

Relaxed Sugar Donor Selectivity of a *Sinorhizobium meliloti* Ortholog of the *Rhizobium leguminosarum* Mannosyl Transferase LpcC

ROLE OF THE LIPOPOLYSACCHARIDE CORE IN SYMBIOSIS OF RHIZOBIACEAE WITH PLANTS*

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The *lpcC* gene of *Rhizobium leguminosarum* and the *lpsB* gene of *Sinorhizobium meliloti* encode protein orthologs that are 58% identical over their entire lengths of about 350 amino acid residues. LpcC and LpsB are required for symbiosis with pea and *Medicago* plants, respectively. *S. meliloti* *lpsB* complements a mutant of *R. leguminosarum* defective in *lpcC*, but the converse does not occur. LpcC encodes a highly selective mannosyl transferase that utilizes GDP-mannose to glycosylate the inner 3-deoxy-D-manno-octulosonic acid (Kdo) residue of the lipopolysaccharide precursor Kdo₂-lipid IV_A. We now demonstrate that LpsB can also efficiently mannosylate the same acceptor substrate as does LpcC. Unexpectedly, however, the sugar nucleotide selectivity of LpsB is greatly relaxed compared with that of LpcC. Membranes of the wild-type *S. meliloti* strain 2011 catalyze the glycosylation of Kdo₂-[4'-³²P]lipid IV_A at comparable rates using a diverse set of sugar nucleotides, including GDP-mannose, ADP-mannose, UDP-glucose, and ADP-glucose. This complex pattern of glycosylation is due entirely to LpsB, since membranes of the *S. meliloti* *lpsB* mutant 6963 do not glycosylate Kdo₂-[4'-³²P]lipid IV_A in the presence of any of these sugar nucleotides. Expression of *lpsB* in *E. coli* using a T7lac promoter-driven construct results in the appearance of similar multiple glycosyl transferase activities seen in *S. meliloti* 2011 membranes. Constructs expressing *lpcC* display only mannosyl transferase activity. We conclude that LpsB, despite its high degree of similarity to LpcC, is a much more versatile glycosyltransferase, probably accounting for the inability of *lpcC* to complement *S. meliloti* *lpsB* mutants. Our findings have important implications for the regulation of core glycosylation in *S. meliloti* and other bacteria containing LpcC orthologs.

As shown in the preceding paper (1), LpcC of *Rhizobium leguminosarum* is a highly selective mannosyl transferase that

utilizes the sugar nucleotide GDP-mannose for the enzymatic glycosylation of the acceptor Kdo₂-lipid IV_A. These findings are consistent with earlier structural studies by Carlson and co-workers (2, 3) demonstrating that mannose is the sole sugar attached to the inner Kdo moiety of the core oligosaccharide of *R. leguminosarum* or *Rhizobium etli* lipopolysaccharide (LPS)¹ (see Fig. 1 of the preceding paper). Furthermore, mutants of *R. leguminosarum* lacking *lpcC* are deficient in mannosyl transferase activity, are unable to form functional root nodules on *Pisum sativum*, and synthesize a truncated LPS molecule lacking the usual core sugar residues beyond the Kdo disaccharide (4).

Detailed structural studies of the LPS core of *Sinorhizobium meliloti* have not yet been reported.² The *S. meliloti* core is complex, and it may contain glucose and galactose residues (5).² The recent sequencing of the *S. meliloti* genome (6) and independent genetic studies by Lagares *et al.* (7) have demonstrated that a gene encoding an ortholog of LpcC is present in *S. meliloti*. Mutant 6963, which harbors a Tn5 insertion in this gene (designated *lpsB*), is defective in symbiosis with *Medicago* spp. (8, 9). Initially, mutant 6963 derived from strain 2011 was shown to be delayed in nodulation and poorly competitive for nodule occupancy on alfalfa (*Medicago sativa*) (8). In later studies, the same mutant was found to be unable to fix nitrogen (Fix⁻) on *Medicago truncatula* (9). Clear signs of plant defense were also observed with an abnormal pattern of infection as judged by electron microscopy (9). More recently, independent *lpsB* mutants generated with the related strain *S. meliloti* 1021 were reported to be Fix⁻ in alfalfa. However, since 2011 and 1021 are both streptomycin-resistant derivatives of the wild type strain SU47, it may be that the defective nitrogen fixation phenotype seen in alfalfa with the *lpsB* mutant of strain 1021 is the consequence of both the *lpsB* lesion and some other as yet unidentified mutation in 1021 that is absent in strain 2011. Whatever the explanation, it is apparent that mutants defective in *lpsB* provide an interesting model system with which to probe the participation of LPS in symbiosis.

As shown in Fig. 1, *S. meliloti* LpsB is 351 amino acid residues long and displays 58% identity and 72% similarity to LpcC over the full length of the entire protein. Accordingly, LpsB, like *R. leguminosarum* LpcC, would be expected to func-

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¹ The abbreviations used are: LPS, lipopolysaccharide; Kdo, 3-deoxy-D-manno-octulosonic acid; MALDI-TOF, matrix-assisted laser desorption/ionization time-of-flight.

² R. Carlson, personal communication.

LpcC:	1	MPDIRDVEIIAPNFKRRLSGVSTSTIVQLIPCQIRLGIKATILGFLPEDLFLKRWQLLG 60
		M.DIRDVE+IAPNFK+RLSGVSTSTI+QL+P.Q...LGKIA.LGFLP+L.LP.+++R L+.
LpsB:	1	MVDIRDVEVIAPNFKRQLSGVSTSTIIVLQVQVQALGKQIAVLGFLGPKLSLPSVFRFLDIH 60
LpcC:	61	LWRPPARRRRRVRHARRNNEMAVGILLRHLTRMPLKLLFTSAQRRTTAYTKWLIRRMDA 120
		LW+.P...R..RVWHARRN.EM...ILLR.L.RM.L+++FTSA+QRRHT.++K+LIRRMDA
LpsB:	61	LWKRPEGRPCRVHARRNEMVLAIFALLRDLRMLKRLIVFTSASQRRHTGSKFLIRRMDA 120
LpcC:	121	VIATSDRSRGSFLEVPHTVIQHGVDLALFHPPEAAEDGIAATGL-PGRHLVGCGRVRRHQK 179
		VIATSR.R++L+VP+TVI.HG+D...F.PP+...A.GL.P...+VGCGRVRRHQK
LpsB:	121	VIATSGRTAAALDVPNTVILEGIDTRKRFPPFDKTEAKKALGLDPAKFFVGCGRVRRHQK 180
LpcC:	180	GTDLFLVRAMITELLPQHTEWAVTSVGRVTAEHVAFADKXADVVAAGLSDRILFLGEVVDI 239
		GTDLFLV.+MI.LLP...+W.A+V+GR.T..B+AF...+LK...V..AGL+DRILF+GE...+I
LpsB:	181	GTDLFDVMSIALPCCRDPGWAIVAGRATGPHLAFSESELKERVAKAGLADRILFVGEHNTI 240
LpcC:	240	KIWRRLTLYVAPSRNEGFLTPLEAMASRTAVVADAGAYAEILVTG--ETGSVVAASD 297
		..WYR.L.L+VAF.R.EGFLTPLEAMA+...VVA+D.GA++EL+..G...ETG.++AA.D
LpsB:	241	PDWYRALDLFVAPQRWEGFLTPLEAMATGVVAVTDVGFSELVGTGGSEETGLIIAADD 300
LpcC:	298	GEALTRAIAPYIADPALAVARGENALRHVRANFALEREASATGAVYNSLL 347
		..+A+..A.A++..D....A...N.L.....NFA+E+EA.AL.VY..SL+
LpsB:	301	LKAMVDAAAAFMDRPRLAASANGRLARTSKNFATIEQEARATAAVYESLM 350

FIG. 1. Sequence similarity comparison of *R. leguminosarum* LpcC and *S. meliloti* LpsB. The middle line indicates identities and similarities. The proteins show 58% identity and 72% similarity over 350 of the 352 amino acid residues in LpcC. The *E* value (28) determined against the nonredundant data base is $\sim 10^{-103}$.

tion as a mannosyl transferase. The *lpsB* gene is, in fact, able to complement the truncation of lipopolysaccharide assembly and the nodulation deficiency seen in strain RSKnH, an *lpcC* mutant of *R. leguminosarum* (7). However, *lpcC* is not able to restore normal lipopolysaccharide assembly to the *S. meliloti* *lpsB* mutant 6963 (7).

To understand the biochemical basis for this genetic anomaly, we have now characterized the glycosyl transferase activity of the parental strain *S. meliloti* 2011 and its LPS core biosynthesis/nodulation-deficient mutant 6963. We have also cloned the *lpsB* gene behind the *T7lac* promoter and expressed it in *Escherichia coli*. The results show conclusively that LpsB, like LpcC, can function as an effective mannosyl transferase using GDP-mannose as the donor and Kdo₂-[4'-³²P]lipid IV_A as the acceptor substrate. However, unlike LpcC, LpsB shows comparable activity with several other sugar nucleotides, including ADP-glucose and UDP-glucose. These additional activities of LpsB may account for the observation that *lpcC* cannot substitute for *lpsB* in cells of *S. meliloti* (7). Our findings further suggest that the most important functions of LpsB for *S. meliloti* biology are those related to the utilization of the sugar nucleotides other than GDP-mannose. How the LpsB protein acquires its relaxed sugar nucleotide selectivity when compared with LpcC can now be studied at the level of enzymology and structure.

EXPERIMENTAL PROCEDURES

Chemicals and Materials—Sugar nucleotides, Kdo, and HEPES were obtained from Sigma. Bicinchoninic assay reagents and Triton X-100 were purchased from Pierce. Yeast extract and peptone-tryptone were from Difco. Silica Gel 60 thin layer chromatography plates were obtained from Merck. Reagent grade pyridine, chloroform, and methanol were purchased from VWR. [γ -³²P]ATP was purchased from PerkinElmer Life Sciences.

Bacterial Strains, Media, and Growth Conditions—The bacterial strains and plasmids used in this study are listed in Table I. *S. meliloti* strains have been described previously (7). *S. meliloti* strains were

grown in TY medium (10 g of NaCl, 10 g of peptone-tryptone, and 5 g of yeast extract per liter, supplemented with 10 mM calcium chloride) at 30 °C. When necessary, the cultures were also supplemented with streptomycin (400 μ g/ml) and/or neomycin (30 μ g/ml). *E. coli* strain BLR(DE3)/pLysS (Novagen) was grown in LB broth (10) at 37 °C.

Molecular Biology Techniques—Plasmids were prepared using the Qiagen Mini Prep Kit (Qiagen). Restriction endonucleases, *Pfu* polymerase (Stratagene), shrimp alkaline phosphatase, and T4 ligase (Invitrogen) were used according to the manufacturers' instructions. Competent cells of *E. coli* were prepared for transformation using the calcium chloride method (11, 12). Introduction of plasmids into *S. meliloti* was carried out by conjugation as previously described (7).

Construction of a Plasmid Expressing LpsB behind the T7lac Promoter—PCR primers were designed to retrieve the *S. meliloti* *lpsB* gene from plasmid pAL100 (Table I) (7) and to introduce *NdeI* and *BamHI* restriction sites at the ends of the PCR product, as indicated. The primers used to obtain *lpsB* have the following sequences: forward, 5'-GAATTCATATGGTTCGATATCCGCGACGTC-3'; reverse, 5'-GAA-TTCGGATCCTCAACGCATCAGGCTTTTCGTAAAC-3'. The conditions for the PCR were as follows: 45 s of denaturation at 98 °C and then 25 cycles of denaturation (45 s, 98 °C), annealing (45 s, 55 °C), and extension (1.5 min, 72 °C). After the last cycle, an additional 10-min extension at 72 °C was performed. The resulting 1056-base pair DNA product was digested with the *NdeI* and *BamHI* restriction enzymes and ligated into an expression vector, pET28b, that had been digested with the same enzymes. The resulting ligation mixture was used to transform *E. coli* XL1 Blue, selecting for kanamycin resistance (30 μ g/ml). The presence of the proper inserts in plasmids from several colonies was verified by restriction mapping. The desired construct, designated pMKRM, was then transferred into *E. coli* BLR(DE3)/pLysS for high level heterologous expression of *lpsB*.

Preparation of Membranes for Assay of LpcC and LpsB—Each *S. meliloti* strain shown in Table I was grown from a single colony in 1 liter of TY medium containing the appropriate antibiotics to an optical density of 600 = 1.4 (late log phase). *E. coli* BLR(DE3)/pLysS/pMKRM was grown from a single colony in 1 liter of LB medium containing 30 μ g/ml chloramphenicol and 30 μ g/ml kanamycin at 37 °C until the A₆₀₀ reached ~ 0.5 . The culture was induced with 1 mM isopropyl-1-thio- β -D-galactopyranoside and incubated with shaking for an additional 3 h at 30 °C.

Cells from the above 1-liter cultures were harvested at 4 °C by centrifugation at 4,000 \times g for 10 min, were each resuspended in 30 ml of 50 mM HEPES, pH 7.5, and were broken by one passage through a French pressure cell at 18,000 p.s.i. Debris and inclusion bodies were removed by centrifugation at 4,000 \times g for 15 min. Membranes were prepared by ultracentrifugation at 100,000 \times g for 60 min. The membranes were resuspended in 20 ml of the same buffer, and the centrifugation was repeated a second time at 100,000 \times g to remove any remaining cytosol. Protein concentration was determined by the bicinchoninic method (13) (Pierce). The membrane pellet was resuspended at 10–15 mg of protein/ml in 50 mM HEPES, pH 7.5, and stored at -80 °C until further use.

Mannosyl Transferase Assays—The mannosyl transferases LpcC and LpsB were assayed using the lipopolysaccharide precursor Kdo₂-[4'-³²P]lipid IV_A, which was synthesized, isolated, and stored as a frozen aqueous dispersion, as described previously (14). Prior to each use, the radiolabeled substrate was subjected to ultrasonic irradiation in a water bath for 1 min. The mannosyl transferase was assayed as follows. The reaction mixture (10- μ l final volume) contained 50 mM HEPES, pH 7.5, 0.1% Triton X-100, 1 mM GDP-mannose, and 10 μ M Kdo₂-[4'-³²P]lipid IV_A (3000–6000 cpm/nmol). The reaction was started by the addition of an appropriate amount of enzyme (usually at a final concentration of 1.0 mg/ml *S. meliloti* membranes or 0.02 mg/ml washed membranes of *E. coli* cells overexpressing *lpsB* behind the *T7lac* promoter) and incubated for the indicated times at 30 °C. The reactions were stopped by spotting 4- μ l samples directly onto a silica gel thin layer plate. After drying the spots at room temperature, the plate was developed in the solvent chloroform, pyridine, 88% formic acid, water (30:70:16:10, v/v/v). Following removal of the solvent with a hot air stream, the plate was analyzed using an Amersham Biosciences PhosphorImager (Storm 840), equipped with ImageQuant software. Other glycosyl transferase activities of LpsB were assayed in the same way but in the presence of the indicated alternative sugar nucleotides present at 1 mM.

Purification of LPS from *S. meliloti* *lpsB* Mutant 6963—LPS (*i.e.* Kdo₂-lipid A) was extracted from a 100-ml culture of late log phase cells (OD = 1.4) with a single phase Bligh-Dyer mixture, consisting of chloroform/methanol/water (1:2:0.8, v/v/v). Following phase partitioning,

TABLE I
Sinorhizobium meliloti strains and plasmids used in this study

Bacterial strains	Genotype	LPS properties	Source or reference
<i>S. meliloti</i> 1021	Parental wild type	Full-length	Ref. 25
<i>S. meliloti</i> 2011	Parental wild type	Full-length	Ref. 26
<i>S. meliloti</i> 6963	lpsB::Tn5	Truncated inner core	Ref. 8
<i>S. meliloti</i> 6963-B	<i>S. meliloti</i> 6963 (pJB1psSme)	Full-length	This work
<i>S. meliloti</i> 6963-C	<i>S. meliloti</i> 6963 (pPN120)	Truncated as in 6963	This work
Plasmids	Relevant characteristics		
pJB3Tc19	Derivative of the minimal replicon of plasmid RK2, Ap ^r , Tc ^r		Ref. 27
pAL100	pACYC184-Gm- <i>mob</i> carrying in <i>DraI</i> a 5.5-kb <i>SstI</i> DNA fragment from <i>S. meliloti</i> 2011 harboring <i>lpsB</i>		Ref. 8
pJB1psSme	pJB3Tc19 carrying a 4.9-kb <i>SstI</i> - <i>HindIII</i> fragment with the region <i>greA-lpsB-lpsE-lpsD</i> from <i>S. meliloti</i> 2011 cloned into the polylinker of the vector between <i>EcoRI</i> - <i>HindIII</i> , Km ^r		Ref. 7
pPN120	pLAFR-1 with a 4.4 kb <i>EcoRI</i> fragment containing <i>greA</i> , <i>lpcC</i> , <i>dctA</i> , and the 5' region of <i>dctB</i> from <i>R. leguminosarum</i> bv. <i>viciae</i> , Tc ^r		Refs. 4 and 7

the LPS was purified by chromatography on a DEAE-cellulose column, prepared in chloroform/methanol/water (2:3:1, v/v/v) (1).

Mass Spectrometry—Spectra were acquired in the negative mode using a matrix-assisted laser desorption/ionization time of flight (MALDI-TOF) Kompact 4 mass spectrometer (Kratos Analytical, Manchester, UK), equipped with a 337-nm nitrogen laser and set at a 20-kV extraction voltage. Conditions were the same as described in the preceding paper (1).

RESULTS

Complex Glycosylation of Kdo₂-lipid IV_A in Membranes of *S. meliloti*—As shown in Fig. 2A (lane 5), membranes of *R. leguminosarum* 3841 (15) glycosylate Kdo₂-[4'-³²P]lipid IV_A in a highly selective manner using GDP-mannose as the preferred donor substrate. The nonphysiological analog ADP-mannose, a natural product found in corn (16, 17), can substitute for GDP-mannose (18), albeit not quite as effectively (Fig. 2A, lane 4). Other core sugars cannot be incorporated prior to the addition of the first mannose residue to Kdo₂-lipid IV_A (data not shown) (4, 19). The mannosyl transferase activity of *R. leguminosarum* is catalyzed by LpcC, which displays very little transferase activity with other sugar nucleotide donors, such as ADP-glucose or UDP-glucose (Fig. 2A, lanes 3 and 9, respectively) (1, 4).

In earlier studies with *S. meliloti* 1021 (18), little or no mannosyl transferase activity was detected in cell extracts or membranes prepared from cells grown on TY medium. *S. meliloti* 1021 cells were therefore used for the expression cloning and characterization of *lpcC* from *R. leguminosarum* (4). However, when grown in a more controlled manner to late log phase, as described above, membranes of *S. meliloti* 1021 do in fact catalyze rapid mannosylation of Kdo₂-[4'-³²P]lipid IV_A (Fig. 2B, lanes 12 and 13). Interestingly, other glycosylations of Kdo₂-[4'-³²P]lipid IV_A are observed with membranes of *S. meliloti* 1021 (Fig. 2B), which are absent in *R. leguminosarum* 3841 (Fig. 2A), as judged by the shift of the radioactive substrate band to more slowly migrating species with ADP-glucose, UDP-galactose, and UDP-glucose (Fig. 2B, lanes 11, 14, and 17, respectively). There is even the formation of some secondary glycosylation products of Kdo₂-[4'-³²P]lipid IV_A with ADP-glucose, UDP-galactose, and UDP-glucose (Fig. 2B, lanes 11, 14, and 17), which are not seen when ADP-mannose or GDP-mannose are employed as the donor substrates (Fig. 2B, lanes 12 and 13). Under matched conditions, ADP-glucose, UDP-galactose, and UDP-glucose do not support any glycosylation of Kdo₂-[4'-³²P]lipid IV_A in membranes of *R. leguminosarum* 3841 (Fig. 2A) or *R. etli* CE3 (data not shown). Neither *R. leguminosarum* nor *S. meliloti* 1021 membranes catalyze efficient glycosylation of Kdo₂-[4'-³²P]lipid IV_A with UDP-GalUA or UDP-GlcNAc as sugar donors (Fig. 2, A (lanes 7 and 8) and B (lanes 15 and 16)). The formation of inorganic phosphate by membranes of *R. leguminosarum* is

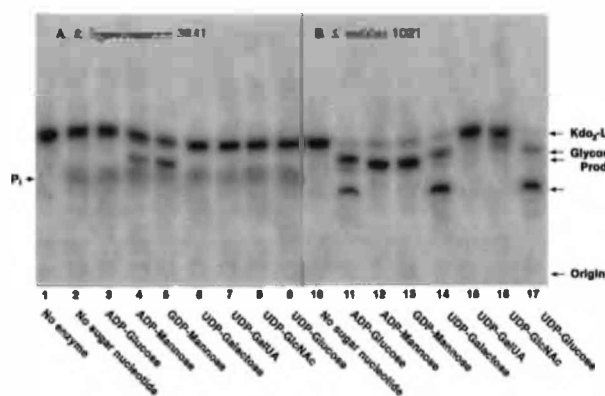


FIG. 2. Distinct patterns of Kdo₂-lipid IV_A glycosylation with membranes of *R. leguminosarum* 3841 versus *S. meliloti* 1021. Reactions were carried out under standard conditions at pH 7.5 using a 1 mM concentration of each of the sugar nucleotides, as indicated. Membranes were used at 1 mg/ml. The incubation was carried out at 30 °C for 60 min.

attributed to the 4'-phosphatase activity, which is absent in *S. meliloti* (Fig. 2, A versus B) (14, 20).

The completed genome of *S. meliloti* 1021 (6) and intriguing recent studies of the isogenic *S. meliloti* 2011 (7) have revealed the presence of a significant ortholog of LpcC (58% identity and 72% similarity over the full length of the protein) in both of these bacterial strains (Fig. 1). In the case of strain *S. meliloti* 2011, this LpcC ortholog (which was termed LpsB) (7) is required both for core oligosaccharide assembly and for symbiosis with *Medicago* spp., as judged by the analysis of mutant 6963, which contains a Tn5 insertion in *lpsB*. We therefore decided to examine the glycosylation of Kdo₂-[4'-³²P]lipid IV_A using membranes of *S. meliloti* 2011 and mutant 6963. When membranes of *S. meliloti* 2011 are assayed at 1 mg/ml for their ability to glycosylate Kdo₂-[4'-³²P]lipid IV_A, essentially the same complex pattern of sugar additions is observed (Fig. 3) as with *S. meliloti* 1021 membranes (Fig. 2B).

Absence of Kdo₂-Lipid IV_A Glycosylation in Membranes of the *lpsB* Mutant 6963—When comparable washed membranes of *S. meliloti* mutant 6963 are assayed, no glycosylation of Kdo₂-[4'-³²P]lipid IV_A of any kind is observed (data not shown). This striking result suggests that the diverse glycosylations seen in Figs. 2B and 3 are all the result of a single enzyme with relaxed sugar nucleotide substrate selectivity, when contrasted with LpcC. Based on this finding alone, however, some kind of pleiotropic regulatory effect secondary to the Tn5 insertion in the *lpsB* gene of mutant 6963 cannot be excluded.

Restoration of Mannosyl Transferase Activity in Mutant 6963 by DNA Fragments Harboring either *lpsB* or *lpcC*—We have

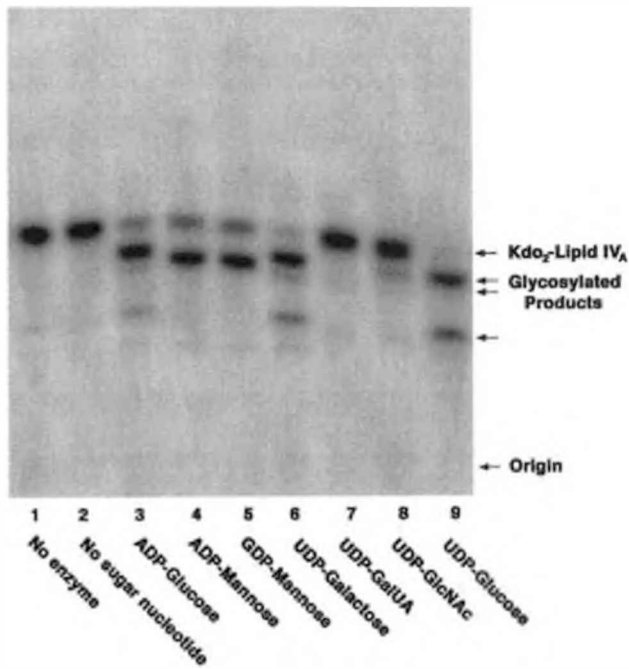


FIG. 3. Glycosylation of Kdo₂-lipid IV_A catalyzed by membranes of *S. meliloti* 2011. Reactions were carried out under standard conditions at pH 7.5 using a 1 mM concentration of each of the sugar nucleotides, as indicated. Membranes were used at 1 mg/ml. The incubation was carried out at 30 °C for 60 min.

previously described the plasmids pJBlpsSme, which contains *lpsB* on a 4.9-kb fragment of *S. meliloti* 2011 DNA, and pPN120, which harbors *lpcC* on a 4.4-kb fragment of *R. leguminosarum* DNA (7). As shown in Fig. 4, introduction of either of these plasmids into mutant 6963 (lanes 4 and 5) restores full mannosyl transferase activity, as judged by assay of cell membranes with Kdo₂-[4'-³²P]lipid IV_A in the presence of GDP-mannose. In fact, all of the other glycosylations of Kdo₂-[4'-³²P]lipid IV_A are also restored to mutant 6963 by plasmid pJBlpsSme (Fig. 5). In contrast, when membranes from mutant 6963 cells harboring the *lpcC* containing plasmid pPN120 are assayed with the other sugar nucleotide donors (Fig. 6), no activity is seen with ADP-glucose, UDP-galactose, or UDP-glucose, consistent with the substrate specificity of native *R. leguminosarum* membranes (Fig. 2A) and of purified *LpcC* (1). These results (Fig. 6) exclude the possibility that additional enzymes present in crude extracts of *S. meliloti* 2011 are interconverting all of the various sugar nucleotides added to the assay system to account for the diverse glycosylations seen in Figs. 2B, 3, and 5.

Heterologous Expression and Substrate Specificity of *LpsB* in *E. coli*—The *lpsB* gene of *S. meliloti* 2011 was cloned behind the T7lac promoter of the expression plasmid pET28b to generate pMKRM and transformed into the *E. coli* strain BLR(DE3)/pLysS. Washed membranes were then assayed from cells that had been induced in late log phase with isopropyl-1-thio-β-D-galactopyranoside for 3 h. As shown in Fig. 7A (upper panel), membranes of cells harboring the vector control (at 0.02 mg/ml) catalyze little or no glycosylation of Kdo₂-[4'-³²P]lipid IV_A, with the possible exception of a small amount of activity seen with ADP-mannose, which might be attributed to the endogenous WaaC activity of the *E. coli* host (1, 21). In contrast, overexpression of *lpsB* behind the T7lac promoter results in robust glycosylation of Kdo₂-[4'-³²P]lipid IV_A (Fig. 7B, lower panel) with a similar broad range of sugar nucleotide substrate utilization as observed in membranes of *S. meliloti* 1021 or 2011 (Figs. 2B and 3). Again, ADP-mannose, GDP-mannose,

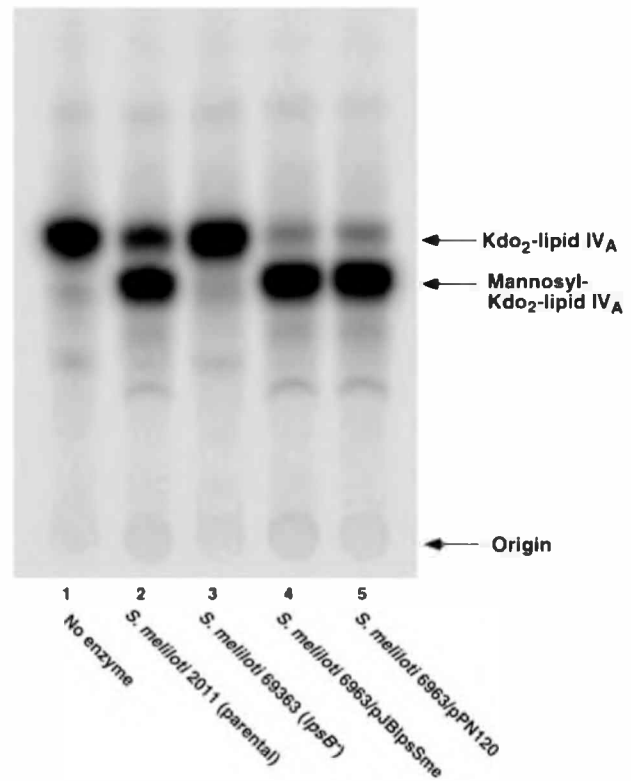


FIG. 4. Restoration of Kdo₂-lipid IV_A mannosylation in membranes of *lpsB* mutant 6963 expressing either *R. leguminosarum lpcC* or *S. meliloti lpsB*. Reactions were carried out under standard conditions at pH 7.5 using 1 mM GDP-mannose as the sugar donor. Membranes were used at 1 mg/ml. The incubation was carried out at 30 °C for 60 min. Lane 1, no enzyme; lane 2, *S. meliloti* 2011; lane 3, *S. meliloti* 6963; lane 4, *S. meliloti* 6963/pJBlpsSme (*lpsB*); lane 5, *S. meliloti* 6963/pPN120 (*lpcC*).

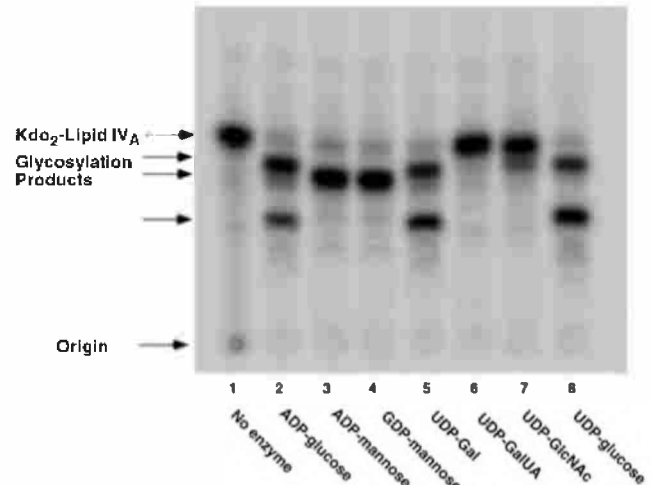


FIG. 5. Restoration of other Kdo₂-lipid IV_A glycosylation reactions in membranes of *lpsB* mutant 6963 expressing *S. meliloti lpsB*. Reactions were carried out under standard conditions at pH 7.5 using a 1 mM concentration of each of the sugar nucleotides, as indicated. Membranes were used at 1 mg/ml. The incubation was carried out at 30 °C for 60 min.

ADP-glucose, and UDP-glucose all function as efficient donor substrates and at comparable rates (Fig. 7B), whereas UDP-GalUA and UDP-GlcNAc are virtually inactive. No further secondary glycosylations are observed in this heterologous system (Fig. 7B versus Figs. 2B, 3, and 5), indicating that additional enzymes present only in *S. meliloti* must catalyze these reactions. Interestingly, UDP-galactose is not used very effec-

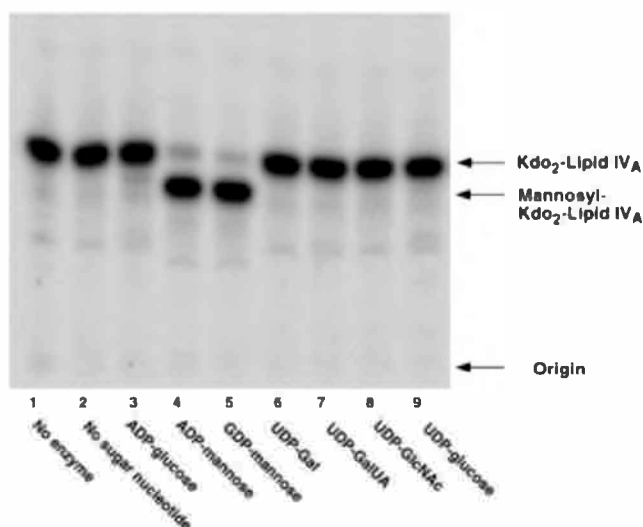


FIG. 6. No Kdo₂-lipid IV_A glycosylation with sugars other than mannose in membranes of *lpsB* mutant 6963 expressing *R. leguminosarum* *lpcC*. Reactions were carried out under standard conditions at pH 7.5 using a 1 mM concentration of each of the sugar nucleotides, as indicated. Membranes were used as the enzyme source at 1 mg/ml. The incubation was carried out at 30 °C for 60 min.

tively as a sugar donor when LpsB is expressed in *E. coli* (Fig. 7B, lane 4). This finding suggests that the robust activity seen with UDP-galactose in *S. meliloti* membranes (Figs. 2B, 3, and 5) may be due to more effective 4'-epimerization of UDP-galactose to form UDP-glucose in *S. meliloti* membranes than in *E. coli* membranes.

The results of Fig. 7B establish unequivocally that *S. meliloti* LpsB can employ a much broader set of sugar nucleotides as donor substrates than can its ortholog LpcC of *R. leguminosarum*. These findings exclude pleiotropic regulatory effects as the explanation for the loss of all of the different glycosylation activities in membranes of mutant 6963. Supporting this result, we had previously suggested the absence of polar effects of the Tn5 insertion in 6963, since the open reading frames located immediately downstream of *lpsB* are transcribed in the opposite direction. Most importantly, our enzymatic data provide a straightforward biochemical rationale for the ability of *S. meliloti* *lpsB* to complement the *R. leguminosarum* *lpcC* mutation but not the reverse (7).

Mass Spectrometry of Truncated LPS Extracted from Mutant 6963—Chloroform/methanol extraction of the phospholipids and core-deficient LPS of mutant 6963, followed by chromatography on DEAE-cellulose, yields a purified preparation consisting of lipid A derivatized with two Kdo residues. As shown in the negative mode MALDI-TOF mass spectrum of this compound (Fig. 8A), the peak at m/z 2493.3 may be interpreted as the $[M - H]^-$ ion of a lipid A species that is acylated with two 3-OH-C14 chains, two 3-OH-C18 chains, and one secondary 29-OH-C30 moiety capped with a β -hydroxybutyrate residue (Fig. 8B). This pattern is consistent with the overall fatty acid composition of lipid A that was recently reported for lipid A of *Sinorhizobium* sp. NGR234 (22). Smaller peaks within the same cluster near m/z 2493.3 that differ by 14 atomic mass units might arise from the presence of shorter odd chain fatty acyl moieties and/or from shorter acyl chains in conjunction with partial *O*-methylation of the β -hydroxybutyrate residue in some molecular species (not shown) (22). The cluster of peaks around m/z 2392.3 might arise from one of the shorter chain lipid A species lacking the β -hydroxybutyrate group (or its methylated version) (22). The peaks around m/z 2052.6 are due to loss of the Kdo disaccharide from the parent compound. The

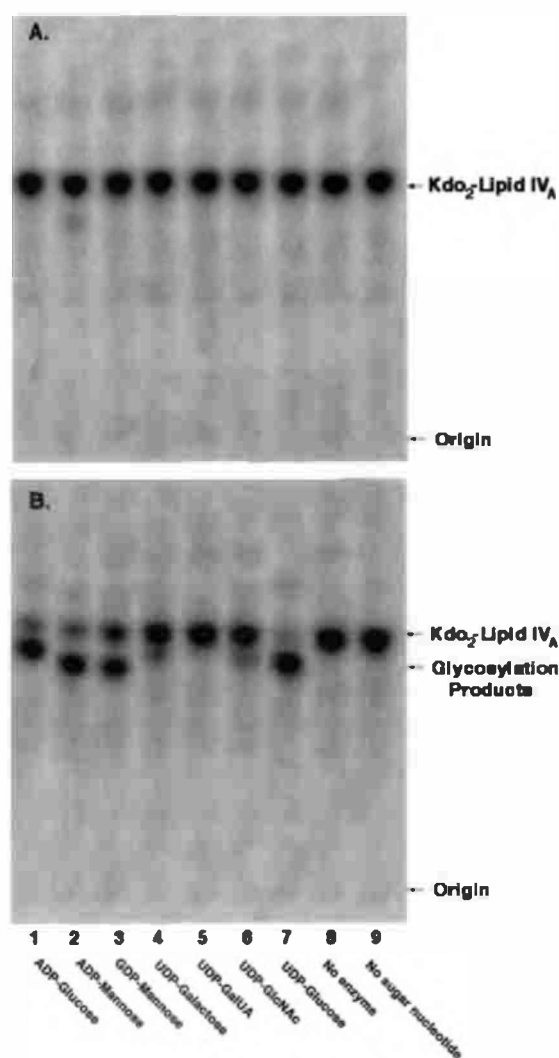


FIG. 7. Glycosylations of Kdo₂-lipid IV_A catalyzed by membranes of *E. coli* expressing *lpsB* behind the T7lac promoter. Reactions were carried out under standard conditions at pH 7.5 using a 1 mM concentration of each of the sugar nucleotides, as indicated. Enzyme was used at 0.02 mg/ml. The incubation was carried out at 30 °C for 60 min. A, membranes of BLR(DE3)/pLysS/pET28B; B, membranes of BLR(DE3)/pLysS/pMKRM.

peaks at m/z 2265.8, 1458.0, and 1003.2 could not be assigned and might be due to impurities.

The mass spectrum of the LPS recovered from mutant 6963 supports the idea that LpsB incorporates the first sugar of the inner core beyond Kdo in a major portion of the LPS molecules of *S. meliloti* and that further core glycosylation of these LPS molecules requires the presence of the sugar incorporated by LpsB (7).

DISCUSSION

R. leguminosarum LpcC (4) and *S. meliloti* LpsB (7) are protein orthologs that display 58% identity and 72% similarity over their entire lengths (Fig. 1). In other metabolic pathways, such as lipid A biosynthesis (23, 24), extensive similarity at this level is predictive of functional identity. Accordingly, LpsB of *S. meliloti* is classified as a mannosyl transferase in genomic databases (6), and it can indeed catalyze the mannosylation of Kdo₂-lipid IV_A *in vitro* (Fig. 7B). However, LpsB can also utilize several other sugar nucleotide substrates, namely ADP-glucose and UDP-glucose (Fig. 7B), which LpcC does not recognize effectively (Fig. 6) (1). We also know that whereas the *lpsB* gene can complement the *lpcC* mutation of *R. leguminosarum*,

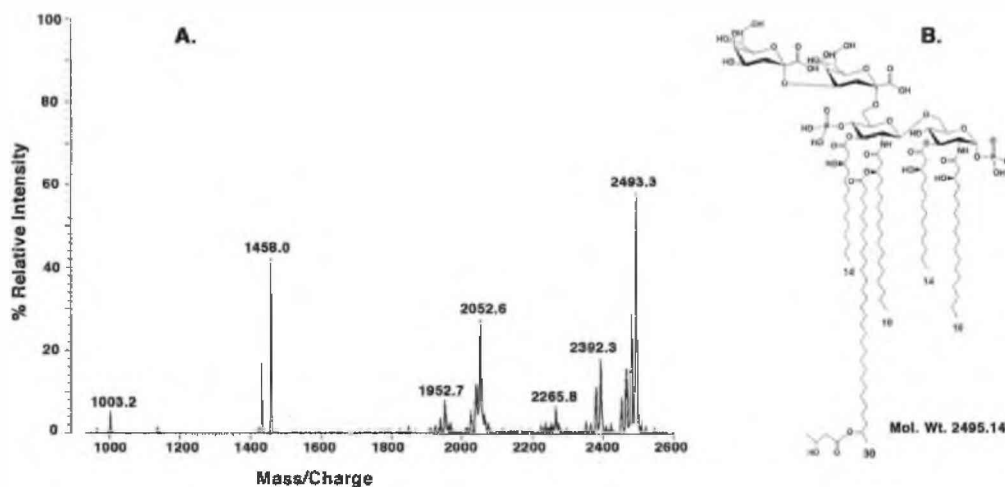


FIG. 8. Negative mode MALDI-TOF mass spectrum of LPS purified from mutant 6963 by chloroform/methanol extraction. The proposed LPS molecular species that could account for the major peaks observed in A is shown in B. This structure is consistent with the overall fatty acid composition recently reported for lipid A of *Sinorhizobium* sp. NGR234 (22).

lpcC cannot complement *lpsB*-deficient strains of *S. meliloti* (7). These findings, taken together, demonstrate that LpsB donor substrates other than GDP-mannose (Fig. 7B) (*i.e.* most likely UDP-glucose or ADP-glucose) are critical for proper LPS core assembly in *S. meliloti*.

The results shown in Fig. 2B suggest that LpsB-catalyzed mannose incorporation (lanes 12 and 13) may not generate a core biosynthetic intermediate suitable for further glycosylation, as is observed when ADP-glucose or UDP-glucose is added as the sugar donor for LpsB (lower migrating bands in Fig. 2B, lanes 11 and 17). Additional evidence for the idea that mannosylation may prevent further core assembly in *S. meliloti* comes from the observation that *lpcC* expressed in the *lpsB* mutant 6963 does not significantly alter the migration of the truncated LPS species that is present in this strain (7), whereas *lpcC* and *lpsB* both restore full-length O-antigen to *lpcC* mutants of *R. leguminosarum* (7). LpcC presumably incorporates mannose into the nascent LPS of the *S. meliloti* mutant 6963, but the resulting product is not a substrate for further glycosylation.

The distinctly different sugar donor selectivity of LpcC versus LpsB demonstrates the need to conduct biochemical and enzymatic studies in conjunction with sequence comparisons when proposing *in vivo* functions for putative glycosyl transferases. Although LpsB does in fact catalyze the incorporation of mannose into Kdo₂-lipid IV_A *in vitro*, the role of LpsB within cells of *S. meliloti* is most likely the transfer of glucose units to the core of nascent LPS. The results of the present study reveal the difficulties associated with the correct functional assignment and data base annotation of glycosyl transferases. Thousands of diverse glycosyl transferase orthologs for which biochemical selectivity and function cannot be assigned are present in both procaryotic and eucaryotic genomes. LpcC and LpsB are members of a very large family of glycosyl transferases, most of which are probably membrane proteins (4). In *E. coli*, RfaG (the first glucosyl transferase of the core domain) is very distantly related to the LpsB/LpcC family (24). Although an x-ray structure has not been reported for any member of this enzyme family, it is likely that LpcC and LpsB will show similar protein folds. The unusual sugar nucleotide promiscuity of LpsB versus LpcC could be explained by subtle differences in the shapes of their donor substrate binding sites. Structural information might be helpful in redesigning the selectivity of glycosyl transferases such as LpsB with potential applications in the field of large scale carbohydrate synthesis. In that context, it remains to be determined whether or not

LpsB actually glycosylates Kdo₂-lipid IV_A in the same position as does LpcC and generates the same anomeric configurations when using different sugar nucleotides as donor substrates.

A discrepancy between the results of Fig. 2B and our earlier studies (18) is the detection of significant mannosyl transferase activity in membranes of *S. meliloti* 1021. We previously reported that such preparations are inactive (18). It may be that the previously reported cultures were not harvested at the same time in late log phase. Furthermore, the cells were not grown in the presence of antibiotics in the previous studies (18, 19). Perhaps the mannosyl transferase is expressed in a regulated manner in response to growth phase or the composition of the medium. Whatever the explanation, the broad substrate selectivity of LpsB might allow for regulation of *S. meliloti* core sugar composition in response to changes in the relative intracellular concentrations of GDP-mannose versus UDP-glucose and/or ADP-glucose. The covalent structure of the *S. meliloti* core has not been reported. It appears to be complex² compared with the well characterized core of *R. leguminosarum* (2, 3). Peculiar characteristics of the *S. meliloti* core were also suggested by earlier immunological studies. For instance, in contrast to most other Gram-negative bacteria, the immunodominant LPS epitopes of *S. meliloti* LPS are not located in the O-antigen domain but are in the core region (8). Interestingly, these core-associated epitopes are missing in the symbiotically defective *lpsB* mutant 6963 (8). Additional structural and physiological studies clearly will be necessary to explore the role of the *S. meliloti* core during symbiosis with plants.

Like the *lpsB* mutants of *S. meliloti*, the *lpcC* mutants of *R. leguminosarum* show aberrant nodulation (4). The LPS in the *lpcC* mutant RSKnH is entirely devoid of any residual species containing O-antigen (7), and both the *lpcC* and the *lpsB* genes can restore O-antigen to RSKnH (7). Similarly, the *lpsB* mutant 6963 contains an altered, rapidly migrating LPS species, not present in the wild type, which we believe consists of Kdo₂-lipid A (7) (Fig. 8). This substance is soluble in chloroform/methanol mixtures, as described under "Experimental Procedures." However, 6963 differs from RSKnH in that it retains some LPS molecules that are still derivatized with an O-antigen-like polymer or some other kind of polysaccharide (7). This more hydrophilic material would not be extracted with chloroform/methanol under our conditions. These complexities could explain why the total LPS, which was extracted with phenol from the *lpsB* mutant described by Campbell *et al.* (5), contains additional sugars besides glucosamine and Kdo. Xy-

lose is reported by Campbell *et al.* (5) as a major component of the LPS in their *lpsB* mutant but is not present in our preparations (Fig. 8). As with the structure of the core domain of *S. meliloti*,³ the final resolution of these issues will require the development of better methods for LPS purification as well as direct structural analysis of intact material by mass spectrometry and NMR spectroscopy.

Many interesting questions remain regarding the relaxed substrate selectivity of LpsB versus LpcC. Is this phenomenon associated with other LpcC orthologs seen in organisms like *Agrobacterium tumefaciens*, *Brucella melitensis*, or *Francisella tularensis*?³ Is the mannosyl transferase activity of *S. meliloti* LpsB necessary under certain physiological circumstances when a truncated LPS core would be desirable? Could a mutant of LpsB be isolated that is unable to utilize GDP-mannose but that still functions normally with UDP- and ADP-glucose? Are the secondary glycosylation reactions that follow glucose incorporation (Fig. 2B, lanes 11 and 17) catalyzed by LpsC or LpsD (7)? The further biochemical and genetic analysis of this important class of glycosyl transferases should help to answer these questions and to advance our understanding of core assembly and function in the *Rhizobiaceae* and other Gram-negative bacteria. Molecular insights into the biochemistry of diverse LPS cores could prove useful in the development of new vaccines and in the improvement of agriculturally important crops.

REFERENCES

- Kanipes, M. I., Ribeiro, A. A., Lin, S., Cotter, R. J., and Raetz, C. R. H. (2003) *J. Biol. Chem.* **278**, 16356–16364
- Bhat, U. R., Krishnaiah, B. S., and Carlson, R. W. (1991) *Carbohydr. Res.* **220**, 219–227
- Carlson, R. W., Reuhs, B., Chen, T. B., Bhat, U. R., and Noel, K. D. (1995) *J. Biol. Chem.* **270**, 11783–11788
- Kadmas, J. L., Allaway, D., Studholme, R. E., Sullivan, J. T., Ronson, C. W., Poole, P. S., and Raetz, C. R. H. (1998) *J. Biol. Chem.* **273**, 26432–26440
- Campbell, G. R., Reuhs, B. L., and Walker, G. C. (2002) *Proc. Natl. Acad. Sci. U. S. A.* **99**, 3938–3943
- Galibert, F., Finan, T. M., Long, S. R., Puhler, A., Abola, P., Ampe, F., Barloy-Hubler, F., Barnett, M. J., Becker, A., Boistard, P., Bothe, G., Boutry, M., Bowser, L., Buhrmester, J., Cadieu, E., Capela, D., Chain, P., Cowie, A., Davis, R. W., Dreano, S., Federspiel, N. A., Fisher, R. F., Gloux, S., Godrie, T., Goffeau, A., Golding, B., Gouzy, J., Gurjal, M., Hernandez-Lucas, I., Hong, A., Huizar, L., Hyman, R. W., Jones, T., Kahn, D., Kahn, M. L., Kalman, S., Keating, D. H., Kiss, E., Komp, C., Lelaure, V., Masuy, D., Palm, C., Peck, M. C., Pohl, T. M., Portetelle, D., Purnelle, B., Ramsperger, U., Surzycki, R., Thebault, P., Vandenbol, M., Vorholter, F. J., Weidner, S., Wells, D. H., Wong, K., Yeh, K. C., and Batut, J. (2001) *Science* **293**, 668–672
- Lagares, A., Hozbor, D. F., Niehaus, K., Otero, A. J., Lorenzen, J., Arnold, W., and Puhler, A. (2001) *J. Bacteriol.* **183**, 1248–1258
- Lagares, A., Caetano-Anolles, G., Niehaus, K., Lorenzen, J., Ljunggren, H. D., Puhler, A., and Favelukes, G. (1992) *J. Bacteriol.* **174**, 5941–5952
- Niehaus, K., Lagares, A., and Puhler, A. (1998) *Mol. Plant Microbe Interact.* **11**, 906–914
- Miller, J. R. (1972) *Experiments in Molecular Genetics*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY
- Bergmans, H. E., van Die, I. M., and Hoekstra, W. P. (1981) *J. Bacteriol.* **146**, 564–570
- Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor, Cold Spring Harbor, NY
- Smith, P. K., Krohn, R. I., Hermanson, G. T., Mallia, A. K., Gartner, F. H., Provenzano, M. D., Fujimoto, E. K., Goeke, N. M., Olson, B. J., and Klenk, D. C. (1985) *Anal. Biochem.* **150**, 76–85
- Basu, S. S., York, J. D., and Raetz, C. R. H. (1999) *J. Biol. Chem.* **274**, 11139–11149
- Poole, P. S., Schofield, N. A., Reid, C. J., Drew, E. M., and Walshaw, D. L. (1994) *Microbiology* **140**, 2797–2809
- Dankert, M., Passeron, S., Recondo, E., and Leloir, L. F. (1964) *Biochim. Biophys. Res. Commun.* **14**, 358–362
- Passeron, S., Recondo, E., and Dankert, M. (1964) *Biochim. Biophys. Acta* **89**, 372–374
- Kadmas, J. L., Brozek, K. A., and Raetz, C. R. H. (1996) *J. Biol. Chem.* **271**, 32119–32125
- Brozek, K. A., Kadmas, J. L., and Raetz, C. R. H. (1996) *J. Biol. Chem.* **271**, 32112–32118
- Price, N. J. P., Jeyaretnam, B., Carlson, R. W., Kadmas, J. L., Raetz, C. R. H., and Brozek, K. A. (1995) *Proc. Natl. Acad. Sci. U. S. A.* **92**, 7352–7356
- Kadmas, J. L., and Raetz, C. R. H. (1998) *J. Biol. Chem.* **273**, 2799–2807
- Gudlavalleti, S. K., and Forsberg, L. S. (2002) *J. Biol. Chem.* **278**, 3957–3968
- Raetz, C. R. H. (1990) *Annu. Rev. Biochem.* **59**, 129–170
- Raetz, C. R. H., and Whitfield, C. (2002) *Annu. Rev. Biochem.* **71**, 635–700
- Meade, H. M., Long, S. R., Ruvkun, G. B., Brown, S. E., and Ausubel, F. M. (1982) *J. Bacteriol.* **149**, 114–122
- Casse, F., Boucher, C., Julliot, J. S., Michell, M., and Dénarié, J. (1979) *J. Gen. Microbiol.* **113**, 229–242
- Blatny, J. M., Brautaset, T., Winther-Larsen, H. C., Haugan, K., and Valla, S. (1997) *Appl. Environ. Microbiol.* **63**, 370–379
- Altschul, S. F., Madden, T. L., Schaffer, A. A., Zhang, J., Zhang, Z., Miller, W., and Lipman, D. J. (1997) *Nucleic Acids Res.* **25**, 3389–3402

³ The *F. tularensis* genome is available on the World Wide Web at artedi.ebc.uu.se/Projects/Francisella/.

Relaxed Sugar Donor Selectivity of a *Sinorhizobium meliloti* Ortholog of the *Rhizobium leguminosarum* Mannosyl Transferase LpcC: ROLE OF THE LIPOPOLYSACCHARIDE CORE IN SYMBIOSIS OF RHIZOBIACEAE WITH PLANTS

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