

Contents lists available at [ScienceDirect](http://ScienceDirect.com)

## International Journal for Parasitology

journal homepage: [www.elsevier.com/locate/ijpara](http://www.elsevier.com/locate/ijpara)

## Invited Review

## Genome-based vaccine design: the promise for malaria and other infectious diseases



Denise L. Doolan\*, Simon H. Apte, Carla Proietti

Infectious Diseases Programme, QIMR Berghofer Medical Research Institute, Brisbane, QLD 4029, Australia

## ARTICLE INFO

## Article history:

Received 15 June 2014

Received in revised form 30 July 2014

Accepted 31 July 2014

Available online 6 September 2014

## Keywords:

Vaccine

Rational vaccine design

Genome-based

Malaria

Infectious diseases

'Omics'

## ABSTRACT

Vaccines are one of the most effective interventions to improve public health, however, the generation of highly effective vaccines for many diseases has remained difficult. Three chronic diseases that characterise these difficulties include malaria, tuberculosis and HIV, and they alone account for half of the global infectious disease burden. The whole organism vaccine approach pioneered by Jenner in 1796 and refined by Pasteur in 1857 with the “isolate, inactivate and inject” paradigm has proved highly successful for many viral and bacterial pathogens causing acute disease but has failed with respect to malaria, tuberculosis and HIV as well as many other diseases. A significant advance of the past decade has been the elucidation of the genomes, proteomes and transcriptomes of many pathogens. This information provides the foundation for new 21st Century approaches to identify target antigens for the development of vaccines, drugs and diagnostic tests. Innovative genome-based vaccine strategies have shown potential for a number of challenging pathogens, including malaria. We advocate that genome-based rational vaccine design will overcome the problem of poorly immunogenic, poorly protective vaccines that has plagued vaccine developers for many years.

© 2014 The Authors. Published by Elsevier Ltd. on behalf of Australian Society for Parasitology Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/3.0/>).

## 1. Vaccines – the global need

The term ‘vaccine’ is derived from *Variolae vaccinae*, literally smallpox of the cow, and was coined by Edward Jenner in 1798 in an article describing the protective effect of cowpox against smallpox (Jenner, 1798; Cohen et al., 1961; Baxby, 1999; Tuells, 2012). Since then, vaccines have been established as one of the most efficient and cost-effective interventions for the control and eradication of disease, and the prevention of morbidity and mortality worldwide. No other modality has had such a major effect on reducing mortality and improving public health, except for water sanitation (World Health Organization, 2006). Moreover, vaccinology is the only science that has eradicated an infectious disease (Andre, 2003), with the landmark achievement in 1977 of the eradication of smallpox (Fenner, 1982), a disease that plagued humankind and shaped our history since earliest civilisation (Fenner et al., 1988). It is anticipated that poliomyelitis will soon be eradicated, although some challenges remain (Pallansch and Sandhu, 2006).

Infectious diseases are responsible for one-third of all deaths worldwide, killing at least 15 million people each year ([\[www.who.int/healthinfo/global\\\_burden\\\_disease/gbd/en/\]\(http://www.who.int/healthinfo/global\_burden\_disease/gbd/en/\)\). They are clearly established as the leading cause of death of children globally and are responsible for 64–68% of deaths in children under 5 years of age, approximately 5 million children each year \(Black et al., 2010; Liu et al., 2012\). It is estimated that at least 3 million deaths per year are prevented by licensed vaccines currently in use \(World Health Organization, 2007, 2009\). Mass smallpox vaccination of children became compulsory in the United Kingdom \(UK\) in 1853 and vaccines are now available for most viral and bacterial diseases common in children including diphtheria \(1923\), whooping cough \(1926\), tetanus \(1937\), influenza \(1942\), pertussis \(1949\), polio \(1958 and 1961\), measles \(1963\), mumps \(1967\), rubella \(1969\), bacterial meningitis \(1974\), pneumonia \(1983\), varicella \(1995\) and rotavirus \(1998\) \(\[http://www.who.int/immunization/policy/position\\\_papers/en/\]\(http://www.who.int/immunization/policy/position\_papers/en/\)\). However, many of these existing vaccines are underutilised and the World Health Organization has estimated that 2.5 million children under the age of 5 years die from vaccine-preventable diseases each year, more than 6800 child deaths every day \(World Health Organization, 2007\). There are also many serious pathogens for which effective vaccines are not yet available including hepatitis C virus \(HCV\), human immunodeficiency virus \(HIV\), Dengue, respiratory syncytial virus \(RSV\) and cytomegalovirus \(CMV\); bacteria \(e.g. \*Mycobacterium tuberculosis\* \(TB\), Group A streptococcus \(GAS\), Group B](http://</a></p>
</div>
<div data-bbox=)

\* Corresponding author. Address: QIMR Berghofer Medical Research Institute, 300 Herston Road, QIMR Locked Bag 2000, Royal Brisbane Hospital, Brisbane, QLD 4029, Australia. Tel.: +61 7 3362 0382; fax: +61 7 3362 0111.

E-mail address: [Denise.Doolan@qimrberghofer.edu.au](mailto:Denise.Doolan@qimrberghofer.edu.au) (D.L. Doolan).

*Streptococcus* (GBS), *Staphylococcus aureus*, *Meningococcus* Group B (MenB), *Shigella*, pathogenic *Escherichia coli*); and parasites (e.g. *Plasmodium*, *Leishmania*, *Schistosoma*, *Trypanosoma*) and these are estimated to claim in excess of 3 million more lives each year (World Health Organization, 2005, 2007).

It is noteworthy that despite the demonstrated success of vaccines in preventing illness caused by viral and bacterial pathogens, there are not yet any licensed vaccines for parasitic infections of humans or for any chronic infections by complex pathogens (World Health Organization, 2006; Moorthy and Kienny, 2010). The pathogens causing these diseases have adapted to long-term coexistence with the human immune system and have evolved sophisticated immune evasion strategies. Many express hundreds or thousands of potential antigenic targets, often in distinct phases of their life cycles, so it is perhaps not surprising that vaccine efforts to develop vaccines based on only a limited number of antigens, often selected on an ad hoc basis and without knowledge of the antigenic repertoire of the organisms, have not been successful.

Moreover, all currently licensed vaccines for infectious diseases are prophylactic, preventing the effects of a future infection by the target pathogen, and there are no licensed therapeutic vaccines for any chronic or acute infectious disease. The only therapeutic vaccine approved by the United States Food and Drug Administration (US FDA) is for a certain type of metastatic prostate cancer (Provenge<sup>®</sup>, DendreonCorp, USA; approved in 2010). Prophylactic vaccines have been developed to prevent human papilloma virus (HPV) and hepatitis B virus (HBV) which cause chronic infections and in some cases cancer (Plotkin, 2008; Levine and Esparza, 2009). However, effective vaccines against the three pathogens responsible for more than half of the global burden of infectious diseases (malaria, HIV, TB) (World Health Organization, 2006) will need to be therapeutic, given the chronicity of these infections in endemic regions.

## 2. History of vaccines

The field of vaccinology originated on 14 May 1796 when Edward Jenner inoculated an 8 year old boy named James Phipps with vaccinia virus contained in pus from lesions on the hand of a milkmaid with cowpox and showed that Phipps did not become infected with smallpox when subsequently variolated (inoculated, or in today's parlance – challenged, with smallpox). Jenner's seminal study predated formal evidence for the germ theory of disease (microbial origin) obtained by Louis Pasteur in 1857 and Robert Koch in 1876 (D'Argenio and Wilson, 2010) which provided the foundation of empirical vaccine development. A century after Jenner's observation, proof-of-concept was established with the development by Louis Pasteur of an attenuated vaccine for chicken cholera in 1879, an anthrax vaccine in 1881, and a rabies vaccine in 1885 <http://www.historyofvaccines.org>.

These and subsequent “first generation” vaccines (e.g. Bacillus Calmette Guerin (BCG)) consisted of live-attenuated pathogens (typhoid, pertussis, measles, mumps, rubella) or inactivated killed pathogens (polio, rabies, cholera, hepatitis A, bubonic plague) and were developed according to the classical “isolation, inactivation and injection of disease-causing pathogen” approach to vaccine development established by Pasteur, and using Koch's postulates as a general guide.

In the second half of the 20th Century, significant advances in many fields including cell culture (enabling the growth of viruses in vitro), polysaccharide chemistry, recombinant DNA technology and immunology, allowed the development of “second generation” vaccines which comprised purified pathogen components such as protein antigens or polysaccharides (e.g. polio, measles, mumps, rubella, chickenpox, tetanus, diphtheria, anthrax, influenza, hepatitis

A, hepatitis B, rotavirus, influenza, pneumonia and human papilloma-virus (Hilleman, 1999; Finco and Rappuoli, 2014; Rhee, 2014). Maurice Hilleman is widely recognised as a pioneer and the most prolific developer of vaccines in this period, developing more than 40 vaccines (Hilleman, 1999; <http://www.historyofvaccines.org>).

It was in this era that Jonas Salk, who developed the first (inactivated) vaccine against polio, defined vaccinology as “the application of the basic requirements for effective immunisation” which include (i) stimulation with a sufficient quantity of antigen, (ii) use of a suitably specific antigen, and (iii) the induction of an appropriate immune response for the prevention of the pathological consequences of infection; also noting that vaccinology “requires an understanding of the etiologic agents, the pathogenic mechanisms, and the epidemiology of the individual diseases” (Salk and Salk, 1977). The majority of currently-licensed vaccines consist of either killed (inactivated) or live attenuated pathogens, or pathogen-related biomolecules including toxoids or polysaccharides (Grimm and Ackerman, 2013; U.S. Food and Drug Administration, 2014). It is notable that most currently licensed vaccines target pathogens with a relatively low degree of antigen variability and work mainly by eliciting functional antibodies (De Gregorio and Rappuoli, 2012).

For many significant pathogens, the generation of broadly protective vaccines has remained elusive. Such pathogens often present technical obstacles to the vaccinologist that include their inability to be cultured in vitro (e.g. *Mycobacterium leper*, papilloma virus type), have antigenic hypervariability (e.g. serogroup B meningococcus, HIV, HCV), or whose life cycles have an intracellular phase that puts them out of the reach of antibodies and therefore require a cellular immune response – controlled predominantly by T cells (e.g. malaria, tuberculosis); furthermore, traditional approaches to vaccine design and development do not allow the rapid development of new vaccines for pandemic agents (Finco and Rappuoli, 2014). Most second generation vaccines do not target these types of pathogens or elicit the correct types of responses and we have reached a point where most of the low-hanging fruit has been taken. These challenges are significant and explain why we have reached the end of the age of second generation vaccine development.

Advances in genomics and other “omics” over the past two decades have given rise to a “third generation” of vaccines (e.g. Meningococcus group B, group A streptococcus, group B streptococcus, *S. aureus*, *E. coli*, *Clostridium difficile*) based on technologies such as reverse vaccinology pioneered by Rappuoli (2000), structural biology and synthetic vaccines (Delany et al., 2014; Finco and Rappuoli, 2014). This activity has resulted in vaccines that protect against an increased range of vaccine-preventable diseases, that are multivalent and target different serotypes, or highly purified vaccines with an improved safety profile, and replace the more reactogenic whole cell vaccines (Rappuoli et al., 2011). These advances, combined with knowledge gained from successes with vaccine development against acute diseases in the 20th Century, provide the foundation for the development of vaccines that have thus far proved elusive, including those that are therapeutic (for chronic diseases), require cellular immunity for protection, for pregnant women and elderly or immunocompromised people, or for new indications such as autoimmune disease and cancer (Poland and Barrett, 2009; Rappuoli et al., 2011). They also enable more efficient pathways for vaccine development and new technologies for assessment of vaccine safety, which are especially pertinent given increased public scrutiny of adverse events associated with vaccination and stringent regulatory requirements for vaccine approval (Rappuoli et al., 2011).

On May 8, 1980, the World Health Assembly certified the world free of naturally occurring smallpox, representing a landmark achievement in the history of vaccinology (Fenner, 1982). The

Global Alliance for Vaccines and Immunisation (GAVI) was launched at the World Economic Forum on 31 January, 2000 with the involvement of the major players in global immunisation, including representatives and leaders from key United Nations (UN) agencies, bilateral aid agencies, the pharmaceutical industry, major philanthropic foundations and government bodies, all partnering with the aim to deliver vaccines to millions of the world's poorest children and boost immunisation coverage (<http://www.gavialliance.org/>). A decade later, on 29 January 2010, Bill and Melinda Gates issued a challenge to participants at the World Economic Forum's annual meeting to make this the "Decade of Vaccines" with a commitment of \$10 billion over the next 10 years to help research, develop and deliver vaccines for the world's poorest countries, with a specific goal to prevent the deaths of approximately 8 million children under the age of 5 years from 2010 to 2019 ([http://www.gatesfoundation.org/Media-Center/Press-Releases/2010/01/Bill-and-Melinda-Gates-Pledge-\\$10-Billion-in-Call-for-Decade-of-Vaccines](http://www.gatesfoundation.org/Media-Center/Press-Releases/2010/01/Bill-and-Melinda-Gates-Pledge-$10-Billion-in-Call-for-Decade-of-Vaccines)). In December 2010, stakeholders from the global health community, including the World Health Organization (WHO), the United Nations Children's Fund (UNICEF), the US National Institute of Allergy and Infectious Diseases (NIAID) and the Bill & Melinda Gates Foundation, formally launched the Decade of Vaccines Collaboration to generate a Global Vaccine Action Plan (GVAP) 2011–2020 ([World Health Organization, 2013](http://www.who.int/vaccines-strategy/action-plan)). Following extensive consultation, the GVAP was endorsed by the 194 Member States of the World Health Organization Assembly on 11 May 2012 to achieve the Decade of Vaccines vision by delivering universal access to immunisation and preventing millions of deaths by 2020 and beyond. The GVAP outlines six strategic objectives to strengthen routine immunisation, and develop and introduce new and improved vaccines and vaccine technologies ([Keith et al., 2013](http://www.who.int/vaccines-strategy/action-plan)).

### 3. Empirical to rational vaccine development

Historically, vaccine development has been based on empirical trial-and-error approaches designed to mimic, by vaccination, the immunity induced by natural infection, which originated in the late 17th Century with Jenner. With this classical "isolate, inactivate and inject" approach, protection against a wide range of bacterial and viral pathogens (but not parasitic pathogens of humans) has been achieved, and whole-organism vaccines still represent a large proportion of the vaccines licensed today.

There are both advantages and disadvantages associated with whole-organism vaccines ([Zepp, 2010](#); [Rappuoli et al., 2011](#); [Grimm and Ackerman, 2013](#)). A major advantage is that they do not require prior knowledge of immune responses associated with protection, or of the pathogen genome or proteome. Moreover, they typically induce sustained protective immune responses without the requirement for adjuvant. Importantly, whole-organism vaccines provide an extensive repertoire of pathogen-derived B cell and T cell epitopes for recognition by the immune system of the vaccinee in the context of multiple genetic backgrounds found in heterogeneous human populations, although only a fraction of these epitopes may be important targets of protective immunity. In contrast, highly purified antigens likely present only a limited number of epitopes and therefore may not be recognised by all individuals (immunological non-responsiveness).

Despite these advantages, whole-organism killed, inactivated or live attenuated vaccines are typically complex and their poorly characterised products have often raised safety concerns including reversion to virulence for live attenuated pathogens, induction of autoimmunity, or unacceptable reactogenicity associated with the induced inflammatory response ([Rappuoli et al., 2011](#)).

Additionally, critical conformational epitopes may be disrupted by the inactivation process ([Grimm and Ackerman, 2013](#)). The most infamous example of whole-organism virulence may be the "Cutter Incident" in 1955 when thousands of American children were inadvertently immunised with live polio virus contained in what was supposed to be an inactivated polio vaccine. This tragically resulted in a number of deaths and many cases of paralysis ([Brown, 1993](#)), and damaged the public's perception of vaccine safety. Some of these problems have been overcome with improved technology and quality control, but even in recent years, unanticipated outcomes have resulted from the use of whole-organism vaccines. Recently, in Australia, an unexpectedly high number of adverse events in children was associated with receipt of the 2010 trivalent influenza vaccine (FluVax) manufactured by CSL Biotherapies (Australia), possibly due to suboptimal virus splitting ([Maraskovsky et al., 2012](#)). Even with the application of more advanced technologies to genetically attenuate the pathogen, the potential for the pathogen to circumvent the attenuating mutation has been noted ([Mikolajczak et al., 2014](#)). Moreover, increased public and regulatory scrutiny of vaccine safety has raised the bar for what constitutes acceptable risk for vaccines ([Kennedy and Poland, 2011](#)) and indeed many, if not most, of the first generation whole-organism vaccines that have proved successful and overwhelmingly safe (and saved millions of lives) would not be approved for use today.

Additionally, the whole-organism approach is almost exclusively restricted to pathogens that can be cultured *in vitro*, and although it has worked for a number of "simple" pathogens with relatively low antigen variability, it has not succeeded for many complex pathogens including those that cause chronic infections and are capable of evading or misdirecting the host immune response. Also, traditional vaccine design is based on the premise of mimicking the immunity induced by natural exposure, but for many pathogens this is suboptimal and robust sustained protection may require inducing immunity that is better than nature, with minimal adverse effects associated with stimulating the inflammatory response ([Zepp, 2010](#)). This is especially true for a chronic infection, where the pathogen is able to co-exist with the host for an indefinite period of time despite the presence of immune responses induced by the host and targeted against the pathogen. Finally, for effective long-term protection, it is important that the vaccine-induced immunity be sustained even in the absence of pathogen exposure to allow for rapid recall of the required immune memory.

To overcome these challenges, vaccine design has become more tailored, focusing on the antigen moieties targeted by protective immune responses ([Zepp, 2010](#)) with the conceptual framework changing from a reductionist view to a holistic view, with a broad perspective of the pathogen and its interaction with the host immune system ([D'Argenio and Wilson, 2010](#)). Thus, the modern era of vaccinology is multidisciplinary, exploiting enhanced knowledge or improved understanding and technological innovations in basic immunology, molecular biology, microbiology, host-pathogen biology, microbial pathogenesis, genetics, biotechnology, bioinformatics and computational science, as well as other fields ([D'Argenio and Wilson, 2010](#); [Kennedy and Poland, 2011](#)). As noted by Rino Rappuoli, "Vaccines have progressed from the crude preparations used to prevent smallpox to one of the most technologically advanced and effective public health interventions devised by man" ([Rappuoli et al., 2011](#)).

### 4. Rational vaccine development in the modern era

The ideal vaccine would induce robust and sustained immune responses in all individuals in all populations, regardless of age or immune status, in the absence of any adverse effects, and would

be effective against all strains and species even for complex pathogens (modified from Zepp, 2010). Rational vaccine design offers the potential to achieve this goal, facilitated by advances in three core areas: (1) conceptual advances, with improved understanding of basic immunology, host–pathogen interactions and pathogenesis; (2) technological innovations in molecular biology, biotechnology, delivery systems and other enabling technologies; and (3) emerging knowledge in ‘omics’ and other data-intensive sciences, combined with bioinformatics and computational biology capability as well as cross-disciplinary approaches.

#### 4.1. Conceptual advances

##### 4.1.1. Learning from nature

Historically, vaccines have been designed to mimic the immunity induced by natural exposure to the target pathogen (Zepp, 2010). In a number of cases, the phenotype of the critical responding cells, mechanisms of immunity and immunological memory, and immune correlates of protection are unknown. As an example, the BCG vaccine for tuberculosis developed in 1927 has been administered approximately four billion times but its mechanism of protective immunity is still not known (Kaufmann et al., 2014). The premise of rational vaccine design is to induce the desired immune response against the key pathogen antigen(s) or epitopes which are targeted by protective immune responses. Dissecting the immune response to the target pathogen following natural infection in humans is a logical step to identify the immune response(s) that should be induced by vaccination and the pathogen antigens targeted by that response(s). A number of factors should be considered.

**4.1.1.1. Immune activation – innate immunity, adaptive immunity, and the innate/adaptive interface.** The requisite steps leading to in vivo immune activation have been reviewed in detail elsewhere (Moser and Leo, 2010; Zepp, 2010; Six et al., 2012). Although traditional vaccine design has focused on the adaptive immune response, it is now recognised that innate immune responses and the innate-adaptive interface are critical for induction of robust protective immune responses. The current state of knowledge indicates that the rate limiting step for vaccination is antigen processing and presentation by dendritic cells, which ultimately directs the differentiation of functionally distinct subsets of immune cells with different effector functions. Soluble proteins appear to be particularly poor in inducing dendritic cell maturation because, presumably, they lack the so-called “danger signal” but this deficiency can be overcome at least in part by adjuvant formulations (Moser and Leo, 2010). The choice of adjuvant has been demonstrated to significantly impact vaccine efficacy (e.g. RTS,S vaccine; (Stewart et al., 2006)). Nonetheless, until recently, Alum was the only adjuvant globally approved for human use (Eisenbarth et al., 2008), but MF59 and monophosphoryl lipid A-containing formulations have received regulatory approval in defined human vaccines (Rueckert and Guzman, 2012). Considerable efforts over the past few decades have been devoted to adjuvant discovery, and some promising adjuvants are now moving through the regulatory pipeline (Reed et al., 2013).

**4.1.1.2. Activation of multiple arms of the immune system.** Almost all licensed vaccines currently available, as well as those that were previously available but have been removed or replaced, rely on the induction of functional antibodies which neutralise or opsonise the target pathogen (Six et al., 2012; Delany et al., 2014). Some (e.g. tetanus) work by neutralisation of the toxin. Although some of these vaccines do induce cellular immune responses (e.g. pertussis, (Mills et al., 1998); hepatitis B (Ren et al., 2003)), the protection is thought to be antibody-mediated and, to the best of our

knowledge, there are no licensed vaccines that are specifically designed to induce protective cellular immune responses. However, for many pathogens, including intracellular pathogens (Seder and Hill, 2000), cellular immune responses play a critical role in protection, either directly via CD8<sup>+</sup> and CD4<sup>+</sup> T cell cytotoxicity targeting pathogen-infected cells or indirectly via cytokines or other mediators produced by the T cells. Additionally, indirectly, helper T cells are required for optimal B cell activation and high affinity antibody responses, and influence the antibody isotype as well as the development of immune memory (Moser and Leo, 2010; Zepp, 2010; Delany et al., 2014). Indeed, the development of vaccines against those pathogens that have thus far proved elusive will likely require activation of all arms of the immune system with functional antibodies as well as efficient CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses, with appropriate selection of pathogen antigen relevant for each immune response. Since conventional technologies have proved ineffective in generating robust cell-mediated immunity, innovative vaccine delivery systems will be crucial (see Section 4.2).

**4.1.1.3. Quality of the immune response is not the same as quantity.** Conventional assessment of vaccine-induced immune responses or correlates of immunity has typically focused only on the frequency and magnitude, or quantity, of the immune response; e.g. ELISA titres as a measure of the peak antibody response; cytotoxic capacity or frequency of antigen-specific T cells as a measure of the T cell response. However, the quality of the immune response is at least as important as the quantity, and quality can be a key determinant of protection (Zepp, 2010). For example, the standard ELISA assay for antibody responses quantifies antibodies that bind to the pathogen but this provides no indication of the capacity of those antibodies to be functional (Plotkin, 2008). In the case of antibody responses, quality can be assessed via affinity, avidity, diversity and biological function, as well as Ig subclass (Plotkin, 2008; Courtin et al., 2009; Feng et al., 2009). For T cell responses, quality can be assessed via phenotypic markers, differentiation state (central memory, effector memory, effector T cells), profile of secreted cytokines and poly-functionality, TCR avidity and TCR repertoire diversity (Germain, 2010; Sallusto et al., 2010). For both antibodies and T cells, the generation and maintenance of immunological memory is critical to ensure rapid reactivation upon pathogen exposure. Recent advances in immune monitoring and immune profiling, and establishment of consortia such as the Human Immunology Project Consortium (<http://www.immuneprofile.org>), provide the foundation for comprehensive evaluation of vaccine-associated immune parameters, and identification of novel immune mediators and pathways (Newell et al., 2013; Poland et al., 2013; Brusic et al., 2014).

**4.1.1.4. Immune evasion, antigenic polymorphism and chronic diseases.** With the conceptual and technological advances of the past decades, we have an improved understanding of antigenic polymorphism and immune evasion, and the associated challenges for vaccine development, especially for chronic diseases. It is now recognised that some pathogens are able to evade host immunity by mutating key antigens or epitopes with evolutionary pressure or during the course of disease. For example, the *Plasmodium* spp. parasite has co-evolved with the human host over millions of years with selection of polymorphisms within the human genome that represent some of the most common monogenic human disorders, such as  $\alpha$ -thalassemia and hemoglobinopathies including sickle-cell trait (Mackinnon and Marsh, 2010; Taylor et al., 2013). Concomitantly, on the parasite side, polymorphisms within antigens or epitopes targeted by host immune responses have been demonstrated within relatively restricted time frames as a means



of circumventing the host immune response (Doolan et al., 1991; Barry et al., 2009; Takala and Plowe, 2009). In addition to polymorphism at the nucleotide level, allelic polymorphism whereby more than one allele exists for specific regions of a protein (e.g., *Plasmodium falciparum* merozoite surface protein 1 or apical membrane antigen 1; *Plasmodium vivax* Duffy Binding Protein), as well as antigenic variation of the asexual blood stages of *Plasmodium* where the parasite expresses at any given time only one member of a family of multi-copy variant surface antigens (e.g., PfEMP1), are also well established (Takala and Plowe, 2009; Kirkman and Deitsch, 2012). Examples of immune escape through antigenic polymorphism or allelic variation have been also demonstrated for other pathogens, with well known examples including influenza (Yewdell, 2013), HIV (Moore et al., 2002; Boutwell et al., 2010; Julien et al., 2012) and hepatitis C virus (Petrovic et al., 2012). Such variation poses significant challenges for vaccine development, since a vaccine that has high strain-specific efficacy against the antigen included in the vaccine would nonetheless be poorly effective against variant circulating strains in the field (Moorthy and Kieny, 2010). This has been demonstrated experimentally with a candidate three-component blood-stage vaccine against malaria known as Combination B (Genton et al., 2002) and subsequent field studies with recombinant merozoite surface protein 1 (MSP1) (Ogutu et al., 2009; Otsyula et al., 2013) and apical membrane antigen 1 (AMA1) vaccines (Ouattara et al., 2010, 2013; Thera et al., 2011). However, recent technological advances (see below) have allowed the development of multivalent vaccines against pathogens with multiple strains or serotype (e.g. the licensed 7-valent, 10-valent or 13-valent conjugate vaccines for *Pneumococcus* or the 4-valent vaccine for *Meningococcus* (De Gregorio and Rappuoli, 2012). Other approaches have demonstrated potential in preclinical evaluation, including chimeric vaccines (e.g. *P. falciparum*, Dutta et al., 2007; Faber et al., 2013) mosaic vaccines (e.g. HIV, Corey and McElrath, 2010), or structure-based vaccine design (see Section 4.3.3).

**4.1.1.5. Not all antigens are equal.** It is now recognised that not all antigens or pathogen-derived epitopes are equal in terms of their capacity to be recognised by the host immune response. This phenomenon, where pathogen-specific immune responses target only a small fraction of the full range of possible antigens or peptide epitopes, is known as immunodominance (Sercarz et al., 1993; Allen et al., 2001). The factors underlying the hierarchy of pathogen-specific epitope recognition are largely unknown, especially in the context of a complex pathogen, but could result from features intrinsic to the protein antigen such as protein abundance, kinetics of protein expression during infection, and rate of synthesis or decay. It could also result from properties intrinsic to the epitope(s), including efficiency of antigen processing and presentation, epitope abundance, major histocompatibility complex (MHC) binding affinity or precursor frequency (Yewdell and Bennink, 2001; Assarsson et al., 2007; Kotturi et al., 2008; Tan et al., 2011). The identification within this hierarchy of antigens (or epitopes) that are key targets of protective immune responses and that will stimulate effective immunity against the target pathogen is a key component of rational vaccine design (Rueckert and Guzman, 2012). There is currently no algorithm or universal approach to identify the important antigens and epitopes, and historically antigens for inclusion in subunit vaccines have been selected empirically, often based on antigen density and cell surface accessibility. In some cases, cell surface proteins have proved to be effective targets of neutralising antibodies; e.g. hepatitis B surface antigen (Schillie and Murphy, 2013). However, in many cases, antigens identified empirically may not be the optimal target of protective immune responses, may be poorly immunogenic, or may have associated safety concerns (Rappuoli et al., 2011).

Advances in the genomic era, with the sequencing of pathogen genomes and systems immunology approaches to define the requisite immune response, offer the potential to overcome these limitations, particularly when the genome of the pathogen is large.

#### 4.1.2. Improving on nature

As stated above, traditional vaccines aim to mimic the immunity induced naturally following infection with the pathogen. However, this approach has not been successful for many diseases, including those caused by pathogens where immunity is typically slow to induce and non-sterilising (e.g. malaria, RSV, *Pseudomonas aeruginosa*) or where infections are chronic or have a latent phase (e.g. HIV, hepatitis C virus, *S. aureus*) (Delany et al., 2014). Thus, modern vaccinology must improve on nature. There are a number of examples which demonstrate that this is possible, specifically: (i) higher titres of neutralising antibodies can be induced in adults by vaccination with a human papilloma virus virus-like particle (VLP) vaccine adjuvanted with AS04 than are induced naturally (Schwarz and Leo, 2008; Zepp, 2010). (ii) Human papilloma virus VLP vaccines can induce a systemic protective immune response, even though the virus enters only via the mucosal system and remains localised to the basal cell layer of the squamous epithelium (Schwarz and Leo, 2008; Zepp, 2010). (iii) Higher titres of neutralising antibodies can be induced in children by vaccination with *Haemophilus influenzae* type b, pneumococcal or meningococcal polysaccharide-protein conjugate vaccines that convert the induced antibody responses from T cell-independent to T cell-dependent responses than are induced naturally (D'Argenio and Wilson, 2010). (iv) A candidate vaccine against *Chlamydia trachomatis* infection comprising the major outer membrane protein (nMOMP) solubilised with amphipols can induce more robust chlamydia-specific humoral and cell-mediated immune responses and protective immunity in mice, by stabilising the protein and increasing epitope accessibility (Tifrea et al., 2014). Additionally, recent advances in approaches such as structural vaccinology are specifically tailored to design vaccines that do better than nature (see Section 4.3.3).

#### 4.2. Technological innovations

Rational vaccine design in the modern era is facilitated by technological innovations in many areas including: (i) technical advances in recombinant DNA technology and biotechnology; (ii) manufacturing improvements including fermentation, production, purification and consistency in manufacturing scale-up; and (iii) enabling technologies which include novel vaccine platforms (adjuvants, viral vectors, DNA vaccines, RNA vaccines, virus-like particles (e.g. human papilloma virus vaccine, liposomes, nanoparticles, mosaic vaccines)) as well as delivery systems (skin patches, electroporation). These have been reviewed in detail elsewhere (Hilleman, 1999; Zepp, 2010; Grimm and Ackerman, 2013).

#### 4.3. Emerging knowledge from genome-scale datasets

Perhaps the most important influences on rational vaccine design have been the tremendous advances made in the genomic era, which began with the sequencing of the complete genome of *H. influenzae* in 1995 (Fleischmann et al., 1995). Since then, the genomes, proteomes and transcriptomes of many pathogens have been elucidated (<http://www.genomesonline.org/>). This information provides the foundation for novel 21st Century approaches to identify target antigens for the development of vaccines, drugs and diagnostic tests; for the development of innovative vaccine delivery systems; and for the development and application of methods to identify immune correlates of protection. These data-rich sets of information have given rise to a large number of new scientific fields

known as “omics” (Baker, 2013; <http://www.genomicglossaries.com/content/omes.asp>), as well as revolutionary conceptual and technological advances which are largely cross-disciplinary. Those most relevant to vaccine design include reverse vaccinology, immunomics, structural vaccinology and systems immunology. These, and others related to the field, have been reviewed elsewhere by us (Doolan, 2011; Schussek et al., 2014) and others (Dormitzer et al., 2008, 2012; Pulendran et al., 2010; De Gregorio and Rappuoli, 2012; Grimm and Ackerman, 2013; Thomas and Luxon, 2013; Delany et al., 2014; Finco and Rappuoli, 2014), so will be only briefly summarised here to provide context.

#### 4.3.1. Reverse vaccinology

Reverse vaccinology, pioneered by Rappuoli and colleagues in 1999 (Rappuoli, 2000), is considered a milestone in vaccinology and biotechnology because it exploits the power of in silico analysis to identify, from the complete genome of the target pathogen, a subset of genes encoding proteins with characteristics associated with vaccine-induced immunity (e.g. location on cell surface) which are then systematically evaluated for capacity to induce the desired immune response/immunogenicity (Rappuoli, 2000). Proof-of-concept for reverse vaccinology was established with *Neisseria meningitidis* (Pizza et al., 2000), identifying in just 18 months more surface exposed antigens than had been discovered in 40 years of conventional vaccinology; a vaccine based on three of these novel antigens and outer membrane vesicles is now licensed in 30 countries (Gorringe and Pajon, 2012).

#### 4.3.2. Immunomics

Immunomics is the study of the subset of pathogen-derived proteins or their epitopes that are recognised by the host immune system. In contrast to reverse vaccinology, immunomics-based identification of vaccine candidates relies not only on in silico prediction algorithms, but also takes advantage of biological samples from humans or animals with immunity to the disease of interest, to define the set of antigens or epitopes that interface with the host immune system (Doolan, 2011; Sette et al., 2005). Thus, immunomics specifically addresses the interface between the host immune system and the pathogen proteome (Klysik, 2001). No vaccines derived from immunomics have yet reached the stage of clinical testing but a number of promising candidate antigens have been identified by us in the malaria model using antibody-based (Doolan et al., 2008; Trieu et al., 2011) or T cell-based (Doolan et al., 2003; Doolan, 2011) approaches.

#### 4.3.3. Structure-based vaccine design or structural vaccinology

Most recently, advances in three-dimensional structural biology enabling determination of protein tertiary structure and potential conformational B cell epitopes has provided the foundation for structural vaccinology or structure-based vaccine design (Dormitzer et al., 2008, 2012; Thomas and Luxon, 2013; Delany et al., 2014). The underlying rationale is that protective epitopes (rather than complete proteins) should be sufficient to induce immune responses and provide protection against pathogens, and constructs can be engineered such that they are more stable, expose hidden cryptic epitopes, or enable broadly cross-protective responses against pathogen variants. Proof-of-concept has been demonstrated in preclinical studies for a number of pathogens, including the highly variant *N. meningitidis* serogroup B surface-exposed factor H-binding protein, the group B *Streptococcus* pilus protein, the F glycoprotein of RSV and influenza HA (Dormitzer et al., 2012).

#### 4.3.4. Systems immunology

Systems immunology (systems vaccinology) falls under the broad umbrella of systems biology and aims to study the immune

system in an integrated perspective rather than studying isolated components, in order to identify immune correlates of protection or signatures of immunogenicity (Querec et al., 2009; Pulendran et al., 2010; Kaufmann et al., 2014). This approach, which successfully predicted the immunogenicity of the highly efficacious yellow fever vaccine in humans (Querec et al., 2009), incorporates a holistic view of the overall host-pathogen interaction as opposed to the simple conventional readouts of vaccine-induced immune responses such as ELISA or ELISpot assays (Six et al., 2012). Such studies would be expected to provide a comprehensive understanding of the host-pathogen interaction and its regulation; to identify novel immune mediators and pathways, and correlates of vaccine efficacy, facilitating vaccine evaluation in the clinic.

## 5. Malaria vaccine development

### 5.1. Challenges and feasibility

The *Plasmodium* parasite has a large, complex genome encoding thousands of potential antigenic targets expressed at different stages of a complex life cycle. It has developed a range of effective strategies for evading the human immune response, including allelic and antigenic variation, a predominantly intracellular existence, and the ability to down-regulate the host's immune response. Current vaccine approaches have failed to yield an effective *P. falciparum* vaccine in spite of 50 years of dedicated effort. Nevertheless, two experimental human vaccine models suggest that a malaria vaccine is indeed a feasible goal.

Sterile infection-blocking immunity can be induced in mice, monkeys and humans by immunisation with *Plasmodium* sporozoites attenuated by radiation such that they can invade the host hepatocyte but do not fully develop (Nussenzweig and Nussenzweig, 1989; Hoffman et al., 2002). T cell responses (in particular CD8<sup>+</sup> T cells) directed against parasite antigens expressed in the infected hepatocyte are considered the primary immune effectors, although antibodies also play a role in inhibiting sporozoite invasion of the hepatocyte (Good and Doolan, 2010). Secondly, individuals surviving past early childhood in areas of high malaria transmission develop substantial clinical immunity and rarely die from malaria, although they are frequently infected (Baird, 1998). Antibodies directed against antigens exposed on the surfaces of merozoites or infected erythrocytes, or released from apical organelles at the moment of invasion, are thought to be the critical effectors since passive transfer of purified immunoglobulin from individuals with lifelong exposure to endemic malaria results in a marked decrease in blood-stage parasitemia and resolution of symptoms in the recipients (Cohen et al., 1961; Langhorne et al., 2008). The human models represented by irradiated sporozoite immunisation or naturally acquired immunity are powerful models for the development of a malaria vaccine to completely prevent infection or death and severe disease, respectively.

More recently, a practical demonstration of the feasibility of a malaria vaccine has been shown by the impact of the RTS,S vaccine. Although phase 3 results with RTS,S/AS01 suggest that efficacy is very low in the target age group and is not sustained (Duncan and Hill, 2011; Olotu et al., 2013), the vaccine may nonetheless receive regulatory approval for implementation through the routine Expanded Program for Immunisation (Bouchie, 2013) since modelling suggests that it may have an impact on the number of “deaths averted” (Smith et al., 2012; Nunes et al., 2013). These data are especially encouraging since the vaccine is based on only a single target antigen (*P. falciparum* circumsporozoite protein) amongst the more than 5,000 expressed by the parasite. However these poor efficacy results highlight the need to include more antigens and induce a broad multi-

pronged immune response involving CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses as well as antibody responses.

### 5.2. Progress, or lack thereof

Malaria vaccine development has focused on subunit vaccines against a very limited number of target antigens (World Health Organization, 2006; Schwartz et al., 2012). Nearly all *Plasmodium* antigens present in the genome have the potential to be targets for effective vaccines, drugs or diagnostic tests and remain so until each is systematically assessed. What distinguishes one protein from another for candidacy as a target for a new drug or as an immunogen for a new vaccine is often determined empirically. Selection of vaccine targets has been based on a variety of criteria which, while not irrational, are not systematic. For example, although some asexual blood-stage antigens were identified by probing a cDNA expression library with antibodies present in inhibitory sera from malaria-exposed individuals (Kemp et al., 1983), other antigens have come to attention due to historical reasons related to the ease with which murine or rabbit antisera were generated against them. Their selection as candidate vaccine targets may be validated by immunoepidemiological evidence of associations between immune responses and clinical immunity, neutralisation or adoptive transfer experiments in animal systems, or induction of protective immunity in animal models with homologous antigens from murine or simian malaria species.

For most of the past half century, malaria researchers attempting to develop a pre-erythrocytic stage malaria vaccine have focused almost exclusively on a single antigen, the circumsporozoite protein (CSP). Several lines of evidence supported the initial focus on the CSP as a pre-erythrocytic stage vaccine candidate (reviewed in Doolan and Martinez-Alier, 2006). In 1997, 16 years after the CSP was identified and 15 years after GlaxoSmithKline (UK) and the Walter Reed Army Institute of Research (WRAIR, US) entered a collaborative agreement to produce a malaria vaccine, it was demonstrated that the RTS,S vaccine could protect ~40% of malaria-naïve volunteers against sporozoite challenge (Stoute et al., 1997; Cohen et al., 2010). A review of the history of the RTS,S vaccine is sobering since in the development effort nearly a dozen other constructs based on the CSP were tested pre-clinically and as many as six different vaccines were tested in the clinic in Phase 1, Phase 1/2a challenge studies and up to Phase 2b studies in malaria endemic regions; some were efficacious but the efficacy was marginal and significant research and development (including almost 40 different clinical trials) was required before the final vaccine candidate was identified (Cohen et al., 2010). After 30 years of dedicated commitment from the pharmaceutical partner, GSK, phase 3 data are finally available which show that the vaccine is far less efficacious in the target population than anticipated and that the effect on clinical malaria is short-lived (Olotu et al., 2013). Consistent with this, a number of studies have established that although the CSP is the dominant sporozoite surface protein, it is not a major component of the protective immune responses induced by immunisation with radiation attenuated *Plasmodium* sporozoites (Doolan et al., 2003; Gruner et al., 2007; Kumar et al., 2006; Trieu et al., 2011).

In the case of erythrocytic stage vaccines, there has been a similar, if slightly less restrictive, focus on a handful of candidate antigen targets, particularly MSP1 (Fowkes et al., 2010) and AMA1 (Remarque et al., 2008), with a marked lack of success in the clinic (Ogutu et al., 2009; Sagara et al., 2009).

There is reason to believe that such a narrow focus is inadequate and misplaced. A possible explanation for the unsatisfactory efficacy of CSP-based vaccines, for example, comes from analysis of immune responses in animals and human volunteers protected by immunisation with irradiated sporozoites. While such volunteers

do generate antibody and T cell responses to CSP, they are often very weak responses, substantially weaker than those induced by immunisation with CSP itself (Doolan and Hoffman, 1997, 2000). This discrepancy suggests that immune responses to CSP are not a major component of the robust protection induced by immunisation with radiation-attenuated sporozoites, and is similar to the low-level responses elicited in the volunteers to other candidate pre-erythrocytic stage vaccine antigens, including SSP2/TRAP and LSA1 (Doolan et al., 1997; Doolan and Hoffman, 2000). These findings suggest two possibilities: either the protective immunity induced by radiation attenuated sporozoites depends on a strong response to a limited number of not-yet-identified antigens, or it depends on the summation of a large number of weak responses to many antigens. In either case, a focus on the large number of other potential pre-erythrocytic stage vaccine candidates present in the genome is warranted.

To date, few proteins in the *Plasmodium* spp. proteome have been eliminated from consideration as potential vaccine candidates, and many are being advanced as vaccine components in clinical trials without rigorous evaluation of efficacy. Moreover, subunit vaccines based on a single or few antigens may elicit a narrow breadth of response, providing neither optimal protection nor protection on genetically diverse backgrounds. This protein-by-protein approach has resulted in a long and largely unsuccessful history of malaria vaccine development, with many candidate vaccine immunogens being in the development pipeline for decades and many failures (World Health Organization, 2007; Schwartz et al., 2012). It is likely that failure to develop a malaria vaccine despite decades of effort can be attributed to this limited and arbitrary list of antigens, and that only by overcoming this restriction will it be possible to identify vaccine targets suitable for inducing protective humoral and cellular immune responses protecting against chronic infections such as malaria. The requirement is both to select optimally protective antigens and to ascertain whether a few or many such antigens underlie the protective immunity induced by immunisation with irradiated sporozoites (pre-erythrocytic stage immunity) or by repeated exposure to infection (blood-stage immunity). In our opinion, genome-based rational vaccine design is the most logical, and only, way forward.

### 5.3. Whole parasite vaccine trend

The long and challenging history of malaria vaccine development has caused a resurgence of interest in the whole-organism approach established by Jenner in the 18th Century. This interest builds on experimental evidence that whole parasite approaches utilising either radiation-attenuated sporozoites or parasitised red blood cells can protect in animal models of malaria, reviewed extensively elsewhere (Butler et al., 2012; Epstein and Richie, 2013; Good, 2013; Good et al., 2013; Loukas and Good, 2013; Vaughan and Kappe, 2013).

Briefly, the core approaches are: (i) metabolically active radiation attenuated sporozoites being championed by Sanaria (Hoffman et al., 2010; Seder et al., 2013); (ii) immunisation with infectious *P. falciparum* sporozoites under chloroquine chemoprophylaxis (CPS) (Roestenberg et al., 2009, 2011; Bijker et al., 2013); (iii) genetically attenuated liver stage and/or blood stage parasites (Mueller et al., 2005; Spring et al., 2013; Mikolajczak et al., 2014); and (iv) chemically attenuated blood stage parasites (Good et al., 2013).

Although some promising results have been obtained in pre-clinical models, it remains to be seen whether the many technical, logistical and regulatory hurdles can be overcome (Menard, 2005; Ballou and Cahill, 2007; Anders, 2011). In our opinion, the difficulties associated with progressing whole-organism malaria vaccines to the clinic are likely to be insurmountable and compel us towards



a modern genome-based rational vaccine design, rather than a return to the approach established over 200 years ago.

#### 5.4. Genome-based rational vaccine design for malaria

Malaria represents an excellent model for genome-based vaccine design against complex pathogens because: (i) it is a significant public health problem in the developing world; (ii) there are two human models of protective immunity that establish the feasibility of developing a malaria vaccine (immunisation by radiation-attenuated *Plasmodium* sporozoites, relying on the generation of protective cellular responses against pre-erythrocytic stage antigens; and naturally acquired immunity, relying on the generation of protective antibody responses against blood stage antigens); (iii) candidate antigens can be assessed either in vitro (using an appropriate immune readout), in animal model challenge systems (using the corresponding *Plasmodium* spp.), or in humans by challenging with *P. falciparum* in a well-validated human challenge model; (iv) both cellular and humoral immune responses to several antigens simultaneously will be likely required, in accordance with the two models of protection in humans; (v) sub-optimal protection has already been achieved using current molecular vaccine technologies, suggesting their feasibility and potential for improvement; (vi) vaccines so generated can be tested for safety and protection against sporozoite challenge in the USA or Europe, and transitioned as appropriate to field testing; and (vii) malaria, similar to many other important human pathogens in the developing world, is a parasitic disease and thus has developed mechanisms to avoid or modulate the host immune system, meaning that the technological solutions developed for malaria vaccine development should be translatable to other chronic infectious agents.

##### 5.4.1. *Plasmodium* 'omics'

The complete genomic sequences of the human parasites *P. falciparum* (3D7 clone), *P. vivax* (Sal I strain), *Plasmodium yoelii* (17XNL strain) and *Plasmodium knowlesi* (H strain) have been elucidated (Gardner et al., 2002; Carlton et al., 2002; Pain et al., 2008). Genome sequence data from nine *Plasmodium* strains are in various stages of completion and are available to the research community at PlasmoDB (<http://www.plasmodb.org/>) (Carlton et al., 2005, 2008; Aurecochea et al., 2009) together with partial genome sequence for an additional 16 geographically diverse *Plasmodium* parasites (15 *P. falciparum*, one *Plasmodium reichenowi*). With the advent of next generation sequencing technology, the number of completed and ongoing sequencing projects is rapidly increasing. The sequencing of at least 105 geographically diverse *Plasmodium* strains/isolates including 75 *P. falciparum* isolates, nine *P. vivax* isolates, *Plasmodium malariae* and *Plasmodium ovale*, representative simian malaria parasites (*P. reichenowi*, *Plasmodium cynomolgi*, *Plasmodium inui*, *Plasmodium coatneyi*, *Plasmodium fragile*), rodent parasites (*Plasmodium chabaudi*, *P. yoelii*, and *Plasmodium berghei*), an avian parasite (*Plasmodium relictum*), and a reptile parasite (*Plasmodium mexicanum*) is in progress (Cai et al., 2012).

In addition to this genomic data, high throughput transcriptomics and proteomics technologies, including microarray DNA chip, mass-spectrometry, yeast two-hybrid (Y2H) screening and most recently RNA-seq, NSR-seq and CHIP-seq, have been used to characterise and profile the expression, regulation and interaction of *Plasmodium* genes at the level of the transcriptome, proteome, metabolome and interactome (Winzeler, 2006; Cai et al., 2012). Large-scale proteomic datasets are also available for several life cycle stages (*P. falciparum*, *P. berghei* and *P. yoelii*) (Florens et al., 2002, 2004; Hall et al., 2005; Khan et al., 2005; Tarun et al., 2008). Transcriptomic data are now available from multiple life-cycle stages or gene knock-out mutants of *P. falciparum* and *P. berghei*

(Hall et al., 2005) as well as multiple stages of *P. yoelii* (mosquito, erythrocytic and liver stages) (Tarun et al., 2008; Mikolajczak et al., 2008; Zhou et al., 2008; Lasonder et al., 2008), *P. vivax* isolates (Bozdech et al., 2008) and other *P. falciparum* strains (Rovira-Graells et al., 2012). Other datasets include expressed sequence tag (EST) data from over 130 libraries (*P. falciparum*, *P. vivax*, *P. berghei* and *P. yoelii*) (Watanabe et al., 2007; Tarun et al., 2008; Florent et al., 2009) and serial analysis of gene expression (SAGE) data (*P. falciparum* only) (Patankar et al., 2001; Gunasekera et al., 2003, 2004). Most recently, transcriptome analysis by next generation sequencing (RNA-seq) has provided information about the abundance of all transcripts in the transcriptome by considering the real number and the level of different transcript isoforms (Otto et al., 2010; Lopez-Berragan et al., 2011; Lindner et al., 2013). Moreover the combination of transcriptomics analysis with ChIP-seq data has provided new insights to unravel the regulation of gene expression (Lopez-Barragan et al., 2011; Hoeijmakers et al., 2012). Other studies have provided transcriptomics data on clinical isolates using NSR-seq (Vignali et al., 2011). Expression profiling data throughout the intraerythrocytic cycle (oligonucleotide-based microarrays in both glass slide and Affymetrix formats) as well as single nucleotide polymorphism (SNP) analysis for 20 *P. falciparum* strains and 100 *P. falciparum* isolates (Jeffares et al., 2007; Mu et al., 2007; Volkman et al., 2007) are also available. Additional functional data sets include evidence of protein–protein interactions (Y2H and predicted interactome) (LaCount et al., 2005; Date and Stoeckert, 2006); Genome Ontology (GO) (Ashburner et al., 2000) and InterPro domain (Mulder et al., 2005) annotations for *P. falciparum*, *P. vivax*, *P. berghei*, *P. yoelii*, *P. knowlesi* and *P. chabaudi*; Enzyme Commission (EC) number (Ashburner et al., 2000) annotations for *P. falciparum*, *P. yoelii* and *P. knowlesi* (Ginsburg, 2006); and metabolic pathway assignments for *P. falciparum* (Ginsburg, 2006). Predictions of protein subcellular localisation (Bendtsen et al., 2004) and transmembrane domains (Krogh et al., 2001) for *P. falciparum*, *P. vivax*, *P. berghei*, *P. yoelii*, *P. knowlesi* and *P. chabaudi* are available, as well as parasite-specific predictions (*P. falciparum* only) for apicoplast localisation (Foth et al., 2003) and for export to the host cell (Hiller et al., 2004; Marti et al., 2004, 2005).

These analyses of genomics, proteomics, transcriptomics, interactomics and, most recently, next generation sequencing, allow for investigation at the molecular level of drug resistance, immune evasion and pathogenesis, and provide the foundation for antigen discovery and prioritisation on a genome-wide or proteome-wide scale. Web-based databases such as PlasmoDB (<http://plasmodb.org/>) and GeneDB (<http://www.genedb.org/>) providing public access to the data and advanced statistical and bioinformatics algorithms and analytical tools such as BioGRID (<http://thebiogrid.org/>), Pathway Commons (<http://www.pathwaycommons.org/>) and STRING (<http://string-db.org/>), allow the researcher to visualise, analyse, integrate and interrogate the data. Overall, these datasets provide a comprehensive foundation for genome-based vaccine design.

##### 5.4.2. Applied 'omics'

With this wealth of genomic, proteomic, transcriptomic and related data, the tools to exploit these data, and biological samples from human volunteers, we have an unprecedented opportunity to develop and implement a rational approach to target antigen selection and vaccine design. In our view, the "post-genomics" era of vaccine research should focus on leveraging these genome-scale datasets for objective evidence-based assessment of candidate targets for rational vaccine development. Rational and systematic genome-based strategies overcome the deficiencies of the current ad hoc approach to target selection. Importantly, they take advantage of the two human models of immunity which depend on



exposure to whole *Plasmodium* organisms expressing hundreds to thousands of potential immune targets, while aiming to “improve on nature”.

Already, exploitation of genomic data with systematic approaches to rational drug design and target selection (chemogenomics) has led to the identification of new classes of antimicrobial drugs (Bhattacharjee et al., 2013). Progress towards genome-based vaccine design is well under way and showing great potential. In our laboratory, we are using the power of immunomics to address two situations: one (predominantly pre-erythrocytic, based on the irradiated sporozoite model) in which T cell targets are selected, and one (predominantly erythrocytic, based on naturally acquired immunity) in which antibody targets are selected. We are using biological specimens (T cells and plasma/sera) from individuals immunised with irradiated sporozoites or naturally exposed to malaria, or immunoglobulin from clinically immune individuals, for in vitro immune screening to assess the capacity of the antigen to be recognised by recall of *Plasmodium*-specific immune responses in protective human models using clinically relevant selection criteria. We have generated unique ‘omic’-scale datasets of proteome-wide T cell responses and antibody responses to *Plasmodium* by systematically screening the *Plasmodium* proteome using protein microarrays and epitope prediction algorithms, with specimens from humans, non-human primates or rodents naturally or experimentally exposed to malaria. (Doolan et al., 2008; Crompton et al., 2010; Barry et al., 2011). Analysis of these comprehensive datasets has revealed that the target antigens of antibody or T cell responses are not randomly distributed throughout the proteome, and that only approximately 30% of the proteome is recognised. We have also established that a large number of newly identified antigens are more immunogenic than antigens identified by traditional methods. Furthermore, antigens that are highly reactive for T cells are not serodominant for antibody responses (Doolan, D.L., unpublished data), suggesting that different approaches are required to identify the most effective targets of T cell and antibody responses. These data establish proof-of-concept for both T cell- and antibody-based approaches to identify antigens and epitopes which represent promising candidates for next generation malaria vaccine development. They also support the premise that omics-scale approaches offer promising solutions to the challenge of vaccine development for complex pathogens.

## 6. Conclusion

There is currently no accepted rational approach to vaccine design and target selection for malaria or any other complex pathogen. We advocate a rational approach to malaria vaccine design capable of exploiting the wealth of genomic, proteomic and bioinformatic information that has been developed in the genomic and post-genomic eras. We believe that rational genome-based vaccine design, enabling the selection of the best possible targets by prioritising antigens according to clinically relevant criteria (frequency and magnitude of clinically relevant immune response and/or biological function), will overcome the problem of poorly immunogenic, poorly protective vaccines that has plagued malaria vaccine developers for the past 25 years. Clearly, additional work will be required to bring rationally-designed vaccines to the clinic, including pre-clinical safety testing, optimisation of immunogenicity, phased human clinical trials, and large scale production and distribution. However, appropriate approaches and standard protocols for these steps are well-established and these remaining steps are not part of the critical roadblock to malaria vaccine development. There remain economic roadblocks to production and licensing of an effective vaccine even after efficacy and safety are demonstrated. We believe, however, that once an effective malaria

vaccine has been developed, private and governmental sources will be persuaded to provide adequate resources to support production and distribution and, once distributed, the vaccine is likely to meet with wide acceptance in Africa. Extension of this approach to other pathogens will require a substantial research investment, however such effort will undoubtedly be worthwhile.

## Acknowledgements

D.L.D. is supported by a National Health and Medical Research Council (NHMRC, Australia) Principal Research Fellowship. Support from a Pfizer Australia Senior Research Fellowship is also gratefully acknowledged.

## References

- Allen, T.M., Mothe, B.R., Sidney, J., Jing, P., Dzuris, J.L., Liebl, M.E., Vogel, T.U., O'Connor, D.H., Wang, X., Wussow, M.C., Thomson, J.A., Altman, J.D., Watkins, D.I., Sette, A., 2001. CD8(+) lymphocytes from simian immunodeficiency virus-infected rhesus macaques recognize 14 different epitopes bound by the major histocompatibility complex class I molecule mamu-A\*01: implications for vaccine design and testing. *J. Virol.* 75, 738–749.
- Anders, R.F., 2011. The case for a subunit vaccine against malaria. *Trends Parasitol.* 27, 330–334.
- Andre, F.E., 2003. Vaccinology: past achievements, present roadblocks and future promises. *Vaccine* 21, 593–595.
- Ashburner, M., Ball, C.A., Blake, J.A., Botstein, D., Butler, H., Cherry, J.M., Davis, A.P., Dolinski, K., Dwight, S.S., Eppig, J.T., Harris, M.A., Hill, D.P., Issel-Tarver, L., Kasarskis, A., Lewis, S., Matese, J.C., Richardson, J.E., Ringwald, M., Rubin, G.M., Sherlock, G., 2000. Gene ontology: tool for the unification of biology. *Nat. Genet.* 25, 25–29.
- Assarsson, E., Sidney, J., Oseroff, C., Pasquetto, V., Bui, H.H., Frahm, N., Brander, C., Peters, B., Grey, H., Sette, A., 2007. A quantitative analysis of the variables affecting the repertoire of T cell specificities recognized after vaccinia virus infection. *J. Immunol.* 178, 7890–7901.
- Aurrecochea, C., Brestelli, J., Brunk, B.P., Dommer, J., Fischer, S., Gajria, B., Gao, X., Gingle, A., Grant, G., Harb, O.S., Heiges, M., Innamorato, F., Iodice, J., Kissinger, J.C., Kraemer, E., Li, W., Miller, J.A., Nayak, V., Pennington, C., Pinney, D.F., Roos, D.S., Ross, C., Stoeckert Jr., C.J., Treatman, C., Wang, H., 2009. PlasmoDB: a functional genomic database for malaria parasites. *Nucleic Acids Res.* 37, D539–D543.
- Baird, J.K., 1998. Age-dependent characteristics of protection v. susceptibility to *Plasmodium falciparum*. *Ann. Trop. Med. Parasitol.* 92, 367–390.
- Baker, M., 2013. Big biology: The ‘omes puzzle’. *Nature* 494, 416–419.
- Ballou, W.R., Cahill, C.P., 2007. Two decades of commitment to malaria vaccine development: GlaxoSmithKline Biologicals. *Am. J. Trop. Med. Hyg.* 77, 289–295.
- Barry, A.E., Schultz, L., Buckee, C.O., Reeder, J.C., 2009. Contrasting population structures of the genes encoding ten leading vaccine-candidate antigens of the human malaria parasite, *Plasmodium falciparum*. *PLoS One* 4, e8497.
- Barry, A.E., Trieu, A., Fowkes, F.J., Pablo, J., Kalantari-Dehaghi, M., Jasinskas, A., Tan, X., Kayala, M.A., Tavul, L., Siba, P.M., Day, K.P., Baldi, P., Felgner, P.L., Doolan, D.L., 2011. The stability and complexity of antibody responses to the major surface antigen of *Plasmodium falciparum* are associated with age in a malaria endemic area. *Mol. Cell. Proteomics* 10 (M111), 008326.
- Baxby, D., 1999. Edward Jenner's Inquiry; a bicentenary analysis. *Vaccine* 17, 301–307.
- Bendtsen, J.D., Nielsen, H., von Heijne, G., Brunak, S., 2004. Improved prediction of signal peptides: SignalP 3.0. *J. Mol. Biol.* 340, 783–795.
- Bhattacharjee, B., Simon, R.M., Gangadharaiah, C., Karunakar, P., 2013. Chemogenomics profiling of drug targets of peptidoglycan biosynthesis pathway in *Leptospira interrogans* by virtual screening approaches. *J. Microbiol. Biotechnol.* 23, 779–784.
- Bijker, E.M., Bastiaens, G.J., Teirlinck, A.C., van Gemert, G.J., Graumans, W., van de Vegte-Bolmer, M., Siebelink-Stoter, R., Arens, T., Teelen, K., Nahrendorf, W., Remarque, E.J., Roeffen, W., Jansens, A., Zimmerman, D., Vos, M., van Schaijk, B.C., Wiersma, J., van der Ven, A.J., de Mast, Q., van Lieshout, L., Verweij, J.J., Hermsen, C.C., Scholzen, A., Sauerwein, R.W., 2013. Protection against malaria after immunization by chloroquine prophylaxis and sporozoites is mediated by preerythrocytic immunity. *Proc. Natl. Acad. Sci. U.S.A.* 110, 7862–7867.
- Black, R.E., Cousens, S., Johnson, H.L., Lawn, J.E., Rudan, I., Bassani, D.G., Jha, P., Campbell, H., Walker, C.F., Cibulskis, R., Eisele, T., Liu, L., Mathers, C., 2010. Global, regional, and national causes of child mortality in 2008: a systematic analysis. *Lancet* 375, 1969–1987.
- Bouchie, A., 2013. GSK plows ahead with EMA malaria vaccine submission. *Nat. Biotechnol.* 31, 1066.
- Boutwell, C.L., Rolland, M.M., Herbeck, J.T., Mullins, J.I., Allen, T.M., 2010. Viral evolution and escape during acute HIV-1 infection. *J. Infect. Dis.* 202 (Suppl. 2), S309–S314.
- Bozdech, Z., Mok, S., Hu, G., Imwong, M., Jaidee, A., Russell, B., Ginsburg, H., Nosten, F., Day, N.P., White, N.J., Carlton, J.M., Preiser, P.R., 2008. The transcriptome of

- Plasmodium vivax* reveals divergence and diversity of transcriptional regulation in malaria parasites. *Proc. Natl. Acad. Sci. U.S.A.* 105, 16290–16295.
- Brown, F., 1993. Review of accidents caused by incomplete inactivation of viruses. *Dev. Biol. Stand.* 81, 103–107.
- Brusic, V., Gottardo, R., Kleinstein, S.H., Davis, M.M. HIPC Steering Committee, 2014. Computational resources for high-dimensional immune analysis from the Human Immunology Project Consortium. *Nat. Biotechnol.* 32, 146–148.
- Butler, N.S., Vaughan, A.M., Hartly, J.T., Kappe, S.H., 2012. Whole parasite vaccination approaches for prevention of malaria infection. *Trends Immunol.* 33, 247–254.
- Cai, H., Zhou, Z., Gu, J., Wang, Y., 2012. Comparative genomics and systems biology of malaria parasites *Plasmodium*. *Curr. Bioinform.* 7 (4).
- Carlton, J., Silva, J., Hall, N., 2005. The genome of model malaria parasites, and comparative genomics. *Curr. Issues Mol. Biol.* 7, 23–37.
- Carlton, J.M., Adams, J.H., Silva, J.C., Bidwell, S.L., Lorenzi, H., Caler, E., Crabtree, J., Angiuoli, S.V., Merino, E.F., Amedeo, P., Cheng, Q., Coulson, R.M., Crabb, B.S., Del Portillo, H.A., Essien, K., Feldblyum, T.V., Fernandez-Becerra, C., Gilson, P.R., Gueye, A.H., Guo, X., Kang'a, S., Kooij, T.W., Korsinczky, M., Meyer, E.V., Nene, V., Paulsen, I., White, O., Ralph, S.A., Ren, Q., Sargeant, T.J., Salzberg, S.L., Stoeckert, C.J., Sullivan, S.A., Yamamoto, M.M., Hoffman, S.L., Wortman, J.R., Gardner, M.J., Galinski, M.R., Barnwell, J.W., Fraser-Liggett, C.M., 2008. Comparative genomics of the neglected human malaria parasite *Plasmodium vivax*. *Nature* 455, 757–763.
- Carlton, J.M., Angiuoli, S.V., Suh, B.B., Kooij, T.W., Perlea, M., Silva, J.C., Ermolaeva, M.D., Allen, J.E., Selengut, J.D., Koo, H.L., Peterson, J.D., Pop, M., Kosack, D.S., Shumway, M.F., Bidwell, S.L., Shallom, S.J., van Aken, S.E., Riedmuller, S.B., Feldblyum, T.V., Cho, J.K., Quackenbush, J., Sedegah, M., Shoaibi, A., Cummings, L.M., Florens, L., Yates, J.R., Raine, J.D., Sinden, R.E., Harris, M.A., Cunningham, D.A., Preiser, P.R., Bergman, L.W., Vaidya, A.B., van Lin, L.H., Janse, C.J., Waters, A.P., Smith, H.O., White, O.R., Salzberg, S.L., Venter, J.C., Fraser, C.M., Hoffman, S.L., Gardner, M.J., Carucci, D.J., 2002. Genome sequence and comparative analysis of the model rodent malaria parasite *Plasmodium yoelii yoelii*. *Nature* 419, 512–519.
- Cohen, J., Nussenzweig, V., Nussenzweig, R., Vekemans, J., Leach, A., 2010. From the circumsporozoite protein to the RTS, S/AS candidate vaccine. *Hum. Vaccin.* 6, 90–96.
- Cohen, S., Mc, G.I., Carrington, S., 1961. Gamma-globulin and acquired immunity to human malaria. *Nature* 192, 733–737.
- Corey, L., McElrath, M.J., 2010. HIV vaccines: mosaic approach to virus diversity. *Nat. Med.* 16, 268–270.
- Courtin, D., Oesterholt, M., Huismans, H., Kusi, K., Milet, J., Badaut, C., Gaye, O., Roeffen, W., Remarque, E.J., Sauerwein, R., Garcia, A., Luty, A.J., 2009. The quantity and quality of African children's IgG responses to merozoite surface antigens reflect protection against *Plasmodium falciparum* malaria. *PLoS ONE* 4, e7590.
- Crompton, P.D., Kayala, M.A., Traore, B., Kayentao, K., Ongoiba, A., Weiss, G.E., Molina, D.M., Burk, C.R., Waisberg, M., Jasinskas, A., Tan, X., Doumbo, S., Doumtabe, D., Kone, Y., Narum, D.L., Liang, X., Doumbo, O.K., Miller, L.H., Doolan, D.L., Baldi, P., Felgner, P.L., Pierce, S.K., 2010. A prospective analysis of the Ab response to *Plasmodium falciparum* before and after a malaria season by protein microarray. *Proc. Natl. Acad. Sci. U.S.A.* 107, 6958–6963.
- D'Argenio, D.A., Wilson, C.B., 2010. A decade of vaccines: integrating immunology and vaccinology for rational vaccine design. *Immunity* 33, 437–440.
- Date, S.V., Stoeckert Jr., C.J., 2006. Computational modeling of the *Plasmodium falciparum* interactome reveals protein function on a genome-wide scale. *Genome Res.* 16, 542–549.
- De Gregorio, E., Rappuoli, R., 2012. Vaccines for the future: learning from human immunology. *Microb. Biotechnol.* 5, 149–155.
- Delany, I., Rappuoli, R., De Gregorio, E., 2014. Vaccines for the 21st century. *EMBO Mol. Med.* 6, 708–720.
- Doolan, D.L., 2011. *Plasmodium* immunomics. *Int. J. Parasitol.* 41, 3–20.
- Doolan, D.L., Hoffman, S.L., 1997. Pre-erythrocytic-stage immune effector mechanisms in *Plasmodium* spp. infections. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 352, 1361–1367.
- Doolan, D.L., Hoffman, S.L., 2000. The complexity of protective immunity against liver-stage malaria. *J. Immunol.* 165, 1453–1462.
- Doolan, D.L., Hoffman, S.L., Southwood, S., Wentworth, P.A., Sidney, J., Chesnut, R.W., Keogh, E., Appella, E., Nutman, T.B., Lal, A.A., Gordon, D.M., Oloo, A., Sette, A., 1997. Degenerate cytotoxic T cell epitopes from *P. falciparum* restricted by multiple HLA-A and HLA-B supertype alleles. *Immunity* 7, 97–112.
- Doolan, D.L., Houghten, R.A., Good, M.F., 1991. Location of human cytotoxic T cell epitopes within a polymorphic domain of the *Plasmodium falciparum* circumsporozoite protein. *Int. Immunol.* 3, 511–516.
- Doolan, D.L., Martinez-Alier, N., 2006. Immune response to pre-erythrocytic stages of malaria parasites. *Curr. Mol. Med.* 6, 169–185.
- Doolan, D.L., Mu, Y., Unal, B., Sundaresh, S., Hirst, S., Valdez, C., Randall, A., Molina, D., Liang, X., Freilich, D.A., Oloo, J.A., Blair, P.L., Aguiar, J.C., Baldi, P., Davies, D.H., Felgner, P.L., 2008. Profiling humoral immune responses to *P. falciparum* infection with protein microarrays. *Proteomics* 8, 4680–4694.
- Doolan, D.L., Southwood, S., Freilich, D.A., Sidney, J., Graber, N.L., Shatney, L., Bebris, L., Florens, L., Dobano, C., Whitney, A.A., Appella, E., Hoffman, S.L., Yates 3rd, J.R., Carucci, D.J., Sette, A., 2003. Identification of *Plasmodium falciparum* antigens by antigenic analysis of genomic and proteomic data. *Proc. Natl. Acad. Sci. U.S.A.* 100, 9952–9957.
- Dormitzer, P.R., Grandi, G., Rappuoli, R., 2012. Structural vaccinology starts to deliver. *Nat. Rev. Microbiol.* 10, 807–813.
- Dormitzer, P.R., Ulmer, J.B., Rappuoli, R., 2008. Structure-based antigen design: a strategy for next generation vaccines. *Trends Biotechnol.* 26, 659–667.
- Duncan, C.J., Hill, A.V., 2011. What is the efficacy of the RTS, S malaria vaccine? *BMJ* 343, d7728.
- Dutta, S., Lee, S.Y., Batchelor, A.H., Lanar, D.E., 2007. Structural basis of antigenic escape of a malaria vaccine candidate. *Proc. Natl. Acad. Sci. U.S.A.* 104, 12488–12493.
- Eisenbarth, S.C., Colegio, O.R., O'Connor, W., Sutterwala, F.S., Flavell, R.A., 2008. Crucial role for the Nalp3 inflammasome in the immunostimulatory properties of aluminium adjuvants. *Nature* 453, 1122–1126.
- Epstein, J.E., Richie, T.L., 2013. The whole parasite, pre-erythrocytic stage approach to malaria vaccine development: a review. *Curr. Opin. Infect. Dis.* 26, 420–428.
- Faber, B.W., Younis, S., Remarque, E.J., Rodriguez Garcia, R., Riasat, V., Walraven, V., van der Werff, N., van der Eijk, M., Cavanagh, D.R., Holder, A.A., Thomas, A.W., Kocken, C.H., 2013. Diversity covering AMA1-MSP119 fusion proteins as malaria vaccines. *Infect. Immun.* 81, 1479–1490.
- Feng, J., Gulati, U., Zhang, X., Keitel, W.A., Thompson, D.M., James, J.A., Thompson, L.F., Air, G.M., 2009. Antibody quantity versus quality after influenza vaccination. *Vaccine* 27, 6358–6362.
- Fenner, F., 1982. A successful eradication campaign. *Global eradication of smallpox. Rev. Infect. Dis.* 4, 916–930.
- Fenner, F., Henderson, D.A., Arita, I., Jezek, Z., Ladnyi, I.D., 1988. *Smallpox and Its Eradication*. World Health Organisation, Geneva, <<http://whqlibdoc.who.int/smallpox/9241561106.pdf>>.
- Finco, O., Rappuoli, R., 2014. Designing vaccines for the twenty-first century society. *Front. Immunol.* 5, 12.
- Fleischmann, R.D., Adams, M.D., White, O., Clayton, R.A., Kirkness, E.F., Kerlavage, A.R., Bult, C.J., Tomb, J.F., Dougherty, B.A., Merrick, J.M., et al., 1995. Whole-genome random sequencing and assembly of *Haemophilus influenzae* Rd. *Science* 269, 496–512.
- Florens, L., Liu, X., Wang, Y., Yang, S., Schwartz, O., Peglar, M., Carucci, D.J., Yates 3rd, J.R., Wu, Y., 2004. Proteomics approach reveals novel proteins on the surface of malaria-infected erythrocytes. *Mol. Biochem. Parasitol.* 135, 1–11.
- Florent, I., Porcel, B.M., Guillaume, E., Da Silva, C., Artiguenave, F., Marechal, E., Brehelin, L., Gascuel, O., Charneau, S., Wincker, P., Grellier, P., 2009. A *Plasmodium falciparum* FcB1-schizont-EST collection providing clues to schizont specific gene structure and polymorphism. *BMC genomics* 10, 235.
- Florens, L., Washburn, M.P., Raine, J.D., Anthony, R.M., Grainger, M., Haynes, J.D., Moch, J.K., Muster, N., Sacchi, J.B., Tabb, D.L., Whitney, A.A., Wolters, D., Wu, Y., Gardner, M.J., Holder, A.A., Sinden, R.E., Yates, J.R., Carucci, D.J., 2002. A proteomic view of the *Plasmodium falciparum* life cycle. *Nature* 419, 520–526.
- Foth, B.J., Ralph, S.A., Tonkin, C.J., Struck, N.S., Fraunholz, M., Roos, D.S., Cowman, A.F., McFadden, G.L., 2003. Dissecting apicoplast targeting in the malaria parasite *Plasmodium falciparum*. *Science* 299, 705–708.
- Fowkes, F.J., Richards, J.S., Simpson, J.A., Beeson, J.G., 2010. The relationship between anti-merozoite antibodies and incidence of *Plasmodium falciparum* malaria: a systematic review and meta-analysis. *PLoS Med.* 7, e1000218.
- Gardner, M.J., Hall, N., Fung, E., White, O., Berriman, M., Hyman, R.W., Carlton, J.M., Pain, A., Nelson, K.E., Bowman, S., Paulsen, I.T., James, K., Eisen, J.A., Rutherford, K., Salzberg, S.L., Craig, A., Kyes, S., Chan, M.S., Nene, V., Shallom, S.J., Suh, B., Peterson, J., Angiuoli, S., Perlea, M., Allen, J., Selengut, J., Haft, D., Mather, M.W., Vaidya, A.B., Martin, D.M., Fairlamb, A.H., Fraunholz, M.J., Roos, D.S., Ralph, S.A., McFadden, G.L., Cummings, L.M., Subramanian, G.M., Mungall, C., Venter, J.C., Carucci, D.J., Hoffman, S.L., Newbold, C., Davis, R.W., Fraser, C.M., Barrell, B., 2002. Genome sequence of the human malaria parasite *Plasmodium falciparum*. *Nature* 419, 498–511.
- Genton, B., Betuela, I., Felger, I., Al-Yaman, F., Anders, R.F., Saul, A., Rare, L., Baisor, M., Lorry, K., Brown, G.V., Pye, D., Irving, D.O., Smith, T.A., Beck, H.P., Alpers, M.P., 2002. A recombinant blood-stage malaria vaccine reduces *Plasmodium falciparum* density and exerts selective pressure on parasite populations in a phase 1–2b trial in Papua New Guinea. *J. Infect. Dis.* 185, 820–827.
- Germain, R.N., 2010. Vaccines and the future of human immunology. *Immunity* 33, 441–450.
- Ginsburg, H., 2006. Progress in in silico functional genomics: the malaria Metabolic Pathways database. *Trends Parasitol.* 22, 238–240.
- Good, M.F., 2013. Immunology. Pasteur approach to a malaria vaccine may take the lead. *Science* 341, 1352–1353.
- Good, M.F., Doolan, D.L., 2010. Malaria vaccine design: immunological considerations. *Immunity* 33, 555–566.
- Good, M.F., Reiman, J.M., Rodriguez, I.B., Ito, K., Yanow, S.K., El-Deeb, I.M., Batzloff, M.R., Stanisic, D.L., Engwerda, C., Spithill, T., Hoffman, S.L., Lee, M., McPhun, V., 2013. Cross-species malaria immunity induced by chemically attenuated parasites. *J. Clin. Invest.* 123, 3353–3362.
- Gorringe, A.R., Pajon, R., 2012. Bexsero: a multicomponent vaccine for prevention of meningococcal disease. *Hum. Vaccin. Immunother.* 8, 174–183.
- Grimm, S.K., Ackerman, M.E., 2013. Vaccine design: emerging concepts and renewed optimism. *Curr. Opin. Biotechnol.* 24, 1078–1088.
- Gunasekera, A.M., Patankar, S., Schug, J., Eisen, G., Kissinger, J., Roos, D., Wirth, D.F., 2004. Widespread distribution of antisense transcripts in the *Plasmodium falciparum* genome. *Mol. Biochem. Parasitol.* 136, 35–42.
- Gunasekera, A.M., Patankar, S., Schug, J., Eisen, G., Wirth, D.F., 2003. Drug-induced alterations in gene expression of the asexual blood forms of *Plasmodium falciparum*. *Mol. Microbiol.* 50, 1229–1239.
- Gruner, A.C., Mauduit, M., Tewari, R., Romero, J.F., Depinay, N., Kayibanda, M., Lallemand, E., Chavatte, J.M., Crisanti, A., Sinnis, P., Mazier, D., Corradin, G.,

- Snounou, Renia, L., . Sterile protection against malaria is independent of immune responses to the circumsporozoite protein. *PLoS ONE* 2, e1371.
- Hall, N., Karras, M., Raine, J.D., Carlton, J.M., Kooij, T.W., Berriman, M., Florens, L., Janssen, C.S., Pain, A., Christophides, G.K., James, K., Rutherford, K., Harris, B., Harris, D., Churcher, C., Quail, M.A., Ormond, D., Doggett, J., Trueman, H.E., Mendoza, J., Bidwell, S.L., Rajandream, M.A., Carucci, D.J., Yates 3rd, J.R., Kafatos, F.C., Janse, C.J., Barrell, B., Turner, C.M., Waters, A.P., Sinden, R.E., 2005. A comprehensive survey of the *Plasmodium* life cycle by genomic, transcriptomic, and proteomic analyses. *Science* 307, 82–86.
- Hilleman, M.R., 1999. Personal historical chronicle of six decades of basic and applied research in virology, immunology, and vaccinology. *Immunol. Rev.* 170, 7–27.
- Hiller, N.L., Bhattacharjee, S., van Ooij, C., Liolios, K., Harrison, T., Lopez-Estrano, C., Haldar, K., 2004. A host-targeting signal in virulence proteins reveals a secretome in malarial infection. *Science* 306, 1934–1937.
- Hoeymakers, W.A., Flueck, C., Francoijs, K.J., Smits, A.H., Wetzel, J., Volz, J.C., Cowman, A.F., Voss, T., Stunnenberg, H.G., Bartfai, R., 2012. *Plasmodium falciparum* centromeres display a unique epigenetic makeup and cluster prior to and during schizogony. *Cell. Microbiol.* 14, 1391–1401.
- Hoffman, S.L., Billingsley, P.F., James, E., Richman, A., Loyevsky, M., Li, T., Chakravarty, S., Gunasekera, A., Chattopadhyay, R., Li, M., Stafford, R., Ahumada, A., Epstein, J.E., Sedegah, M., Reyes, S., Richie, T.L., Lyke, K.E., Edelman, R., Laurens, M.B., Plowe, C.V., Sim, B.K., 2010. Development of a metabolically active, non-replicating sporozoite vaccine to prevent *Plasmodium falciparum* malaria. *Hum. Vaccin.* 6, 97–106.
- Hoffman, S.L., Goh, L.M., Luke, T.C., Schneider, I., Le, T.P., Doolan, D.L., Sacci, J., de la Vega, P., Dowler, M., Paul, C., Gordon, D.M., Stoute, J.A., Church, L.W., Sedegah, M., Heppner, D.G., Ballou, W.R., Richie, T.L., 2002. Protection of humans against malaria by immunization with radiation-attenuated *Plasmodium falciparum* sporozoites. *J. Infect. Dis.* 185, 1155–1164.
- Jeffares, D.C., Pain, A., Berry, A., Cox, A.V., Stalker, J., Ingle, C.E., Thomas, A., Quail, M.A., Siebenthall, K., Uhlemann, A.C., Kyes, S., Krishna, S., Newbold, C., Dermitzakis, E.T., Berriman, M., 2007. Genome variation and evolution of the malaria parasite *Plasmodium falciparum*. *Nat. Genet.* 39, 120–125.
- Jenner, E., 1798. An inquiry into causes and effects of variolae vaccinae, a disease, discovered in some of the western countries of England, particularly Gloucestershire, and know by the name of cow pox. Reprinted by Milan: R Lier & Co, 1923, 84.
- Julien, J.P., Lee, P.S., Wilson, I.A., 2012. Structural insights into key sites of vulnerability on HIV-1 Env and influenza HA. *Immunol. Rev.* 250, 180–198.
- Kaufmann, S.H., Juliana McElrath, M., Lewis, D.J., Del Giudice, G., 2014. Challenges and responses in human vaccine development. *Curr. Opin. Immunol.* 28C, 18–26.
- Keith, J.A., Agostini Bigger, L., Arthur, P.A., Maes, E., Daems, R., 2013. Delivering the promise of the Decade of Vaccines: opportunities and challenges in the development of high quality new vaccines. *Vaccine* 31 (Suppl. 2), B184–B193.
- Kennedy, R.B., Poland, G.A., 2011. The top five “game changers” in vaccinology: toward rational and directed vaccine development. *OMICS* 15, 533–537.
- Kemp, D.J., Coppel, R.L., Cowman, A.F., Saint, R.B., Brown, G.V., Anders, R.F., 1983. Expression of *Plasmodium falciparum* blood-stage antigens in *Escherichia coli*: detection with antibodies from immune humans. *Proc. Natl. Acad. Sci. U.S.A.* 80, 3787–3791.
- Khan, S.M., Franke-Fayard, B., Mair, G.R., Lasonder, E., Janse, C.J., Mann, M., Waters, A.P., 2005. Proteome analysis of separated male and female gametocytes reveals novel sex-specific *Plasmodium* biology. *Cell* 121, 675–687.
- Kirkman, L.A., Deitsch, K.W., 2012. Antigenic variation and the generation of diversity in malaria parasites. *Curr. Opin. Microbiol.* 15, 456–462.
- Klysik, J., 2001. Concept of immunomics: a new frontier in the battle for gene function? *Acta. Biotheor.* 49, 191–202.
- Kotturi, M.F., Scott, I., Wolfe, T., Peters, B., Sidney, J., Cheroutre, H., von Herrath, M.G., Buchmeier, M.J., Grey, H., Sette, A., 2008. Naive precursor frequencies and MHC binding rather than the degree of epitope diversity shape CD8+ T cell immunodominance. *J. Immunol.* 181, 2124–2133.
- Krogh, A., Larsson, B., von Heijne, G., Sonnhammer, E.L., 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J. Mol. Biol.* 305, 567–580.
- Kumar, K.A., Sano, G., Boscardin, S., Nussenzweig, R.S., Nussenzweig, M.C., Zavala, F., Nussenzweig, V., 2006. The circumsporozoite protein is an immunodominant protective antigen in irradiated sporozoites. *Nature* 444, 937–940.
- LaCount, D.J., Vignali, M., Chettier, R., Phansalkar, A., Bell, R., Hesselberth, J.R., Schoenfeld, L.W., Ota, I., Sahasrabudhe, S., Kurschner, C., Fields, S., Hughes, R.E., 2005. A protein interaction network of the malaria parasite *Plasmodium falciparum*. *Nature* 438, 103–107.
- Langhorne, J., Ndungu, F.M., Sponaas, A.M., Marsh, K., 2008. Immunity to malaria: more questions than answers. *Nat. Immunol.* 9, 725–732.
- Lasonder, E., Janse, C.J., van Gemert, G.J., Mair, G.R., Vermunt, A.M., Douradinha, B.G., van Noort, V., Huynen, M.A., Luty, A.J., Kroeze, H., Khan, S.M., Sauerwein, R.W., Waters, A.P., Mann, M., Stunnenberg, H.G., 2008. Proteomic profiling of *Plasmodium* sporozoite maturation identifies new proteins essential for parasite development and infectivity. *PLoS Pathog.* 4, e1000195.
- Levine, M.M.L., Esparza, R.J., 2009. Vaccines and vaccination in historical perspective. In: Levine, M.M., Dougan, G., Good, M.F., Liu, M.A., Nabel, G.J., Nataro, J.P., Rappuoli, R. (Eds.), *New Generation Vaccines*, Fourth ed. CRC Press, Boca Raton, FL, USA, pp. 1–11.
- Lindner, S.E., Mikolajczak, S.A., Vaughan, A.M., Moon, W., Joyce, B.R., Sullivan Jr, W.J., Kappe, S.H., 2013. Perturbations of *Plasmodium* Puf2 expression and RNA-seq of Puf2-deficient sporozoites reveal a critical role in maintaining RNA homeostasis and parasite transmissibility. *Cell. Microbiol.* 15, 1266–1283.
- Liu, L., Johnson, H.L., Cousens, S., Perin, J., Scott, S., Lawn, J.E., Rudan, I., Campbell, H., Cibulskis, R., Li, M., Mathers, C., Black, R.E., 2012. Global, regional, and national causes of child mortality: an updated systematic analysis for 2010 with time trends since 2000. *Lancet* 379, 2151–2161.
- Lopez-Barragan, M.J., Lemieux, J., Quinones, M., Williamson, K.C., Molina-Cruz, A., Cui, K., Barillas-Mury, C., Zhao, K., Su, X.Z., 2011. Directional gene expression and antisense transcripts in sexual and asexual stages of *Plasmodium falciparum*. *BMC Genomics* 12, 587.
- Loukas, A., Good, M.F., 2013. Back to the future for antiparasite vaccines? *Expert Rev Vaccines* 12, 1–4.
- Mackinnon, M.J., Marsh, K., 2010. The selection landscape of malaria parasites. *Science* 328, 866–871.
- Maraskovsky, E., Rockman, S., Dyson, A., Koernig, S., Becher, D., Morelli, A.B., Barnden, M., Camuglia, S., Bodle, J., Vandenberg, K., Wang, I.M., Cristescu, R., Loboda, A., Citron, M., Fontenot, J., Hung, D., Schoofs, P., Pearse, M., 2012. Scientific investigations into febrile reactions observed in the paediatric population following vaccination with a 2010 Southern Hemisphere Trivalent Influenza Vaccine. *Vaccine* 30, 7400–7406.
- Marti, M., Baum, J., Rug, M., Tilley, L., Cowman, A.F., 2005. Signal-mediated export of proteins from the malaria parasite to the host erythrocyte. *J. Cell Biol.* 171, 587–592.
- Marti, M., Good, R.T., Rug, M., Knuepfer, E., Cowman, A.F., 2004. Targeting malaria virulence and remodeling proteins to the host erythrocyte. *Science* 306, 1930–1933.
- Menard, R., 2005. Medicine: knockout malaria vaccine? *Nature* 433, 113–114.
- Mikolajczak, S.A., Silva-Rivera, H., Peng, X., Tarun, A.S., Camargo, N., Jacobs-Lorena, V., Daly, T.M., Bergman, L.W., de la Vega, P., Williams, J., Aly, A.S., Kappe, S.H., 2008. Distinct malaria parasite sporozoites reveal transcriptional changes that cause differential tissue infection competence in the mosquito vector and mammalian host. *Mol. Cell. Biol.* 28, 6196–6207.
- Mikolajczak, S.A., Lakshmanan, V., Fishbaugher, M., Camargo, N., Harupa, A., Kaushansky, A., Douglass, A.N., Baldwin, M., Healer, J., O'Neill, M., Phuong, T., Cowman, A., Kappe, S.H., 2014. A next generation genetically attenuated *Plasmodium falciparum* parasite created by triple gene deletion. *Mol. Ther.* <http://dx.doi.org/10.1038/mt.2014.85>.
- Mills, K.H., Ryan, M., Ryan, E., Mahon, B.P., 1998. A murine model in which protection correlates with pertussis vaccine efficacy in children reveals complementary roles for humoral and cell-mediated immunity in protection against *Bordetella pertussis*. *Infect. Immun.* 66, 594–602.
- Moore, C.B., John, M., James, I.R., Christiansen, F.T., Witt, C.S., Mallal, S.A., 2002. Evidence of HIV-1 adaptation to HLA-restricted immune responses at a population level. *Science* 296, 1439–1443.
- Moorthy, V.S., Kienny, M.P., 2010. Reducing empiricism in malaria vaccine design. *Lancet Infect. Dis.* 10, 204–211.
- Moser, M., Leo, O., 2010. Key concepts in immunology. *Vaccine* 28 (Suppl. 3), C2–C13.
- Mu, J., Awadalla, P., Duan, J., McGee, K.M., Keebler, J., Seydel, K., McVean, G.A., Su, X.Z., 2007. Genome-wide variation and identification of vaccine targets in the *Plasmodium falciparum* genome. *Nat. Genet.* 39, 126–130.
- Mueller, A.K., Labaied, M., Kappe, S.H., Matuschewski, K., 2005. Genetically modified *Plasmodium* parasites as a protective experimental malaria vaccine. *Nature* 433, 164–167.
- Mulder, N.J., Apweiler, R., Attwood, T.K., Bairoch, A., Bateman, A., Binns, D., Bradley, P., Bork, P., Bucher, P., Cerutti, L., Copley, R., Courcelle, E., Das, U., Durbin, R., Fleischmann, W., Gough, J., Haft, D., Harte, N., Hulo, N., Kahn, D., Kanapin, A., Krestyaninova, M., Lonsdale, D., Lopez, R., Letunic, I., Madera, M., Maslen, J., McDowell, J., Mitchell, A., Nikolskaya, A.N., Orchard, S., Pagni, M., Ponting, C.P., Quevillon, E., Selengut, J., Sigrist, C.J., Silventoinen, V., Studholme, D.J., Vaughan, R., Wu, C.H., 2005. InterPro, progress and status in 2005. *Nucleic Acids Res.* 33, D201–D205.
- Newell, E.W., Sigal, N., Nair, N., Kidd, B.A., Greenberg, H.B., Davis, M.M., 2013. Combinatorial tetramer staining and mass cytometry analysis facilitate T-cell epitope mapping and characterization. *Nat. Biotechnol.* 31, 623–629.
- Nunes, J.K., Cardenas, V., Loucq, C., Maire, N., Smith, T., Shaffer, C., Maseide, K., Brooks, A., 2013. Modeling the public health impact of malaria vaccines for developers and policymakers. *BMC Infect. Dis.* 13, 295.
- Nussenzweig, V., Nussenzweig, R.S., 1989. Rationale for the development of an engineered sporozoite malaria vaccine. *Adv. Immunol.* 45, 283–334.
- Ogutu, B.R., Apollo, O.J., McKinney, D., Okoth, W., Siangla, J., Dubovsky, F., Tucker, K., Waitumbi, J.N., Diggs, C., Wittes, J., Malkin, E., Leach, A., Soisson, L.A., Milman, J.B., Otieno, L., Holland, C.A., Polhemus, M., Remich, S.A., Ockenhouse, C.F., Cohen, J., Ballou, W.R., Martin, S.K., Angov, E., Stewart, V.A., Lyon, J.A., Heppner, D.G., Withers, M.R., 2009. Blood stage malaria vaccine eliciting high antigen-specific antibody concentrations confers no protection to young children in Western Kenya. *PLoS One* 4, e4708.
- Olutu, A., Fegan, G., Wambua, J., Nyangweso, G., Awuondo, K.O., Leach, A., Lievens, M., Lebouilleux, D., Njuguna, P., Peshu, N., Marsh, K., Bejon, P., 2013. Four-year efficacy of RTS, S/AS01E and its interaction with malaria exposure. *N. Engl. J. Med.* 368, 1111–1120.
- Otsyula, N., Angov, E., Bergmann-Leitner, E., Koeh, M., Khan, F., Bennett, J., Otieno, L., Cummings, J., Andagalu, B., Tosh, D., Waitumbi, J., Richie, N., Shi, M., Miller, L., Otieno, W., Otieno, G.A., Ware, L., House, B., Godeaux, O., Dubois, M.C., Ogutu, B., Ballou, W.R., Soisson, L., Diggs, C., Cohen, J., Polhemus, M., Heppner Jr, D.G., Ockenhouse, C.F., Spring, M.D., 2013. Results from tandem Phase 1 studies



- evaluating the safety, reactogenicity and immunogenicity of the vaccine candidate antigen *Plasmodium falciparum* FVO merozoite surface protein-1 (MSP1(42)) administered intramuscularly with adjuvant system AS01. *Malar. J.* 12, 29.
- Otto, T.D., Wilinski, D., Assefa, S., Keane, T.M., Sarry, L.R., Bohme, U., Lemieux, J., Barrell, B., Pain, A., Berriman, M., Newbold, C., Llinas, M., 2010. New insights into the blood-stage transcriptome of *Plasmodium falciparum* using RNA-Seq. *Mol. Microbiol.* 76, 12–24.
- Ouattara, A., Mu, J., Takala-Harrison, S., Saye, R., Sagara, I., Dicko, A., Niangaly, A., Duan, J., Ellis, R.D., Miller, L.H., Su, X.Z., Plowe, C.V., Doumbo, O.K., 2010. Lack of allele-specific efficacy of a bivalent AMA1 malaria vaccine. *Malar. J.* 9, 175.
- Ouattara, A., Takala-Harrison, S., Thera, M.A., Coulibaly, D., Niangaly, A., Saye, R., Tolo, Y., Dutta, S., Heppner, D.G., Soisson, L., Diggs, C.L., Vekemans, J., Cohen, J., Blackwelder, W.C., Dube, T., Laurens, M.B., Doumbo, O.K., Plowe, C.V., 2013. Molecular basis of allele-specific efficacy of a blood-stage malaria vaccine: vaccine development implications. *J. Infect. Dis.* 207, 511–519.
- Pain, A., Bohme, U., Berry, A.E., Mungall, K., Finn, R.D., Jackson, A.P., Mourier, T., Mistry, J., Pasini, E.M., Aslett, M.A., Balasubramanian, S., Borgwardt, K., Brooks, K., Carret, C., Carver, T.J., Cherevach, I., Chillingworth, T., Clark, T.G., Galinski, M.R., Hall, N., Harper, D., Harris, D., Hauser, H., Ivens, A., Janssen, C.S., Keane, T., Larke, N., Lapp, S., Marti, M., Moule, S., Meyer, I.M., Ormond, D., Peters, N., Sanders, M., Sanders, S., Sargeant, T.J., Simmonds, M., Smith, F., Squares, R., Thurston, S., Tivey, A.R., Walker, D., White, B., Zuijderwijk, E., Churcher, C., Quail, M.A., Cowman, A.F., Turner, C.M., Rajandream, M.A., Kocken, C.H., Thomas, A.W., Newbold, C.I., Barrell, B.G., Berriman, M., 2008. The genome of the simian and human malaria parasite *Plasmodium knowlesi*. *Nature* 455, 799–803.
- Pallansch, M.A., Sandhu, H.S., 2006. The eradication of polio – progress and challenges. *N. Engl. J. Med.* 355, 2508–2511.
- Patankar, S., Munasinghe, A., Shoaibi, A., Cummings, L.M., Wirth, D.F., 2001. Serial analysis of gene expression in *Plasmodium falciparum* reveals the global expression profile of erythrocytic stages and the presence of anti-sense transcripts in the malarial parasite. *Mol. Biol. Cell* 12, 3114–3125.
- Petrovic, D., Dempsey, E., Doherty, D.G., Kelleher, D., Long, A., 2012. Hepatitis C virus – T-cell responses and viral escape mutations. *Eur. J. Immunol.* 42, 17–26.
- Pizza, M., Scarlato, V., Masignani, V., Giuliani, M.M., Arico, B., Comanducci, M., Jennings, G.T., Baldi, L., Bartolini, E., Capocchi, B., Galeotti, C.L., Luzzi, E., Manetti, R., Marchetti, E., Mora, M., Nuti, S., Ratti, G., Santini, L., Savino, S., Scarselli, M., Storni, E., Zuo, P., Broecker, M., Hundt, E., Knapp, B., Blair, E., Mason, T., Tettelin, H., Hood, D.W., Jeffries, A.C., Saunders, N.J., Granoff, D.M., Venter, J.C., Moxon, E.R., Grandi, G., Rappuoli, R., 2000. Identification of vaccine candidates against serogroup B meningococcus by whole-genome sequencing. *Science* 287, 1816–1820.
- Plotkin, S.A., 2008. Vaccines: correlates of vaccine-induced immunity. *Clin. Infect. Dis.* 47, 401–409.
- Poland, G., Barrett, A., 2009. The old and the new: successful vaccines of the 20th century and approaches to making vaccines for the important diseases of the 21st century. *Curr. Opin. Immunol.* 21, 305–307.
- Poland, G.A., Quill, H., Tognias, A., 2013. Understanding the human immune system in the 21st century: the Human Immunology Project Consortium. *Vaccine* 31, 2911–2912.
- Pulendran, B., Li, S., Nakaya, H.I., 2010. Systems vaccinology. *Immunity* 33, 516–529.
- Querec, T.D., Akondy, R.S., Lee, E.K., Cao, W., Nakaya, H.I., Teuwen, D., Pirani, A., Gernert, K., Deng, J., Marzolf, B., Kennedy, K., Wu, H., Bennouna, S., Oluoch, H., Miller, J., Vencio, R.Z., Mulligan, M., Aderem, A., Ahmed, R., Pulendran, B., 2009. Systems biology approach predicts immunogenicity of the yellow fever vaccine in humans. *Nat. Immunol.* 10, 116–125.
- Rappuoli, R., 2000. Reverse vaccinology. *Curr. Opin. Microbiol.* 3, 445–450.
- Rappuoli, R., Black, S., Lambert, P.H., 2011. Vaccine discovery and translation of new vaccine technology. *Lancet* 378, 360–368.
- Reed, S.G., Orr, M.T., Fox, C.B., 2013. Key roles of adjuvants in modern vaccines. *Nat. Med.* 19, 1597–1608.
- Remarque, E.J., Faber, B.W., Kocken, C.H., Thomas, A.W., 2008. Apical membrane antigen 1: a malaria vaccine candidate in review. *Trends Parasitol.* 24, 74–84.
- Ren, F., Hino, K., Yamaguchi, Y., Funatsuki, K., Hayashi, A., Ishiko, H., Furutani, M., Yamasaki, T., Korenaga, K., Yamashita, S., Konishi, T., Okita, K., 2003. Cytokine-dependent anti-viral role of CD4-positive T cells in therapeutic vaccination against chronic hepatitis B viral infection. *J. Med. Virol.* 71, 376–384.
- Rhee, J.H., 2014. Towards Vaccine 3.0: new era opened in vaccine research and industry. *Clin. Exp. Vaccine Res.* 3, 1–4.
- Roestenberg, M., McCall, M., Hopman, J., Wiersma, J., Luty, A.J., van Gemert, G.J., van de Vegte-Bolmer, M., van Schaijk, B., Teelen, K., Arens, T., Spaarman, L., de Mast, Q., Roeffen, W., Snounou, G., Renia, L., van der Ven, A., Hermsen, C.C., Sauerwein, R., 2009. Protection against a malaria challenge by sporozoite inoculation. *N. Engl. J. Med.* 361, 468–477.
- Roestenberg, M., Teirlinck, A.C., McCall, M.B., Teelen, K., Makamop, K.N., Wiersma, J., Arens, T., Beckers, P., van Gemert, G., van de Vegte-Bolmer, M., van der Ven, A.J., Luty, A.J., Hermsen, C.C., Sauerwein, R.W., 2011. Long-term protection against malaria after experimental sporozoite inoculation: an open-label follow-up study. *Lancet* 377, 1770–1776.
- Rovira-Graells, N., Gupta, A.P., Planet, E., Crowley, V.M., Mok, S., Ribas de Pouplana, L., Preiser, P.R., Bozdech, Z., Cortes, A., 2012. Transcriptional variation in the malaria parasite *Plasmodium falciparum*. *Genome Res.* 22, 925–938.
- Rueckert, C., Guzman, C.A., 2012. Vaccines: from empirical development to rational design. *PLoS Pathog.* 8, e1003001.
- Sagara, I., Dicko, A., Ellis, R.D., Fay, M.P., Diawara, S.I., Assadou, M.H., Sissoko, M.S., Kone, M., Diallo, A.I., Saye, R., Guindo, M.A., Kante, O., Niamele, M.B., Miura, K., Mullen, G.E., Pierce, M., Martin, L.B., Dolo, A., Diallo, D.A., Doumbo, O.K., Miller, L.H., Saul, A., 2009. A randomized controlled phase 2 trial of the blood stage AMA1-C1/Alhydrogel malaria vaccine in children in Mali. *Vaccine* 27, 3090–3098.
- Salk, J., Salk, D., 1977. Control of influenza and poliomyelitis with killed virus vaccines. *Science* 195, 834–847.
- Sallusto, F., Lanzavecchia, A., Araki, K., Ahmed, R., 2010. From vaccines to memory and back. *Immunity* 33, 451–463.
- Schillie, S.F., Murphy, T.V., 2013. Seroprotection after recombinant hepatitis B vaccination among newborn infants: a review. *Vaccine* 31, 2506–2516.
- Schussek, S., Trieu, A., Doolan, D.L., 2014. Genome- and proteome-wide screening strategies for antigen discovery and immunogen design. *Biotechnol. Adv.* 32, 403–414.
- Schwartz, L., Brown, G.V., Genton, B., Moorthy, V.S., 2012. A review of malaria vaccine clinical projects based on the WHO rainbow table. *Malar. J.* 11, 11.
- Schwarz, T.F., Leo, O., 2008. Immune response to human papillomavirus after prophylactic vaccination with AS04-adjuvanted HPV-16/18 vaccine: improving upon nature. *Gynecol. Oncol.* 110, S1–S10.
- Seder, R.A., Chang, L.J., Enama, M.E., Zephir, K.L., Sarwar, U.N., Gordon, I.J., Holman, L.A., James, E.R., Billingsley, P.F., Gunasekera, A., Richman, A., Chakravarty, S., Manoj, A., Velmurugan, S., Li, M., Ruben, A.J., Li, T., Eappen, A.G., Stafford, R.E., Plummer, S.H., Hendel, C.S., Novik, L., Costner, P.J., Mendoza, F.H., Saunders, J.G., Nason, M.C., Richardson, J.H., Murphy, J., Davidson, S.A., Richie, T.L., Sedegah, M., Sutamihardja, A., Fahle, G.A., Lyke, K.E., Laurens, M.B., Roederer, M., Tewari, K., Epstein, J.E., Sim, B.K., Ledgerwood, J.E., Graham, B.S., Hoffman, S.L., Team, V.R.C.S., 2013. Protection against malaria by intravenous immunization with a nonreplicating sporozoite vaccine. *Science* 341, 1359–1365.
- Seder, R.A., Hill, A.V., 2000. Vaccines against intracellular infections requiring cellular immunity. *Nature* 406, 793–798.
- Sercarz, E.E., Lehmann, P.V., Ametani, A., Benichou, G., Miller, A., Moudgil, K., 1993. Dominance and crypticity of T cell antigenic determinants. *Ann. Rev. Immunol.* 11, 729–766.
- Sette, A., Fleri, W., Peters, B., Sathiamurthy, M., Bui, H.H., Wilson, S., 2005. A roadmap for the immunomics of category A-C pathogens. *Immunity* 22, 155–161.
- Six, A., Bellier, B., Thomas-Vaslin, V., Klatzmann, D., 2012. Systems biology in vaccine design. *Microb. Biotechnol.* 5, 295–304.
- Smith, T., Ross, A., Maire, N., Chitnis, N., Studer, A., Hardy, D., Brooks, A., Penny, M., Tanner, M., 2012. Ensemble modeling of the likely public health impact of a pre-erythrocytic malaria vaccine. *PLoS Med.* 9, e1001157.
- Spring, M., Murphy, J., Nielsen, R., Dowler, M., Bennett, J.W., Zarling, S., Williams, J., de la Vega, P., Ware, L., Komisar, J., Polhemus, M., Richie, T.L., Epstein, J., Tamminga, C., Chuang, I., Richie, N., O’Neil, M., Heppner, D.G., Healer, J., O’Neill, M., Smithers, H., Finney, O.C., Mikolajczak, S.A., Wang, R., Cowman, A., Ockenhouse, C., Krzych, U., Kappe, S.H., 2013. First-in-human evaluation of genetically attenuated *Plasmodium falciparum* sporozoites administered by bite of *Anopheles* mosquitoes to adult volunteers. *Vaccine* 31, 4975–4983.
- Stewart, V.A., McGrath, S.M., Walsh, D.S., Davis, S., Hess, A.S., Ware, L.A., Kester, K.E., Cummings, J.F., Burge, J.R., Voss, G., Delchambre, M., Garcon, N., Tang, D.B., Cohen, J.D., Heppner Jr., D.G., 2006. Pre-clinical evaluation of new adjuvant formulations to improve the immunogenicity of the malaria vaccine RTS, S/AS02A. *Vaccine* 24, 6483–6492.
- Stoute, J.A., Slaoui, M., Heppner, D.G., Momin, P., Kester, K.E., Desmons, P., Wellde, B.T., Garcon, N., Krzych, U., Marchand, M., 1997. A preliminary evaluation of a recombinant circumsporozoite protein vaccine against *Plasmodium falciparum* malaria. *N. Engl. J. Med.* 336, 86–91.
- Takala, S.L., Plowe, C.V., 2009. Genetic diversity and malaria vaccine design, testing and efficacy: preventing and overcoming ‘vaccine resistant malaria’. *Parasite Immunol.* 31, 560–573.
- Tan, A.C., La Gruta, N.L., Zeng, W., Jackson, D.C., 2011. Precursor frequency and competition dictate the HLA-A2-restricted CD8+ T cell responses to influenza A infection and vaccination in HLA-A2.1 transgenic mice. *J. Immunol.* 187, 1895–1902.
- Tarun, A.S., Peng, X., Dumpit, R.F., Ogata, Y., Silva-Rivera, H., Camargo, N., Daly, T.M., Bergman, L.W., Kappe, S.H., 2008. A combined transcriptome and proteome survey of malaria parasite liver stages. *Proc. Natl. Acad. Sci. U.S.A.* 105, 305–310.
- Taylor, S.M., Cerami, C., Fairhurst, R.M., 2013. Hemoglobinopathies: slicing the Gordian knot of *Plasmodium falciparum* malaria pathogenesis. *PLoS Pathog.* 9, e1003327.
- Thera, M.A., Doumbo, O.K., Coulibaly, D., Laurens, M.B., Ouattara, A., Kone, A.K., Guindo, A.B., Traore, K., Traore, I., Kouriba, B., Diallo, D.A., Diarra, I., Daou, M., Dolo, A., Tolo, Y., Sissoko, M.S., Niangaly, A., Sissoko, M., Takala-Harrison, S., Lyke, K.E., Wu, Y., Blackwelder, W.C., Godeaux, O., Vekemans, J., Dubois, M.C., Ballou, W.R., Cohen, J., Thompson, D., Dube, T., Soisson, L., Diggs, C.L., House, B., Lanar, D.E., Dutta, S., Heppner Jr., D.G., Plowe, C.V., 2011. A field trial to assess a blood-stage malaria vaccine. *N. Engl. J. Med.* 365, 1004–1013.
- Thomas, S., Luxon, B.A., 2013. Vaccines based on structure-based design provide protection against infectious diseases. *Expert Rev. Vaccines* 12, 1301–1311.
- Tifrea, D.F., Pal, S., Popot, J.L., Cocco, M.J., de la Maza, L.M., 2014. Increased immunoreactivity of MOMP epitopes in a vaccine formulated with amphiphols may account for the very robust protection elicited against a vaginal challenge with *Chlamydia muridarum*. *J. Immunol.* 192, 5201–5213.
- Trieu, A., Kayala, M.A., Burk, C., Molina, D.M., Freilich, D.A., Richie, T.L., Baldi, P., Felgner, P.L., Doolan, D.L., 2011. Sterile protective immunity to malaria is associated with a panel of novel *P. falciparum* antigens. *Mol. Cell. Proteomics* 10 (M111), 007948.

- Tuell, J., 2012. Vaccinology: the name, the concept, the adjectives. *Vaccine* 30, 5491–5495.
- U.S. Food and Drug Administration, 2014. Complete List of Vaccines Licensed for Immunization and Distribution in the U.S.. U.S. Department of Health and Human Services, Silver Spring, USA, <<http://www.fda.gov/biologicsbloodvaccines/vaccines/approvedproducts/ucm093833.htm>>.
- Vaughan, A.M., Kappe, S.H., 2013. Vaccination using radiation- or genetically attenuated live sporozoites. *Methods Mol. Biol.* 923, 549–566.
- Vignali, M., Armour, C.D., Chen, J., Morrison, R., Castle, J.C., Biery, M.C., Bouzek, H., Moon, W., Babak, T., Fried, M., Raymond, C.K., Duffy, P.E., 2011. NSR-seq transcriptional profiling enables identification of a gene signature of *Plasmodium falciparum* parasites infecting children. *J. Clin. Invest.* 121, 1119–1129.
- Volkman, S.K., Sabeti, P.C., DeCaprio, D., Neafsey, D.E., Schaffner, S.F., Milner Jr., D.A., Daily, J.P., Sarr, O., Ndiaye, D., Ndir, O., Mboup, S., Duraisingh, M.T., Lukens, A., Derr, A., Stange-Thomann, N., Waggoner, S., Onofrio, R., Ziaugra, L., Mauceli, E., Gnerre, S., Jaffe, D.B., Zainoun, J., Wiegand, R.C., Birren, B.W., Hartl, D.L., Galagan, J.E., Lander, E.S., Wirth, D.F., 2007. A genome-wide map of diversity in *Plasmodium falciparum*. *Nat. Genet.* 39, 113–119.
- Watanabe, J., Wakaguri, H., Sasaki, M., Suzuki, Y., Sugano, S., 2007. Comparasite: a database for comparative study of transcriptomes of parasites defined by full-length cDNAs. *Nucleic Acids Res.* 35, D431–D438.
- Winzeler, E.A., 2006. Applied systems biology and malaria. *Nat. Rev. Microbiol.* 4, 145–151.
- World Health Organization, 2005. GIVS – Global Immunization Vision and Strategy 2006–2015. Geneva (WHO/IVB/05.05). <[http://www.who.int/vaccines-documents/DocsPDF05/GIVS\\_Final\\_EN.pdf](http://www.who.int/vaccines-documents/DocsPDF05/GIVS_Final_EN.pdf)>.
- World Health Organization, 2006. State of the Art of New Vaccine Research and Development. Geneva (WHO/IVB/06.01). <[http://whqlibdoc.who.int/hq/2006/WHO\\_IVB\\_06.01\\_eng.pdf](http://whqlibdoc.who.int/hq/2006/WHO_IVB_06.01_eng.pdf)>.
- World Health Organization, 2007. GFIMS – Global Framework for Immunization Monitoring, and Surveillance. Geneva (WHO/IVB/07.06). <[http://whqlibdoc.who.int/hq/2007/WHO\\_IVB\\_07.06\\_eng.pdf](http://whqlibdoc.who.int/hq/2007/WHO_IVB_07.06_eng.pdf)>.
- World Health Organization, 2009. WHO, UNICEF, World Bank. State of the World's Vaccines and Immunization, 3rd ed. Geneva, <[http://www.unicef.org/media/files/SOWVI\\_full\\_report\\_english\\_LR1.pdf](http://www.unicef.org/media/files/SOWVI_full_report_english_LR1.pdf)>.
- World Health Organization, 2013. Global Vaccine Action Plan (GVAP) 2011–2020. ISBN 978 92 4 150498 0. <[http://www.who.int/immunization/global\\_vaccine\\_action\\_plan/GVAP\\_doc\\_2011\\_2020/en/](http://www.who.int/immunization/global_vaccine_action_plan/GVAP_doc_2011_2020/en/)>.
- Yewdell, J.W., 2013. To dream the impossible dream: universal influenza vaccination. *Curr. Opin. Virol.* 3, 316–321.
- Yewdell, J.W., Bennink, J.R., 2001. Cut and trim: generating MHC class I peptide ligands. *Curr. Opin. Immunol.* 13, 13–18.
- Zepp, F., 2010. Principles of vaccine design – lessons from nature. *Vaccine* 28 (Suppl. 3), C14–C24.
- Zhou, Y., Ramachandran, V., Kumar, K.A., Westenberger, S., Refour, P., Zhou, B., Li, F., Young, J.A., Chen, K., Plouffe, D., Henson, K., Nussenzweig, V., Carlton, J., Vinetz, J.M., Duraisingh, M.T., Winzeler, E.A., 2008. Evidence-based annotation of the malaria parasite's genome using comparative expression profiling. *PLoS ONE* 3, e1570.