptor II and soluble receptor for advanced glycation end products, are related to the cognitive impairment in Alzheimer's disease and mild cognitive impairment patients. *Neuroimmunomodulation* 14(3–4):163–167
81. Hasegawa Y et al (2000) Increased soluble tumor necrosis factor receptor levels in the serum of elderly people. *Gerontology* 46(4):185–188
82. Craig-Schapiro R et al (2011) Multiplexed immunoassay panel identifies novel CSF biomarkers for Alzheimer's disease diagnosis and prognosis. *PLoS One* 6(4):e18850
83. Buchhave P et al (2010) Soluble TNF receptors are associated with Abeta metabolism and conversion to dementia in subjects with mild cognitive impairment. *Neurobiol Aging* 31(11):1877–1884
84. Ait-ghezala G et al (2008) Diagnostic utility of APOE, soluble CD40, CD40L, and Abeta1-40 levels in plasma in Alzheimer's disease. *Cytokine* 44(2):283–287
85. Mocali A et al (2004) Increased plasma levels of soluble CD40, together with the decrease of TGF beta 1, as possible differential markers of Alzheimer disease. *Exp Gerontol* 39(10):1555–1561
86. Doecke JD et al (2012) Blood-based protein biomarkers for diagnosis of Alzheimer disease. *Arch Neurol* 16:1–8
87. Pirttila T et al (1994) Alpha 1-antichymotrypsin and IL-1 beta are not increased in CSF or serum in Alzheimer's disease. *Neurobiol Aging* 15(3):313–317
88. Licastro F et al (2000) Blood levels of alpha-1-antichymotrypsin and risk factors for Alzheimer's disease: effects of gender and apolipoprotein E genotype. *Dement Geriatr Cogn Disord* 11(1):25–28
89. Blum-Degen D et al (1995) Interleukin-1 beta and interleukin-6 are elevated in the cerebrospinal fluid of Alzheimer's and de novo Parkinson's disease patients. *Neurosci Lett* 202(1–2):17–20
90. Forlenza OV et al (2009) Increased serum IL-1beta level in Alzheimer's disease and mild cognitive impairment. *Dement Geriatr Cogn Disord* 28(6):507–512
91. Gomez-Tortosa E et al (2003) Cerebrospinal fluid markers in dementia with Lewy bodies compared with Alzheimer disease. *Arch Neurol* 60(9):1218–1222
92. Martinez M et al (2000) Increased cerebrospinal fluid fas (Apo-1) levels in Alzheimer's disease. Relationship with IL-6 concentrations. *Brain Res* 869(1–2):216–219

93. Martinez M, Frank A, Hernanz A (1993) Relationship of interleukin-1 beta and beta 2-microglobulin with neuropeptides in cerebrospinal fluid of patients with dementia of the Alzheimer type. *J Neuroimmunol* 48(2):235–240
94. Tarkowski E et al (2003) Cerebral pattern of pro- and anti-inflammatory cytokines in dementias. *Brain Res Bull* 61(3):255–260
95. Licastro F et al (2000) Increased plasma levels of interleukin-1, interleukin-6 and alpha-1-antichymotrypsin in patients with Alzheimer's disease: peripheral inflammation or signals from the brain? *J Neuroimmunol* 103(1):97–102
96. Harigaya Y et al (1995) Alpha 1-antichymotrypsin level in cerebrospinal fluid is closely associated with late onset Alzheimer's disease. *Intern Med* 34(6):481–484
97. Licastro F et al (1995) Increased serum alpha 1-antichymotrypsin in patients with probable Alzheimer's disease: an acute phase reactant without the peripheral acute phase response. *J Neuroimmunol* 57(1–2):71–75
98. Licastro F et al (1995) Acute phase reactant alpha 1-antichymotrypsin is increased in cerebrospinal fluid and serum of patients with probable Alzheimer disease. *Alzheimer Dis Assoc Disord* 9(2):112–118
99. Lawlor BA et al (1996) Acute phase reactants in Alzheimer's disease. *Biol Psychiatry* 39(12):1051–1052
100. Mulder SD et al (2009) CSF levels of PSA and PSA-ACT complexes in Alzheimer's disease. *Ann Clin Biochem* 46(Pt 6):477–483
101. Angelis P et al (1998) Serum interleukin-6 and interleukin-6 soluble receptor in Alzheimer's disease. *Neurosci Lett* 244(2):106–108
102. Cojocaru IM et al (2011) Study of interleukin-6 production in Alzheimer's disease. *Rom J Intern Med* 49(1):55–58
103. Hampel H et al (1997) Interleukin-6 is not altered in cerebrospinal fluid of first-degree relatives and patients with Alzheimer's disease. *Neurosci Lett* 228(3):143–146
104. Helmy AA et al (2012) Role of interleukin 6 and alpha-globulins in differentiating Alzheimer and vascular dementias. *Neurodegener Dis* 9(2):81–86
105. Marz P et al (1997) Interleukin-6 (IL-6) and soluble forms of IL-6 receptors are not altered in cerebrospinal fluid of Alzheimer's disease patients. *Neurosci Lett* 239(1):29–32
106. Murase K et al (1993) NGF level of is not decreased in the serum, brain-spinal fluid, hippocampus, or parietal cortex of individuals with Alzheimer's disease. *Biochem Biophys Res Commun* 193(1):198–203
107. Rosler N, Wichart I, Jellinger KA (2001) Clinical significance of neurobiochemical profiles in the lumbar cerebrospinal fluid of Alzheimer's disease patients. *J Neural Transm* 108(2):231–246
108. Singh VK, Guthikonda P (1997) Circulating cytokines in Alzheimer's disease. *J Psychiatr Res* 31(6):657–660
109. Teunissen CE et al (2003) Combination of serum markers related to several mechanisms in Alzheimer's disease. *Neurobiol Aging* 24(7):893–902
110. van Duijn CM, Hofman A, Nagelkerken L (1990) Serum levels of interleukin-6 are not elevated in patients with Alzheimer's disease. *Neurosci Lett* 108(3):350–354
111. Bonaccorso S et al (1998) Serotonin-immune interactions in elderly volunteers and in patients with Alzheimer's disease (DAT): lower plasma tryptophan availability to the brain in the elderly and increased serum interleukin-6 in DAT. *Aging (Milano)* 10(4):316–323
112. Hampel H et al (1999) Discriminant power of combined cerebrospinal fluid tau protein and of the soluble interleukin-6 receptor complex in the diagnosis of Alzheimer's disease. *Brain Res* 823(1–2):104–112
113. Yamada K et al (1995) Decreased interleukin-6 level in the cerebrospinal fluid of patients with Alzheimer-type dementia. *Neurosci Lett* 186(2–3):219–221
114. Hampel H et al (1998) Decreased soluble interleukin-6 receptor in cerebrospinal fluid of patients with Alzheimer's disease. *Brain Res* 780(2):356–359
115. Ojala J et al (2009) Expression of interleukin-18 is increased in the brains of Alzheimer's disease patients. *Neurobiol Aging* 30(2):198–209
116. Lee KS et al (2008) Bioplex analysis of plasma cytokines in Alzheimer's disease and mild cognitive impairment. *Immunol Lett* 121(2):105–109
117. Lindberg C et al (2005) Soluble interleukin-1 receptor type II, IL-18 and caspase-1 in mild cognitive impairment and severe Alzheimer's disease. *Neurochem Int* 46(7):551–557
118. Malaguarnera L et al (2006) Interleukin-18 and transforming growth factor-beta 1 plasma levels in Alzheimer's disease and vascular dementia. *Neuropathology* 26(4):307–312
119. Reale M et al (2012) Relationship between inflammatory mediators, Abeta levels and ApoE genotype in Alzheimer disease. *Curr Alzheimer Res* 9(4):447–457
120. Eikelenboom P et al (2010) Neuroinflammation—an early event in both the history and pathogenesis of Alzheimer's disease. *Neurodegener Dis* 7(1–3):38–41
121. Galimberti D et al (2003) Chemokines in serum and cerebrospinal fluid of Alzheimer's disease patients. *Ann Neurol* 53(4):547–548
122. Galimberti D et al (2006) Intrathecal chemokine synthesis in mild cognitive impairment and Alzheimer disease. *Arch Neurol* 63(4):538–543
123. Correa JD et al (2011) Chemokines in CSF of Alzheimer's disease patients. *Arq Neuropsiquiatr* 69(3):455–459
124. Tuppo EE, Arias HR (2005) The role of inflammation in Alzheimer's disease. *Int J Biochem Cell Biol* 37(2):289–305
125. Chao CC et al (1994) Transforming growth factor beta in Alzheimer's disease. *Clin Diagn Lab Immunol* 1(1):109–110
126. De Servi B et al (2002) Decrease of TGF-beta1 plasma levels and increase of nitric oxide synthase activity in leukocytes as potential biomarkers of Alzheimer's disease. *Exp Gerontol* 37(6):813–821
127. Juraskova B et al (2010) Transforming growth factor beta and soluble endoglin in the healthy senior and in Alzheimer's disease patients. *J Nutr Health Aging* 14(9):758–761
128. Rota E et al (2006) Increased intrathecal TGF-beta1, but not IL-12, IFN-gamma and IL-10 levels in Alzheimer's disease patients. *Neurol Sci* 27(1):33–39
129. Tarkowski E et al (2002) Increased intrathecal levels of the angiogenic factors VEGF and TGF-beta in Alzheimer's disease and vascular dementia. *Neurobiol Aging* 23(2):237–243
130. Zetterberg H, Andreasen N, Blennow K (2004) Increased cerebrospinal fluid levels of transforming growth factor-beta1 in Alzheimer's disease. *Neurosci Lett* 367(2):194–196
131. Rodriguez-Rodriguez E et al (2007) Serum levels and genetic variation of TGF-beta1 are not associated with Alzheimer's disease. *Acta Neurol Scand* 116(6):409–412
132. Lieberman J et al (1995) Serum alpha 1-antichymotrypsin level as a marker for Alzheimer-type dementia. *Neurobiol Aging* 16(5):747–753
133. DeKosky ST et al (2003) Plasma and cerebrospinal fluid alpha1-antichymotrypsin levels in Alzheimer's disease: correlation with cognitive impairment. *Ann Neurol* 53(1):81–90

134. Han Y et al (2012) Combination of plasma biomarkers and clinical data for the detection of sporadic Alzheimer's disease. *Neurosci Lett* 516(2):232–236
135. Licastro F et al (2001) Alpha-1-antichymotrypsin and oxidative stress in the peripheral blood from patients with probable Alzheimer disease: a short-term longitudinal study. *Alzheimer Dis Assoc Disord* 15(1):51–55
136. O'Bryant SE et al (2009) Brain-derived neurotrophic factor levels in Alzheimer's disease. *J Alzheimers Dis* 17(2):337–341
137. Li G et al (2009) Cerebrospinal fluid concentration of brain-derived neurotrophic factor and cognitive function in non-demented subjects. *PLoS One* 4(5):e5424
138. Angelucci F et al (2010) Alzheimer's disease (AD) and mild cognitive impairment (MCI) patients are characterized by increased BDNF serum levels. *Curr Alzheimer Res* 7(1):15–20
139. Laske C et al (2007) BDNF serum and CSF concentrations in Alzheimer's disease, normal pressure hydrocephalus and healthy controls. *J Psychiatr Res* 41(5):387–394
140. O'Bryant SE et al (2011) Serum brain-derived neurotrophic factor levels are specifically associated with memory performance among Alzheimer's disease cases. *Dement Geriatr Cogn Disord* 31(1):31–36
141. Parkitny L et al (2013) Multiplex cytokine concentration measurement: how much do the medium and handling matter? *Mediat Inflamm* 2013:890706
142. Jack CR Jr et al (2013) Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol* 12(2):207–216