

Contribution to the phylogeography of the nose-horned viper (*Vipera ammodytes*, Linnaeus, 1758) in the Central Balkan Peninsula

Tijana Čubrić^{1,*}, Gorana Stamenković², Marija Ilić³ and Jelka Crnobrnja-Isailović^{1,4}

¹Department of Biology and Ecology, Faculty of Science and Mathematics, University of Niš, Višegradska 33, 18 000 Niš, Serbia

²Department of Genetics, Institute for Biological Research “Siniša Stanković” – National Institute of the Republic of Serbia, University of Belgrade, Despota Stefana 142, 11000 Belgrade, Serbia

³Department of Hydroecology and Water Protection, Institute for Biological Research “Siniša Stanković” – National Institute of the Republic of Serbia, University of Belgrade, Despota Stefana 142, 11000 Belgrade, Serbia

⁴Department of Evolutionary Biology, Institute for Biological Research “Siniša Stanković” – National Institute of the Republic of Serbia, University of Belgrade, Despota Stefana 142, 11000 Belgrade, Serbia

*Corresponding author: tijana.cubric@pmf.edu.rs

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Abstract: Seven genetic clades have been recognized within the species *Vipera ammodytes* (nose-horned viper); however, the precise phylogenetic position of many Balkan populations is unknown. We used Bayesian analysis of the mtDNA sequences from the 16S rRNA mtDNA gene obtained from 47 individuals (26 novel samples sequenced in this study and 21 sequences available from GenBank). Our results show that sampled nose-horned vipers from localities in Serbia are clustered within three clades: the northeastern (23 individuals), the northwestern (two individuals) and the southeastern (one individual). Results revealed an overlapping distribution of the northeastern and the northwestern clades in two populations. We have revealed that the northeastern clade extends further south than previously suggested, to the Ohrid/Prespa lakes in North Macedonia. Our findings contribute to the knowledge of the genetic diversity of this species in Serbia and help to clarify the geographical distributions of mtDNA-defined clades.

Keywords: *Vipera ammodytes*; nose-horned viper; phylogeography; 16S rRNA; Balkan Peninsula

INTRODUCTION

Phylogeographic studies serve as a valuable tool in conservation biology [1-4], not only to help understand the evolutionary history of a species but for providing insight into a species' genetic diversity, which further enables proper assessment of the impact of various threats [5,6]. The use of mitochondrial DNA (mtDNA) has been applied in molecular analyses to aid conservation studies to investigate the genetic diversity and genetic structure of a species and identify any evolutionarily significant units [7-12]. Furthermore, phylogeographic studies are used to understand the role of paleogeographical events and demographic processes in shaping diversification within a species and the geographical distribution of the lineages [13].

Vipera ammodytes (Squamata: Serpentes: Viperidae: Viperinae) is a venomous viper with a distribu-

tion ranging from southern Austria and northern Italy, through the Balkans to Asia Minor [14]. This snake inhabits dry rocky habitats including woodland and scrub, sand dunes, hillsides, screes, stone walls, and sometimes cultivated areas with an elevation range limit of up to 2500 m a.s.l. [14,16,17]. Seven subspecies of *V. ammodytes* were recognized in the past on the basis of phenotypic characters [14,18] before contemporary multivariate analyses of morphology synonymized three, and suggested the validity of only four subspecies, *V. a. ammodytes*, *V. a. meridionalis*, *V. a. montandoni* and *V. a. transcaucasiana* [19].

Using a phylogeographic approach, seven main genetic clades within *V. ammodytes* were defined [15] as follows: (1) a northwestern clade that inhabits Italy, Austria, Slovenia, Croatia and Bosnia and Herzegovina; (2) a northeastern clade from western, central and

eastern Serbia and western Bulgaria; (3) a Montenegrin clade from Montenegro; (4) a southwestern clade from Albania and northwestern Greece; (5) a southeastern clade from the southernmost part of Serbia, most of the Republic of North Macedonia, central and eastern Bulgaria, northern and central Greece, Turkey, Armenia and Georgia; (6) a Peloponnesian clade from the Peloponnesian peninsula; (7) a Cyclades clade from the Cyclades islands. However, the distribution boundaries of most of these clades have not been clearly defined due to the lack of samples throughout the range and from boundary zones of the clades, and because of the absence of nuclear markers. More information on the contact zones would be of great importance for our understanding of the current distribution and evolutionary integrity of these lineages.

The main aim of this study was to identify the position of some populations of *V. ammodytes* in Serbia from localities that have not been included in previous phylogeographic studies and are geographically positioned in proposed delineation zones of the main clades [15]. For analysis, we used mtDNA sequences from the 16S rRNA gene as several sequences of the same gene were already available for this species [15]; this is a common and reliable universal barcoding marker used for vertebrate studies [20].

MATERIALS AND METHODS

Ethics statement

All samples were collected with permits obtained from the Ministry of Agriculture and Nature Protection (No. 353-01-170/2016-17 and No.353-01-2666/2016-17). At present, vipers are not protected by law in Montenegro, while the sample from the border between Albania and the Republic of North Macedonia was collected opportunistically from a road-kill specimen.

Sample collection

This study was based on a combination of 16S rRNA sequences from novel samples and sequences obtained from GenBank. In total, we analyzed 47 16S rRNA sequences. Tissues were collected from tail tips (preserved in 96% ethanol) from 26 individuals sampled at 15 localities in Serbia, one locality in Montenegro and

one locality in the Republic of North Macedonia directly bordering Albania. The samples in Serbia were mostly collected during 2016 and 2017, whereas samples from Montenegro and Albania/North Macedonia were collected in 2010. Twenty-one 16S rRNA sequences submitted to GenBank by Ursenbacher et al. [15] were included. GenBank accession numbers for previously published sequences and from this study with the localities for collected samples are given in Supplementary Table S1. Four additional 16S rRNA sequences from GenBank of four different species of vipers (*Vipera latastei*, *Vipera aspis*, *Montivipera xanthina* and *Vipera berus*) from GenBank were used as outgroups in the phylogenetic analysis.

PCR and sequencing

Tissue preserved in ethanol (5 mm of snake tail) was used for DNA extraction. DNA was extracted using Accuprep® Genomic DNA Extraction kit according to manufacturer's instructions (Bioneer, Daejeon, South Korea). A 544-bp fragment of 16S rRNA was amplified by the polymerase chain reaction (PCR) using primers 16SA (5'-CGCCTGTTTATCAAAAACAT-3') and 16SB (5'-CCGG TCTGAACTCAGATCACGT-3'), in an established PCR protocol [21]. Briefly, about 100 ng of extracted DNA was amplified in a 25- μ L total volume of the reaction solution by PCR as follows: initial denaturation: 95°C, 3 min; 40 cycles: denaturation 95°C, 30 s; annealing 52°C, 30 s; extension: 72°C, 40 s. The amplified products were sequenced using the Sanger protocol in both directions on automated equipment by Macrogen (Seoul, South Korea).

Phylogenetic analysis

Sequences obtained in this study were visually inspected by FinchTV (Geospiza, Inc., Seattle, WA) and aligned with published sequences of the same mtDNA region using ClustalW [22]. Bayesian phylogenetic inference was performed with MrBayes software v.3.2 [23], using TPM3uf (three parameter model) [24] + I model, which was selected by the JmodelTest program v.2.1.4 [25]. A Markov Chain Monte Carlo (MCMC) search was made for 3×10^6 generations using tree sampling every 100th generation; 25% of the trees were discarded as 'burn-in' before generating a consensus tree. Consensus trees were viewed and edited in FigTree

v. 1.4.0. Genetic distances between novel sequences were calculated using neighbor-joining method implemented in PAUP v. 4.0b10.

RESULTS

The analysis of 26 sequences of the 16S rRNA gene revealed eight unique haplotypes. In Serbia, six haplotypes were detected: the first was shared by 15 samples, the second was shared by two (15KRP_RS and 3SVL_RS), and each of the remaining detected haplotypes were unique to one sample each (19RKO_RS, 24POZ_RS, 26LJU_RS and 27MGT_RS). The remaining two haplotypes represented samples from Albania/North Macedonia (21ALM_MK) and another from Montenegro (18MON_ME). All samples sequenced in this work had a deletion at position 2206 nt of mtDNA in the reference sequence NC 036956, except for the sample from Montenegro (18_MON_ME), which had thymine at the position, as did a few other GenBank sequences (MO6_ME, MO2_ME, SW3_GR, SW2_GR, SW1_AL, SE5_BG and SE1_RS, SE3_AR, SE4_TR, CY4_GR, CY3_GR, CY2_GR, PE2_GR, PE1_GR, PE3_GR).

The phylogenetic tree (Fig. 1) was concordant with the previous study on *V. ammodytes* [15] regarding the main genetic clades. In two localities (in western and central Serbia) we confirmed the overlapping distribution of two distinctive clades (northeastern and northwestern). One of the two individuals belonging to the same local population in western Serbia (code 4KRP_RS in Fig. 1) clustered with the northeastern clade, the other (code 15KRP_RS) clustered with the northwestern clade. The genetic distance between these two vipers was estimated as 0.01 (1%). At the other locality from central Serbia, six of the seven analyzed individuals from the same local population clustered with the northeastern clade, while individual 3SVL_RS clustered with the northwestern clade (see Fig. 1). The genetic distance between the “northwestern” individual (code 3SVL_RS) and another six from that same population was estimated as 0.01 (1%). All the other novel samples from Serbia clustered with the northeastern clade (Fig. 1). The specimen from Montenegro (code 18MON_ME) clustered with the Montenegrin branch of the northeastern clade. The nose-horned viper collected on the Albanian-North Macedonian border (code 21ALM_MK) clustered

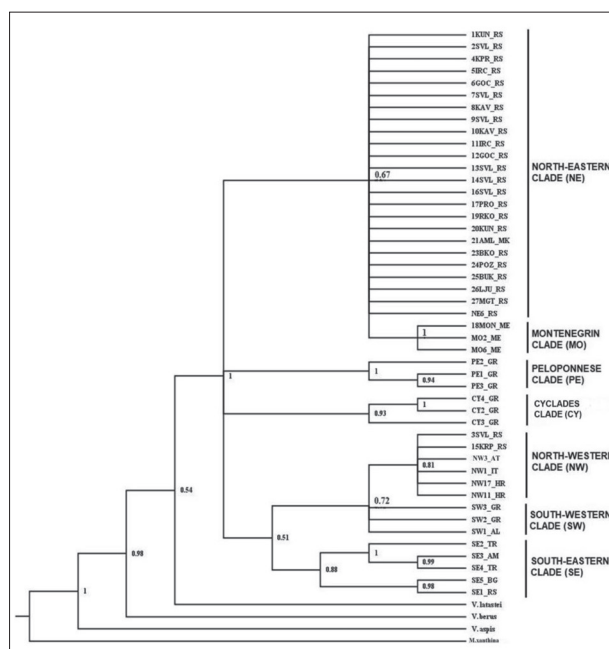


Fig. 1. Phylogenetic analyses of 16S rRNA sequences from 47 samples of *Vipera ammodytes* including four outgroup sequences from *Vipera aspis*, *Vipera latastei*, *Vipera berus* and *Montivipera xanthina*. Phylogenetic tree inferred by Bayesian statistics integrated in MrBayes v.3.2. software. Bootstrap values are shown in nodes.

with the northeastern clade. We have presented the geographical positions of the identified genetic clades for the samples analyzed in this study in Fig. 2, together with the surrounding sequences from GenBank showing their affiliation to the specific clade.

DISCUSSION

We recognize the presence of three main *V. ammodytes* mtDNA-defined clades in Serbia instead of the two that were previously identified [15]: (1) populations from most of Serbia are part of the northeastern clade; (2) in the extreme southeast of Serbia, the populations are part of the southeastern clade; (3) we have also identified a couple from the northwestern clade in western and central Serbia, where they overlap with the northeastern clade. Western Serbia has been mentioned [26] as part of the eastern border for the northwestern clade, and it was assumed that specific orographic formations and aquatic barriers prevented the further spread of the northwestern clade east and southeast into the Balkans. It is still not clear which barriers contributed to the distribution of these two clades in Serbia. The detec-

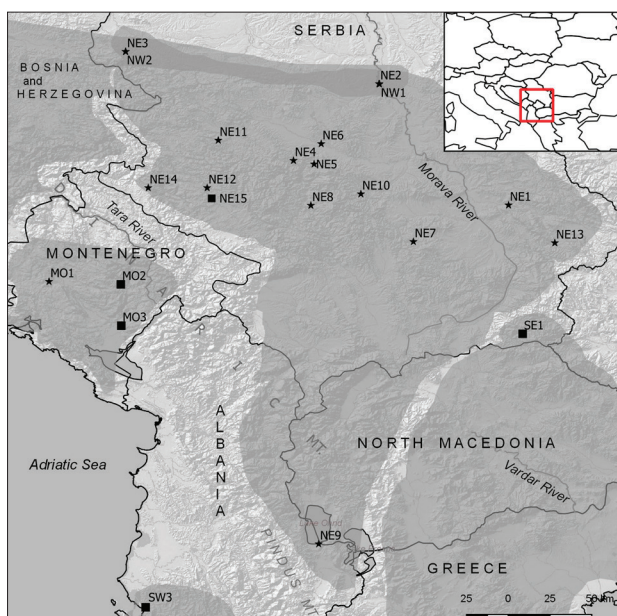


Fig. 2. Map of the sampling localities of identified genetic clades (this study) with surrounding samples used from GenBank. Multiple samples from the same location that belong to the same clade are represented with a single mark for the following: NE2 (6), NE5 (2), NE6 (2). Novel samples are marked with stars while the used GenBank samples are marked with squares. NE corresponds to the northeastern clade, NW corresponds to the northwestern clade, MO corresponds to the Montenegrin clade, SE corresponds to the southeastern clade, SW corresponds to the southwestern clade; these abbreviations correspond to the abbreviations of clades in Fig. 1. Shaded areas suggest the updated potential geographic distribution of the genetic clades in the Central Balkan Peninsula.

tion of populations in the westernmost part of Serbia that have individuals clustering with the northwestern clade suggests that the Drina River, which represents the natural border between Bosnia and Herzegovina and Serbia, has not been a strong enough barrier to isolate the northwestern clade in the west.

We assume that the presence of the northwestern clade in western and central Serbia is probably due to a natural transition zone between the two clades. However, we must take into consideration information obtained from our interviews with local inhabitants at the location in central Serbia where one of the seven samples clustered with the northwestern clade (3_SVL_RS); two farmers stated that 10 to 20 years ago there was a deliberate release of *V. ammodytes* collected from different localities in former Yugoslavia for the purpose of venom milking and production of anti-venom components. Therefore, the presence of genes corresponding to the northwestern clade could

be the result of persisting haplotypes originating from individuals released outside of their place of origin.

Another contribution of this study is the more detailed understanding of the geographic distribution of the main genetic clades of *V. ammodytes* in the Central Balkans. Our results suggest that the northeastern clade extends further southward into the Balkans than previously defined [15]. According to the molecular clock [15], the first splits within *V. ammodytes* occurred during the early Pliocene (3.4-4.9 Mya), separating the Montenegrin and northeastern clade from the northwestern, the southwestern from the Peloponnese, Cyclades and southeastern clades. However, the authors could not determine where the split occurred. There were extensive inflows of the sea during the Pliocene at the area of the Axios and Strymon river basins that reached northwards almost to the Central Balkans along the Vardar and perhaps the Morava River valleys [27]. The presence of the Dinaric, Pindus and Rhodope massifs could explain the distribution of the northeastern and the southeastern clades on the territory of North Macedonia. These orographic formations and aquatic barriers isolated the southeastern, the southwestern and the northeastern clades in this area. As postulated for *V. ammodytes* [15], *Ichthyosaura alpestris* [28] and *Lissotriton vulgaris* [29], the distribution of the southwestern clade corresponds to regions of the “refugia-within-refugia” isolated by the Dinaric and Pindus Mts. Nevertheless, as there is a lack of samples from most of the territory of Kosovo, North Macedonia and Albania, in order to clearly define the boundary zones between the northeastern and the southeastern clades, further analyses of additional populations from these areas are needed.

The precise shape of the western boundary of the Montenegrin clade has not been defined [15] and implies the necessity of analyzing populations in the area extending from Dubrovnik in the north to the border between Montenegro and Croatia in the south. Sampling should also include populations distributed from the border zone between Bosnia and Herzegovina and Montenegro north of the Tara River canyon and further west to the Neretva River, which is known to be one of the biogeographic barriers in this part of the Balkan Peninsula [30].

Further investigations include an analysis of other nuclear genetic markers apart from mtDNA, in order to

identify the level of gene flow between these clades as it is possible that nuclear gene exchange does occur [31]. This species has low dispersal capability. Together with the results of the investigation of Ursenbacher et al. [15] based only on mtDNA, it would appear that sex-biased dispersal contributed to the obtained pattern of haplotype distribution. If males tend to disperse further than females during the mating season, which is expected and confirmed for most snake species [32-34], genetic analysis of nuclear genes would be necessary for further clarification of the contact zones between clades. To explain the distribution of nose-horned viper haplotypes in the studied area, we should not ignore the possible impact of occasional human-mediated transport.

The results of this study support future conservation plans for the nose-horned viper in the Central Balkans. This species is legally protected in Serbia because of uncontrolled overexploitation in the past [35], and it is still subject to illegal collecting at one locality (personal communication from a local collector). Even sustainable exploitation can lead to alteration and possible loss of genetic variation [36], and maintaining a species' genetic diversity in areas where it is exposed to illegal collecting is crucial [2].

To conclude, the results of this study provide insight into the phylogenetic positions of nose-horned viper populations that inhabit the boundary areas of mtDNA-identified clades in the Central Balkans. Our findings underline the importance of analyzing multiple samples from the same local population in genetic studies, especially if those populations are located in the supposed transition zones between distinctive genetic clades. To obtain a more complete insight into species' or population genetic structures, it is necessary to conduct comprehensive analyses using both mitochondrial and nuclear genes and to include a range of samples from locations in the supposed contact zones of the main genetic clades.

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Author contributions: Tijana Čubrić was responsible for study design, sample collecting, part of the genetic analysis, interpretation of the results, writing of the manuscript and technical preparation of the manuscript. Gorana Stamenković was responsible for the genetic analysis, partly for the interpretation of the results and for reviewing the draft version of the manuscript. Marija Ilić participated in the genetic analysis. Jelka Crnobrnja-Isailović was responsible for part of the sample collecting, interpretation of the results and review of the draft version of manuscript.

Conflict of interest disclosure: The authors declare that there are no conflicts of interest.

REFERENCES

1. Frankham R, Ballou JD, Briscoe, DA: A primer of Conservation Genetics. 4th ed. Cambridge: Cambridge University Press; 2004.
2. Frankham R, Briscoe, DA, Ballou, JD: Introduction to conservation genetics. 1st ed. Cambridge: Cambridge University Press; 2002.
3. Atnaf M, Yao N, Martina K, Dagne K, Wegary D, Tesfaye K. Molecular genetic diversity and population structure of Ethiopian white lupin landraces: Implications for breeding and conservation. PLoS ONE. 2017;12(11):e0188696.
4. Ralls K, Ballou JD, Dudash, MR, Eldridg MD, Fenster CB, Lacy RC, Sunnucks P, Frankham R. Call for a paradigm shift in the genetic management of fragmented populations. Conserv Lett. 2018;11:e12412.
5. Teixeira J, Gonçalves H, Ferrand N, García-París M, Recuero E. Mitochondrial phylogeography of the Iberian endemic frog *Rana iberica*, with implications for its conservation. Curr Zool. 2018;64(6):755-64.
6. Kheng V, Zichello JM, Lumbantobing DN, Lawalata SZ, Andayani N, Melnick DJ. Phylogeography, Population Structure, and Conservation of the Javan Gibbon (*Hylobates moloch*). Int J Primatol. 2018;39(1):5-26.
7. Moritz C. Applications of mitochondrial DNA analysis in conservation: a critical review. Mol Ecol. 1994;3(4):401-11.
8. Hui M, Nuryanto A, Kochzius M. Concordance of microsatellite and mitochondrial DNA markers in detecting genetic population structure in the boring giant clam *Tridacna crocea* across the Indo-Malay Archipelago. Mar Ecol. 2016;38(1):e12389.
9. Berger C, Štambuk A, Maguire I, Weiss S, Füreder L. Integrating genetics and morphometrics in species conservation—A case study on the stone crayfish, *Austropotamobius torrentium*. Limnologica. 2017;69:28-38.
10. Faulks LK, Kerezszy A, Unmack PJ, Johnson JB, Hughes JM (2017). Going, going, gone? Loss of genetic diversity in two critically endangered Australian freshwater fishes, *Scaturiginichthys vermeilipinnis* and *Chlamydogobius squamigenus*, from Great Artesian Basin springs at Edgbaston, Queensland, Australia. Aquat Conserv. 2017;27(1):39-50.
11. Richmond J Q, Wood DA, Westphal MF, Vandergast AG, Leaché AD, Saslaw LR, Butterfield HS, Fisher, RN. Persistence of historical population structure in an endangered

- species despite near-complete biome conversion in California's San Joaquin Desert. *Mol Ecol.* 2017;26(14):3618-35.
12. Hostim-Silva M, Bertoncini AA, Borgonha M, Leite JR, Freitas MO, Daros FA, Bueno LS, Farro CAP, Koenig, C. C. The Atlantic Goliath Grouper: Conservation Strategies for a Critically Endangered Species in Brazil. In: Rossi-Santos M, Finkl C, editors. *Advances in Marine Vertebrate Research in Latin America*. Vol 22. Springer, Cham: Coastal Research Library; 2018. p. 367-405.
 13. Rodriguez-Robles JA, Jezkova T, Garcia MA. Evolutionary relationships and historical biogeography of *Anolis desechensis* and *A. monensis*, two lizards endemic to small islands in the eastern Caribbean Sea. *J. Biogeogr.* 2007; 34:1546-58.
 14. Crnobrnja-Isailović J, Haxhiu I. *Vipera ammodytes*. In: Gasc JP, Cabela A, Crnobrnja-Isailovic J, Dolmen D, Grosenbacher K, Haffner P, Lescure J, Martens H, Martínez Rica JP, Maurin H, Oliveira ME, Sofianidou TS, Veith M, Zuidervijk A, editors. *Atlas of Amphibians and Reptiles in Europe*. Paris: Societas Europaea Herpetologica and Museum National d' Histoire Naturelle; 1997. P. 384-5.
 15. Ursenbacher S, Schweiger S, Tomović Lj, Crnobrnja-Isailović J, Fumagalli L, Mayer W. Molecular phylogeography of the nose-horned viper (*Vipera ammodytes*, Linnaeus (1758)): evidence for high genetic diversity and multiple refugia in the Balkan peninsula. *Mol Phylogenet Evol.* 2008;46(3):1116-28.
 16. Agasyan A, Avci A, Tuniyev B, Crnobrnja Isailović J, Lymberakis P, André H, Cogalniceanu D, Wilkinson J, Üzümlü N, Orlov N, Podloucky R., Tuniyev S, Kaya U, Sindaco R, Böhme W, Ajtici R, Ugurtas IH, Sevinç M, Tomović Lj, Crochet PA, Haxhiu I, Joger U, Sterijovski B, Jelić D. *Vipera ammodytes*. The IUCN Red List of Threatened Species. [Internet]. 2009 [cited 2018 Oct 20]. Available from: <http://dx.doi.org/10.2305/IUCN.UK.2009.RLTS.T62255A12584303.en.e.T62255A12584303>.
 17. Tuniyev B, Nilson G, Kasaka Y, Avci A, Agasyan A, Orlov NL, Tuniyev S. *Vipera transcaucasiana*. The IUCN Red List of Threatened Species. [Internet]. 2009 [cited 2018 Oct 20]. Available from: <http://dx.doi.org/10.2305/IUCN.UK.2009.RLTS.T164708A5919515.en.e.T164708A114549008>.
 18. Golay P, Smith H, Broadley D, Dixon J, McCarthy C, Rage J-C, Schatti B, Toriba M. *Endoglyphs and other major venomous snakes of the World*. 1st ed. Geneva: Azemiops SA Herpetological Data Center; 1993.
 19. Tomović Lj. Systematics of the nose-horned viper (*Vipera ammodytes*, Linnaeus, 1758). *Herpetol J.* 2006;16(2):191-201.
 20. Vences M, Thomas M, van der Meijden A, Chiari Y, Vieites DR. Comparative performance of the 16S rRNA gene in DNA barcoding of amphibians. *Front Zool.* 2005;2:5.
 21. Ilić M, Stamenković G, Nikolić V, Marković V, Marinković N, Paunović M, Crnobrnja-Isailović, J. Identification of syntopic anuran species in early tadpole stages: correspondence between morphometric and genetic data. *Appl Ecol Environ Res.* 2016;14(2):381-97.
 22. Thompson JD, Higgins DG, Gibson TJ. Clustal W. improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 1994;22(22):4673-80.
 23. Ronquist F, Huelsenback JP. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics.* 2003;19:1572-4.
 24. Kimura M. Estimation of evolutionary distances between homologous nucleotide sequences. *Proc Natl Acad Sci U S A.* 1981;78:454-8.
 25. Darriba D, Taboada GL, Doallo R, Posada D. jModelTest 2: more models, new heuristics and parallel computing. *Nat Methods.* 2012;9(8):772.
 26. Tomović Lj. *Sistematika i biogeografija poskoka Vipera ammodytes* (Linnaeus, 1758) (Viperidae, Serpentes). [dissertation]. [Belgrade]: Biološki fakultet, Univerzitet u Beogradu. 2005.
 27. Yilmaz PO, Norton I, Leary D, Chuchla RJ. Tectonic evolution and paleogeography of Europe. In: Ziegler P.A, Horvath F, editors. *Pery-Tethys memoir 2: structure and prospects of Alpine Basins and Forelands*. Paris: Memoires du Museum National d'Histoire Naturelle; 1996. p. 47-60.
 28. Sotiropoulos K, Eleftherakos K, Džukić G, Kalezić ML, Legakis A, Polymeni RM. Phylogeny and Biogeography of the alpine newt *Mesotriton alpestris* (Salamandridae, Caudata), inferred from mtDNA sequences. *Mol. Phylogent. Evol.* 2007;45:211-26.
 29. Babik W, Branicki W, Crnobrnja-Isailović J, Cogalniceanu D, Sas I, Olgun K, Poyarkov NA, Garcia-Paris M, Arntzen JW. Phylogeography of two European newt species-discordance between mtDNA and morphology. *Mol. Ecol.* 2005;14:2475-91.
 30. Krystufek B, Buzan EV, Hutchinson WF, Hnafling, B. Phylogeography of the rare Balkan endemic Martino's vole, *Dinaromys bogdanovi*, reveals strong differentiation within the western Balkan Peninsula. *Mol Ecol.* 2007;16(6):1221-32.
 31. Gibbs HL, Corey SJ, Blouin-Demers G, Prior KA, Weatherhead PJ. Hybridization between mtDNA - defined phylogeographic lineages of black ratsnakes (*Pantherophis* sp.). *Mol Ecol.* 2006;15(12):3755-67.
 32. Paquin MM, Wylie GD, Routman EJ. Population structure of the giant garter snake, *Thamnophis gigas*. *Conserv Genet.* 2006;7(1):25-36.
 33. Dubey S, Brown GP, Madsen T, Shine R. Male-biased dispersal in a tropical Australian snake (*Stegonotus cucullatus*, Colubridae). *Mol Ecol.* 2008;17(15):3506-14.
 34. Lukoschek V, Waycott M, Keogh JS. Relative information content of polymorphic microsatellites and mitochondrial DNA for inferring dispersal and population genetic structure in the olive sea snake, *Aipysurus laevis*. *Mol Ecol.* 2008;17(13):3062-77.
 35. Ajtici R. Nose-horned viper (*Vipera ammodytes*) conservation problems in Serbia. *Protection of Nature.* 2009;60:319-26.
 36. Allendorf FW, England PR, Luikart G, Ritchie PA, Ryman N. Genetic effects of harvest on wild animal populations. *Trends Ecol Evo.* 2008;23(6):327-37.

Supplementary Data

Supplementary Table S1.

Available at: http://serbiosoc.org.rs/NewUploads/Uploads/Cu-bric%20et%20al_3548_Supplementary%20Table%20S1.pdf